

Design and evaluation of moxifloxacin hydrochloride ocular inserts

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The objective of the present investigation was to prepare and evaluate ocular inserts of moxifloxacin. An ocular insert was made from an aqueous dispersion of moxifloxacin, sodium alginate, polyvinyl alcohol, and dibutyl phthalate by the film casting method. The ocular insert (5.5 mm diameter) was cross-linked by CaCl₂ and was coated with Eudragit S-100, RL-100, RS-100, E-100 or L-100. The *in vitro* drug drainage/permeation studies were carried out using an all-glass modified Franz diffusion cell. The drug concentration and mucoadhesion time of the ocular insert were found satisfactory. Cross-linking and coating with polymers extended the drainage from inserts. The cross-linked ocular insert coated with Eudragit RL-100 showed maximum drug permeation compared to other formulations.

Keywords: moxifloxacin, ocular insert, goat cornea

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Eyes are among the most readily accessible organs in terms of location in the body. The physiological constraints of the eye render this organ impervious to foreign substances, thus presenting a constant challenge to the formulator to circumvent the protective barriers of the eye without causing permanent tissue damage (1). All ocular therapeutics have been mostly administered to the eye as simple aqueous eye drops. About 90 % of the dose applied topically from such solutions is lost due to pre-corneal losses (lacrimation and drainage), which leads to poor availability and frequent dosing is required for the instillation to achieve an adequate level and therapeutic effect (2). The standard treatment of severe bacterial keratitis requires frequent administration of fluoroquinolone eye drops (3). However, this regimen is not only disruptive to the patient and usually necessitates hospitalization, but it has also been associated with toxicity to the corneal epithelium (4, 5). To overcome these limitations, ocular inserts seem promising by prolonging the contact time with improved efficiency of the therapy and patient compliance. Ocular inserts offer many advantages over conventional dosage forms, like

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increased ocular residence, possibility of releasing drugs at a slow and constant rate, accurate dosing, and avoidance of toxicity due to preservative and increased shelf life (6). Ocular inserts of ciprofloxacin hydrochloride (7) and ofloxacin (8) have been designed to improve ocular availability.

Moxifloxacin, [1-cyclopropyl-6-fluoro-1,4-dihydro-8-methoxy-7-[(4*aS*,7*aS*)-octahydro-6*H*-pyrrolol (3,4*b*) pyridin-6-yl]-4-oxo-3-quinoline carboxylic acid], is a fourth generation fluoroquinolone with a methoxy group at the C-8 position and a bulky C-7 side chain (9). Moxifloxacin has increased activity against *S. aureus* compared to second and third generation fluoroquinolones. Bactericidal activity of moxifloxacin is mediated by the inhibition of DNA gyrase (topoisomerase II) and topoisomerase IV, essential enzymes involved in bacterial DNA replication, transcription, repair and recombination (10). Effects of the formulation factor on *in vitro* corneal permeation of moxifloxacin from aqueous drops have been investigated and the results showed that moxifloxacin 0.5 % (*m/V*) ophthalmic solution (pH 7.2) containing benzalkonium chloride (0.01 %) (BAK) and EDTA (0.01 %) provides maximum *in vitro* ocular availability through all excised mammalian corneas (11). The *in vitro* permeation profile of moxifloxacin oil drops has been reported (12). Oil drops containing castor oil with benzyl alcohol provided maximum *in vitro* permeation through all the corneas. Aqueous and oily drops of moxifloxacin showed better permeation characteristics but these formulations require frequent instillation to achieve therapeutic concentration due to shorter precorneal residence time. Taking the above information in account, the purpose of the current study was to formulate ocular inserts of moxifloxacin that would be capable of prolonging the contact time, thereby potentially enhancing intra-corneal delivery of ophthalmic medicament.

EXPERIMENTAL

Materials

Moxifloxacin hydrochloride (99.97 % purity on anhydrous basis) was obtained from Ranbaxy Laboratories (India) as a gift. Sodium alginate, Eudragit S-100 (anionic methacrylic acid-methyl methacrylate copolymer), Eudragit RL-100 (acrylic acid and methacrylic acid ester copolymer containing 10 % trimethylammonium methacrylate chloride), Eudragit RS-100 (acrylic acid methacrylic acid ester copolymer containing 5 % trimethylammonium methacrylate chloride), Eudragit E-100 (cationic polymer based on dimethylaminoethyl methacrylate and other neutral methacrylic acid esters), Eudragit L-100 (anionic copolymer based on methacrylic acid and methyl methacrylate) were obtained from Jubilant Organosys (India) as gift samples. Polyvinyl alcohol (PA) and dibutyl phthalate were obtained from the Central Drug House (India) and S.D. Fine chemicals Ltd (India), respectively. All other chemicals were of analytical grade. Fresh goat eyeball was obtained from a butcher's shop. The method of cornea dissection and the apparatus used in permeation studies were the same as described previously (13).

Preparation of ocular inserts

The moxifloxacin control ocular insert (MIF1) was made from an aqueous dispersion (40 mL) of moxifloxacin hydrochloride (400 mg), sodium alginate (0.25 g), polyvinyl alcohol (PVA) (1.5 g), and dibutyl phthalate (0.3 mL) by the film casting method. Ocular insert (5.5 mm diameter) was cross-linked (MIF2) by CaCl_2 (0.2 %, *m/V*) by dipping and drying 5 times. Similarly, the cross-linked insert was coated with different grades of Eudragit polymer 0.2 % (*m/V*) in isopropyl alcohol and acetone by the dip and dry method (Table I). The polymers used were Eudragit S-100 (MIF3), Eudragit RL-100 (MIF4), Eudragit RS-100 (MIF5), Eudragit E-100 (MIF6) and Eudragit L-100 (MIF7). The mass gain of the ocular insert after coating with polymer was found to be in the range 23.4–29.6 %.

Table I. Compositions of moxifloxacin insert formulations for ocular delivery

Formulation	Polymer composition (18+12+70 %, <i>m/m</i>)	Plasticizer (dibutyl phthalate, %, <i>V/V</i>)	Cross-linking agent CaCl_2 (%, <i>m/V</i>)	Coating material (0.2 %, <i>m/V</i>)
MIF1	MOX + SA + PVA	0.12	–	–
MIF2	MOX + SA + PVA	0.12	0.2	–
MIF3	MOX + SA + PVA	0.12	0.2	Eudragit S-100
MIF4	MOX + SA + PVA	0.12	0.2	Eudragit RL-100
MIF5	MOX + SA + PVA	0.12	0.2	Eudragit RS-100
MIF6	MOX + SA + PVA	0.12	0.2	Eudragit E-100
MIF7	MOX + SA + PVA	0.12	0.2	Eudragit L-100

MOX – moxifloxacin, SA – sodium alginate, PVP– polyvinyl alcohol

Physicochemical evaluation of ocular inserts

Thickness uniformity. – Insert thickness (5.5 mm diameter) was measured at five different points using a Micrometer screw gauge (Mitutoyo Co., Japan) and the mean insert thickness was noted ($n = 3$).

Drug content uniformity. – Three inserts were taken out of the film and drug concentration determined. The uncoated ocular insert was dissolved in 10 mL of 0.1 mol L⁻¹ HCl while the coated insert was dissolved in 10 mL of acetone. The volume was made up to 100 mL with distilled water and the solution was filtered. The drug in the filtrate was analyzed by measuring absorbance at 291 nm in a spectrophotometer 1601 (Shimadzu, Japan). The experiment was done in triplicate.

Mucoadhesion time

The mucoadhesion time was determined (in triplicate) after application of coated or uncoated ocular inserts (5.5 mm diameter, 0.40–0.45 mm thickness) on a freshly cut goat eyelid. The eyelid was fixed on the bottom of a beaker with cyanoacrylate glue. Ocular insert (coated or uncoated) was attached to the mucosal surface of the eyelid by apply-

ing a light force with a fingertip for 20 s. The beaker was filled with 800 mL of bicarbonate Ringer solution pH 7.4 and stirred at a rate of 150 rpm at room temperature (14). The time needed for complete detachment of the insert from the mucosal surface was considered as mucoadhesion time.

Drug permeation

The *in vitro* drug permeation studies were carried out by putting the ocular insert (5.5 mm in diameter) on Millipore membrane filter (0.15 μm); between the donor and receptor compartments of an all-glass modified Franz diffusion cell. The Millipore membrane filter was used to simulate the corneal epithelial barrier as isolated cornea will not remain viable beyond 4 h. To simulate the tear flow, the donor compartment was infused with bicarbonate Ringer, pH 7.4, at a flow rate of 20 $\mu\text{L min}^{-1}$ throughout the study. The drained sample was monitored for moxifloxacin at 291 nm every hour. Similarly, a 1.0-mL sample was withdrawn at hourly intervals from the receptor compartment (containing 10 mL bicarbonate Ringer, pH 7.4, under stirring at 37 °C) and the drug permeated was measured. Each withdrawn sample was replaced with an equal volume of fresh bicarbonate Ringer. Drug permeation experiments were also carried out using freshly excised goat cornea for 4 h. At the end of the experiment, each cornea (freed from adhering sclera) was weighed, soaked in 1 mL of methanol, dried overnight at 90 °C and reweighed. Corneal hydration was calculated from the difference in masses.

The drug permeation data was plotted according to zero-order, first-order kinetics, Higuchi equation and Korsmeyer-Peppas equation (15).

Fourier transform infrared (FTIR) spectroscopy

The FTIR spectra of the pure drug and physical mixture (moxifloxacin, polyvinyl alcohol and sodium alginate) were taken as KBr pellets in the range of 4000–650 cm^{-1} (Perkin Elmer Model 1600 FT-IR spectrophotometer, USA). The infrared analysis of all inserts was carried out in the same range by ATR-IR spectroscopy (Perkin Elmer Model 1600 FT-IR spectrophotometer with ATR mode (Pike Miracle ATR Accessory, Perkin Elmer, USA).

In vivo study

Selected ocular inserts (MIF4 and MIF5) sterilized using γ -irradiation were used for *in vivo* drug release studies. The protocol on the use of animals (albino rabbits) was approved by the Institutional Ethics Committee. Two groups of ten healthy albino rabbits were used to study the drug release *in vivo* from formulations that showed satisfactory *in vitro* drug release (16). Each rabbit was kept under good hygienic conditions in order to avoid vulnerability to any disease, including ophthalmic types. Selected ocular inserts were placed in the *cul-de-sac* of each rabbit while the other eye served as a control. At periodic intervals (1–10 h) the inserts were taken out carefully from the *cul-de-sac* of each rabbit and analyzed for the remaining drug content. The remaining drug was subtracted from the initial drug content of the insert, which gave the amount of drug released in the rabbit eye.

Stability of ocular inserts

Stability studies were carried out on ocular insert formulations according to ICH guidelines (17). A sufficient number of ocular inserts (packaged in aluminum foils) were stored in a humidity chamber with a relative humidity of $75 \pm 5\%$ and temperatures of $40 \pm 2\text{ }^\circ\text{C}$ or at room temperature. Samples were withdrawn at time 0, 3 weeks, 6 weeks, 3 months and 6 months. Ocular inserts were also evaluated for their physical characteristics (color) and analyzed for drug concentration. The degradation rate constant was determined from the plot of logarithm of the remaining drug *vs.* time.

RESULTS AND DISCUSSION

Physicochemical evaluation and mucoadhesion time of ocular inserts

The thickness of ocular inserts was found to be from 0.40 ± 0.00 to 0.45 ± 0.06 millimeters. Drug concentration of the ocular insert without cross-linking and uncoated (control MIF1) was maximum (*i.e.*, 0.149 mg mm^{-2}), followed by the cross-linked ocular insert (*i.e.*, 0.122 mg mm^{-2}) and coated ocular insert (*i.e.*, 0.115 mg mm^{-2}) (Table II). Inserts were supposed to be retained in the lower conjunctival *cul-de-sac* and accordingly the mucoadhesive property of the inserts was evaluated. The mucoadhesion time of various ocular inserts was found to be 0.32–1.07 h. Control ocular inserts showed maximum mucoadhesion, which appears to be due to the presence of hydroxyl groups in the inserts (contributed by PVA and alginate), which could form hydrogen bonding with the mucosa. Cross-linking by CaCl_2 reduced the mucoadhesion. On coating with Eudragit polymers, cross-linked insert further reduced mucoadhesion as the surface hydroxyl group of the insert got coated with polymer having a carboxylic group and/or quaternary ammonium group. Thus, taking both mucoadhesion and sustained drug release into consideration, the cross-linked ocular insert coated with Eudragit RL-100 appears to be promising (Table II).

Fourier transform infrared (FTIR) spectroscopy

FTIR spectra of moxifloxacin hydrochloride (Fig. 1) showed aromatic C=C stretching at 1621 , 1515 and 1454 cm^{-1} and C–H bending for substituted benzene at 873 cm^{-1} . Besides, spectra also showed carboxylic acid C=O stretching at 1705 cm^{-1} , C–N stretching at 1350 cm^{-1} , stretching of monofluorobenzene at 1183 cm^{-1} . Spectra of a physical mixture of the drug, PVA and sodium alginate showed peaks at 3520 cm^{-1} due to stretching of the hydroxyl group, 2920 cm^{-1} due to C–H stretching and 1023 cm^{-1} due to C–O–C stretching (which appears to be contributed by PVA and alginate), along with drug peaks. The ATR spectra of the blank insert of PVA-alginate showed shifting of the hydroxyl peak from 3520 to 3306 cm^{-1} due to intermolecular hydrogen bonding, C–H stretching at 2913 and C–O–C stretching at 1027 cm^{-1} . Uncross-linked and cross-linked inserts showed hydroxyl stretching, C–H stretching along with asymmetric C–O–C stretching at 1247 cm^{-1} and symmetric C–O–C stretching at 1090 and 1023 cm^{-1} , indicating branching on the carbon atom adjacent to the oxygen atom. Besides, ATR spectra showed

Table II. Amount of moxifloxacin permeated and drained from ocular insert through Millipore membrane filter and goat cornea

Formulation	Thickness (mm) ^a	Muco-adhesive time (h) ^a	Drug concentration (mg mm ⁻²) ^a	Millipore filter membrane			Goat cornea		
				Drug permeated (%)	Drug drained (%)	Drug permeated (%)	Drug drained (%)	Drug permeated (%)	Drug drained (%)
MIF1	0.404 ± 0.003	1.07 ± 0.03	0.149 ± 0.03	34.95 ± 2.45 (5 h)	40.61 ± 0.47 (5 h)	15.57 ± 0.54 (4 h)	24.30 ± 0.34 (4 h)	76.02 ± 0.12	
MIF2	0.403 ± 0.003	0.90 ± 0.03	0.122 ± 0.001	49.63 ± 1.21 (6 h)	39.81 ± 0.85 (6 h)	17.42 ± 0.20 (4 h)	21.47 ± 0.39 (4 hrs)	76.61 ± 0.47	
MIF3	0.440 ± 0.003	0.35 ± 0.03	0.115 ± 0.002	56.77 ± 2.21 (7 h)	28.83 ± 0.81 (7 h)	17.47 ± 0.19 (4 h)	17.70 ± 0.04 (4 h)	76.72 ± 0.58	
MIF4	0.444 ± 0.055	0.32 ± 0.02	0.115 ± 0.001	68.42 ± 1.83 (9 h)	30.38 ± 0.24 (9 h)	18.21 ± 0.11 (4 h)	20.66 ± 0.13 (4 h)	76.99 ± 0.85	
MIF5	0.442 ± 0.003	0.33 ± 0.02	0.115 ± 0.001	59.98 ± 0.19 (7 h)	34.05 ± 0.70 (7 h)	17.51 ± 0.08 (4 h)	21.16 ± 0.51 (4 h)	76.58 ± 0.44	
MIF6	0.441 ± 0.003	0.42 ± 0.02	0.115 ± 0.001	52.43 ± 2.19 (6 h)	35.61 ± 1.57 (6 h)	17.20 ± 0.27 (4 hrs)	23.76 ± 0.33 (4 h)	76.02 ± 0.12	
MIF7	0.453 ± 0.003	0.35 ± 0.03	0.115 ± 0.001	53.29 ± 2.17 (5 h)	37.14 ± 0.55 (5 h)	17.22 ± 0.01 (4 h)	21.91 ± 0.18 (4 h)	76.39 ± 0.25	

^a Mean ± SD, n = 3.

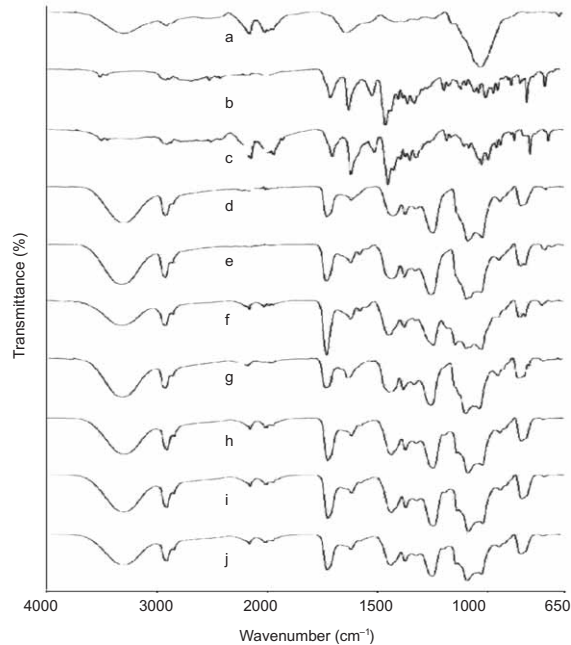


Fig. 1. FTIR spectra of: a) blank insert (without moxifloxacin), b) moxifloxacin, c) physical mixture of the drug, PVA and sodium alginate, d) control insert (not cross-linked and uncoated), MIF1, e) cross linked insert (CaCl_2), MIF2, f) optimal formulation (Eudragit RL-100), MIF4, g) Eudragit RS-100, MIF5, h) Eudragit S-100, MIF3, i) Eudragit E-100, MIF6, j) Eudragit L-100, MIF7.

aromatic C=C stretching, C=O stretching and C–N stretching. All the coated inserts showed aromatic C=C stretching at usual positions, indicating incorporation of moxifloxacin and peaks for ester at 1730 cm^{-1} , since acrylate polymers are esters. It would be important to mention here that ATR spectra provide information on the surface functional groups. Major characteristic peaks of moxifloxacin were found in the entire coated ocular insert, confirming the presence of the drug in the polymer without interaction.

In vitro/ex vivo permeation study

The results are shown in Table II, indicating that the *in vitro* drug release from the plain ocular insert (control) was sustained for 5 h. The drug release from MIF2 was extended up to 6 h. Formulation MIF6 did not have any effect on drug release, while MIF3 and MIF5 could sustain drug release up to 7 h. After coating with different grades of Eudragit, drug release from ocular inserts was found to be sustained due to the presence of polymeric film. Among all formulations, MIF4 showed the best sustaining effect. The MIF4 formulation gave 68.4 % drug permeated through the Millipore membrane filter after 9 hours.

To mimic real life conditions, permeation studies were then conducted with freshly excised goat cornea and the results are shown in Table II. Considering cornea viability, the experiment was conducted for 4 h and the drug permeation from inserts ranged between 15.6 and 17.5 %, which was less than the permeation observed with the Millipore membrane filter in 4 h. Millipore membrane filter acts as a mechanical barrier to drug diffusion while cornea [made of epithelium (lipophilic), stroma (hydrophilic), and endothelium (less lipophilic than epithelium)] acts as a lipophilic-hydrophilic barrier and the drug will have to partition through the barrier for corneal penetration. Accordingly, permeation through the cornea would be lower compared to that across the Millipore membrane filter. The corneal hydration level was found to be below 80 % in each experiment, indicating the inserts would not cause any damage to the cornea.

The release profiles of ocular inserts were treated with the Korsmeyer-Peppas equation and slope values were > 0.89 , indicating super case II type of drug release from ocular inserts through the Millipore membrane filter and excised goat cornea (15). Thus, all formulations followed the Korsmeyer-Peppas kinetics model (Table III).

In vivo drug release study

Formulations MIF4 and MIF5 were selected for *in vivo* drug release studies on the basis of maximum mucoadhesion and better sustained behavior of the drug among all insert formulations. Ocular inserts were removed carefully at hourly intervals (from

Table III. Kinetics of *in vitro* drug release from ocular inserts through Millipore membrane filter and goat cornea

Formulation	R^2								Mechanism of drug release ^a
	Zero-order ^a		First-order ^a		Higuchi ^a		Korsmeyer-Peppas ^a		
	Millipore membrane	Goat cornea	Millipore membrane	Goat cornea	Millipore membrane	Goat cornea	Millipore membrane ^a	Goat cornea ^a	
MIF1	0.8482	0.9938	0.9432	0.9979	0.9877	0.9511	0.9988	0.9997	Non-Fickian
MIF2	0.9781	0.9919	0.9622	0.9969	0.9871	0.9536	0.9992	0.9999	Non-Fickian
MIF3	0.9444	0.9930	0.9634	0.9975	0.9831	0.9514	0.9963	1.000	Non-Fickian
MIF4	0.9653	0.9926	0.9625	0.9983	0.9742	0.9515	0.9982	0.9993	Non-Fickian
MIF5	0.9626	0.9922	0.9748	0.9978	0.9891	0.9537	0.9991	0.9999	Non-Fickian
MIF6	0.9688	0.9889	0.9654	0.9950	0.9712	0.9577	0.9964	0.9999	Non-Fickian
MIF7	0.9642	0.9938	0.9678	0.9979	0.9702	0.9511	0.9964	0.9997	Non-Fickian

^a $n = 3$.

1–10 hours) and analyzed for residual drug content. The drug was subtracted from the initial content of the insert and gave the amount of drug released in the rabbit eye. Fig. 3 presents the cumulative drug release (%) from formulations MIF4 and MIF5 at different time intervals. The *in vivo* study suggests the formulation MIF4 showed maximum cumulative drug release of 85.0 % and release was sustained up to 10 h. Formulation MIF5 showed 72.1 % cumulative release which was sustained up to 8 h. Also, formulation MIF4 had more tendency to stick to ocular mucosa due to the presence of higher concentration of quaternary ammonium groups (10 %) in Eudragit RL-100 as compared to Eudragit RS-100 (5 %). The presence of quaternary ammonium groups renders positive charge to the polymers by which it can interact with anionic mucin and thereby increase its residence on corneal surface.

Stability

Finally, accelerated stability studies at elevated temperature and humidity revealed no significant changes in color in insert formulations. The moxifloxacin concentration in all formulations at accelerated and room temperature are shown in Fig. 2. The degradation rate constants (k_{cal}) and shelf life (t_{90}) values for MIF1, MIF3, MIF5–MIF7 at room temperature were found to range between 2.12 and 1.69 days⁻¹, and 490–617 days. MIF2 and MIF4 showed lower degradation constants of 1.12 and 1.41 day⁻¹ and longer shelf life of 926 and 739 days, resp. The stability study concluded that formulations MIF2 and MIF4 showed the lowest degradation and maximum shelf life as per the ICH guidelines (17) suggest there is no need to add overages to ensure 2 year shelf life.

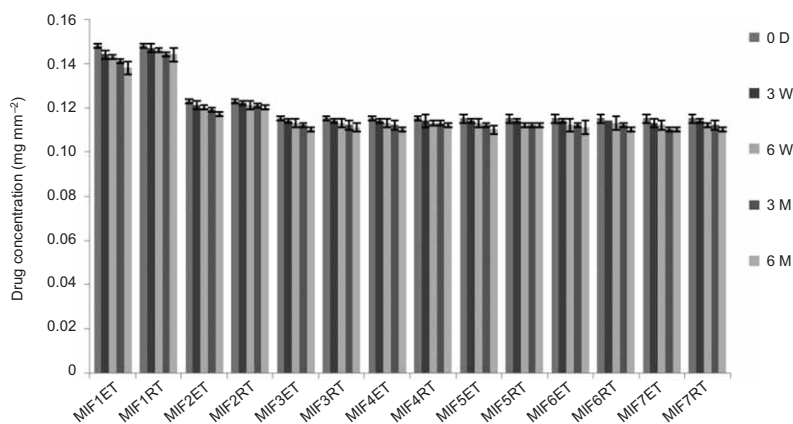


Fig. 2. Stability of moxifloxacin ocular inserts under accelerated condition and room temperature. Mean \pm SD ($n = 3$). ET – elevated temperature (40 °C), RT – room temperature, D – days, W – weeks, M – months.

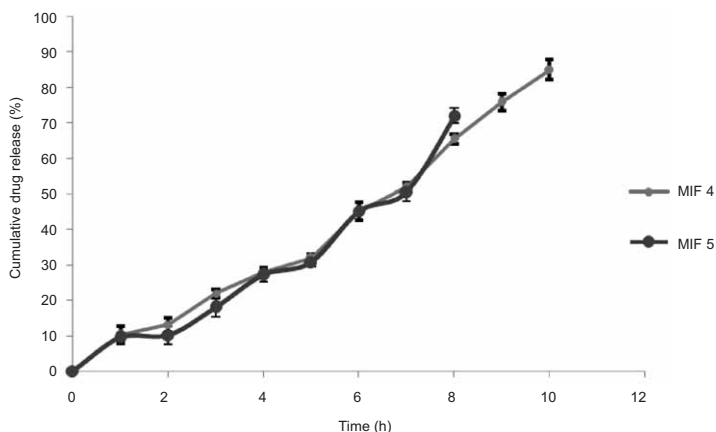


Fig. 3. *In vivo* moxifloxacin release profile from ocular inserts.

CONCLUSIONS

In conclusion, we suggest that the ocular insert formulation of moxifloxacin can be a promising vehicle for topical ocular administration of antibiotics. Its application could possibly replace the use of fortified solutions of antimicrobials and reduce the necessity for repeated drug administration at frequent intervals, thereby potentially lowering corneal toxicity and increasing patient compliance.

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S A Ž E T A K

Dizajniranje i vrednovanje okularnih umetaka moksifloksacin hidroklorida

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Cilj rada bio je priprava i evaluacija okularnih umetaka moksifloksacina. Okularni umetak izrađen je od vodene suspenzije moksifloksacina, natrijevog alginata, polivinilnog alkohola i dibutil-ftalata metodom odlijevanja filma. Okularni umetak (promjera 5,5 mm) umrežen je pomoću CaCl₂ i obložen Eudragitom S-100, RL-100, RS-100, E-100 ili Eudragit L-100. *In vitro* drenaža/permeacija lijeka proučavana je koristeći staklenu modificiranu Franzovu difuzijsku ćeliju. Koncentracija lijeka i vrijeme mukoadhezije oku-

larnih umetaka bili su zadovoljavajući. Umrežavanje i oblaganje polimerima produjilo je drenažu iz umetaka. Umreženi okularni umetci obloženi s Eudragit RL-100 pokazali su veću permeaciju lijeka u odnosu na ostale pripravke.

Ključne riječi: moksifloksacin, okularni umetak, kornea koze

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