



Serotonin

DOROTEA MÜCK-ŠELER
NELA PIVAC

Division of Molecular Medicine,
Ruđer Bošković Institute, Bijenička 54, 10000
Zagreb, Croatia

Correspondence:

Dorotea Mück-Šeler
Laboratory for Molecular Neuropsychiatry
Division of Molecular Medicine
Ruđer Bošković Institute
Bijenička 54, 10000 Zagreb
E-mail: seler@irb.hr

Abbreviations:

alpha-MTrp;
alpha-methyl tryptophan;
AD, Alzheimer's disease;
ADHD attention-deficit/hyperactivity disorder;
BBB, blood-brain barrier;
CSF cerebrospinal fluid;
MAO, monoamine oxidase;
PTSD, posttraumatic stress disorder;
5-HIAA, 5-hydroxyindoleacetic acid;
5-HTT, serotonin transporter;
TPH, tryptophan hydroxylase

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Abstract

Serotonin is a monoamine that could be found in plants, animals and human body. The homeostasis of serotonin is maintained by the series of interdependent processes that include synthesis, storage, transport and removal/degradation. In the human body serotonin is synthesized in two independent compartments that are separated by brain-blood barrier. The majority of serotonin is synthesized in enterochromaffin cells of the gastrointestinal tract, released in the blood stream and stored in blood platelets. About 5% of serotonin is synthesized in the brain within serotonergic neurons. As a neurotransmitter serotonin plays an important role in the regulation of physiological functions like body temperature, sleep, vomiting, sexuality, appetite, behaviour and cognitive functions such as learning and memory. The dysfunction of the serotonergic system has been implicated in the aetiology of a variety of psychiatric (depression, schizophrenia, alcoholism) and neurological (migraine, Alzheimer's disease, epilepsy) disorders. Recent genetic association studies of the neuropsychiatric disorders have focused on functional polymorphisms i.e. DNA sequence variations that alter the expression and/or functioning of the gene product in the loci encoding different genes. Some of them are genes for tryptophan hydroxylase, serotonin transporter and serotonergic receptors.

THE BEGINNINGS

Serotonin (5-hydroxytryptamine, 5-HT) was discovered 60 years ago in blood, peripheral tissues and central nervous system (1). It was first identified as a vasoconstrictor substance that is released from platelets during the coagulation of blood, and later as a monoamine neurotransmitter in the brain. It has been established that gastrointestinal tract, blood platelets and brain were the main locations of serotonin in the mammal's body. In addition, serotonin could be also found in plants (bananas, walnuts, tomatoes, hickories, pineapples), mushrooms, octopuses, and in poison of insects (spiders, scorpions, wasps).

Synthesis and metabolism

Serotonin is a tryptamine that consists of an indole ring with a hydroxide group on the fifth C atom, and a carboxyl-amide side chain (Figure 1).

The main precursor of serotonin is the essential amino acid L-tryptophan that must be provided by food. L-tryptophan and the other precursor, 5-hydroxytryptophan, are transported from blood to brain by the active carrier system located in the blood-brain barrier (BBB). It is believed that serotonin, due to its chemical properties, does not cross BBB, but new evidence (2) suggests that serotonin might cross endothelial cells of the BBB using serotonin transporter. The serotonin synthesis occurs in a two-step enzymatic procedure (Figure 2). The first

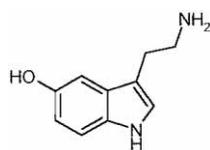


Figure 1. The chemical structure of serotonin or 3-(2-aminoethyl)-5-hydroxyindole.

and rate-limited step is hydroxylation of the tryptophan to 5-hydroxytryptophan. This reaction is catalyzed by tryptophan hydroxylase, a specific enzyme located only in the serotonergic neurons (3). The 5-hydroxytryptophan is decarboxylated by a nonspecific enzyme, aromatic amino acid decarboxylase, into serotonin. Serotonin levels could be determined in the brain and various body fluids including serum/plasma, platelets, cerebro-

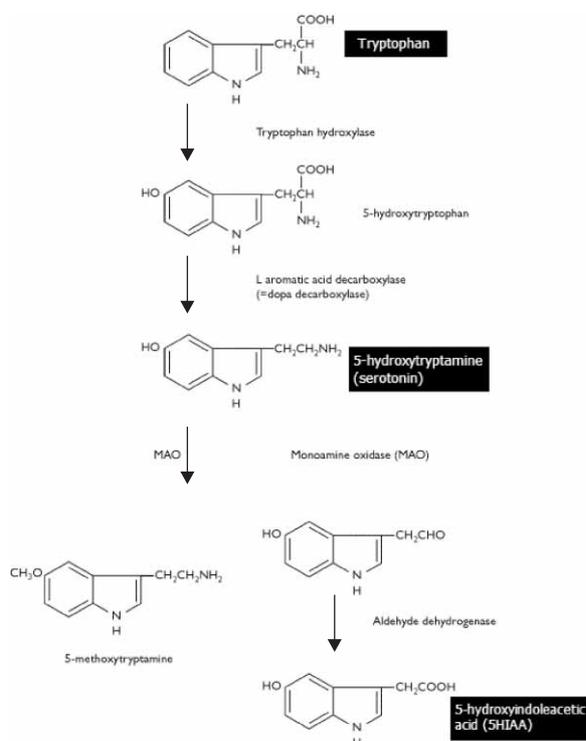


Figure 2. Synthesis and degradation of serotonin.

spinal fluid (CSF) and urine. The flavine-containing mitochondrial enzyme monoamine oxidase (MAO) is the most important enzyme for degradation of serotonin. This process has two steps: first step is a degradation of serotonin into 5-hydroxyindole acetaldehyde, and the second one is degradation through aldehyde dehydrogenase regulated conversion into 5-hydroxyindoleacetic acid (5-HIAA) as the main metabolite of serotonin.

In the human body, serotonin is synthesized and located in two compartments that are separated by BBB. The first compartment, called »peripheral« compartment, contains the majority (about 95%) of serotonin in the body. Peripheral serotonin is synthesized in enterochromaffin cells of the gastrointestinal tract. The second, i.e. »central« compartment of serotonin is the central nervous system that synthesizes serotonin within serotonergic neurons. Recent studies (3, 4) revealed that tryptophan hydroxylase exists in two isoforms with different location within the body of mammals (Table 2). Tryptophan hydroxylase type 1 (TPH1) is responsible for the synthesis of the peripheral serotonin (4), while tryptophan hydroxylase type 2 (TPH2) is the predominant isoform in the brain.

Serotonergic receptors

The complex functions of serotonergic system would be impossible without a large number of serotonergic receptors (5). Serotonergic receptors are classified in seven different groups or »families« called 5-HT₁, 5-HT₂, 5-HT₃, 5-HT₄, 5-HT₅, 5-HT₆, 5-HT₇, and several subtypes (Table 1) that differ in terms of structure, action, and location. Serotonergic receptors are distributed on the presynaptic and postsynaptic neurons in the central nervous system and on the different peripheral cells and organs (Table 1). The majority of serotonin receptors are G protein-coupled receptors. The exception is 5-HT₃ receptor that belongs to the ligand-gated ion channel receptors. Serotonergic receptors activate an intracellular second messenger (cAMP, IP₃, DAG) cascade and produce an excitatory or inhibitory response. Serotonin receptors are very important sites of action for different classes of psychotropic drugs, like antidepressant drugs (5), atypical antipsychotic drugs (olanzapine, risperidone) and psychoactive compounds (LSD, DMT).

TABLE 1

The serotonergic receptors.

Type	Subtype	Distribution	Intracellular response
5-HT-1	1A, 1B, 1D, 1E, 1F	CNS, blood vessels	Inhibitory
5-HT-2	2A, 2B, 2C	CNS, platelets, blood vessels, smooth muscle	Excitatory
5-HT-3	3A, 3B	CNS, GI tract	Excitatory
5-HT-4		CNS	Excitatory
5-HT-5		CNS	Inhibitory
5-HT-6		CNS	Excitatory
5-HT-7		CNS, GI tract, blood vessels	Excitatory

Serotonin in the central nervous system

Serotonergic neurons are widely distributed throughout the mammalian brain and serotonergic system is the largest single system in the brain. The nine groups of serotonergic cell bodies are located mainly in the area of brain stem raphe nuclei. Serotonergic nerve terminals could be found in nearly all other regions of the central nervous system. The widespread distribution of the raphe projections suggests a highly collateralized axon system (6). The communication of serotonergic system with other important neurotransmitter systems like catecholaminergic system (7) is well established, although the mechanisms of interaction are not yet completely understood. Significant amounts of data have demonstrated that these interactions are very important in the mechanisms of action of antidepressant and anxiolytic drugs.

Serotonin is implicated in many physiological (body temperature, sleep, vomiting, sexuality, appetite), behavioural (aggression, mood) and cognitive (learning, memory) functions (8). In addition, serotonin has an important role in the growth of the central nervous system during development. It plays a critical role as a growth factor in the immature brain, directing both proliferation and maturation. This is supported by the higher serotonin turnover rate in the immature mammalian brain than at any other time in life. Recent data (9) suggested that an overload of serotonin during cortical development could induce abnormal distribution and incorrect positioning of cortical interneurons.

There are several methods for the determination of serotonin synthesis rate *in vivo*: a) pharmacological manipulation, i.e. after the administration of compound that inhibits enzyme aromatic acid decarboxylase like NSD-1015, b) the use of radiolabel tryptophan, c) the use of radiolabel alpha-methyl tryptophan (alpha-MTrp) as an analogue of tryptophan. The limitation of the pharmacological methods is that the effect of the tested compound on the serotonin synthesis could be in part influenced by the pre-pharmacological manipulation itself. Radiolabel tryptophan is an essential amino acid incorporated in proteins, while radiolabel tryptophan metabolites, like serotonin and 5-HIAA, are lost very rapidly from the brain. The use of alpha-MTrp labelled with ^3H or ^{14}C and the determination of serotonin synthesis by an autoradiographic method (10) permits the measurement of serotonin synthesis in the rat brain with high anatomical resolution and without any pharmacological pre-treatment. The disadvantages of this method are the need for special equipment and a long procedure.

The alteration of the serotonergic system has been related with the aetiology of different neurological (migraine, Alzheimer's disease, epilepsy) and psychiatric (depression, schizophrenia, mood disorders, alcoholism, ADHD, PTSD) disorders.

Peripheral serotonin

Peripheral serotonin is synthesized in enterochromaffin cells of the gastrointestinal tract. The synthesis is reg-

ulated by TPH1 (4). From gut serotonin is released in the blood stream and then stored mostly in blood platelets. The other peripheral cells that contain serotonin are macrophages and mast cells. Peripheral serotonin is metabolized in the liver by a MAO-A to 5-HIAA. The 5-HIAA is filtrated and excreted by the kidney. The vast increase in urine excretion of 5-HIAA was found in carcinoid syndrome, due to the pronounced production of serotonin by carcinoid cells.

In humans a direct association between neurotransmitters in the brain and those excreted in the urine is not yet defined. New evidence suggests (2) that neurotransmitters excretion in the urine might be used as possible biomarkers of the central nervous system activity. The study in rats treated with 5-hydroxytryptophan has shown simultaneous changes in the activity of brain serotonergic system and urinary serotonin levels.

Serotonin has an important role in the vascular biology. It is involved in the control of vascular resistance, blood pressure, haemostasis and platelet function (11). One of the most important functions of the peripheral serotonin is the promotion of platelet aggregation and blood clotting. Activated 5-HT_{2A} receptors on platelet membrane also stimulate platelet activation and aggregation.

Blood platelets and serotonergic neurons

The function of central serotonin in mood, state of mind, and behaviour, as well as its role in cognition and memory are very difficult to establish. Literature data suggest that blood platelets can be used as an easy obtainable peripheral model for the some processes in the central serotonergic neurons (12–14) (Table 2). The kinetic and pharmacological characteristics of the active transport of serotonin from plasma to platelets are similar to the reuptake of serotonin from synaptic cleft into presynaptic neurons. In addition, platelet MAO type B activity corresponds to MAO-B activity in presynaptic part of neurons. There are also 5-HT_{2A} receptors (14), α_2 -adre-

TABLE 2

Similarities and differences between blood platelets and serotonergic synaptosomes.

Brain serotonergic neuron	Blood platelets
Similarities	
Serotonin stored in dense bodies or vesicles	
Active transport (uptake) of serotonin	
Binding sites for ^3H -paroxetine, ^3H -imipramine	
Receptors: 5-HT _{2A} and α_2 -adrenergic	
Monoamine oxidase type B	
Differences	
Serotonin synthesis	No synthesis
Function: Neurotransmission	Function: aggregation, vasoactive compound

nergic receptors (15) and binding sites for ^3H -imipramine and ^3H -paroxetine on the platelet membrane, which can be used as peripheral markers for the evaluation of pharmacologic and kinetic characteristics of equivalent central nervous receptors and binding sites on presynaptic or postsynaptic part of the serotonergic neurons in psychiatric and neurodegenerative disorders.

Numerous studies have tested platelet serotonin concentrations in various psychiatric disorders. Altered platelet serotonin concentrations were found in patients with different psychiatric and neurological disorders like major depression (16), subtype of major depression with psychotic symptoms (17), bipolar disorder (18), schizophrenia (16, 19–21), postpartum depression (22), post-traumatic stress disorders (PTSD) with comorbid depression (23), PTSD with psychotic symptoms (24), alcoholism (25), attention-deficit/hyperactivity disorder (ADHD) with impulsive symptoms (26), migraine (27), and Alzheimer's disease (28, 29).

SEROTONIN AND NEUROPSYCHIATRIC DISORDERS

Preclinical and clinical investigations suggest that serotonin could be related to the aetiology and treatment of different neuropsychiatric disorders including depression, schizophrenia, PTSD, alcoholism, ADHD and Alzheimer's disease (AD).

Depression

Depression is a severe and devastating mental disorder with a high prevalence worldwide, and with twice higher risk in women than in men. Although, the pathogenesis and treatment of depression were the topics of intensive preclinical and clinical research, the complex neurobiological basis of depression is still unclear (30). The first hypothesis of depression was established almost forty years ago (31). It postulated that depression is a consequence of the low serotonin and/or noradrenalin levels and the dysfunction in the central serotonergic, noradrenergic and dopaminergic systems. These neurotransmitters interact with each other and regulate mood, sleep, anhedonia, appetite, concentration and motivation, suicidal behaviour, cognitive and autonomic functions (8) that are frequently disturbed in depression.

In following years numerous attempts were made to identify the reproducible neurochemical alterations in the nervous systems of patients with depression, but mostly with negative or inconsistent results. Concentrations of 5-HIAA, the major 5-HT metabolite, in CSF have been extensively studied in depressed subjects. A significant reduction in CSF 5-HIAA was found in depressed patients (32), but several other studies (33) were not able to confirm the difference in 5-HIAA levels between depressed patients and healthy controls. Recent study (34) described elevated brain serotonin turnover rate in medication-free patients with depression, particularly in those carrying the short form (s allele) of the gene for serotonin transporter. However it was not clear if the

elevated brain serotonin turnover is a consequence of increased neuronal activity, enhanced vesicular leakage and subsequent intraneuronal metabolism or reduced brain serotonin transporter availability.

Platelet serotonin levels, platelet 5-HTT and platelet 5-HT_{2A} receptors were also investigated in depression. The decreased (35) or unaltered (19, 36) platelet serotonin levels were found in depressive patients as well as altered number of platelet 5-HT_{2A} receptors and the number of platelet serotonin uptake sites (37).

The majority of antidepressant drugs in current use, act by affecting the neurotransmitters (serotonin, noradrenalin, and dopamine), their receptors and enzymes involved in their synthesis or degradation (38). However, the clinical improvement after antidepressant therapy is usually observed 2–3 weeks after the beginning of the treatment. It is believed that this therapeutic delay depends on the antidepressant-induced desensitization of serotonergic or noradrenergic receptors.

Schizophrenia

Schizophrenia is a complex and multifactorial mental disorder with the prevalence of 1% worldwide. The alterations of the dopaminergic system and their receptors are the basis for the »dopaminergic hypothesis« of schizophrenia. Since this hypothesis is not able to explain the complex symptoms of disease and efficacy of atypical antipsychotics, with a higher affinity for 5-HT_{2A} than to D₂ receptors, it is supposed that schizophrenia might be associated with the dysfunction of other neurotransmitter systems like serotonergic system (39). In addition serotonergic system regulates some physiological and behavioural functions that are disturbed in schizophrenia (8). In line with this »serotonin hypothesis of schizophrenia« the alteration of serotonergic activity was found in the brain of schizophrenic patients, with the decreased serotonin neurotransmission in cortical regions, and increased in the putamen, accumbens and pallidus. The decreased or unaltered density of 5-HT₂ receptors in frontal cortex (39), decreased density of 5-HT_{1A} receptors and unchanged 5-HT₆ receptor binding (40) were also found in schizophrenic patients.

An increase in platelet serotonin concentration was observed in schizophrenic patients with predominantly chronic time course (41), with paranoid symptoms (19), positive symptoms (42) and in schizophrenic patients born in winter (20). Although there is no direct evidence that platelets serotonin concentration correlates with central serotonin levels, an abnormal tomographic brain scans were found in schizophrenic patients with high platelet serotonin levels (43).

Posttraumatic stress disorder

Posttraumatic stress disorder (PTSD) is a severe psychiatric and polygenetic disease that appears in some people that survived the extremely dangerous traumatic life event like natural disasters, war or sexual abuse. PTSD could be associated with alterations in different

neuroendocrine (44) and neurotransmitter (45) systems. Despite the intensive neurobiological research, the role of serotonin in the pathophysiology of PTSD is still unclear. Some studies (46) suggested altered serotonergic function in PTSD that might contribute to cognitive disturbances, depressive symptoms and many physiological and pathological behaviours, such as aggression, that frequently arises in PTSD.

It is believed that serotonergic neurons from dorsal nuclei raphe with nerve terminals in hippocampus and amygdala are responsible for anxiogenic response to stress via 5-HT_{2A} receptors, while neurons from median raphe have anxiolytic effect achieved through 5-HT_{1A} receptors. However, a positron emission tomography study (47) did not find altered 5-HT_{1A} receptors binding in patients with PTSD compared to healthy subjects. Indirect peripheral evidences for the altered efficacy of serotonergic system in PTSD are decreased serum serotonin concentration and altered number of platelet serotonin transporters (48). Platelet serotonin concentration was higher in PTSD veterans with psychotic subtype of PTSD compared to platelet serotonin concentration in non-psychotic war veterans or in healthy controls (24). Since platelet serotonin concentrations correlated with the severity of delusions, the core psychotic symptoms, these data confirmed that platelet serotonin concentration might be used as a trait marker of psychotic symptoms in PTSD.

Alcoholism

Literature data suggest that alcohol dependence and alcohol abuse could be associated with the disturbance in serotonergic system (49). The role of serotonin in alcoholism is based on the data showing alterations in measures of the serotonergic function in the brain, as well as in CSF, blood precursor availability, uptake of serotonin in blood platelets and challenge studies.

Post-mortem brain analyses have found reduced serotonin transporter binding in the hippocampus (50) or in dorsal striatum (51), and decreased density of 5-HT_{1A} receptors (52) in patients with alcoholism compared to non-alcoholic controls. Reduce activity of serotonergic transporter in the brain of abstinent alcoholics was confirmed in vivo using SPECT (53). The values of 5-HIAA concentration in CSF are in line with central serotonergic disturbances in patients with alcoholism. Low levels of 5-HIAA in CSF were found in early-onset alcoholics (54), in abstinent alcoholics (55) and in alcoholic impulsive offenders (56).

Decreased plasma tryptophan levels and low serotonin precursor availability suggest impaired serotonin synthesis in alcoholism (49). The results on the blood platelets serotonin transporter activity in patients with alcoholism are inconsistent. Lower (57), increased (58), or unaltered (59) serotonin uptake into platelets were found in alcoholics when compared to healthy controls. Alcoholism-induced fall of serotonin transporter activity has been related to a decreased platelet serotonin content observed

in male and female alcoholic patients, independently on the presence of comorbid psychiatric disorders (60).

It has been shown that prolactin or cortisol response to administration of serotonergic drugs like fenfluramine, m-chlorophenylpiperazine, 6-chloro-2-1-piperazinylpyrazine, and adrenocorticotrophic hormone (ACTH) response to m-chlorophenylpiperazine, was lower in alcoholics than in non-alcoholic patients (61), suggesting also altered central serotonergic function in alcoholism.

ADHD

Serotonin dysfunction has been implicated also in ADHD, although the primary neurotransmitter that is altered in ADHD is dopamine, and to a lesser extent, noradrenalin (62). However, besides the classical characteristics symptoms of ADHD (such as hyperactivity, inattention and impulsivity), aggression, as well as disturbances in the cognition, are also frequent in ADHD. These findings confirmed also the role of serotonin in ADHD (63). The concentration of serotonin was found to be lower in ADHD (64), or unaltered in ADHD (26). Since ADHD is a multifactorial and clinically heterogeneous psychiatric disorder, platelet serotonin concentration was found to be increased in children with ADHD with pronounced impulsive symptoms (26) suggesting that higher platelet serotonin concentration in impulsive compared to non-impulsive children with ADHD might be used as a possible trait marker of impulsivity in ADHD. The impulsivity is associated with serotonin function in non-clinical sample (65). There is a significant correlation between impulsivity and lower serotonergic function (66). On the other hand, the opposite data exist, and therefore impulsivity has been associated also with the increased serotonergic functioning (67) in children and adolescents.

Alzheimer's disease

Alzheimer's disease (AD) is a neurodegenerative disorder characterized with a progressive loss of cognitive functions such as learning and memory. The aetiology and pathophysiology of AD is still unclear. The neurobiological alterations in AD include accumulations of amyloid plaques outside and neurofibrillary tangles inside neurons and the dysfunctions of cholinergic, catecholaminergic and serotonergic systems. The decrease in the brain concentration of serotonin and 5-HIAA was found in AD (68). The loss of presynaptic somatodendritic 5-HT_{1A} autoreceptors and postsynaptic 5-HT_{1A} heteroreceptors (68), and 5-HT₂ receptors in cerebral cortex (69), were also found in patients with AD. It seems that the development of behavioural and psychological symptoms in AD (70) is related to the genetic variants of 5-HT_{2A} and 5-HT_{2C} receptors. In addition, daily living and functioning was improved in patients with AD treated with combination of rivastigmine and selective serotonin reuptake inhibitor, fluoxetine.

The reduced serotonin concentration in platelets (29), CSF (71) and brain (72) of patients with AD would sug-

gest the decrease in serotonin synthesis. The main factors that influence serotonin synthesis are plasma availability of its precursor tryptophan and the activity of the rate-limiting enzyme TPH. Plasma level of tryptophan depends on the dietary intake and feeding behaviour that could be also changed in AD. In patients with AD low tryptophan concentrations in serum (73), plasma (74) and CSF (71) was found in some, but not all studies (75). The alterations in serotonergic and kynuramine pathways of tryptophan metabolism have been connected to pathophysiology of AD (76), suggesting that low plasma tryptophan concentration in AD might be also a consequence of the enhanced tryptophan degradation via the kynuramine pathway (73). In addition, tryptophan depletion in healthy volunteers (77) or in patients with mild to moderate AD (78) induced changes in cognitive performance.

Altered serotonin synthesis in AD (72) might be a consequence of reduced TPH activity in particular brain areas of patients with AD (79) or the lack of the TPH cofactors tetrahydrobiopterin and folic acid. Since TPH activity is sensitive to reactive oxygen species, the tetrahydrobiopterin deficiency could also impair serotonin synthesis through oxidative damage of TPH (80).

A decreased platelet serotonin concentration observed in patients with AD in the late phase of disease (29), might be related to the reduced serotonin active transport through platelet membrane. This finding is in line with the decrease in the maximum number (V_{max}) of serotonin transporters found in severely ill patients with AD compared to both patients with mild AD and healthy controls (81).

CANDIDATE GENES OF THE SEROTONERGIC SYSTEM

Recent genetic association studies of the neuropsychiatric disorders have focused on functional polymorphisms i.e. DNA sequence variations that alter the expression and/or functioning of the gene product in the loci encoding different genes. Some of them are genes for TPH, serotonin transporter (5-HTT) and serotonin type 1A (5-HT1A), 1B (5-HT1B), 2A (5HT2A) receptors.

Tryptophan hydroxylase

Tryptophan hydroxylase (EC 1.14.16.4) is a key enzyme in the synthesis of serotonin. It metabolizes the essential amino acid L-tryptophan, by hydroxylation at the position 5, to the serotonin precursor L-5-hydroxytryptophan (Figure 3). The cofactors are O_2 (dioxygen), BH_4 / (6R)-L-erythro-5,6,7,8-tetrahydrobiopterin/ and Fe^{2+} . TPH is a specific and selective enzyme that is only located in the serotonin producing cells like serotonergic neurons, pineal gland and enterocromaffine cells of the gut. It exists in two isoforms (3), with different location within the body of mammals (Table 3). One isoform is TPH1 that is responsible for the synthesis of the peripheral serotonin (4), while TPH2 is the predominant isoform in the brain.

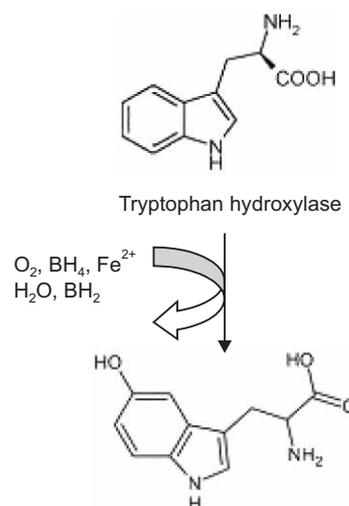


Figure 3. Tryptophan hydroxylase catalyzes tryptophan to 5-hydroxytryptophan.

In humans, the genes for TPH1 (MIM *191060) and TPH2 (MIM *607478) are located on chromosomes 11 at position 11p14-p15.3 and 12 (12q21), respectively. The human TPH2 gene spans 97 kilobases (kb) and consists of 11 exons. The sequence identity between two homologous TPH is 71% (121). In TPH1 knockout mice the concentration of peripheral serotonin was reduced to 6% (blood), 1.5% (jejunum) and 0.1% (colon) as compared to serotonin concentration in wild type mice (3, 82). However, genetically modified mice without TPH1 isoform had normal serotonin concentration in hippocampus and frontal cortex (82), and similar development, appearance and behaviour as mice with TPH1 gene. Those results suggested that serotonin synthesis in the brain depends mostly on the TPH2 isoform activity. Recently, Savelieva *et al.* (3) reported the phenotypic evaluation of TPH2 knockout mice and double knockout mice without both TPH1 and TPH2 gene. They have found the pronounced decrease in serotonin levels in cortex, thalamus/hypothalamus, olfactory bulb, cerebellum hippocampus, brainstem and striatum in both groups of knockout mice compared to wild type mice. The lowest serotonin concentration was determined in double knockout mice without TPH1 and TPH2 gene. Although genetically modified mice were similar in appearance, or histological analysis with no loss of the serotonergic cell bodies in raphe nucleus, there were differences in the body weight, body size and percent of body fat in male, and percent of body fat in female double knockout mice compared to wild type mice (3). There were a few effects of *TPH* genotype on behaviour. Mutant mice had similar exploratory behaviour and locomotor activity and showed increased anxiety-like behaviour (3).

Several studies analyzed the association between genetic variants of the TPH in neuropsychiatric disorders and emotion-related personality traits, but with inconsistent results. Zhang *et al.* (83) reported that allele A of the functional polymorphism G1463A at TPH2 gene could

TABLE 3

Two tryptophan hydroxylase isoforms, their location and function of the corresponding central or peripheral serotonin.

Tryptophan hydroxylase isoform	
TPH1	TPH2
Location	
Peripheral tissue	Brain
Serotonin functions	
Migraine	Migraine
Vasoconstriction	Sleep
Haemostasis	Behaviour
Immune system	Mood
Intestinal motility	Food intake and body weight
Melatonin synthesis	Neuropsychiatric disorders

be related to the low TPH2 activity and consequently to the impaired brain serotonin synthesis in depressed patients. Their suggestion that 1463A allele could be a risk factor for unipolar major depression were not confirmed in the larger number of patients of the West European Caucasian origin with affective disorders (84), or in Chinese population with unipolar depression (85). Recent, haplotype analysis of the eight polymorphisms: rs4448731 (intergenic), rs4565946 (intron 2), rs11179000 (intron 4), rs7955501 (intron 5), rs10506645 (intron 7), rs4760820 (intron 8), rs1487275 (intron 8) and rs10879357 (intron 8) of the human TPH2 (86) has shown no association between TPH2 polymorphisms and bipolar depression and suicidal behaviour in Brazilian population. There were also no significant differences in genotype or allele frequencies of the TPH2 -703G/T polymorphism between bipolar patients and healthy subjects of the Korean origin (87). The meta-analysis of TPH1 and TPH2 gene variants in the large number of subjects with ADHD from four independent European Caucasian samples have shown no consistent evidence for common genetic variants in the TPH1 and TPH2 regions in ADHD (88).

It has been hypothesized that smaller volumes of the amygdala and hippocampus are related to the presence of the T allele of the TPH2 (rs4570625; G-703T) polymorphism in Japanese subjects. This was the first study that compared personality traits (harm avoidance, reward dependence, novelty seeking, persistence, self directedness, cooperativeness, self-transcendence) assessed with the Temperamental and Character Inventory (89), with the volume of amygdala and hippocampus (determined using magnetic resonance imaging) in T allele carriers and GG individuals. The results suggested that regional brain volume could be a brain structural intermediate phenotype between genetic variations in TPH2, i.e. between serotonin synthesis and personality traits related to mood or anxiety disorders (89).

Serotonin Transporter-Linked Polymorphic Region

The serotonin transporter (5-HTT) is an important protein responsible for the active transport of serotonin into neurons, enterochromaffin cells and platelets. In the brain 5-HTT is localized in membrane of presynaptic nerve terminals and in dendritic arbors close to serotonergic cell bodies. 5-HTT regulates serotonin levels in the synaptic cleft following neuronal stimulation, and consequently the magnitude and duration of its effect on postsynaptic serotonergic receptors. It terminates the action of serotonin by rapid reuptake of released serotonin from synaptic cleft into presynaptic neuron by means of an active transport process that depends upon maintenance of ion gradients across the cell membrane by $\text{Na}^+\text{-K}^+\text{-ATPase}$ (90). The *in vitro* and *in vivo* (91) studies have shown that 5-HTT is not only a protein with important role in the homeostatic regulation of the serotonergic function, but also a site of action for several classes of antidepressant drugs including classical tricyclic compounds and novel selective serotonin reuptake inhibitors (91).

Among a number of genes involved in the synthesis of protein related to metabolism and function of serotonergic system, gene encoding the 5-HTT (*SLC6A4*) is the most extensively investigated. The 5-HTT gene (OMIM *182138) is located on chromosome 17 at position 17q11.1-17q12 and consisted of fifteen exons encoding a protein of 630 amino acids with 12 transmembrane domains (90, 92). The most studied variants in the 5-HTT gene are the serotonin transporter-linked polymorphic region (5-HTT gene-linked polymorphic region, 5-HTTLPR; rs795541) and a functional variable number of tandem repeats (VNTR) polymorphism in intron 2. 5HTTLPR is a repetitive element of varying length in the 5' flanking region located 1.4kb upstream of the transcription start site that modulates transcriptional activity of human 5-HTT (90). A deletion/insertion in the 5-HTTLPR creates two alleles (each of 20 to 23 bp): short or S allele and long or L allele made up of fourteen and sixteen repeated elements, respectively. Some studies have found that long allele results in higher serotonin transporter mRNA transcription in human cell lines. The uptake of serotonin is two-fold higher in cells containing the homozygous L/L form of the *SLC6A4* than either the L/S or S/S forms. On the other hand, S allele is associated with reduced transcriptional efficiency and decreased 5-HT expression and uptake (92).

It has been suggested that this 5-HTTLPR polymorphism alters the promoter activity and consequently serotonergic functions (93). In this respect, a significantly higher maximal number (B_{max}) of platelet serotonin uptake sites was found in subjects carrying the L/L genotype as compared with B_{max} of platelet serotonin uptake sites in individuals with L/S or S/S genotype (94). Additionally, the uptake of serotonin is approximately two-fold higher in cells containing the homozygous L/L form of the 5-HTT, while S allele is related to reduced transcriptional efficiency and therefore decreased serotonin uptake (92). Volumetric neuroimaging studies have

shown that the *S* allele is associated with reduced grey matter volume in the limbic system and disrupted amygdala-cingulate coupling (95).

The intron 2 VNTR contains nine, 10, or 12 copies of a sixteen- or seventeen-base pair repeat (96). A stronger expression and greater enhancer activity was observed by the 12-copy allele than by the 10-copy allele in the hind-brain of transgenic embryonic mice (97) and in embryonic stem cells (98) suggesting its functionality. Very poor linkage disequilibrium was found between 5HTTLPR and the intron 2 VNTR (99).

A lot of studies conducted in healthy individuals and in patients with psychiatric and neurological disorders suggested that 5-HTTLPR could be considered as a candidate gene for depression (99), mood disorders (100), alcoholism, autism and stress related psychiatric disorders, while other studies did not replicate these data (101). An association between early stressful life event (childhood maltreatment, abuse, lack of social support) and increasing risk for the development of depression in subjects carrying short alleles was found (102). In addition, there is a growing body of literature suggesting the connection between stressful life events and occurrence of depression that may be dependent on variation at the 5-HTTLPR locus of the *5-HTT* (102). However, recent meta-analysis (103) did not confirm that the 5-HTTLPR genotype and stressful life events, alone or in combination, are risk factors for the development of depression in both male and female subjects.

Different findings related to the association between 5HTTLPR and psychiatric disorders might be induced by differences in population genetic structure and sub-structure between cases and controls, and to ancestry differences corresponding to ethnic groups. There are significant differences in the allele frequencies in 5-HTTLPR between Caucasian and Asian populations, since *S* allele is found in 42% of Caucasians and in 79% of Asians (104). In the large groups of healthy subjects from Croatia and the Russian federation (Russians, Bashkirs and Tatars) there were significant ethnic differences in allele and genotype frequencies of the 5-HTTLPR (104). These findings might explain the contradictory results showing the positive or negative or no association between various psychiatric disorders, treatment outcomes and 5-HTTLPR across different populations.

The association between 5HTTLPR and s platelet serotonin concentration was also studied, but the findings were contradictory, showing positive, negative or a lack of association (105–107). The study including large groups of healthy male and smaller groups of female Caucasian subjects of Croatian origin, free of neuro-psychiatric disorders, showed also a lack of association between 5-HTTLPR genotypes and platelet 5-HT concentration, and failed to detect the functional relevance of the 5-HTTLPR variants on platelet 5-HT concentration (107). Therefore, since a positive significant association between *L/L* and *L/S* genotypes and increased blood serotonin levels was detected in patients with obsessive-com-

pulsive disorder, the results of the lack of association between 5HTTLPR and platelet serotonin concentration suggest that genotype-induced changes in 5-HTT transcription and consequent changes in platelet serotonin concentration might be differently regulated in healthy subjects and psychiatric patients (107).

The relationship between the changes in serotonin uptake and 5-HTT gene in patients with AD is inconsistent. A study (108) failed to find the difference in the allelic distribution on the deletion/insertion polymorphism of the 5-HTT gene between patients and controls. In contrast, an association between long allele of the 5-HTT gene and development of aggressive behaviour in AD was found (109).

Serotonin type 1A receptor

Serotonergic receptor type 1A (5-HT_{1A}) is one of the most investigated and characterized serotonergic receptors. The 5-HT_{1A} receptors were found in a variety of human brain regions (110). They are subdivided according to their location in a) presynaptic somatodendritic 5-HT_{1A} autoreceptors located on cell bodies and dendrites of serotonergic neurons in dorsal and median raphe nuclei and b) 5-HT_{1A} heteroreceptor positioned postsynaptically in the pyramidal cells and interneurons of cortico-limbic regions (hippocampus, cerebral cortex and lateral septum) that received serotonergic input from the raphe nuclei (5). The activation of presynaptic 5-HT_{1A} autoreceptors by serotonin or its agonists inhibits the firing rate of the serotonergic neuron and reduces serotonin synthesis, turnover and release (111) and thus affects the serotonergic activity in projection areas. In addition, 5-HT_{1A} receptors have an important role in the neurodevelopmental processes such as synapse formation, neurite outgrowth and neuronal migration (112). Due to their wide distribution and multitude of functions, 5-HT_{1A} receptors are associated with aetiology and treatment of mental disorders (113), especially major depressive disorder.

Post-mortem evaluation of the number and/or affinity of 5-HT_{1A} receptors in the brain of depressed patients revealed different results among studies. Reduced 5-HT_{1A} ligand binding or receptor gene expression was found in cortico-limbic regions of the suicide depressed victims with or without medication at the time of death (114). The increase in 5-HT_{1A} receptor binding (115) and negative results (116) were also reported. This lack of consistency between studies can be due to the variety of factors including sex, comorbidities (substance abuse, alcohol dependence and other psychiatric diagnoses), post-mortem delay and to the different methods used for the measurement of the 5-HT_{1A} ligand binding. The desensitization of presynaptic 5-HT_{1A} autoreceptors, but not postsynaptic receptors was observed after chronic treatment with antidepressants like serotonin reuptake inhibitors and monoamine oxidase inhibitors.

The 5-HT_{1A} receptor is coded by the *HTR1A* gene (OMIM *109760) located on chromosome 5 at position

5q11.2-q13. Recently, the 27 single nucleotide polymorphisms of the *HTR1A* gene were described (113). Among them the most investigated, and the most prevalent in normal human population is a functional polymorphism C-1019G (rs6295), located within the promoter region of the gene. It has been shown that this polymorphism regulates gene expression (117). Genetic studies suggested that the G allele of the C-1019G is associated with an increase in presynaptic 5-HT1A receptor expression and lowered expression of the postsynaptic 5-HT1A receptor. These opposite effects could be related to two transcription factors: epidermal autoregulatory factor-1 (Deaf-1 or NUDR) and Hes5 that are co-localized on both, presynaptic and postsynaptic 5-HT1A receptors (117, 118). Deaf-1 is a repressor at somatodendritic 5-HT1A receptor, but enhances the transcription in non-serotonergic neurons that express postsynaptic 5-HT1A receptors.

Several studies investigated the association between C-1019G *HT1A* promoter polymorphism and regional binding potential of two selective 5-HT1A antagonists ¹¹C-WAY100635 or ¹⁸F-MPPF using Positron Emission Tomography (PET) in healthy subjects, but with inconsistent results (119). Concerning response to treatment, the C-1019G variant seems to be of primary interest in antidepressant response: C allele carriers generally show a better response to treatment, especially in Caucasian samples (113).

Serotonin type 1B receptor

Serotonergic receptors type 1B (5HT1B) were found to be rodent specific and similar to the human serotonergic receptor type 1D (5-HT1D). Since these two types of 5-HT1 receptors share similar brain distribution, transductional features and function, it has been suggested that they are species homologues with 97% overall sequence homology (5).

Serotonergic receptors type 1B (5HT1B) are terminal autoreceptors and postsynaptic heteroreceptors (5), located in the central nervous system in the basal ganglia, striatum, hippocampus and cortex. 5-HT1B receptors were also found on the variety of vascular tissues including cerebral arteries. The main function of autoreceptors located on the nerve terminals is the control of serotonin release, but they may act as terminal heteroreceptors controlling the release of other neurotransmitters like catecholamine and GABA (5).

A recent study (120) has shown the co-localization and interaction of 5-HT1B receptors and protein p11 at the cell surface in vitro. The p11 knockout mice exhibited a depression-like phenotype and had reduced responsiveness to 5HT1B receptor agonists and reduced behavioural reactions to antidepressants. The decrease in brain p11 expression was found in the animal model of depression and in the brain tissue from depressed patients, while an over expression in p11 and the increase in 5HT1B receptor function was observed in rodent brains after antidepressants or electroconvulsive therapy (120).

Animal studies suggested that 5-HT1B receptors have a role in aggressive behaviour related to alcoholism. Knockout mice showed increased alcohol intake and more propensities to aggressive behaviour, although other studies (121) did not confirm the relationship between 5-HT1B receptors and alcoholism. The 5-HT1B receptor gene has been postulated to play a modulatory role in alcohol consumption and alcohol dependence, and was considered as candidate gene for alcoholism (122).

In humans 5-HT1B receptor is encoded by the gene *HTR1B* (OMIM*182131) located on chromosome 6 within the region 6q13-q26 at position 6q14.1. Intronless *HTR1B* gene consists of a single exon, encoding a 390 amino acid peptide (123). A number of polymorphisms has been discovered in the coding sequence and surrounding 5' and 3' untranslated regions. *5HTR1B* gene is an attractive candidate for studies of the genetic basis of ADHD (124). The most widely studied polymorphism of the *HTR1B* gene in and around the *HTR1B* locus is relatively common synonymous G861C polymorphism (124).

Serotonin type 2A receptor

Serotonin type 2A receptor (5HT2A) is a G protein coupled serotonergic receptor located on the membrane of postsynaptic serotonergic neurons. High concentrations of 5-HT2A receptors were found on the apical dendrites of pyramidal cells in layer V in cortex (prefrontal, parietal, somatosensory), claustrum and basal ganglia (5). In the brain 5-HT2A receptors mediate hormone secretion, mood and perception, and regulate different behaviours. 5-HT2A receptors are highly expressed in blood platelets, fibroblasts, and many cell types of the cardiovascular system. Widely distributed peripheral 5-HT2A receptors are involved in the platelet aggregation (14), capillary permeability and vascular smooth muscles contraction. 5-HT2A receptors are molecular target for many atypical antipsychotic drugs like olanzapine or risperidone.

The loci encoding the serotonin type 2A receptor (*HTR2A*) are located on the long arm of the chromosome 13q14-q21 in man, and on chromosome 14 in the mouse (125). The *HTR2A* gene consists of 3 exons separated by 2 introns and spans over 20 kb (126). A number of polymorphisms encoding for *HTR2A* gene was found, including A-1438G (rs6311), T102C (rs6313) and His452Tyr (rs6314). Several genetic studies investigated the association of *HTR2A* genetic variants and vulnerability to psychiatric disorders like schizophrenia (127), suicide (128), panic disorder, alcoholism (129) and AD (130), with inconsistent and mostly negative results. Several studies reported that particular polymorphism in *HTR2A* gene may, to some extent, account for the difference in treatment response to risperidone (131), clozapine (132) and antidepressants (133).

IN CONCLUSION

Since serotonin discovery in the gastrointestinal tract (1), the comprehensive investigations enlarge its first role as a hormone to the neurotransmitter function in the central nervous system that has a myriad of central and peripheral functions. However, serotonin has an important role in the modulation of the effects of other neurotransmitters. In the words of Thomas Carew, a Yale researcher, »Serotonin is only one of the molecules in the orchestra. But rather than being the trumpet or the cello player, it is the band leader who choreographs the output of the brain.«

REFERENCES

- ERSPAMER V, ASERO B 1952 Identification of enteramine, specific hormone of enterochromaffine cell, as 5-hydroxytryptamine. *Nature* 169: 800–801
- MARC D J, AILTS J W, AILTS CAMPEAU D C, BULL M J, OLSON K L 2010 Neurotransmitters excreted in the urine as biomarkers of nervous system activity: Validity and clinical applicability. *Neurosci Biobehav Rev* doi:10.1016/j.neurobiorev.2010.07.007
- SAVELIEVA K V, ZHAO S, POGORELOV V M, RAJAN I, YANG Q, CULLINAN E, THOMAS H, LANTHORN T H 2008 Genetic disruption of both tryptophan hydroxylase genes dramatically reduces serotonin and affects behaviour in models sensitive to antidepressants. *PLoS ONE* 3(10): e3301, doi:10.1371/journal.pone.0003301
- Q, Q, SUN W, P, W, X-Q, Z, YU W, B, R, V, M, E, R, E, PLATT, K, A, D, A, B, Z-C 2008 Discovery and characterization of novel tryptophan hydroxylase inhibitors that selectively inhibit serotonin synthesis in the gastrointestinal tract. *J Pharmacol Exp Therapy* 325: 47–55
- HOYER D, HANNON J P, MARTIN G R 2002 Molecular, pharmacological and functional diversity of 5-HT receptors. *Pharmacol Biochem Behav* 71: 533–554
- JACOBS B L, AZMITIA E C 1992 Structure and function of the brain serotonin system. *Physiol Review* 72: 165–229
- D'SOUZA UM, CRAIG IW 2006 Functional polymorphisms in dopamine and serotonin pathway genes. *Human Mutation* 27: 1–13
- LUCKII 1998 The spectrum of behaviours influenced by serotonin. *Biol Psychiatry* 44: 151–162
- RICCIO D, POTTER G, WALZER C, VALLET P, SZABO G, VUTSKITS L, KISS J Z, DAYER A G 2009 Express of serotonin affects embryonic of the serotonin receptor 6. *Mol Psychiatry* 14: 280–290
- DIKSIC M, NAGAHIRO S, SOURKES T L, YAMAMOTO Y L 1990 A new method to measure brain serotonin synthesis in vivo. I. Theory and basic data for a biological model. *J Cereb Blood Flow Metab* 10: 1–12
- WATTS S W 2005 5-HT_{1B} in systemic hypertension: Foe, friend or fantasy. *Clin Chem* 108: 399–412
- STAHL S M 1985 Platelets as pharmacological models for the receptors and biochemistry of monoaminergic neurons. In: Longenecker G L (ed) Platelets: Physiology and Pharmacology, Academic Press, New York, p 307–340
- ANDRES A H, RAO M A, OSTROWITZKI S, ENZIAN W 1993 Human brain cortex and platelet serotonin₂ receptor binding properties and their regulation by endogenous serotonin. *Life Sci* 52: 313–321
- MENDELSON S C 2000 The current status of the platelet 5-HT_{2A} receptor in depression. *J Affect Disord* 57: 13–24
- PILETZ J E, SCUBERT D S P, HALARIS A 1986 Evaluation of studies on platelet alpha₂-adrenoreceptors in depressive illness. *Life Sci* 39: 1589–1616
- MUCK-SELER D, PIVAC N, MUSTAPIĆ M, CRNČEVIĆ Ž, JAKOVLJEVIĆ M, ŠAGUD M 2004 Platelet serotonin and plasma prolactin and cortisol in healthy, depressed and schizophrenic women. *Psychiatry Res* 127: 217–226
- MUCK-SELER, D, JAKOVLJEVIĆ M, PIVAC N 1996 Platelet 5-HT concentrations and suicidal behaviour in recurrent major depression. *J Affect Disord* 39: 73–80
- SAGUD M, PIVAC N, MUSTAPIC M, NEDIC G, MIHALJEVIC-PELES A, KRAMARIC M, JAKOVLJEVIC M, MUCK-SELER D 2008 The influence of lamotrigine on platelet serotonin concentration in patients with bipolar depression. *Psychopharmacology* 197: 683–685
- MUCK-SELER D, JAKOVLJEVIC M, DEANOVIC Z 1991 Platelet serotonin in subtypes of schizophrenia and unipolar depression. *Psychiatry Res* 38: 105–113
- MUCK-SELER D, PIVAC N, JAKOVLJEVIC M 1999 Sex differences, season of birth and platelet 5-HT levels in schizophrenic patients. *J Neural Transm* 106: 337–347
- MUCK-SELER D, PIVAC N, JAKOVLJEVIĆ M, BRZOVIĆ Z 1999 Platelet 5-HT, plasma cortisol and dexamethasone suppression test in schizophrenic patients. *Biol Psychiatry* 45: 1433–1439
- MAURER-SPUREJ E, PITTENDREIGH C, MISRI S 2006 Platelet serotonin levels support depression scores for women with postpartum depression. *J Psychiatry Neurosci* 32: 23–29
- MUCK-SELER D, PIVAC N, JAKOVLJEVIĆ M, ŠAGUD M, MIHALJEVIĆ-PELEŠ A 2003 Platelet 5-HT concentration and comorbid depression in war veterans with or without posttraumatic stress disorder. *J Affect Disord* 75: 171–179
- PIVAC N, KOZARIC-KOVACIC D, MUSTAPIC M, DEZELJIN M, BOROVECKI A, GRUBISIC-ILIC M, MUCK-SELER D 2006 Platelet serotonin in combat related posttraumatic stress disorder with psychotic symptoms. *J Affect Disord* 93: 223–227
- PIVAC N, MUCK-ŠELER D, MUSTAPIC M, NENADIC-ŠVIGLIN K, KOZARIC-KOVACIC D 2004 Platelet serotonin concentration in alcoholic subjects. *Life Sci* 76: 521–531
- HERCIGONJA-NOVKOVIC V, RUDAN V, PIVAC N, NEDIC G, MUCK-SELER D 2009 Platelet serotonin concentration in children with attention-deficit/hyperactivity disorder. *Neuropsychobiology* 59: 17–22
- MUCK-SELER D, DEANOVIC Z, DUPELJ M 1979 Platelet serotonin (5-HT) releasing factor in plasma of migrainous patients. *Headache* 19: 14–17
- KUMAR A M, SEVUSH S, KUMAR M, RUIZ J, EISDORFER C 1995 Peripheral serotonin in Alzheimer's disease. *Neuropsychobiology* 32: 9–12
- MUCK-SELER D, PRESECKI P, MIMICA N, MUSTAPIC M, PIVAC N, BABIC A, NEDIC G, FOLNEGOVIC SMALC V 2009 Platelet serotonin concentration and monoamine oxidase type B activity in female patients in early, middle and late phase of Alzheimer's disease. *Progr Neuro-Psychopharmacol Biol Psychiatry* 33: 1226–1231
- NESTLER E J, BARROT M, DILEONE R J, EISCH A J, GOLD S J, MONTEGGIA L M 2002 Neuro Depression Rev Neuron 34: 13–25
- COPPEN A 1967 The biochemistry of affective disorders. *Br J Psychiatry* 113: 1237–1264
- ASBERG M, TRASKMAN L, THOREN P 1976 5-HIAA in the cerebrospinal fluid. A biochemical suicide predictor? *Arch Gen Psychiatry* 33: 1193–1197
- PLACIDI GP, OQUENDO MA, MALONE KM, HUANG YY, ELLIS SP, MANN JJ 2001 Aggressivity, suicide attempts, and depression: relationship to cerebrospinal fluid monoamine metabolite levels. *Biol Psychiatry* 50: 783–791
- BARTON D A, ESLER M D, DAWOOD T, LAMBERT E A, HAIKERWA L D, BRENCHLEY C, SOCRATOUS F, HASTINGS J, GUO L, WIESNER G, KAYE D M, BAYLES R, SCHLAICH M P, LAMBER G W 2008 Elevated brain serotonin turnover in patients with depression effect of genotype and therapy. *Arch Gen Psychiatry* 65: 38–46
- OXENKRUG G F 1979 The content and uptake of 5-HT by blood platelets in depressive patients. *J Neural Transm* 45: 285–289
- MANN J J, McBRIDGE P A, ANDERSON G M, MIECKZKOWSKI T A 1992 Platelet and whole blood serotonin content in depressed inpatients: correlations with acute and life-time psychopathology. *Biol Psychiatry* 32: 243–257
- SHELINE Y I, BARDGETT M E, JACKON J L, NEWCOMER J W, CSERNANSKY J G 1995 Platelet serotonin markers and depressive symptomatology. *Biol Psychiatry* 37: 442–447
- TAMMINGA C A, NEMEROFF C B, BLAKELY R D, BRADY L, CARTER C S, DAVIS K L, DINGLELINE R, GORMAN J M, GRIGORIADIS D E, HENDERSON D C, INNIS R B, KILLEN J, LAUGHREN T P, MCDONALD W M, MURPHY G M JR, PAUL S M, RUDORFER M V, SAUSVILLE E, SCHATZBERG A

- F, SCOLNICK E M, SUPPES T 2002 Developing novel treatments for mood disorders: accelerating discovery. *Biol Psychiatry* 52: 589–609
39. ABL-DARGHAM A, LARUELLE M, AGHAJANIAN G K, CHARNEY D, KRYSTAL J 1997 The role of serotonin in the pathophysiology and treatment of schizophrenia. *J Neuropsychiat Clin Neurosci* 9: 1–17
 40. WONG A H C, VAN TOL H H M 2003 Schizophrenia: from phenomenology to neurobiology. *Neurosci Biobehav Reviews* 27: 269–306
 41. MUCK-SELER D, JAKOVLJEVIĆ M, DEANOVIC Z 1988 Time course of schizophrenia and platelet 5-HT level. *Biol Psychiatry* 23: 243–251
 42. PIVAC N, MUCK-SELER D, JAKOVLJEVIĆ M 1997 Platelet 5-HT levels and hypothalamic-pituitary-adrenal axis activity in schizophrenic patients with positive and negative symptoms. *Neuro-psychobiology* 36: 19–21
 43. DELISI L E, NECKERS L M, WEINBERGER D R, WYATT R J 1981 Increased whole blood serotonin concentrations in chronic schizophrenic patients. *Arch Gen Psychiatry* 38: 647–650
 44. FINK G 2011 Stress controversies: Post-traumatic stress disorder, hippocampal volume, gastroduodenal ulceration. *J Neuroendocrinol* 23: 107–117
 45. HEIM C, NEMEROFF C B 2009 Neurobiology of posttraumatic stress disorder. *CNS Spectrums* 14 (Suppl. 1): 13–24
 46. GOVEAS J S, CSERNANSKY J G, COCCARO E F 2004 Platelet serotonin content correlates inversely with life history of aggression in personality-disordered subjects. *Psychiatry Res* 126: 23–32
 47. BONNE O, BAIN E, NEUMEISTER A, NUGENT A C, VYTHILINGAM M, CARSON R E, LUCKDNBAUGH D A, ECKELMAN W, HERSCOCITCH P, DREVETS W C, CHARNEY DS 2005 No change in serotonin type 1A receptor binding in patients with posttraumatic stress disorder. *Am J Psychiatry* 162: 383–385
 48. MELLMAN T A, KUMAR A M 1994 Platelet serotonin measures in posttraumatic stress disorder. *Psychiatry Res* 53: 99–101
 49. JOHNSON B A 2004 Role of the serotonergic system in the neurobiology of alcoholism – Implications for treatment. *CNS Drugs* 18: 1105–1118
 50. CHEN H-T, CASANOVA M F, KLEINMAN J E, ZITO M, GOLDMAN D, LINNOILA M 1991 ³H-Paroxetine binding in brains of alcoholics. *Psychiat Res* 38: 293–299
 51. STORVIK M, TIIHONEN J, HAUKIJÄRVI T, TUPALA E 2006 Lower serotonin transporter binding in caudate in alcoholics. *Synapse* 144–151
 52. DILLON K A, GROSS-ISSEROFF R, ISRAELI M, BIEGON A 1991 Autoradiographic analysis of serotonin 5-HT_{1A} receptor binding in the human brain post mortem: effects of age and alcohol. *Brain Res* 554: 56–64
 53. HEINZ A, MANN K, WEINBERGER D R, GOLDMAN D 2001 Serotonergic dysfunction, negative mood states, and response to alcohol. *Alcohol: Clin Exp Res* 25: 487–495
 54. FILS-AIME M L, ECKARDT M J, GEORGE D T, BEOWN D L, MEFFORD I, LINNOILA M 1996 Early onset alcoholics have lower cerebrospinal fluid 5HIAA levels than late onset alcoholics. *Arch Gen Psychiatry* 53: 211–216
 55. BALLENGER J, GOODWIN F, MAJOR L 1979 Alcohol and central serotonin metabolism in man. *Arch Gen Psychiatry* 36: 224–227
 56. VIRKKUNEN M, GOLDMAN D, NIELSEN DA, LINNOILA M 1995 Low brain serotonin turnover rate (low CSF 5-HIAA) and impulsive violence. *J Psychiatry Neurosci* 20: 271–275
 57. KENT T A, CAMPBELL J L, PAZDERNIK T L, HUNTER R, GUNN W H, GOODWIN D W 1985 Blood platelet uptake of serotonin in men alcoholics. *J Stud Alcohol* 46: 357–359
 58. FARAJ B A, OLKOWSKI L, JACKSON R T 1997 Prevalence of high serotonin uptake in lymphocytes of abstinent alcoholics. *Biochem Pharmacol* 53: 53–57
 59. JAVORS M, TIOURIRINE M, PRIHODA T 2000 Platelet serotonin uptake is higher in early-onset than in late-onset alcoholics. *Alcohol Alcoholism* 35: 390–393
 60. PIVAC N, KOZARIC-KOVACIC D, MUSTAPIC M, DEZELJIN M, NENADIC-SVIGLIN K, MUCK-SELER D 2008 Peripheral biological markers in alcoholism, In: Pivac N (ed). Nova Science Publishers, New York, p 1–93
 61. KRYSTAL J H, WEBB E, COONEY N L, KRANZLER H R, CHARNEY D S 1994 Specificity of ethanol-like effects elicited by serotonergic and noradrenergic mechanisms. *Arch Gen Psychiatry* 51: 898–911
 62. BIEDERMAN J 2005 Attention-Deficit/Hyperactivity Disorder: A Selective Overview. *Biol Psychiatry* 57: 1215–1220
 63. OADES R D 2002 Dopamine may be »hyper« with respect to noradrenaline metabolism, but »hypo« with respect to serotonin metabolism in children with attention deficit hyperactivity disorder. *Behav Brain Res* 130: 97–102
 64. COMINGS D E 1993 Serotonin and the biochemical genetics of alcoholism: lessons from studies of attention deficit hyperactivity disorder (ADHD) and Tourette syndrome. : 237–241
 65. MANUCK S B, FLORY J D, MCCAFFERY J M, MATTHEWS K A, MANN J J, MULDOON M F 1998 Aggression, impulsivity, and central nervous system serotonergic responsivity in a nonpatient sample. 19: 287–299
 66. FLORY J D, NEWCORN J H, MILLER C, HARTY S, HALPERIN J M 2007 Serotonergic function in children with attention-deficit hyperactivity disorder: Relationship to later antisocial personality disorder. *Br J Psychiatry* 190: 410–414
 67. HALPERIN J M, SHARMA V, SIEVER L J, SCHWARTZ S T, MATTER K, WORNELL G, NEWCORN J H 1994 Serotonergic function in aggressive and nonaggressive boys with attention-deficit hyperactivity disorder. *Am J Psychiatry* 151: 243–248
 68. KEPE V, BARRIO J R, HUANG, S-C, ERCOLI L, SIDDARTH P, SHOGHI-JADID K, COLE G M, SATYAMURTHY N, CUMMINGS J L, SMALL G W, PHELPS M E 2006 Serotonin 1A receptors in the living brain of Alzheimer's disease patients. *Proc Natl Acad Sci USA* 103: 702–707
 69. BLIN J, BARON J C, DUBOIS B, CROUZE L C, FIORELLI M, ATTAR-LEVY D, PILLON B, FOURNIER D, VIDALHET M, AGID Y 1993 Loss of brain 5-HT₂ receptors in Alzheimer's disease. *Brain* 116: 497–510
 70. PRITCHARD A L, HARRIS J, PRITCHARD C W, COATES J, HAQUE S, HOLDER R, BENTHAM P, LENDON C L 2008 Role of 5-HT_{2A} and 5-HT_{2C} polymorphisms in behavioural and psychological symptoms of Alzheimer's disease. *Neurobiol Aging* 29: 341–347
 71. TOHGI H, ABE T, TAKAHASHI S, KIMURA M, TAKAHASHI J, KIKUCHI T 1992 Concentrations of serotonin and its related substances in the cerebrospinal fluid in patients with Alzheimer type dementia. *Neurosci Lett* 141: 9–12
 72. GARCIA-ALLOZA M, GIL-BEA F, DIEZ-ARIZA M, CHEN CPL-H, FRANCIS P T, LASHERAS B, RAMIREZ M J 2005 Cholinergic-serotonergic imbalance contributes to cognitive and behavioural symptoms in Alzheimer's disease. *Neuropsychologia* 43: 442–449
 73. WIDNER B, LEBLHUBER F, WALLI J, TILZ G P, DEMEL U, FUCHS D 2000 Tryptophan degradation and immune activator in Alzheimer's disease. *J Neural Transm* 107: 343–353
 74. FEKKES D, VAN DER CAMMON TJM, VAN LOON CMP, VERSCHOOR C, VAN HARSKAMP F, DE KONING I, SCHUDEL W J, PEPPLINKHUIZEN L 1998 Abnormal amino acid metabolism in patients with early stage Alzheimer dementia. *J Neural Transm* 105: 287–294
 75. FONTEH A N, HARRINGTON R J, TSAI A, LIAO P, HARRINGTON M G 2007 Free amino acid and peptide changes in the body fluids from Alzheimer's disease subjects. *Amino Acids* 32: 213–224
 76. RUDDICK J P, EVANS A K, NUTT D J, LIGHTMAN S L, ROOK G A W, LOWRY C A 2006 Tryptophan metabolism in the central nervous system: medical implications. *Expert Rev Mol Med* 8: 1–27
 77. PARK S B, COULL J T, MCSHANE R H, YOUNG A H, SAHAKIAN B J, ROBBINS T W, COWEN P J, 1994 Tryptophan depletion in normal volunteers produces selective impairments in learning and memory. *Neuropharmacology* 33: 575–588
 78. NEWHOUSE P, TATRO A, NAYLOR M, QUEALEY K, DELGADO P 2002 Alzheimer's disease, serotonin systems and tryptophan depletion. *Am J Geriatr Psychiatry* 10: 483–484
 79. GOTTFRIES C G 1990 Disturbance of the 5-hydroxytryptamine metabolism in brains from patients with Alzheimer's dementia. *J Neural Transm* 30(Suppl): 33–43
 80. CASH C D 1998 Why tryptophan hydroxylase is difficult to purify: A reactive oxygen-derived species-mediated phenomenon that may be implicated in human pathology. *Gen Pharmacol* 30: 569–574
 81. ARORA R C, EMERY O B, MELTZER H Y 1991 Serotonin uptake in the blood platelets of Alzheimer's disease patients. *Neurology* 41: 1307–1309

82. WALTHER D J, BADER M 2003 A unique central tryptophan hydroxylase isoform. *Biochem Pharmacol* 66: 1673–1680
83. ZHANG X, GAINETDINOV R R, BEAULIEU J M, SOTNIKOVA T D, BURCH L H, WILLIAMS R B, SCHWARTZ D A, RANGA K, KRISHNAN R, CARON M G 2005 Loss-of-Function mutation in meport tryptophan hydroxylase-2 identified in unipolar major depression. *Neuron* 45: 11–16
84. DELORME R, DURAND C M, BETANCUR C, WAGNER M, RUHRMANN S, GRABE H-J, NYGREN G, GILLBERG C, LEBOYER M, BOURGERON T, COURTET P, JOLLANT F, BURESI C, AUBRY J-M, BAUD P, BONDOLFI G, BERTSCHY G, PERROUD N, MALAFOSSE A 2006 No human tryptophan hydroxylase-2 gene R441H mutation in a large cohort of psychiatric patients and control subjects. *Biol Psychiatry* 60: 202–203
85. XUE K X, FAN C H, LI X L 2009 No association between tryptophan hydroxylase-2 gene G1463A polymorphism and unipolar depression in a Southern Chinese Han population. *Hong Kong J Psychiatry* 1: 1–5
86. CAMPOS S B, MARQUES MIRANDA D, SOUZA B R, FERNANDO SILVA NEVES P A P, BICALHO M A C, CASADEI MELILLO P H, TRAMONTINA J, KAPCZINSKI F, ROMANOSILVA M A, CORREA H 2010 Association of polymorphisms of the tryptophan hydroxylase 2 gene with risk for bipolar disorder or suicidal behaviour. *J Psychiat Res* 44: 271–274
87. CHOI K-Y, YOON H-K, KIM Y-K 2010 Association between Serotonin-Related Polymorphisms in 5HT2A, TPH1, TPH2 Genes and Bipolar Disorder in Korean Population. *Psychiat Invest* 7: 60–67
88. JOHANSSON S, HALMÖY A, MAVROCONSTANTI T, JACOBSEN K K, ELISABETH T, LANDAAS K K, REIF A, JACOB C, BOREATTI-HÜMMER A, KREIKER S, LESCH K-L, KAN C C, SANDRA KOOIJ S J J, KIEMENEY L A, BUITELAAR J K, FRANKE B, RIBASÉS M, BOSCH R, BAYÉS M, CASAS M, RAMOS-QUIROGA J A, CORMAND B, KNAPPSKOG P, HAAVIK J 2010 Common variants in the TPH1 and TPH2 regions are not associated with persistent ADHD in a combined sample of 1,636 adult cases and 1,923 controls from four European populations. *Am J Mol Genetics Part B: Neuropsychiatric Genetics* DOI: 10.1002/ajmg.b.31067.
89. CLONINGER C R, SVRAKIC D M, PRZYBECK T R 1993 A psychobiological model of temperament and character. *Arch Gen Psychiatry* 50: 975–990
90. LESCH K P, GUTKNECHT L 2005 Pharmacogenetics of the serotonin transporter. *Progr Neuro-Psychopharmacol Biol Psychiatry* 29: 1062–1073
91. SCHLOSS P, WILLIAMS D C 1998 The serotonin transporter: a primary target for antidepressant drugs. *J Psychopharmacol* 12: 15–21
92. LESCH K P, MOSSNER R 1998 Genetically driven variation in serotonin uptake: is there a link to affective spectrum, neurodevelopmental, and neurodegenerative disorders? *Biol Psychiatry* 44: 179–192
93. NAKAMURA M, UENO S, SANO A, TANABE H 2000 The human serotonin transporter gene linked polymorphism (5-HTTLPR) shows ten novel allelic variants. *Mol Psychiatry* 5: 32–38
94. GREENBERG B D, TOLLIVER T J, HUANG S J, LI Q, BEN-GEL D, MURPHY D L 1999 Genetic variation in the serotonin transporter promoter region affects serotonin uptake in human blood platelets. *Am J Med Genetics* 88: 83–87
95. PEZAWAS L, MEYER-LINDENBERG A, DRABANT E M, VERCHINSKI B A, MUNOZ K E, KOLACHANA B S, EGAN M F, MATTAY V S, HARIRI A R, WEINBERGER D 2005 5-HTTLPR polymorphism impacts human cingulate-amygdala interactions: A genetic susceptibility mechanism for depression. *Nat Neurosci* 8: 828–834
96. LESCH K P, BALLING U, GROSS J, STRAUSS K, WOŁOZIN B L, MURPHY D L, RIEDERER P 1994 Organization of the human serotonin transporter gene. *J Neural Transm* 95: 157–162
97. MACKENZIE A, QUINN J P 1999 A serotonin transporter gene intron 2 polymorphic region, correlated with affective disorders, has allele dependent differential enhancer-like properties in the mouse embryo. *Proc Nat Acad Sci USA* 96: 15251–15255
98. FISKERSTRAND C E, LOVEJOY E A, QUINN J P 1999 An intronic polymorphic domain often associated with susceptibility to affective disorders has allele dependent enhancer activity in embryonic stem cells. *FEBS Letters* 458: 171–174
99. LEVINSON D F 2006 The genetics of depression: A review. *Biol Psychiatry* 60: 84–92
100. COOK E H, COURCHESNE R, LORD C, COX N J, YAN S, LINCOLN A, HAAS R, COURCHESNE E, LEVENTHAL B L 1997 Evidence of linkage between the serotonin transporter and autistic disorder. *Mol Psychiatry* 2: 247–250
101. MENDLEWICZ J, MASSAT I, SOUERY D, DEL-FAVERO J, ORUC L, NOTHEN M M, BLACKWOOD D, MUIR W, BATTERSBY S, LERER B, SEGMAN R H, KANEVA R, SERRETTI A, LILLI R, LORENZI C, JAKOVljeVIC M, IVEZIC S, RIETSCHEL M, MILANOVA V, VAN BROECKHOVEN C 2004 Serotonin transporter 5-HTTLPR polymorphism and affective disorders: no evidence of association in a large European multicenter study. *Eur J Hum Genetics* 12: 377–382
102. CASPIA, SUGDEN K, MOFFITT T E, TAYLOR A W, CRAIG I, HARRINGTON L, MCCLAY J, MILL J, MARTIN J, BRAITHWAITE A, POULTON R 2003 Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science* 301: 386–389
103. RISCH N, HERRELL R, LEHNER T, LIANG K Y, EAVES L, HOH J, GRIEM A, KOVACS M, OTT J, RIES MERIKANGAS K 2009 Interaction between the serotonin transporter gene (5-HTTLPR), stressful life events, and risk of depression. A Meta-analysis. *J Am Med Association* 301: 2462–2471
104. NOSKOVA T, PIVAC N, NEDIC G, KAZANTSEVA A, GAY-SINA D, FASKHUTDINOVA G, GAREEVA A, KHALILOVA Z, KHUSNUTDINOVA E, KOZARIC KOVACIC D, KOVACIC Z, JOKIC M, MUCK SELER D 2008 Ethnic differences in the serotonin transporter polymorphism (5-HTTLPR) in several European populations. *Prog Neuro-Psychopharmacol Biol Psychiatry* 32: 1735–1739
105. SERRETTI A, CALATI R, MANDELLI L, DE RONCHI D 2006 Serotonin transporter gene variants and behavior: A comprehensive review. *Curr Drug Targ* 7:1659–1669
106. GREENBERG B D, TOLLIVER T J, HUANG S J, LI Q, BEN-GEL D, MURPHY D L 1999 Genetic variation in the serotonin transporter promoter region affects serotonin uptake in human blood platelets. *Am J Med Genet* 88: 83–87
107. PIVAC N, NEDIĆ G, MUSTAPIĆ M, BABIĆ A, STIPČEVIĆ T, BOROVEČKI F, HAJNŠEK S, MUCK-ŠELER D 2009 The lack of genotype-phenotype relationship between platelet serotonin concentration and serotonin transporter gene promoter polymorphism in healthy subjects. *Neurosci Lett* 462: 45–48
108. ZILL P, PADBERG F, DE JONGE S, HAMPPEL H, BURGER K, STUBNER S, BOETSCH T, JURGEN MOLLER H, ACKENHEIL M, BONDY B 2000 Serotonin transporter (5-HTT) gene polymorphism in psychogeriatric patients. *Neurosci Lett* 284: 113–115
109. SUKONICK D L, POLLOCK B G, SWEET R A, MULSANT B H, ROSEN J, KLUNK W E, KASTANGO K B, DEKOSKY S T, FERRELL R E 2001 The 5-HTTPR*^S/^L polymorphism and aggressive behavior in Alzheimer's disease. *Arch Neurol* 58: 1425–1428
110. PAZOS A, PALACIOS J M 1985 Quantitative autoradiographic mapping of serotonin receptors in the rat brain. I. Serotonin-1 receptors. *Brain Res* 346: 205–230
111. LE FRANCOIS B, CZESAK M, STEUBL D, ALBERT P R 2008 Transcriptional regulation at a *HTR1A* polymorphism associated with mental illness. *Neuropharmacol* 55: 977–985
112. WHITTAKER-AZMITIA P M, DRUSE M, WALKER P, LAUDER J M 1996 Serotonin as a developmental signal. *Behav Brain Res* 73: 19–29
113. DRAGO A, DE RONCHI D, SERRETTI A 2008 5-HT_{1A} gene variants and psychiatric disorders: a review of current literature and selection of SNPs for future studies. *Int J Neuropsychopharmacol* 11: 701–721
114. LÓPEZ-FIGUEROA A L, NORTON C S, LÓPEZ-FIGUEROA M O, BURKE S, MEADOR-WOODRUFF J H, LÓPEZ J F, WATSON S J 2004 Serotonin 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{2A} receptor mRNA expression in subjects with major depression, bipolar disorder and schizophrenia. *Biol Psychiatry* 55: 225–233
115. STOCKMEIER C A, SHAPIROLA, DILLEY G E, KOLLI T N, FRIEDMAN L, RAJKOWSKA G 1998 Increase in serotonin-1A autoreceptors in the midbrain of suicide victims with major depression—postmortem evidence for decreased serotonin activity. *J Neurosci* 18: 7394–7401
116. LOWTHER S, DE PAERMENTIER F, CHEETHAM S C, CROMPTON M R, KATONA C L, HORTON R W 1997 5-HT_{1A} receptor binding sites in post-mortem brain samples from depressed suicide and controls. *J Affect Disord* 42: 199–207

117. LEMONDE S, TURECKI G, BAKISH D, DU L, HRDINA P D, BOWN C D, SEQUEIRA A, KUSHWAHA N, MORRIS S J, BASAK A, OU X-M, ALBERT P R 2003 Impaired expression at a 5-Hydroxytryptamine 1A receptor gene polymorphism associated with major depression and suicide. *J Neurosci* 23: 8788–8799
118. SZEWCZYK B, ALBERT P R, BURNS A M, CZESAK M, OVERHOLSER J C, JURJUS G J, MELTZER H Y, KONICK L C, DIETER L, HERBST N, MAY W, RAJKOWSKA G, STOCKMEIER C A, AUSTIN M C 2009 Gender-specific decrease in NUDR and 5-HT_{1A} receptor proteins in the prefrontal cortex of subjects with major depressive disorder. *Int J Neuropsychopharmacol* 12: 155–168
119. LOTHE A, BONI C, COSTES N, BOUVARD S, GORWOOD P, LAVENNE F, ALVAREZ M, RYVLIN P 2010 5-HT_{1A} gene promoter polymorphism and [¹⁸F]MPPF binding potential in healthy subjects: a PET study. *Behav Brain Functions* 6: 37–46
120. SVENNINGSSON P, CHERGUI K, RACHLEFF I, FLAJOLET M, ZHANG X, EL YACOUBI M, VAUGEOIS J.-M, NOMIKOS G G, GREENGARD P 2006 Alterations in 5-HT_{1B} receptor function by p11 in depression-like states. *Science* 311: 77–80
121. GORWOOD P, AISSI F, BATEL P, ADES J, COHEN-SALMON C, HAMON M, BONI C, LANFURMEY L 2002 Reappraisal of the serotonin 5HT_{1B} receptor gene in alcoholism: of mice and men. *Brain Res Bull* 57: 103–107
122. SOYKA M, PREUSS U W, KOLLER G, ZILL P, BONDY B 2004 Association of 5-HT_{1B} receptor gene and antisocial behaviour in alcoholism. *J Neural Transm* 111: 101–109
123. SANDERS A R, CAO Q, TAYLOR J, LEVIN T, BADNER J A, CRAVCHIK A, COMERON J M, NARUYA S, DEL ROSARIO A, SALVI D., WALCZYK K, MOWRY B, LEVINSON D F, CROWE R R, SILVERMAN J M, GEJMAN P V 2001. Genetic diversity of the human serotonin receptor 1B (HTR1B) gene. *Genomics* 72: 1–14
124. SMOLLER J W, BIEDERMAN J, ARBEITMAN L, DOYLE A E, FAGERNESS J, PERLIS J H, SKLAR P, FARAONE S V 2006 Association between the 5HT_{1B} receptor gene (*HTR1B*) and the inattentive subtype of ADHD. *Biol Psychiat* 59: 460–467
125. SPARKES R S, LAN N, KLISAK I, MOHANDAS T, DIEP A, KOJIS T, HEINZMANN C, SHIH J C 1991 Assignment of a serotonin 5HT₂ receptor gene (*HTR2*) to human chromosome 13q14-q21 and mouse chromosome 14. *Genomics* 9: 461–465
126. CHEN K, YANG W, GRIMSBY J, SHIH J C 1992 The human 5-HT₂ receptor is encoded by a multiple intron-exon gene. *Brain Res Mol Brain Res* 14: 20–26
127. ABDOLMALEKY H M, FARAONE S V, GLATT S J, TSUANG M T 2004 Meta-analysis of association between the T102C polymorphism of the 5-HT_{2A} receptor gene and schizophrenia. *Schizophr Res* 67: 53–62
128. VIDETIC A, PUNGERCIC G, ZUPANIC PAJNIC I., ZUPANC T, BALAZIC J, TOMORI M, KOMEL R 2006 Association study of seven polymorphisms in four serotonin receptor genes on suicide victims. *Am J Med Genetics Part B (Neuropsychiatric Genetics)* 141B: 669–672
129. NAKAMURA T, MATSUSHITA S, NISHIGUCHI N, KIMURAM, YOSHINO A, HIGUCHI S 1999 Association of a polymorphism of the 5HT_{2A} receptor gene promoter region with alcohol dependence. *Mol Psychiatry* 4: 85–88
130. NACMIAS B, TEDDE A, FORLEO P 2001 Association between 5-HT_{2A} receptor polymorphism and psychotic symptoms in Alzheimer's disease. *Biol Psychiatry* 50: 472–475
131. YAMANOUCHI Y, IWATA N, SUZUKI T, KITAJAMA T, IKEDA M, OZAKI N 2003 Effect of DRD2, 5-HT_{2A} and COMT genes on antipsychotic response to risperidone. *Pharmacogenomic J* 3: 356–361
132. HARVEY L, REID R E, MA C, KNIGHT P J K, PFEIFER T A, GRIGLIATTI T A 2003 Human genetic variations in the 5-HT_{2A} receptor: a single nucleotide polymorphism identified with altered response to clozapine. *Pharmacogenetics* 13: 107–118
133. MCMAHON F J, BUERVENICH S, CHARNEY D, LIPSKY R, RUSH A J, WILSON A F, SORANT A J M, PAPANICOLAOU G J, LAJE G, FAVA M, TRIVEDI M H, WISNIEWSKI S R, MANJI H 2006 Variation in the gene encoding the serotonin 2A receptor is associated with outcome of antidepressant treatment. *Am J Hum Genetics* 78: 804–814