Diffuse large B-cell lymphoma represents 30-40% of all non-Hodgkin lymphomas in Western countries (1). Tumor cells are peripheral lymphocytes from germinal centers or a postgerminative stage of B-cell development. Very high percentage of the cells express BCL6, with 30-50% cases showing the expression of BCL2 protein. 3q27 region abnormalities, with the most likely candidate being the BCL6 gene, are detected in 30% of the cases.

**Aim** To explore the association between FOXP1, BCL2, and BCL6 gene expression in diffuse large B-cell lymphoma tumor cells and their association with the presence of FOXP3 lymphocytes.

**Methods** Samples of lymph nodes from 53 patients with newly diagnosed diffuse large B-cell lymphoma were taken at the time of the diagnosis and immunostained for CD10, MUM1, BCL6, BCL2, FOXP1, and FOXP3. Fluorescent in situ hybridization analysis was used for the detection of FOXP1, BCL2, and BCL6 gene abnormalities. The χ² test was used for data analysis.

**Results** FOXP1 protein was detected in 28 cases, genetic abnormalities involving the FOXP1 locus were found in 19 cases, and both were present in 13 cases (χ² = 7.157; P = 0.028). FOXP3 positive cells were detected in 37 cases. There was a significant relationship between BCL2 expression and FOXP1 genetic abnormalities (χ² = 5.858; P = 0.016) and between BCL2 expression and BCL2 genetic abnormalities (χ² = 6.349; P = 0.012). There was also an association between BCL6 and FOXP1 genetic abnormalities (χ² = 8.497; P = 0.004).

**Conclusion** Association was observed between additional FOXP1 gene copies and BCL2 protein expression as well as changes on both FOXP1 and BCL2 genes in samples of our DLBCL patients. FOXP3 positive cells showed no association with presence of any of analyzed proteins considered as a prognostic markers in DLBCL neither with changes of their genes.
IgH/BCL2 - t(14;18)(q32;q21) translocation is known to be the oncogenic event underlying the development of follicular lymphoma and is seen in 20-30% of cases of diffuse large B-cell lymphoma (1). Diffuse large B-cell lymphoma shows heterogeneity of its genetic abnormalities and cell type of origin. Many authors have tried to divide diffuse large B-cell lymphoma into groups according to its relationship to normal B-cell counterparts sharing a common gene expression profile (1). The currently accepted subgroups include the germinal center (GC)-like cells subgroup, activated B cell (ABC)-like subgroup, and a third subgroup named simply “type 3,” representing all the diffuse large B-cell lymphoma cases that cannot be specified as either GC or ABC subgroup (2-6).

There is a lack of predictive markers that can be reliably and robustly applied to routine diagnosis of diffuse large B-cell lymphoma. Some studies found that BCL2 and BCL6 proteins have an impact on diffuse large B-cell lymphoma development and outcome, but there is debate as to whether they can be used as good prognostic markers (1). Other studies have shown that overexpression of FOXP1 protein has an impact on diffuse large B-cell lymphoma development (7-9). In normal tissue, FOXP1 expression was observed in scattered cells in germinal centers, in most cells of the mantle zone, and in a few B-cells in the T-cell zone (10). It was found to have stronger expression following B-cell activation (11), to be an essential transcriptional regulator for early B-cell development (12), and to influence RAG expression (13). In tumor cells, FOXP1 shows loss of expression, cytoplasmic localization, or overexpression (10). Translocation of the FOXP1 gene has been demonstrated in extranodal B-cell lymphoma of marginal zone of MALT type and other B-cell non-Hodgkin lymphomas (14-16).

Other members of FOXP family are also known to have a role in tumorigenesis. FOXP3 mutations can cause IPEX syndrome (immune deregulation, polyendocrinopathy, enteropathy, X-linked syndrome) (17,18). This gene has been recently recognized as a significant factor in the regulation of T-cell activation (19) and as an important gene for the development of regulatory T-lymphocytes (20,21). FOXP3 immunostaining can be used for detection of Treg cells (22) and has been used for analyzing the correlation between the number of Tregs and the prognosis in follicular lymphoma patients (23,24).

In this study, we investigated the expression and genetic abnormalities of BCL2, BCL6, and FOXP1 genes in Croatian patients with diffuse large B-cell lymphoma from a single hematopathology center. Our aim was to determine the association between FOXP1 and BCL2, as previously described by Banham et al (8) and Barrans et al (7), and the differences in this association between GC and ABC subgroup of diffuse large B-cell lymphoma. We were especially interested in the association among FOXP1, BCL2, and BCL6, which would indicate the processes in diffuse large B-cell lymphoma tumor cells. The presence of FOXP3 positive cells and their association with prognostic markers and/or gene abnormalities in tumor cells was also explored.

Patients and methods

Patients

We have analyzed 53 samples of formalin-fixed paraffin-embedded lymph nodes taken at the time of a diagnosis from patients with de novo diffuse large B-cell lymphoma. The patients were 25 women and 28 men with median age of 66 years (range, 23-91). The diagnoses were made by pathologists from the Department of Pathology and Cytology, Merkur University
Hospital according to the World Health Organization criteria (1).

This study is a part of a research project approved by the Ethics Committee of the Zagreb University School of Medicine.

**Methods**

**Immunostaining.** Immunostaining was performed on 4 μm-thick formalin-fixed paraffin-embedded lymph nodes sections using the LSAB/HRP kit (Dako, Glostrup, Denmark), according to the manufacturer’s instructions. The JC12 anti-FOXP1 and 236A/E7 anti-FOXP3 antibodies were acquired from Dr Alison Banham, Oxford, UK; BCL2-124 from the LRF Immunodiagnostics Unit, Oxford, UK; and BCL6 from Dako.

Cases with more than 30% of positive tumor cells were considered positive. FOXP1 staining intensity was graded as weak (+), moderate (++) or strong (+++).

All cases with at least some FOXP3-positive cells were scored as positive for the presence of Treg cells.

Diffuse large B-cell lymphomas were divided into GC or ABC types according to the method described by Hans et al (4).

**Fluorescent in situ hybridization.** Fluorescent in situ hybridization (FISH) was performed according to the protocol described by Ventura et al (25). Abnormalities of the FOXP1 gene were detected with a break-apart probe (26), abnormalities of BCL2 gene with a dual color fusion probe (Vysis, Downer’s Grove, IL, USA), and abnormalities of BCL6 gene with a break-apart probe (Vysis). Cut-off values for each FISH probe were determined on 5 sections of non-tumor formalin-fixed paraffin-embedded lymph nodes. Cut-off values for all probes were first found to be in the range between 5% and 7% of tumor cells in a sample with any of the investigated abnormalities (FOXP1 probe 6.3%, BCL2 probe 5.2%; BCL6 probe 6.9%) and then the value was estimated to be 7%.

**Statistical analysis**

Differences between the groups were tested using $\chi^2$ test (Statistical Package for the Social Sciences for Windows, version 14.0; SPSS Inc., Chicago, IL, USA). The level of statistical significance was set at 0.05.

**Results**

**Immunostaining**

BCL2 and BCL6 protein were expressed in 37 and 17 diffuse large B-cell lymphoma cases, respectively (Figure 1D and 1E). Both proteins were expressed in tumor cells of all positive samples with approximately same intensity. FOXP1 protein was expressed in 28 cases (Figure 1F), with weak staining observed in 10 cases, moderate staining in 7 cases, and strong staining in 11 cases (Table 1). FOXP3 positive infiltrating lymphocytes were found in 37 cases. There were 1%-10% of FOXP3 positive cells in the whole tumor section area in 31 cases and more than 30% of FOXP3 positive cells in 6 cases.

Two or more of the investigated proteins were expressed in the same case of diffuse large B-cell lymphoma (FOXP1 and BCL2 in 15 cases; FOXP1 and BCL6 in 3 cases; and FOXP1, BCL2, and BCL6 in 5 cases). Seven cases were negative for all the investigated proteins.

**Fluorescent in situ hybridization**

BCL2 locus abnormalities were observed in 15 cases (Figure 1A). The abnormalities were as follows: 5 cases of translocations (five 2F1R1G), 8 cases of amplifications/aneusomy (one 5R2G; seven 3R2G), and 2 cases of both translocation and amplification/aneusomy (one 2F2R1G; one 2F3R1G). BCL6 abnormalities were observed in 26 cases, as follows:
(Figure 1B); 7 cases of translocations (seven 1F1R1G), 14 cases of amplifications/aneusomy (eight 3F; four 4F; two >5F), and 5 cases of both translocation and amplification/aneusomy (four 2F1R1G; one 3F1R1G). No differentiation was made between amplifications and aneusomy. Only a single case showed a split in the FOXP1 locus (one 1F1R1G), and additional gene copies/aneusomy of the FOXP1 probe were seen in 18 cases (13 3F, 3 4F, 2 5F) (Table 2, Figure 1C). The case with a split in the FOXP1 locus was checked for IGH translocation with an IGH break-apart probe (Vysis) and showed a split signal, indicating the presence of FOXP1/IGH translocation.

More than one gene abnormality was observed in 17 cases as follows: BCL2 and BCL6 in 1 case, FOXP1 and BCL2 in 1 case, FOXP1 and BCL6 in 6 cases, and FOXP1, BCL2, and BCL6 in 9 cases.

FOXPT1 gene abnormalities were usually present together with BCL6 gene abnormalities ($\chi^2 = 8.497, P = 0.004$). A significant association was also observed between the absence

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<th>negative</th>
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*Staining was defined as weak (+), medium (++), and strong (+++) (7,8).

Table 3. Intensity of FOXP1 protein expression in relation to FOXP1 gene abnormalities in 28 patients with FOXP1-positive diffuse large B-cell lymphoma

<table>
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</tr>
<tr>
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<td>0</td>
</tr>
<tr>
<td>Strong</td>
<td>3</td>
<td>6</td>
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</tr>
</tbody>
</table>

*Staining was defined as weak (+), medium (++), and strong (+++) (7,8).
of FOXP1 gene abnormalities and the presence of BCL2 gene abnormalities ($\chi^2 = 10.394, P = 0.001$).

**Association between genetic abnormalities and protein expression**

Cases without FOXP1 gene abnormalities showed either the presence or absence of expression of BCL2 protein. However, cases with FOXP1 gene abnormality showed the expression of BCL2 protein ($\chi^2 = 5.858, P = 0.016$). A significant association between FOXP1 gene abnormalities and FOXP1 protein expression level was also observed. When no gene abnormalities were observed, there was no deregulation of protein expression. Conversely, in cases showing a FOXP1 translocation or additional gene copy, FOXP1 protein was overexpressed ($\chi^2 = 7.157, P = 0.028$) (Table 3).

When there was no BCL2 gene aberration, the BCL2 protein was not expressed, but when there was IGH/BCL2 translocation or additional BCL2 gene copy, BCL2 protein was usually expressed ($\chi^2 = 7.157, P = 0.028$) (Table 3).

Discussion

Our group of diffuse large B-cell lymphoma patients showed additional copy of FOXP1 gene in 36.7% of cases (18 of 49 samples adequate for interpretation) and FOXP1 protein expression in 58% of cases (28 of 50 samples adequate for interpretation), similar to previously published diffuse large B-cell lymphoma data (4,8,27). FOXP3 positive cells were detected in 37 cases.

Cases with positive FOXP1 expression had varying immunostaining intensities, as was expected (10,15). There was a significant association between FOXP1 gene abnormality and protein levels – when there were no gene abnormalities, protein expression was present as weak immunostaining intensity and when there was an additional gene copy, protein expression was present as moderate/strong staining. It is possible that cases with additional gene copies or even chromosome 3 gains are actively transcribing FOXP1 and thus yielding higher amounts of protein through gene dosage effects, although this is not the only known mechanism of FOXp1 expression (de)regulation (27,28).

Only a single case showed FOXP1 translocation. Besides that it also showed IGH translocation, thus possibly forming the FOXP1/IGH translocation. This case showed no FOXP1 expression, since the total count of FOXP1 positive tumor cells was lower than 30%, suggesting that overexpression of FOXP1 protein is not necessarily a result of gene rearrangement (27,29). This was a morphologically clear case of diffuse large B-cell lymphoma which had no BCL2 and/or BCL6 translocations or additional gene copies.

A significant association was found between the presence of FOXP1 and BCL6 gene abnormalities. When there was an additional copy of one of these genes, there was also an additional copy of the other. Since both these genes are located on chromosome 3, this is most likely a result of the aneusomy of chromosome 3 (1).

The association between FOXP1 and BCL2 gene abnormalities requires further in-
vestigation to discover the mechanism underlying this phenomenon. According to our results, if there is no change on one gene there will be no change on the other gene. However, if there is additional $FOXP1$ gene copy then $BCL2$ protein will be expressed. This finding is contradictory to the previously described $BCL2$ and $FOXP1$ association in diffuse large B-cell lymphoma (7) and might be a result of the small number of cases studied in our study.

$BCL2$ gene abnormalities (both translocations and additional copy numbers) appear together with $BCL2$ protein expression, suggesting that $BCL2$ expression in diffuse large B-cell lymphoma is regulated by translocations or copy number changes. $IGH/BCL2$ translocation has been recognized as a cause for constitutive expression of $BCL2$ protein in follicular lymphoma and has been found also in diffuse large B-cell lymphoma (30,31).

We did not find any significant association between GC and ABC group and changes in $FOXP1$ gene/protein. This differs from a previous study (7) and suggests that the division of diffuse large B-cell lymphoma based on the tumor cells origin detected by immunohistochemistry is not informative enough, similar to morphological division (1).

$FOXP3$ expression, indicating the presence of Tregs in a tumor tissue, was also observed, but it did not show any significant association with $BCL2$, $BCL6$, and $FOXP1$. This observation also applies to the abnormalities involving $BCL2$, $BCL6$, and $FOXP1$ genes.

Considering $FOXP3$ positive cells in our diffuse large B-cell lymphoma group, we found no relation between them and any of the analyzed prognostic markers. This suggests a need for further studies that will investigate whether regulatory T cells have any impact on the processes that take place in diffuse large B-cell lymphoma tumor cells.

In conclusion, the results obtained on 53 patients from the Merkur University Hospital showed that there was an association between $FOXP1$ and $BCL2$ in diffuse large B-cell lymphoma patients, but their presence was not related to GC or ABC lymphoma subgroups. $FOXP3$-positive T lymphocytes in these entities were present, but their impact on events in B tumor cells is still not clear.

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References


