

Apoptosis in Psoriasis

Marija Kaštelan, Larisa Prpić-Massari, Ines Brajac

University Department of Dermatology and Venereology, Rijeka University Hospital Center and School of Medicine, Rijeka, Croatia

Corresponding author:

Professor Marija Kaštelan, MD, PhD
University Department of Dermatology
and Venereology
Rijeka University Hospital Center and
School of Medicine
Krešimirova 42
HR-51000 Rijeka, Croatia
marijakastelan@yahoo.com

Received: January 15, 2009

Accepted: June 10, 2009

SUMMARY Apoptosis is a process of programmed cell death that maintains homeostasis of the skin. Apoptotic cell death regulates keratinocyte proliferation and formation of stratum corneum. The process by which keratinocytes undergo apoptosis is a multistep program mediated by binding of specific death ligands to death receptors or by the release of effector cell granules. Dysfunctional apoptosis has an important role in the development of several skin diseases. Psoriasis is a common chronic inflammatory skin disease characterized by hyperproliferation with incomplete differentiation of epidermal keratinocytes and decreased keratinocyte apoptosis. Psoriatic keratinocytes possess an enhanced ability to resist apoptosis, which might be one of the key pathogenetic mechanisms in psoriasis. In addition, psoriasis is nowadays also recognized as the most prevalent autoimmune disease resulting from aberrant activation of both innate and adaptive immunity. However, the role of cell cytotoxicity mediated by cytotoxic CD8+ T cells and NK cells in psoriasis is as yet unclear. Here, we review the role of different apoptotic pathways in psoriasis.

KEY WORDS: apoptosis, Bcl-2 family proteins, Fas/FasL, perforin, psoriasis

INTRODUCTION

Apoptosis is a unique process of programmed cell death that maintains homeostasis of the skin. Kerr *et al.* were the first to describe apoptosis as an active process morphologically characterized by cell shrinkage, nuclear condensation, cellular fragmentation and phagocytosis by neighboring macrophages and dendritic cells (1). Apoptosis plays a critical role in several physiologic functions such as cell deletion during embryonic development, balancing cell number in continuously renewing tissues, and immune system develop-

ment (2). In the skin, apoptotic cell death regulates keratinocyte proliferation and formation of stratum corneum. Balance between cell death and cell proliferation maintains homeostasis of epidermal compartment (3). The gradients of antiapoptotic and apoptotic factors control the timing of apoptosis in the epidermis, thus regulating epidermal growth and differentiation. It seems that terminal differentiation of keratinocytes represents simply a special form of apoptosis (4).

Apoptotic pathways relevant in keratinocyte apoptosis include several mechanisms. The "extrinsic" pathway is triggered by binding of Fas ligand (FasL) or tumor necrosis factor (TNF) to membrane death receptors that recruit adapter molecules leading to the activation of caspase-8 (2,5). The "intrinsic" pathway includes mitochondrial release of cytochrome *c* and along with the cofactor Apaf-1, the formation of an activated caspase-9 apoptosome (2,6). Mitochondria might also trigger apoptosis through the release of Smac/DIABLO blocking inhibitor of apoptosis (IAP) or by apoptosis-inducing factor (AIF) that mediates caspase-independent apoptosis (2). Finally, both pathways end in DNA cleavage followed by the formation of apoptotic bodies and phagocytosis by neighboring cells. Apoptosis is controlled by Bcl-2 family proteins of which some (Bcl-2, Bcl-x_L) may block apoptosis, whereas others (Bax, bak, Bid) stimulate apoptotic process (7).

Dysfunctional apoptosis has an important role in the development of several skin diseases (2,8). Some are characterized by increased keratinocyte apoptosis, e.g., toxic epidermal necrolysis or graft-versus-host disease, whereas others like non-melanoma skin cancers and psoriasis are associated with decreased apoptosis. It seems that diseases with increased apoptosis tend to be acute, whereas those characterized by decreased apoptosis are mostly chronic and associated with epidermal hyperplasia or hyperkeratosis (2).

PSORIASIS AND Bcl-2 FAMILY PROTEINS

Psoriasis is a common chronic inflammatory skin disease characterized by hyperproliferation and incomplete differentiation of epidermal keratinocytes (9). Psoriatic keratinocytes possess an enhanced ability to resist apoptosis, which might be one of the key pathogenetic mechanisms in psoriasis (10). However, up to now limited data exist regarding the underlying mechanisms of this defect in the apoptosis control mechanisms of psoriatic keratinocytes. As mentioned before, the process of apoptosis is controlled by Bcl-2 family proteins including several pro-apoptotic and anti-apoptotic proteins (11). Data regarding expression of Bcl-2 family proteins in psoriatic plaques are controversial. Some groups have reported an overexpression of Bcl-2 protein, whereas others observed no expression of anti-apoptotic Bcl-2 molecule in psoriatic epidermis (12,13). Takahashi *et al.* (14) found higher expression of Bcl-x_L and Bax proteins in psoriatic epidermis, as also reported by

other authors (15,16). Tomkova *et al.* observed diffuse staining of pro-apoptotic Bax molecule in psoriatic lesions, however, along with significant overexpression of apoptosis suppressing Bcl-x_L (15). Therefore, additional studies are necessary to determine whether the psoriatic epidermis is in a pro-apoptotic or antiapoptotic status regarding the expression of Bcl-2 and Bax proteins in psoriasis. Data on the expression of anti-apoptotic Bcl-x_L protein showed its overexpression in all layers of psoriatic epidermis (10,14,17). It seems that up-regulation of Bcl-x_L molecule is at least partly responsible for the increased epidermal thickness, a hallmark of psoriasis. Moreover, psoriatic keratinocytes are extremely resistant to apoptosis compared with normal-skin derived keratinocytes (10). So, it has been suggested that overexpression of Bcl-x_L by psoriatic keratinocytes and their resistance to undergo apoptosis finally lead to epidermal acanthosis in psoriasis. It has been recently shown that tumor necrosis factor- α (TNF- α) stimulates the synthesis of anti-apoptotic Bcl-x and bcl-2 as well as pro-apoptotic Bax protein in psoriatic lesions (12,18). Having in mind the important role of TNF- α in psoriatic inflammatory cascade and the efficacy of anti-TNF treatment in psoriasis, it is likely that TNF- α influences the apoptotic process in psoriatic epidermis through changes in the intracellular Bax/Bcl-2 ratio. Delayed apoptosis of psoriatic keratinocytes and subsequent epidermal hyperplasia are partly due to an overexpression of keratinocyte-derived IL-15 cytokine and its receptor IL-15R in psoriatic epidermis (19). Cytokine IL-15 is also a potent chemoattractant like T cell and NK cell growth factor. It has a role in the influx and activation of neutrophils in psoriatic epidermis (19). So, it seems that keratinocyte-derived IL-15 is responsible for prolonged keratinocyte survival and also for T cell and neutrophil accumulation in psoriatic lesions.

The terminal deoxynucleotidyl transferase-mediated dUTP-biotin nick-end labeling (TUNEL) method is widely used for the detection of apoptotic cells (20). Most keratinocytes in psoriatic lesions are TUNEL-positive cells, however, without morphological evidence of apoptosis (13,21). Moreover, Laporte *et al.* observed a decreased number of apoptotic cells in psoriatic epidermis compared to normal one (21). Kawashima *et al.* report an increase of double-strand DNA breaks in psoriatic lesions as the result of active DNA replication but not apoptosis (17). Therefore, it is likely that abundant TUNEL-positive keratinocytes in psoriatic epidermis are rather proliferating than apoptotic cells.

PSORIASIS AND PERFORIN/GRANZYME OR Fas/FasL APOPTOTIC MECHANISM

Psoriasis is nowadays also recognized as a T-cell mediated disease resulting from aberrant activation of both innate and adaptive immunity (22). The psoriatic inflammatory cascade is orchestrated by proinflammatory CD4+ T cells producing interferon- γ (Th1 cells) or interleukin-17 (Th17 cells) and cytotoxic CD8+ T cells (CTLs) producing Th1 cytokine pattern. In addition, it is also driven by activated keratinocytes that influence T cell activation and trafficking, as well as by activation of natural killer (NK) cells and natural killer-like T (NK-T) cells representing innate immunity (22). Recently, involvement of IL-17 and IL-22 in psoriasis has been also recognized. It is likely that the secretion of cytokines such as IL-22 and IL-17 could result in keratinocyte hyperproliferation, leading to psoriatic skin lesions (23). Apart from T-cells, certain subsets of dendritic cells (DCs) are found to be important in the orchestration of the disease process (24). Proinflammatory CD11c+ DCs, which produce TNF- α , are increased in psoriasis, whereas Langerhans cells from symptomless psoriatic skin show migratory defects (24-26). Moreover, psoriatic dermal DCs promote increased proliferation of autoreactive T cells and production of Th1 cytokines, IFN- γ and IL-2.

The role of cell cytotoxicity mediated by cytotoxic CD8+ T cells and NK cells in psoriasis is as yet unclear. CTLs and NK cells mediate apoptosis *via* the release of cell granules, perforin and granzymes, or by binding of ligands to their death receptors on target cells (27). Upon activation, CTLs and NK cells release perforin, a pore forming molecule, which enables entry of granzyme into the target cell in order to mediate DNA degradation (28). The importance of perforin-mediated cytotoxicity has been demonstrated in several autoimmune diseases and in some inflammatory skin diseases as well (29,30). We have previously reported on the systemic up-regulation of perforin molecule in the exacerbation phase of psoriasis that were mainly related to perforin-positive CD8+P+ cells, a subpopulation with cytotoxic potential and rapid killing cells (31). A similar situation was found in psoriatic lesions where we observed significant accumulation of perforin-positive cells in suprabasal epidermis in close contact with damaged keratinocytes (32). Yawalkar *et al.* also report on up-regulation of perforin and granzyme-B expression in psoriatic lesions (33). It is likely that perforin-positive T cells induce damage to adjacent keratinocytes subsequently triggering

an injury response program and regenerative hyperplasia, a type of hyperplasia programmed in keratinocytes as an injury-repair response pathway (34,35). In addition to the established role for perforin as an effector molecule, it may also act as a regulatory molecule of the immune system (36). In the skin, CTLs might use perforin molecule to regulate antigen presentation by antigen presenting cells, which could result in persistence of the chronic immune system activation (36).

Apoptosis induced by CTLs may also be mediated by the Fas ligand (FasL) binding to Fas (CD95) on target cell (37). In addition, Fas expression on keratinocytes can be up-regulated by Th1 type cytokines that create a microenvironment typical for psoriasis (38). Several groups have reported overexpression of Fas in psoriatic epidermis, so it is likely that Th1 cytokines up-regulate Fas expression in psoriatic lesions (14). Data on FasL expression in psoriatic skin are controversial, as some groups report low FasL expression and the others observed an increase of FasL in all cell layers of psoriatic epidermis (39,40). However, it has been recently suggested that Fas/FasL signaling might induce an alternative pathway promoting the synthesis of inflammatory cytokines, TNF- α and IL-8 instead of apoptosis (41). Assuming the fact that psoriatic keratinocytes are relatively resistant to apoptosis, it is likely that elevated expression of Fas along with anti-apoptotic Bcl-x_L protein in psoriatic epidermis inhibit Fas-mediated apoptosis and induce proinflammatory TNF- α production by keratinocytes. Moreover, it is likely that Fas/FasL pathway is an essential early event in psoriasis induction (41).

In conclusion, all these findings clearly show the importance of apoptotic process in the development and maintenance of psoriatic lesions. Based on these observations, novel apoptosis-based therapies could be directed towards enhancement of apoptotic process in psoriasis.

Acknowledgment. This study was financially supported by the Ministry of Science, Education and Sports of the Republic of Croatia (No. 062039-0197).

References

1. Kerr JFR, Wyllie AH, Currie AR. Apoptosis. A basic biological phenomenon with wide ranging implications in tissue kinetics. *Br J Cancer* 1972;26:239-57.

2. Raj D, Brash DE, Grossman D. Keratinocyte apoptosis in epidermal development and disease. *J Invest Dermatol* 2006;126:243-57.
3. Bowen AR, Hanks AN, Allen SM, Alexander A, Diedrich MJ, Grossman D. Apoptosis regulators and responses in human melanocytic and keratinocytic cells. *J Invest Dermatol* 2003;120:48-55.
4. Allombert-Blaise C, Tamiji S, Mortier L, Fauvel H, Tual M, Delaporte E, *et al.* Terminal differentiation of human epidermal keratinocytes involves mitochondria- and caspase-dependent cell pathway. *Cell Death Differ* 2003;10:850-2.
5. Curtin JF, Cotter TG. Live or let die: regulatory mechanisms in Fas-mediated apoptosis. *Cell Signal* 2003;15:983-92.
6. Van Gurp M, Festjens N, van Loo G, Saelens X, Vandenameele P. Mitochondrial intermembrane proteins in cell death. *Biochem Biophys Res Commun* 2003;304:487-97.
7. Bowen AR, Hanks AN, Murphy KJ, Florell SR, Grossman D. Proliferation, apoptosis, and survivin expression in keratinocytic neoplasms and hyperplasia. *Am J Dermatopathol* 2004;26:177-81.
8. Thompson CB. Apoptosis in the pathogenesis and treatment of disease. *Science* 1995;267:1456-62.
9. McKay IA, Leigh IM. Altered keratinocyte growth and differentiation in psoriasis. *Clin Dermatol* 1995;13:105-14.
10. Wrone-Smith T, Mitra RS, Thompson CB, Jasty R, Castle VP, Nickoloff BJ. Keratinocytes derived from psoriatic plaques are resistant to apoptosis compared with normal skin. *Am J Pathol* 1997;151:1321-9.
11. Adams JM, Cory S. The bcl-2 protein family: arbiters of cell survival. *Science* 1998;281:1322-6.
12. Wrone-Smith T, Johnson T, Nelson B, Boise LH, Thompson CB, Nunez G, *et al.* Discordant expression of Bcl-x and Bcl-2 by keratinocytes *in vitro* and psoriatic keratinocytes *in vivo*. *Am J Pathol* 1995;146:1079-88.
13. Bianchi L, Farrace MG, Nini G, Piacentini M. Abnormal bcl-2 and tissue transglutaminase expression in psoriatic skin. *J Invest Dermatol* 1994;103:829-33.
14. Takahashi H, Manabe A, Ishida-Yamamoto A, Hashimoto Y, Iizuka H. Aberrant expression of apoptosis-related molecules in psoriatic epidermis. *J Dermatol Sci* 2002;28:187-97.
15. Tomkova H, Fujimoto W, Arata J. Expression of the bcl-2 homologue Bax in normal human skin, psoriasis vulgaris and non-melanoma skin cancers. *Eur J Dermatol* 1998;8:256-60.
16. Fukuya Y, Higaki M, Kawashima M. Effect of vitamin D₃ on the increased expression of Bcl-x_L in psoriasis. *Arch Dermatol Res* 2002;293:620-5.
17. Kawashima, Doi H, Ito Y, Shibata MA, Yoshinaka R, Otsuki Y. Evaluation of cell death and proliferation in psoriatic epidermis. *J Dermatol Sci* 2004;35:207-14.
18. Krüger-Krasagakis S, Galanopoulos VK, Giannikaki L, Stefanidou M, Tosca AD. Programmed cell death of keratinocytes in infliximab-treated plaque-type psoriasis. *Br J Dermatol* 2006;154:460-6.
19. Rückert R, Asadullah K, Seifert M, Budagian VM, Arnold R, Trombotto C, *et al.* Inhibition of keratinocyte apoptosis by IL-15: a new parameter in the pathogenesis of psoriasis. *J Immunol* 2000;165:2240-50.
20. Gavrieli Y, Sherman Y, Ben-Saason SA. Identification of programmed cell death *in situ* via specific labeling of nuclear DNA fragmentation. *J Cell Biol* 1992;119:493-501.
21. Laporte M, Galand P, Fokan D, de Graef C, Heene M. Apoptosis in established and healing psoriasis. *Dermatology* 2000;200:314-6.
22. Ghoreschi K, Weigert C, Röcken M. Immunopathogenesis and role of T cells in psoriasis. *Clin Dermatol* 2007;25:574-80.
23. Nograles KE, Zaba LC, Guttman-Yassky E. Th17 cytokines interleukin (IL)-17 and IL-22 modulate distinct inflammatory and keratinocyte response pathways. *Br J Dermatol* 2008;159:1092-102.
24. Boyman O, Conrad C, Tonel G, Gilliet M, Nestle FO. The pathogenic role of tissue-resident immune cells in psoriasis. *Trends Immunol* 2007;28:51-7.
25. Lowes MA, Bowcock AM, Krueger JG. Pathogenesis and therapy of psoriasis. *Nature* 2007;445:866-73.
26. Cumberbatch M, Singh M, Dearman RJ, Young HS, Kimber I, Griffiths CEM. Impaired Langerhans cell migration in psoriasis. *J Exp Med* 2006;203:953-60.
27. Schultz DR, Harrington WJ. Apoptosis: pro-

- grammed cell death at a molecular level. *Semin Arthritis Rheum* 2003;32:345-69.
28. Yagita H, Nakata M, Kawasaki A, Shinkai J, Okumura K. Role of perforin in lymphocyte-mediated cytotoxicity. *Adv Immunol* 1992;51:215-41.
 29. Gulan G, Ravlic-Gulan J, Štrbo N, Sotosek V, Nemeč B, Matovinović D. Systemic and local expression of perforin in lymphocyte subsets in acute and chronic rheumatoid arthritis. *J Rheumatol* 2003;30:660-70.
 30. Teraki Y, Shiohara T. Apoptosis and the skin. *Eur J Dermatol* 1999;9:413-26.
 31. Prpić L, Štrbo N, Sotošek V, Gruber F, Podack ER, Rukavina D. Assessment of perforin expression in peripheral blood lymphocytes in psoriatic patients during exacerbation of disease. *Acta Derm Venereol Suppl (Stockh)* 2000;211:14-6.
 32. Kaštelan M, Prpić-Massari L, Gruber F, Zamolo G, Žauhar G, Čoklo M, *et al.* Perforin expression is up-regulated in the epidermis of psoriatic lesions. *Br J Dermatol* 2004;151:831-6.
 33. Yawalkar N, Schmid S, Braathen LR, Pichler WJ. Perforin and granzyme B may contribute to skin inflammation in atopic dermatitis and psoriasis. *Br J Dermatol* 2001;144:1133-9.
 34. Kaštelan M, Prpić-Massari L, Brajac I. Apoptosis mediated by cytolytic molecules might be responsible for maintenance of psoriatic plaques. *Med Hypotheses* 2006;67:336-7.
 35. Krueger JG, Bowcock A. Psoriasis pathophysiology: current concepts of pathogenesis. *Ann Rheum Dis* 2005;64 (Suppl II):ii30-ii36.
 36. Stepp SE, Porunello MA, Bennett M, de Saint Basile G, Kumar V. Perforin: more than just effector molecule. *Immunol Today* 2000;21:254-6.
 37. Peter ME, Krammer PH. The CD95 (Apo-1/Fas) DISC and beyond. *Cell Death Differ* 2003;10:26-35.
 38. Nickoloff BJ. The immunologic and genetic basis of psoriasis. *Arch Dermatol* 1999;135:1104-10.
 39. Gutierrez-Steil C, Wrone-Smith T, Sun X, Krueger JG, Coven T, Nickoloff BJ. Sunlight-induced basal cell carcinoma tumor cells and ultraviolet-B-irradiated psoriatic plaques express Fas ligand (CD95L). *J Clin Invest* 1998;101:33-9.
 40. Lee SH, Jang JJ, Lee JY, Kim SY, Park WS, Shin MS, *et al.* Fas ligand is expressed in normal skin and some cutaneous malignancies. *Br J Dermatol* 1998;139:186-91.
 41. Gilhar A, Yaniv R, Assy B, Serafimovich S, Ullman Y, Kalish RS. Fas pulls the trigger on psoriasis. *Am J Pathol* 2006;168:170-5.