DETECTION OF THALLIUM AND URANIUM IN WELL WATER AND BIOLOGICAL SPECIMENS OF AN EASTERN CROATIAN POPULATION

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Using inductively-coupled plasma mass spectrometry (ICP-MS), we measured the concentrations of thallium and uranium in local water resources from three villages (Ćelije, Draž, and Potnjani) in eastern Croatia, with the aim to determine if they were associated with the levels of these same elements in the serum, urine, and hair collected from the residents of this area. The exposure of the local population to thallium and uranium through drinking water was generally low. ICP-MS was capable of measuring the levels of both of the elements in almost all of the analysed samples. Although there were differences in the concentrations of both elements in water samples and biological specimens taken from the residents, they did not reach the maximum contaminant level in any of the four sample types studied. Although hair was previously reported as an excellent indicator of occupational and environmental exposure to various elements, our study did not confirm it as a reliable biological material for tracing thallium and uranium levels, mainly due to the very low concentrations of these elements, often well below the detection limit. However, our results have shown that the concentration of thallium and uranium in drinking water can be effectively traced in urine samples.

KEYWORDS: biological specimens, inductively-coupled plasma mass spectrometry, local water resources, toxic metals

Thallium is a naturally occurring trace element, widely distributed at very low concentrations in the earth’s crust. It is not an essential element for life and does not have a known biological use (1), but has been confirmed toxic in studies on animals (2). As one of the most toxic heavy metals, thallium has also been responsible for many accidental, occupational, deliberate, and therapeutic poisonings (3, 4). Its continued use as a rodenticide in many developing countries and its increasing use in technology have raised concerns about the exposure risk for animals and humans (4). Major sources of thallium in the environment are fossil fuels and the smelting of ferrous and non-ferrous ores. Minor sources include oil fraction refinement, cement production, brickworks, potash-derived fertilizers, and the production and uses of thallium (5).

Uranium is also not an essential element for life and can be toxic both to humans and the environment. Due to its chemical and radiological properties, uranium is very useful in industry and commerce. People are continually exposed to uranium through air, water, and food as well as in a lesser extent through the radiation it emits (6, 7). Uranium is quickly absorbed by the intestine and lungs. It enters the bloodstream quickly and is deposited in tissues, mainly kidneys and bones. Its clearance from the bloodstream is relatively rapid (8). Uranium impairs growth and
development and damages DNA (9). Data from experimental studies on animals and human epidemiology studies have shown that uranium has many harmful effects on the kidneys and reproductive organs (7).

The drinking water in eastern Croatia contains high concentrations of toxic elements such as arsenic, manganese, and iron (10-12). However, there is a lack of data about the possibility of the exposure of local inhabitants to thallium and uranium, since the drinking water from the studied sites has never been chemically analysed in such detail.

In this area, people often use private wells as a source of drinking water. Although the analysis of well water and determination of concentrations of potentially toxic elements can reduce the risk of harmful effects to human populations, the private owners of such wells are not obliged by the regulatory authorities to test their water. People using private wells are therefore at a risk of exposure to a variety of contaminants. The inhabitants of the rural areas of eastern Croatia mainly work in agriculture. Crop cultivation involves large amounts of phosphate fertilisers, which along with coal and fuel burning may be a source of both thallium and uranium in drinking water.

Therefore, the main objective of our study was to establish the levels of exposure to uranium and thallium in the drinking water of the aforementioned three villages and determine which biological specimen is most accurate in the detection of thallium and uranium levels.

MATERIALS AND METHODS

Study area

The study was performed between May 2005 and June 2007 in three communities: Ćelije, Draž, and Potnjani, situated in the eastern Croatia regions of Slavonija and Baranja. The locations of the sites are shown in Figure 1.

Water sampling and analysis

In the three communities, the residents use drinking water from wells (tube wells and dug wells) by pumping it into their household water pipes. A total of 90 samples of drinking water were collected: 56 samples in May and June of 2005 and 2006, and 34 samples in May and June of 2007 (Table 1). The wells were selected at random. The study was performed in accordance with the Croatian regulations on the safety of drinking water.

During the sampling of the well water, the water was allowed to run from the tap for about 60 seconds or was taken from the hydraulic accumulator that pumps the well water into the households. It was then sampled in 100 mL polyethylene bottles. After that, 1 mL of 33 % nitric acid was added to each polyethylene bottle containing a sample of the well water, and the bottles were shaken vigorously. The samples of well water were placed in a freezer at -20 °C. After collection, all of the samples were transported in a portable freezer to the Laboratory of General and Inorganic Chemistry, Faculty of Chemical Engineering and Technology, University of Zagreb. Complete analysis of the drinking well water samples was performed by inductively-coupled plasma mass spectrometry (ICP-MS) Perkin Elmer, ELAN DRC-e, according to United States Environmental Protection Agency (US EPA), method 200.8. (13). The operating conditions for all water and biological samples are described below.

Urine, blood, and hair sampling and analysis

The study included 30 sample donors of both sexes aged 35 to 75 years who lived in three village communities and represented a cross-section of the population in these communities. The study was approved by the Ethics Committee of University of Osijek School of Medicine. The purpose of the study was explained to each participant, who then provided their written informed consent.

Venous blood and spot urine samples were taken from 30 subjects from the three communities in May
and June of 2005 and 2006. During May and June of the third year (2007), 36 blood, spot urine, and hair samples were taken from 17 volunteers from Ćelije and Draž only.

The blood and urine samples were taken by a laboratory technician. At least 3 mL of blood was taken from each participant using a Vacuette® Blood Collection Needle (38 mm×0.9 mm) and a Vacuette® Serum Gel Tube (3.5 mL, Greiner Bio-One, Kremsmünster, Austria). The serum was separated into 1.5 mL Eppendorf® microtubes 3810X (3810X, Eppendorf Geratebau, Hamburg, Germany) on the same day.

The spot urine samples were placed in 55 mL urine collection containers (Plastika Moćan, Samobor, Croatia). The hair samples, approximately 0.1 cm wide and 3 cm long, were cut with stainless steel scissors from the occipital scalp of each subject and placed into polyethylene bags. If the hair was short, several samples from the same part of the head were taken.

The serum and urine samples were thawed, acidified with 60 % nitric acid (0.1 mL per 10 mL of sample), mixed well, and 0.5 mL diluted to 10 mL with 1 % nitric acid before analysis. The hair samples were washed with acetone, air-dried for 24 h, weighed to exactly 0.1 g, dissolved in 1 mL of nitric acid, and diluted to 10 mL for analysis. High purity nitric acid (“Suprapure”®, Merck KGaA, Darmstadt, Germany) was used throughout.

The operating conditions of the ICP-MS using nickel sampler/skimmer cones were: RF power 1300 W, plasma gas flow 15 L min⁻¹, auxiliary gas flow 1 L min⁻¹, nebulizer gas flow 0.94 L min⁻¹ to 0.96 L min⁻¹, peristaltic pump speed 1 L min⁻¹. The instrument was calibrated after every twelve samples, using an external “71-Element Group Multi Element Standard Solution” (Inorganic Ventures, Christiansburg, Virginia, USA) and internal standards with elements yttrium (Y), indium (In), terbium (Tb) and bismuth (Bi) (Inorganic Ventures, USA).

### Table 1

<table>
<thead>
<tr>
<th>Parameters*</th>
<th>Local community</th>
<th>Ćelije</th>
<th>Draž</th>
<th>Potnjani</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Thallium</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N(n) § M (25 % to 75 %)*</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Water / μg L⁻¹</td>
<td>40 (34)</td>
<td>&lt;0.001 (0.001 to 0.01)</td>
<td>36 (35)</td>
<td>0.001 (0.01 to 0.023)</td>
<td>14 (14)</td>
</tr>
<tr>
<td>Serum / μg L⁻¹</td>
<td>78 (26)</td>
<td>0.05 (0.05 to 0.1)</td>
<td>82 (48)</td>
<td>&lt;0.05 (0.05 to 0.05)</td>
<td>61 (37)</td>
</tr>
<tr>
<td>Urine / μg L⁻¹</td>
<td>76 (32)</td>
<td>0.07 (0.05 to 0.18)</td>
<td>82 (18)</td>
<td>0.15 (0.05 to 0.27)</td>
<td>63 (33)</td>
</tr>
<tr>
<td>Hair / μg g⁻¹</td>
<td>14 (4)</td>
<td>0.07 (0.04 to 0.28)</td>
<td>22 (3)</td>
<td>0.14 (0.05 to 0.25)</td>
<td>-</td>
</tr>
<tr>
<td><strong>Uranium</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N(n) § M (25 % to 75 %)*</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Water / μg L⁻¹</td>
<td>40 (38)</td>
<td>0.001 (0.001 to 0.012)</td>
<td>36 (13)</td>
<td>0.85 (0.02 to 7.39)</td>
<td>14 (8)</td>
</tr>
<tr>
<td>Serum / μg L⁻¹</td>
<td>78 (46)</td>
<td>&lt;0.05 (0.05 to 0.05)</td>
<td>82 (51)</td>
<td>&lt;0.05 (0.05 to 0.05)</td>
<td>61 (35)</td>
</tr>
<tr>
<td>Urine / μg L⁻¹</td>
<td>76 (61)</td>
<td>&lt;0.05 (0.05 to 0.1)</td>
<td>82 (46)</td>
<td>0.05 (0.05 to 0.09)</td>
<td>63 (52)</td>
</tr>
<tr>
<td>Hair / μg g⁻¹</td>
<td>14 (14)</td>
<td>&lt;0.01 (0.01 to 0.01)</td>
<td>22 (21)</td>
<td>&lt;0.01 (0.01 to 0.02)</td>
<td>-</td>
</tr>
</tbody>
</table>

*Median (interquartile range, 25 % to 75 %); †Kruskal Wallis test; ‡Mann Whitney test; §Number of samples (number of samples below the detection limit)
The analytical methods were validated using standard reference materials: “ICP multi element standard solution X for surface water testing” (CertiPUR®, Merck, Darmstadt, Germany) and standard samples “Trace Elements Urine Blank” and “Trace Elements Urine” (SERO AS, Billingstad, Norway).

Statistical analysis

Normality of distribution was tested with the Kolmogorov-Smirnov test. Median and interquartile ranges were given for variables that were not distributed normally. Since the data were not normally distributed, Kruskal-Wallis test and Mann-Whitney U-test were used to determine the differences in the measured parameters among the three communities. The significance was set at $p=0.05$.

RESULTS

The levels of thallium and uranium measured in the samples of drinking well water and biological specimens (serum, urine, and hair) collected from the residents of the studied communities are shown in Table 1 and Figure 2.

Drinking well water

Concentrations of thallium in the majority of samples of drinking water collected in all three communities were below the detection limit, so these results could not be compared. Concentrations of uranium in the samples of drinking water were the highest in Draž, with a significantly greater number of samples of drinking water that had measurable values of uranium, whereas the samples from the other two villages were mostly free of uranium. The Kruskal-Wallis test confirmed significant differences ($p<0.001$) between the uranium levels measured at all three sampling sites (Table 1, Figure 2). The most prominent differences (Mann-Whitney U-test) were observed between the uranium levels measured in the samples of drinking water from Draž as compared to the levels measured in Ćelije ($p=0.001$), and between Draž and Potnjani ($p=0.005$).

Serum

The thallium concentration measured in the serum samples from all of the three sampling sites differed significantly ($p=0.024$; Kruskal-Wallis test) (Table 1, Figure 2). Thallium concentrations were significantly higher (Mann-Whitney U-test) in serum samples taken from the subjects in Ćelije compared to Draž ($p=0.011$).
and Potnjani \((p=0.009)\). There was no significant difference in the level of uranium.

**Urine**

The thallium concentration measured in the urine samples collected in all of the three villages differed significantly (Kruskall-Wallis test, \(p=0.002\)) (Table 1, Figure 2). Thallium concentrations were significantly higher (Mann-Whitney U-test) in urine samples from Draž compared to the samples taken from Ćelije \((p=0.004)\) and Potnjani \((p=0.002)\).

The uranium concentration was significantly different in the urine samples (Kruskal-Wallis test, \(p<0.001\)) collected in all three villages (Table 1, Figure 2). We found a statistically significant difference (Mann-Whitney test, \(p<0.001\)) in the concentration of uranium in urine between subjects from Draž and Ćelije, and Draž and Potnjani.

**Hair**

There was no statistically significant difference in the concentration of thallium in the hair samples of subjects from the communities Draž and Ćelije (Table 1).

A large number of hair samples from Draž and Ćelije had uranium concentrations below the detection limit, so these results could not be compared (Table 1).

**DISCUSSION**

The present study is the first to report thallium and uranium levels measured in drinking water and three biological specimens in an eastern Croatian population using the ICP-MS method. On the grounds of the obtained data, it intention is to discuss the mutual relationships between these levels. The obtained results confirm that ICP-MS was capable of measuring detectable levels for both elements in almost all of the samples analysed, but that urine was the most accurate.

**Thallium and uranium in drinking water**

The knowledge about the sources, mobility, and transport routes of thallium in the environment is very important to evaluate and reduce the risk of contamination with this element (14). The major sources of thallium in drinking water are leaching from ore-processing sites, as well as discharge from electronics, glass, and pharmaceutical industries (15). Other potential sources are inadequate waste removal and storage and chemical dumps. Thallium compounds easily mix with water, whereas small amounts can be transmitted through the air from coal power plants, cement plants, and smelting plants (16-19). Although thallium is more toxic than cadmium, lead, and mercury, data on its presence in water and biological tissues are scarce, perhaps due to the generally low levels of thallium in the environment and the high analytical detection limit (5). Several recent studies (3, 20-22) have dealt with the effect of thallium on the environment and its hazards to humans. In certain geochemically unfavourable areas, natural processes along with human activities can lead to an increase of thallium in surface water, groundwater, and soil (21-22). The exploitation of sulphide minerals leads to the enrichment of local soil, water, and crops with thallium, which finally results in chronic thallium poisoning (14). Most poisonings in people occur when water contains up to \(3.214 \text{ mg L}^{-1}\) of thallium (23).

The rules on the safety of drinking water in Croatia do not define standard values for thallium (24). However, the United States Environmental Protection Agency (15) set \(2 \text{ μg L}^{-1}\) as the maximum contaminant level (MCL) for thallium in drinking water.

Our results have shown that there are no threats for residents in all of the three communities, since the thallium levels were very low and none of the monitoring sites exceeded the MCL value of \(2 \text{ μg L}^{-1}\).

Ingesting water from drilled wells is a source of exposure to uranium in different populations around the world (8, 25-28). Great variation in the exposure levels of uranium has been reported. In some areas of northern Greece, uranium concentrations in groundwater ranged from \(0.01 \text{ μg L}^{-1}\) to \(10 \text{ μg L}^{-1}\) (29). The uranium concentration in drinking water samples collected in Punjab, India varied from \(1.39 \text{ μg L}^{-1}\) to \(98.25 \text{ μg L}^{-1}\), with a mean value of \(19.84 \text{ μg L}^{-1}\) (30). Uranium levels in samples of Canadian drinking water ranged from undetectable in treated municipal water samples to exceptionally high (\(>700 \text{ μg L}^{-1}\)) in private groundwater supplies (8). Similar elevated levels of uranium (866 \text{ μg L}^{-1} and 1160 \text{ μg L}^{-1}) were detected in the drinking well water of a private owner in rural north-western Connecticut, USA (28). Very high concentrations of naturally occurring uranium, up to 12,000 \text{ μg L}^{-1}, were measured in Finnish groundwater and wells, presumably because the water in those areas runs through bedrock composed mainly of uranium-rich granitoids and granites (31).

The results of our study show that the exposure of the local population to uranium in drinking water was...
generally low. In all of the three communities, the median level of uranium in drinking water was significantly lower than 30 μg L⁻¹, which is the new provisional World Health Organization guideline value for uranium in drinking water (32). Croatian legal provisions on drinking water do not stipulate standards for the concentration of uranium in drinking water (24). Since people in the rural areas of eastern Croatia mainly work in agriculture, using large amounts of phosphate fertilizers for the cultivation of crops, we assume that the dissolution of phosphate fertilizers, together with the combustion of coal and other fuels (8), may have been the source of the uranium detected in the drinking water.

**Thallium in biological specimens**

An important step in the accurate evaluation of the potentially detrimental effects of thallium is the measurement of its content in biological specimens (33). For that purpose, blood, serum, urine, and hair are commonly used (2, 34-37).

In our study, thallium levels were determined in serum samples, and its highest concentration (median 0.05 μg L⁻¹ and interquartile range <0.05 μg L⁻¹ to 0.10 μg L⁻¹) was detected in subjects from Ćelije. However, these values were well below 2 μg L⁻¹, which is considered a normal total blood thallium concentration (38, 39). Furthermore, the levels of thallium measured in the serum of the subjects were quite comparable with the levels reported in two previous Italian studies (Table 2; 38, 40).

Blood thallium levels, however, are not considered reliable measures of exposure, except after extensive exposure (41). The possible reasons for that are the very rapid clearance of thallium from blood in animals and humans (6, 42, 43) and the peculiar kinetic features of thallium ion absorption in the gastrointestinal tract (44).

Thallium is excreted in both animals and humans predominantly by the kidneys and intestine, but also in partly through hair and milk. Studies on acute and chronic poisoning agree that the highest concentration is found in the kidneys (6).

Establishing human urinary thallium concentrations is a quick way to identify human exposure to thallium pollution (14). For a reliable analysis of thallium, the collection of 24-hour urine is advisable, because of the diurnal variation, i.e. peak excretion might vary as a result of a variation in drinking patterns. A thallium concentration higher than 20 μg L⁻¹ is used as evidence of excessive exposure (41), while urine thallium concentrations greater than 200 μg L⁻¹ are considered toxic (41, 45).

Nails and urine in humans contain high concentrations of thallium in comparison with other biological samples (46). Data from the general EU population show thallium concentration in urine of 0.066 μg L⁻¹ and in blood 0.063 μg L⁻¹. The overall correlations between thallium in urine and blood in this study were weak (47). The tentatively proposed reference intervals of thallium are (0.019±0.17) μg L⁻¹ in urine and (0.014±0.19) μg L⁻¹ in blood (47).

Since the concentration of thallium in most samples of drinking water in our study was below the detection limit, the higher concentrations of thallium measured in the urine of subjects from Draž and the higher concentrations of thallium measured in the serum of subjects from Ćelije cannot be attributed to drinking water, thus presumably reflecting exposure to unknown sources. To determine the exact source, we would need to establish the occupational and other exposures of subjects with elevated levels of thallium in the urine and serum.

**Uranium in biological specimens**

Assessment of occupational or environmental exposure to uranium compounds is largely based on urinary analysis (48). ICP-MS has proven to be a fast, reliable and highly sensitive technique for measuring uranium concentrations in urine (49). Spot urine samples are routinely used to monitor occupational exposure to uranium and other toxic heavy metals, such as mercury, lead, and cadmium (50). The diurnal fluctuations of uranium in spot samples and diurnal variations in the uranium concentrations in different urine samples from the same individual could be quite large (48, 50). Therefore, complete 24-hour urine samples give better precision in estimating uranium concentrations at low levels than spot samples (51). In our study, due to certain technical difficulties, spot urine samples were used. To overcome the drawbacks that arise from the analysis of spot urine samples, they were taken in three consecutive years, together with hair samples (51, 52). The results obtained in our study (level of uranium in urine) were similar to the results reported in a previous study conducted in eastern Croatia (Table 2, 53).

A study from northern Italy reported significant regional variations in the concentration of uranium in urine samples of different individuals, which was the result of an unequal distribution of uranium in different regions and differences in dietary habits (49). In a
A study on a Finnish population, where drinking water was the major source of exposure to uranium, its content in different biological specimens (urine, hair, and nails) was correlated with uranium intake, particularly at elevated levels ≥10 μg per day (54). These results clearly demonstrate that uranium found in bioassays can be traced back to drinking water, thus providing a direct link to the source of exposure (55).

Seldén et al. (56) found that subjects exposed to various concentrations of uranium in drinking water (range <0.20 μg L\(^{-1}\) to 470 μg L\(^{-1}\), median 6.7 μg L\(^{-1}\)) had eight times higher levels of urinary uranium than the control subjects, who were not exposed. Hair is an excellent indicator of occupational and environmental exposure to various elements, including uranium. Compared to other biological specimens like blood and urine, which tend to show current and recent body status, hair provides status over a longer period of time (57). Hair is an effective bio-concentrator; the samples can be easily stored, the concentration reflects an integrated value, and finally, the measurement of the \(^{234}\text{U}/^{238}\text{U}\) isotopic ratio in digested hair samples by multicollector-inductively coupled plasma mass spectrometry (MC-ICPMS) is feasible and highly informative (55). Muikku et al. (58) investigated a Finnish working population that was not occupationally exposed to uranium and found that their mean uranium concentration in hair was 3- to 15- fold higher than the values reported in the literature, while the mean uranium concentration in urine was similar to those measured elsewhere in Europe. These large variations can be explained by the variation in uranium concentration in drinking water. Very high concentrations of uranium were also measured in private drilled wells in southern Finland (58).

The majority of hair samples from our study had uranium concentrations below the detection limit and these samples were not reliable biological material for tracing uranium. This was due to the fact that the subjects had obviously not ingested uranium concentrations sufficiently high for detection. However, the urine samples enabled us to work with these small concentrations. That is why we were able to find a statistically significant difference in the concentration of uranium in subjects from Ćelije and Potnjani (\(p<0.001\)), but not Draž.

<table>
<thead>
<tr>
<th>Type of sample</th>
<th>Community / Reference</th>
<th>Value reported</th>
<th>Thallium/μg L(^{-1})</th>
<th>Uranium/μg L(^{-1})</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum</td>
<td>Region of Umbria (38)</td>
<td>Mean±SE</td>
<td>0.03±0.001</td>
<td>0.26±0.004</td>
</tr>
<tr>
<td>Serum</td>
<td>Region of Calabria (38)</td>
<td>Mean±SE</td>
<td>0.04±0.001</td>
<td>0.26±0.005</td>
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<tr>
<td>Serum</td>
<td>Eastern Croatia, areas of warfare (53)</td>
<td>Median (interquartile range)</td>
<td>-</td>
<td>0.10 (0.02 to 0.23)</td>
</tr>
<tr>
<td>Serum</td>
<td>Eastern Croatia, areas of heavy warfare (53)</td>
<td>Median (interquartile range)</td>
<td>-</td>
<td>0.11 (0.04 to 0.28)</td>
</tr>
<tr>
<td>Urine</td>
<td>Eastern Croatia, areas of moderate warfare (53)</td>
<td>Median (interquartile range)</td>
<td>-</td>
<td>0.03 (0.00 to 0.18)</td>
</tr>
<tr>
<td>Urine</td>
<td>Eastern Croatia, areas of heavy warfare (53)</td>
<td>Median (interquartile range)</td>
<td>-</td>
<td>0.02 (0.00 to 0.11)</td>
</tr>
<tr>
<td>Hair</td>
<td>Eastern Croatia, areas of moderate warfare (53)</td>
<td>Median (interquartile range)</td>
<td>-</td>
<td>0.00 (0.00 to 0.01)</td>
</tr>
<tr>
<td>Hair</td>
<td>Eastern Croatia, areas of heavy warfare (53)</td>
<td>Median (interquartile range)</td>
<td>-</td>
<td>0.00 (0.00 to 0.00)</td>
</tr>
<tr>
<td>Urine</td>
<td>Healthy Italians (40)</td>
<td>Mean±SD</td>
<td>0.42±0.09</td>
<td>&lt;0.1</td>
</tr>
<tr>
<td>Serum (or plasma)</td>
<td>Healthy Italians (40)</td>
<td>Mean±SD</td>
<td>0.18±0.009</td>
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</tbody>
</table>

*SE - standard error; SD - standard deviation*
CONCLUSION

The results of this study show that the concentration of thallium and uranium in drinking water can be effectively traced in urine samples. Although hair is an excellent indicator of occupational and environmental exposure to various elements, our study did not confirm it as a reliable biological material for tracing either the thallium or uranium concentration, mainly due to the very low concentrations of both elements.

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Sažetak

ODREĐIVANJE RAZINA TALIJA I URANA U BUNARSKOJ VODI I BIOLOŠKIM UZORCIMA STANOVA ISTOČNOHRVATSKIH SELA

Korišćeni masenu spektrometriju induktivno spregnute plazme (ICP-MS), izmjerili smo koncentracije talija i urana u lokalnim izvorima vode triju istočnohrvatskih sela (Čelije, Draž i Potnjani) radi određivanja jesu li povezane s razinama obaju elemenata u serumu, urinu i kosi stanovnika tih triju sela. Izloženost lokalnog stanovništva taliju i uranu preko pitke vode općenito je bila niska. ICP-MS uspješno je izmjerio razine obaju elemenata u gotovo svim analiziranim uzorcima. Iako je bilo razlika u razinama obaju elemenata u uzorcima vode i biološkim uzorcima stanovnika, nijedan nije prešao maksimalnu dopuštenu granicu. Kosa je prethodno opisana kao izvrstan pokazatelj profesionalne ili okolišne izloženosti različitim elementima, no rezultati našeg ispitivanja nisu potvrdili njenu pouzdanost pri mjerenju talija i urana uslijed vrlo niskih koncentracija obaju elemenata, koje su često bile i ispod razine detekcije. Međutim, pokazali su kako uzorci urina mogu poslužiti kao vrlo dobri indikatori prisutnosti talija i urana u pitkoj vodi.

KLJUČNE RIJEČI: biološki uzorci, lokalni izvori vode, masena spektrometrija induktivno spregnutom plazmom, toksični metali

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