Evaluating unspecific oxidative stress parameters in the sera of patients with irritable bowel syndrome

Abstract

Objective: Irritable bowel syndrome (IBS) is a common global condition characterized by abdominal pain and alterations in bowel habits not caused by other organic diseases and its etiopathogenesis has not been elucidated. In this study, we aimed to evaluate the oxidative stress parameters in patients with IBS.

Materials and methods: Fifty patients diagnosed with IBS using the Rome III criteria and a control group of 50 healthy subjects were included in the study. Oxidative stress parameters including total oxidant status (TOS), total antioxidant status (TAS) and oxidative stress index (OSI) values were analyzed from all study subjects.

Results: Compared to the controls; the TOS and OSI values were significantly higher, and the TAS value was significantly lower in IBS patients (p < 0.001 for all).

Conclusions: In present study we demonstrated that oxidative stress increased and antioxidant capacity decreased in IBS, and antioxidants might be beneficial in the supportive treatment for IBS.

INTRODUCTION

Irritable bowel syndrome (IBS) is a common global condition that impairs quality of life. It is characterized by abdominal pain and altered bowel habits not caused by other known organic diseases. Abdominal pain in IBS is usually described as a crampy sensation with variable intensity and periodic exacerbations. The location and character of the pain can vary widely. Emotional stress and eating may exacerbate the pain, while defecation often provides some relief (1, 2). Altered bowel habits are ranging from diarrhea and constipation or normal bowel habits alternating with either diarrhea and/or constipation. Constipation may last from days to months, with interludes of diarrhea or normal bowel function. Stools are often hard and patients may also experience a sense of incomplete evacuation even when the rectum is empty. Diarrhea is usually occurs during waking hours, most often in the morning or after meals and accompanied by lower abdominal cramps and urgency. Stools generally characterized as frequent loose stools of small to moderate volume with mucus discharging. Numerous factors have been implicated in the etiology of IBS, including genetics, intestinal infections, over-production of intestinal bacteria, increased cytokine response and inflammation, irregularity of serotonergic functions, and psychosocial aspects (3–8). Since it has been regarded as a functional bowel disease because of the etiopathogenesis of IBS has not
been fully illuminated, diagnosis of IBS is made based on the Rome III clinical criteria, last modified in 2006 (9).

The term oxidative stress is used to describe a series of chemical reactions that result in the production of free oxygen radicals and other reactive molecules (10). These free radicals and reactive oxygen molecules are neutralized by the complex structure of the antioxidant system but when the pro-oxidant and antioxidants systems are dysregulated, oxidative stress can damage important cellular components including lipids, proteins and nucleic acids (11, 12). It was already showed that the oxidative stress products increase if there is an endocrine disease that affects the body’s metabolic rate and an inflammatory disease which cause acute or chronic organ damage in previous studies (13-17). But there is limited number of studies investigating the presence of oxidative stress in non organic and functional diseases. Therefore the current study was undertaken and aimed to evaluate the role of oxidative stress in IBS etiopathogenesis by measuring the total oxidative status (TOS), total antioxidant status (TAS) values and calculating oxidative stress index (OSI) by Erel’s method which is easy, stable, reliable, sensitive, inexpensive and fully automated in IBS patients.

**MATERIALS AND METHODS**

The study participants included 50 patients who presented at the Gastroenterology Polyclinic of Harran University Medical Faculty and were diagnosed with IBS, and a control group of 50 healthy individuals. The diagnosis of IBS was made according to the Rome III criteria. Routine laboratory studies (complete blood count, fasting blood glucose, urea, creatinine, alanine amino transferase, aspartate amino transferase, alkaline phosphatase, gama glutamil transferase, sedimentation, C-reactive protein, thyroid stimulating hormone) and abdominal ultrasonographic examination were evaluated while selection of patients to exclude from the other diseases. Patients < 18 years old, addicted to alcohol or medication, and those with a previous diagnosis of hematological, renal, hepatic, cardiovascular, neurological, chronic inflammatory bowel disease, chronic inflammatory connective tissue diseases and malign diseases were excluded from the study. Informed consent, conforming to the Helsinki Declaration of 2008, was obtained from all patients and control subjects. The local ethics committee approved the study protocol.

**Biochemical Analysis**

All blood samples were drawn from a large antecubital vein without interruption of venous flow, following an overnight fasting state into blood tubes and immediately stored on ice at 4 °C. The serum was then separated from the cells by centrifugation at 3000 rpm for 10 min and they were stored until analyzing at −80 °C. Ten milliliters of blood was used for baseline routine laboratory tests. Thyroid stimulating hormone (TSH), levels were analyzed using an electrochemiluminescence immunometric assay

<table>
<thead>
<tr>
<th>Parameters</th>
<th>IBS Group (n=50)</th>
<th>Healthy subjects (n=50)</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>37.2±11.9</td>
<td>39.5±11.7</td>
<td>0.932</td>
</tr>
<tr>
<td>Sex (male/female)</td>
<td>27/23</td>
<td>28/22</td>
<td>0.989</td>
</tr>
<tr>
<td>Hematocrit (%)</td>
<td>41.0±4.6</td>
<td>41.7±4.3</td>
<td>0.646</td>
</tr>
<tr>
<td>Glucose (mg/dL)</td>
<td>99.2±7.1</td>
<td>93.9±4.3</td>
<td>0.277</td>
</tr>
<tr>
<td>Urea (mmol/L)</td>
<td>28.5±8.9</td>
<td>28.0±10.8</td>
<td>0.587</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.6±0.1</td>
<td>0.7±0.1</td>
<td>0.534</td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>24.3±11.6</td>
<td>23.9±14.2</td>
<td>0.219</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>27.5±19.4</td>
<td>29.4±25.3</td>
<td>0.761</td>
</tr>
<tr>
<td>TSH (mIU/L)</td>
<td>2.7±0.7</td>
<td>2.5±0.8</td>
<td>0.511</td>
</tr>
</tbody>
</table>

Abbreviations: IBS: Irritable Bowel Syndrome, AST: aspartate aminotransferase, ALT: alanine aminotransferase, TSH: Thyroid stimulating hormone

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<tbody>
<tr>
<td>TAS (μmol Trolox equivalent/L)</td>
<td>850 ± 80.0</td>
<td>1040 ± 90.0</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>TOS (μmol H₂O₂ equivalent/L)</td>
<td>44.84±1.56</td>
<td>37.87±1.11</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>OSI (arbitrary units)</td>
<td>5.27±0.48</td>
<td>3.64±0.26</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

Abbreviations: IBS: Irritable Bowel Syndrome, TAS: Total Antioxidant Status, TOS: Total Oxidant Status, OSI: Oxidative Stress Index
calculated as follows: the oxidative stress index (OSI). The serum OSI value was
percentage of TOS level to TAS level was regarded as the
levels (18, 19). TAS levels were converted to μmol. The
molar hydrogen peroxide equivalents per liter (μmol
H\textsubscript{2}O\textsubscript{2} equivalent/L).

**Measurement of Total Oxidant Status**

Serum TOS was measured using a novel automated
method developed by Erel (18). Oxidants present in the
sample oxidize the ferrous ion-odianisidine complex to
ferric ion. The oxidation reaction is enhanced by glycerol
molecules, which are abundant in the reaction medium.
The ferric ion generates a colored complex with Xylenol
Orange in an acidic medium. Color intensity, which can
be measured spectrophotometrically (V-530; Jasco\textsuperscript{®}, To-
kyo, Japan), is related to the quantity of oxidant molecules
present in the sample. The assay is calibrated with hydro-
gen peroxide and the results expressed in terms of micro-
moles [8-hydroxy-2'-deoxyguanosine (8-OHdG), reactive
oxygen intermediates (ROIs), lipid peroxidation products
[8-hydroxy-2'-deoxyguanosine (8-OHdG)], reactive
oxygen intermediates (ROIs), lipid peroxidation products
malonaldehyde (MDA) and nitric oxide (NO) were
studied in these studies (27-31). However there is limited
Epidemiological studies have been shown the increased
oxidative stress in inflammatory, degenerative and endo-
crine diseases which affect tissues integrity and meta-
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oxygen intermediates (ROIs), lipid peroxidation products
malonaldehyde (MDA) and 4-hydroxynonenal (HNE), catalase (CAT), glutathione peroxidase (GSH-Px), xan-
thine oxidase (XO), adenosine deaminase (AD) activities,
and malondialdehyde (MDA) and nitric oxide (NO) were
studied in these studies (27-31). However there is limited
number of study evaluating oxidative stress products in
IBS. Mete R. et al. (32) evaluated the plasma concentra-
tions of malondialdehyde (MDA) and nitric oxide (NO)
and the plasma activities of oxidant and antioxidant en-
zymes (Superoxide dismutase (SOD), catalase (CAT),
glutathione peroxidase (GSH-Px), xanthine oxidase (XO),

The OSI was defined as the ratio of the TOS to TAS
levels (18, 19). TAS levels were converted to μmol. The
percentage of TOS level to TAS level was regarded as the
oxidative stress index (OSI). The serum OSI value was
calculated as follows:

\[
\text{OSI (AU)} = \left[\frac{(\text{TOS, } \mu\text{mol H}_2\text{O}_2 \text{ equivalent/L})}{(\text{TAS, } \mu\text{mol Trolox equivalent/L})}\right] \times 100.
\]

The results were expressed as μmol Trolox equivalent/L.

**Measurement of Total Antioxidant Status**

Serum TAS was measured using a novel automated
method developed by Erel (19). In this method, hydroxyl
radical, the most potent biological radical, is produced.
In the assay, ferrous ion solution in reagent 1 is mixed
with hydrogen peroxide present in reagent 2. Sequential-
ly- produced radicals, such as the brown-colored dia-
isidinyl radicalcation produced by the hydroxyl radical,
are also potent radicals. This method allows measuring
the antioxidant effect of the sample against potent free-
radical reactions that are initiated by the hydroxyl radical.
The assay has excellent precision values of more than 97%.
The results are expressed as μmol Trolox equivalent/L.

**Oxidative Stress Index**

The OSI was defined as the ratio of the TOS to TAS
levels (18, 19). TAS levels were converted to μmol. The
percentage of TOS level to TAS level was regarded as the
oxidative stress index (OSI). The serum OSI value was
calculated as follows:

\[
\text{OSI (AU)} = \left[\frac{(\text{TOS, } \mu\text{mol H}_2\text{O}_2 \text{ equivalent/L})}{(\text{TAS, } \mu\text{mol Trolox equivalent/L})}\right] \times 100.
\]

The results were expressed in Arbitrary Units.

**Statistical Analysis**

All statistical analyses were performed using SPSS for
Windows version 17.0 (SPSS Inc., Chicago, IL, USA).
Kolmogorov- Smirnov tests were used to test the normal-
ity of data distribution. The data were expressed as arithmetic
means and standard deviations. Independent sam-
ple T-test was respectively used in normally and
non-normally distributed continuous variables between
groups. Paired test was used to analyze changes within
each group. Two-sided p value < 0.05 was considered sta-
tistically significant.

**RESULTS**

The demographic, clinical and laboratory data of all
study participants are shown in Table 1; no significant
differences between the two groups regarding these
data were noted. All oxidative stress markers (TAS, TOS,
and OSI) were significantly different among the groups
(p < 0.001 for all) (Table 2). Compared to the control
group, the TOS and OSI values of the IBS patients were
determined to be higher and the TAS values were lower.

**DISCUSSION**

The main finding of the present study was that TOS
values were higher and TAS values were lower in patients
with IBS compared to control. In light of this information
we can say there is an oxidative injury in the IBS although
we cannot clear up whether it is a cause or a result.

Traditionally, IBS has been considered as a condition
arising from brain-gut dysregulation and classified as one
of the functional gastrointestinal disorders, hence its
symptoms not explained by structural or biochemical ab-
normalities (4). Supports this fact IBS has been demon-
strated accompanied by depression, anxiety, fibromyalgia
syndrome and gastric esophageal reflux disease in resent
studies (20-26).

Epidemiological studies have been shown the increased
oxidative stress in inflammatory, degenerative and endo-
crine diseases which affect tissues integrity and meta-
bolic rate of body. There were several studies demonstrat-
ing higher concentrations of oxidative stress products and
antioxidants in the chronic inflammatory diseases of gas-
trointestinal tractus such as crohn disease, ulcerative col-
tis and chronic hepatitis. Especially superoxide dismutase
protein carbonyl content (POPs), DNA oxidation prod-
cts [8-hydroxy-2'-deoxyguanosine (8-OHdG)], reactive
oxygen intermediates (ROIs), lipid peroxidation products
malonaldehyde (MDA) and 4-hydroxynonenal (HNE), catalase (CAT), glutathione peroxidase (GSH-Px), xan-
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zymes (Superoxide dismutase (SOD), catalase (CAT),
glutathione peroxidase (GSH-Px), xanthine oxidase (XO),
adenosine deaminase (AD) activities) patients with IBS. Plasma XO and AD activities, lipid peroxidation product MDA and NO concentrations were significantly higher in IBS patients than in controls. The SOD, CAT, and GSH-Px activities in the serum of patients with IBS were significantly lower than that of controls. Oran et al. (33) demonstrated the presence of oxidative stress, a disturbance in prooxidant – antioxidant balance and increased inflammation in patients with IBS by showing the drop in paraoxonase and arylesterase activities accompanied with an increase in conjugated diene levels. Urinary redox potential had also been studied in depression, MDA and SOD in fibromyalgia, which are the other functional diseases related with IBS (34-36). In our study, we evaluated the TAS and TOS values by Erel’s method which has high linearity, rapid, easy, stable, reliable, sensitive, inexpensive and fully automated and the results are highly reproducible (18, 19). We found TOS was increased and TAS was decreased in patients with IBS compared to the controls similar to previous studies that specific oxidative stress markers were evaluated in.

Additionally it was tried to explain the oxidative stress plays a role in the pathogenesis of IBS by evaluating the potential effects of antioxidants in previous experimental studies. Firstly Asadi-Shahmirzadi et al. (37) demonstrated that the severity of stress-induced IBS was diminished by the Aloe vera/German chamomile mixture at all doses used but not dose-dependently, via inhibiting colonic MPO activity and improving oxidative stress status. Later Zhang et al. (38) evaluated the melatonin effects on gastric residual rate, small intestine propulsion rate and regeneration of gastric mucosa broken down by noise stress. Melatonin is effective in reversing the gastrointestinal motility disorder and gastric stress ulcer on noise stress induced rats by its antioxidant and neuroendocrine activity in the circadian organization that previously demonstrated (38). Lastly Garabadu et al. (39) demonstrated that Eugenol protected against restraint stress induced development of IBS-like gastrointestinal dysfunction through modulation of HPA-axis and brain monoaminergic pathways apart from its antioxidant effect. Similar studies were performed in clinical subjects and demonstrated that antioxidants improves IBS symptoms. Kuiken SD et al. (40) argued a hypothesis that nitric oxide (NO) is involved in maintaining visceral hypersensitivity in IBS and NO synthase inhibitor NG-monomethyl-L-arginine (L-NMMA) can restore on rectal resting volume, rectal sensitivity to distension and rectal compliance. Although L-NMMA did not alter rectal resting volume, rectal sensitivity to distension and rectal compliance, significantly increased the threshold for discomfort/pain in IBS patients (40). Melatonin is one of the favorite therapeutic agent plays an important role in gastrointestinal physiology and includes anxiolytic, anti-inflammatory and motility regulatory effects. In recent studies it was demonstrated that its antioxidant effects plays a role to improve IBS symptoms (41, 42).

In conclusion our present study demonstrated that there is increased oxidative stress and decreased antioxidant capacity in patient with IBS; therefore, we should consider that antioxidants might be beneficial in the supportive treatment of IBS. The limitation of the present study is that the TAS, TOS and OSI values were not examined after the IBS treatment and they were not compared to other inflammatory parameters and natural antioxidants such as albumin bilirubin and uric acid. To support our results prospective and randomized controlled trials are necessary.

Conflict of interest: None

References


