

## Hypereosinophilic Syndrome and Proliferative Diseases

Marius A. Ionescu<sup>1,2</sup>, Li Wang<sup>1,2</sup>, Anne Janin<sup>1,2,3</sup>

<sup>1</sup>Inserm, U728; <sup>2</sup>Université Paris 7; <sup>3</sup>AP-HP, Hôpital Saint-Louis, Service de Pathologie, Paris, France

### Corresponding author:

Anne Janin, MD, PhD  
Service de Pathologie and Inserm U728,  
Université Paris VII  
Hôpital Saint Louis  
1 Avenue Claude Vellefaux  
75475, Paris Cedex 10  
France  
anne\_janin@yahoo.com

Received: June 16, 2009

Accepted: November 5, 2009

**SUMMARY** Therapy principles of the last decade and recent advances in the research of polynuclear eosinophil have led to a new approach in the hypereosinophilic syndrome (HES), with important consequences on the development of new and effective therapies. HES is defined by persistent and marked eosinophilia and eosinophil-related organ damage in the absence of any evident cause of hypereosinophilia. Two variants of HES have been characterized, with different prognosis and possible associations with malignant diseases such as myeloid leukemia or T-cell lymphomas. The lymphocytic variant of HES (L-HES) is characterized by the presence of T cell clones, IL-5 expression and possible progression to T-cell lymphoma. Besides steroid therapy, the anti-IL-5 monoclonal antibody mepolizumab is considered as a target therapy for L-HES. The myeloproliferative variant of HES (M-HES) is associated with an increased risk of myeloid leukemia and good response to anti-tyrosine-kinase therapy. The imatinib target is a kinase resulting from an 800-kb deletion on chromosome 4. The fusion gene Fip1-like 1-platelet-derived growth factor receptor alpha (FIP1L1-PDGFR $\alpha$ ) has been validated as a marker of response to anti-tyrosine-kinase therapy. Early identification of HES variants is crucial for the rapid introduction of early and appropriately adjusted therapy.

**KEY WORDS:** hypereosinophilic syndrome, T-cell lymphomas, myeloproliferative disorders

### INTRODUCTION

Hypereosinophilia is commonly associated with atopy, parasitosis, drug hypersensitivity, and malignancies. When there is no underlying disease but multiple organ involvement by eosinophilic infiltrates, the diagnosis of hypereosinophilic syndrome (HES) is proposed (1). HES is a rare disorder with an estimated incidence rate of 0.5 to 1.0 cases *per* 100,000 inhabitants *per* year (2) and is characterized by persistent eosinophilia associated with end-organ damage (3). HES prognosis can be poor because of eosinophilic myocarditis

and fibrosis, or because of associated malignant diseases such as myeloid leukemia or T-cell lymphoma (4-6).

### THE HYPEREOSINOPHILIC SYNDROME: THE EVOLUTION OF THE CONCEPT

Hypereosinophilia with no evident etiology was initially proposed as a syndrome by Hardy and Anderson in 1968 (3). In 1975, Chusid *et al.*

(7) established diagnostic criteria for "idiopathic HES": 1) peripheral eosinophilia ( $>1500$  eosinophils/ $\text{mm}^3$ ) for more than six months; 2) absence of reactive eosinophilia, caused by parasites, allergies, neoplasia; and 3) evidence of end-organ damage related to hypereosinophilia. The clinical spectrum for idiopathic HES was proposed at the National Institute of Health Conference in 1982 (8). In 1994, clonal proliferation of T-cell type helper 2 (Th2) was reported in one case of HES (4).

The lymphocytic variant of HES (L-HES) was later defined and related to an increased interleukin-5 (IL-5) expression by clonal T-cells and activated eosinophils. IL-5 is one of the key mediators for eosinophilic activation and survival (4-6). Based on these findings, L-HES targeted therapies have been recently developed, using anti-IL-5 monoclonal antibodies (mepolizumab, alemtuzumab) associated with systemic steroids (9).

The myeloid variant of HES (M-HES) was described in the last decade, based on HES and chronic myeloid leukemia (CML) cases that responded successfully to tyrosine-kinase inhibitors (imatinib mesylate) (10). In M-HES and CML cases, interstitial deletions on chromosome 4q12 were found, with fusion of the FIP1-like 1 and platelet-derived growth factor receptor alpha genes, and a fusion product encoding a protein with constitutive tyrosine kinase activity (11). Markers of response to imatinib (FIP1L1-PDGFR- $\alpha$  fusion gene) were then identified (Thr 674Ile mutation) (12-16). New generations of tyrosine-kinase inhibitors have now been developed (nilotinib, PKC412).

With characterization of the two HES variants (L-HES and M-HES), "idiopathic HES" is now a temporary diagnosis necessitating further investigations (11,17,18).

### **CLINICAL FEATURES AND LABORATORY FINDINGS IN HES VARIANTS**

Cutaneous and mucosal signs occur in 27% to 64% of HES cases (19-22), in both L-HES and M-HES variants.

#### **Lymphocytic HES variant (L-HES)**

This variant of HES is characterized by pruritus, nonspecific erythematous papules and plaques, urticaria-like lesions, erythroderma, and angioedema. Endomyocardial fibrosis is rarely reported; pulmonary and digestive involvement due to eosinophilic infiltrates can be found. In this variant, clonal CD3- CD4+ T cells of phenotype express

IL-4, IL-5 and IL-13 cytokines (Th2 profile). Progression to T cell lymphomas has been reported (18,23,24). This form is equally distributed in females and males.

#### **Myeloproliferative HES variant (M-HES)**

M-HES shows male predominance with 4-9:1 ratio (25) and is characterized by hepatosplenomegaly; mucosal ulcerations can occur (26). Blood tests show persistent eosinophilia, increased levels of serum tryptase, vitamin B12 and leukocyte alkaline phosphatase, anemia and/or thrombocytopenia, and circulating leukocyte precursors (27). The prognosis of the M-HES variant is poor, with severe cardiac complications, resistance to steroid therapy, and an increased risk of developing myeloid malignancies. Patients with M-HES can develop blast crisis revealing acute eosinophilic or myeloid leukemia. They can also develop granulocytic sarcoma (28). Mucosal lesions such as ulcerations, erosions or aphthous lesions may occur in M-HES and are considered as one of the clinical markers of M-HES with characteristic genetic mutation (2,29). The mucosal lesions can be discrete, round or oval ulcers, located on oral, labial, conjunctival and/or genital mucosa. Oral ulcerations in HES must be differentiated from ulcerations of other causes, mainly aphthae (primarily aphthosis in AIDS), Behçet's disease, local traumas, malignant neoplasms, drug- or irradiation-induced ulcerations, cyclic neutropenia, FAPA (fever, aphthous stomatitis, pharyngitis, adenitis), gluten-sensitive enteropathy, and hematinic deficiencies (iron, zinc, folate, vitamin B) (30). A major complication of severe HES, whatever the variant, is endomyocardial necrosis (often asymptomatic) or endomyocardial thrombosis. This is followed by endomyocardial fibrosis, leading to severe cardiac failure (31). Atrioventricular valves can be involved in the late stage of endomyocardial fibrosis, leading to congestive heart failure. Peripheral thromboembolism may also occur, independently of endomyocardial lesions, due to an increased number of blood eosinophils (32,33).

#### **PATHOLOGY**

Being present in blood ( $>1500/\text{mm}^3$ ), eosinophils can also be localized in the skin or mucosa, inducing similar histopathologic changes in both variants. On hematoxylin-eosin stain, skin biopsies from HES untreated lesions such as plaques or papules show a cellular infiltrate composed of lymphocytes, macrophages and eosinophils that are characterized by bilobate nuclei and eosinophilic

cytoplasm. May-Grünwald-Giemsa stain provides precise assessment of eosinophil count and tissue distribution. Activated eosinophils are often degranulated, with eosinophil granules scattered in the intercellular space within the inflammatory infiltrate. On immunohistochemistry, antibodies directed against eosinophil cationic proteins (eosinophil peroxidase (EPO), major basic protein (MBP), eosinophil cationic protein (ECP) and eosinophil-derived neurotoxin (EDN)) can be used on histologic sections in order to identify eosinophils or their extracellular granules as the result of cellular degranulation. Electron microscopy (EM) allows for precise identification of eosinophils and their degranulation. Intact, non-activated eosinophils are characterized by cytoplasmic granules with a dense matrix and crystallized central core. When eosinophils are activated, they undergo degranulation and have an inverted density of the central cores of the cytoplasmic granules associated with a partially dissociated cytoplasm (34,35). Eosinophil activation can also lead to full cell lysis, with ultrastructural features associating a necrotic nucleus, a disintegrated cytoplasm, and surrounding extracellular free granules (36,37). The ultimate stage of eosinophil lysis results in the constitution of Charcot-Leyden crystals, with a characteristic sharp and dense crystal shape on EM (38). These Charcot-Leyden crystals are resistant markers of eosinophil lysis and can be found within the tissue two to three weeks after eosinophil lysis (39). They can also be detected in the blood and body fluids such as bronchoalveolar lavage or urine (40).

## PATHOPHYSIOLOGY

The activation of polynuclear eosinophilic cells is made first by the release of cationic proteins, through eosinophil degranulation and/or lysis that induce tissue damage (41). Eosinophil infiltrates can be found in the skin and mucosa lesions of HES patients (42,43), in digestive tract of patients with eosinophilic gastroenteritis (44), and in both the skin and duodenal mucosa of patients with dermatitis herpetiformis (45). Hypodense eosinophils are among basic signs of blood eosinophil activation. Tissue eosinophil infiltrates show signs of cellular activation, i.e. the release of cationic proteins, the presence of hypodense granules on ultrastructural examination, and eosinophilic synthesis of IL-5.

Eosinophil activation is also associated with acute symptoms such as flare-ups (46-48) or early recurrences (49,50) of inflammatory diseases, microvascular lesions and necrosis (51,52). The

release of cationic proteins (ECP and MBP) can also lead to lipid membrane disruption and target cell death (53). In hypereosinophilic syndromes, activated eosinophils are observed in early oral ulcerations (20,21) as well as in late endomyocardial fibrosis (54).

Eosinophil trafficking to tissues and activation are also increased in HES. Eosinophils are derived from bone marrow myeloid progenitors (granulocyte-erythrocyte-megakaryocyte-macrophage colony-forming units, GEMM-CFU) through the action of granulocyte-macrophage colony stimulating factor (GM-CSF), interleukin (IL)-3 and IL-5 (55). Eosinophils are then released into the blood stream and migrate within 24-48 hours towards their target tissues (56-58). In physiological conditions, tissue eosinophils survival is short (one or two days) (52,59,60). IL-5 expressed by T cells or/and by eosinophils increases the survival of eosinophils (61,62).

In L-HES, the main role is played by IL-5 and eotaxin-1, eotaxin-2 and eotaxin-3. IL-5 is the main factor for the activation and *in situ* survival of tissue eosinophils (63-65). IL-5 can be produced by lymphocytes, mastocytes, basophils and eosinophils (66-70). T-helper lymphocytes with a cytokine profile type Th2 express IL-5, IL-4 and IL-13. This type of response is linked to hypereosinophilia, high serum IgE levels and polyclonal hypergammaglobulinemia, mainly found in L-HES (71,72). Th2 T cells are found in atopic dermatitis, commonly associated with eosinophilia (73). Clonal Th2 T cells are also found in T-cell lymphomas, often as a complication of L-HES (74). Eosinophil recruitment and activation can be induced by an overproduction of IL-5, IL-3 and GM-CSF (75,76). This is observed mainly in the L-HES variant associated with Hodgkin's disease, T-cell lymphomas or solid tumors (70,77). Eosinophil autocrine expression of IL-5 involves chronic eosinophil infiltration (31,40,44,49).

In M-HES, the eosinophilic recruitment, proliferation, and activation can be induced by an acquired chromosome change within myeloid cells (78,79), followed by an increased hematopoiesis and subsequently associated with myeloproliferative disorders. M-HES characteristic genetic changes are deletions on chromosome 4q12 and induction of FIP1L1-PDGFR- $\alpha$  fusion gene and platelet-derived growth factor receptor alpha genes, with the consequent fusion product encoding a protein that has constitutive tyrosine kinase activity. The marker of anti-tyrosine-kinase therapy response is the presence of FIP1L1-PDGFR- $\alpha$  fusion gene, while the

marker of resistance to this type of therapy is the mutation of Thr 674Ile gene (12-16).

## TREATMENT

Steroids are first line therapy for HES. Topic or systemic corticotherapy decreases dramatically the eosinophilic activation and tissue recruitment by inducing eosinophilic apoptosis (18,28).

### Treatment of L-HES

In the lymphocytic variant, steroids are first line therapy. Interferon alpha (IFN- $\alpha$ ) and cyclosporin A can be co-administered. If signs of malignant transformation are diagnosed, the treatment lines specific for T-cell lymphoma are proposed, with chemotherapy (cyclophosphamide, hydroxydoxorubicin, oncovin, prednisone, i.e. CHOP-like regimen), with or without monoclonal antibodies, fludarabine, 2-chlorodeoxyadenosine (2-CdA). In resistant or recurrent cases, intensive high dose chemotherapy followed by hematopoietic stem cell transplantation can be used. Specific L-HES therapy is based on anti-IL-5 monoclonal antibodies mepolizumab and alemtuzumab (80,81). In a double blind clinical trial in 85 patients with L-HES (FIP1L1-PDGFR $\alpha$  negative), mepolizumab significantly decreased the clinical activity of HES and was corticosteroid-sparing (9).

### Targeted treatment for M-HES

Imatinib mesylate is a tyrosine kinase inhibitor initially introduced for the treatment of chronic myeloid leukemias, and remains the first line treatment in M-HES, leading to dramatic improvement (13). Imatinib target is a kinase resulting from a 800 kb deletion on chromosome 4 cDNA derived from the fusion gene encoded novel protein composed of the kinase domain of platelet-derived growth factor receptor- $\alpha$  (PDGFR $\alpha$ ) linked to the FIP1 gene. This genetic rearrangement was found in the eosinophilic cell line EOL-1 (peripheral blood, leukemia, acute myeloid, DSMZ ACC 386), and was also detected in blood cells from patients with HES (14). In an animal model (Ba/F3 mouse, peripheral blood, pro B cell, DSMZ ACC 300), cells expressing FIP1L1-PDGFR $\alpha$  were 100-times more sensitive to imatinib than cells expressing BCR-ABL (13). In chronic eosinophilic leukemia (n=23) and hypereosinophilic syndrome (n=13), imatinib mesylate provided complete molecular remission in 87% of cases at 12 months (82).

Other drugs used in the treatment of M-HES are hydroxyurea and IFN- $\alpha$ . If signs of malignant

transformation are found, chemotherapy or hematopoietic stem cell transplantation must be proposed.

## CONCLUSIONS

The field of "idiopathic hypereosinophilic syndromes" has been consistently reduced, due to better understanding of the eosinophil biology. Hypereosinophilic syndrome therapy has been dramatically changed by anti-tyrosine kinase derivatives (imatinib) indicated in myeloproliferative forms of HES. The fusion gene FIP1L1-PDGFR $\alpha$  associated with the kinase target of imatinib has been recognized as a biological marker in patients that may benefit from this targeted therapy. In the L-HES variant (FIP1L1-PDGFR $\alpha$  negative patients), anti-IL-5 therapy (mepolizumab, alemtuzumab) has been proved to be active and corticosteroid-sparing. The early identification of the two HES variants is therefore crucial for the rapid introduction of early and appropriately adjusted therapy.

## References

1. Roufosse F, Cogan E. Practical approach to hypereosinophilia. *Rev Med Brux* 2008;29:400-8.
2. Leiferman KM, Gleich GJ. Hypereosinophilic syndrome: case presentation and update. *J Allergy Clin Immunol* 2004;113:50-8.
3. Hardy WR, Anderson RE. The hypereosinophilic syndromes. *Ann Intern Med* 1968;68:1220-9.
4. Cogan E, Schandene L, Crusiaux A, Cochaux P, Velu T, Goldman M. Brief report: clonal proliferation of type 2 helper T cells in a man with the hypereosinophilic syndrome. *N Engl J Med* 1994;330:535-8.
5. Roufosse F, Schandene L, Sibille C, Willard-Gallo K, Kennes B, Eflora A, *et al.* Clonal Th2 lymphocytes in patients with the idiopathic hypereosinophilic syndrome. *Br J Haematol* 2000;109:540-8.
6. Simon HU, Plotz SG, Dummer R, Blaser K. Abnormal clones of T cells producing interleukin-5 in idiopathic eosinophilia. *N Engl J Med* 1999;341:1112-20.
7. Chusid MJ, Dale DC, West BC, Wolff SM. The hypereosinophilic syndrome: analysis of fourteen cases with review of the literature. *Medicine (Baltimore)* 1975;54:1-27.



8. Fauci AS, Harley JB, Roberts WC, Ferrans VJ, Gralnick HR, Bjornson BH. NIH Conference. The idiopathic hypereosinophilic syndrome. Clinical, pathophysiologic, and therapeutic considerations. *Ann Intern Med* 1982;97:78-92.
9. Rothenberg ME, Klion AD, Roufousse FE, Kahn JE, Weller PF, Simon HU, *et al.* Treatment of patients with the hypereosinophilic syndrome with mepolizumab. *N Engl J Med* 2008;358:1215-28.
10. Druker BJ, Talpaz M, Resta DJ, Peng B, Buchdunger E, Ford JM, *et al.* Efficacy and safety of a specific inhibitor of the BCR-ABL tyrosine kinase in chronic myeloid leukemia. *N Engl J Med* 2001;344:1031-7.
11. Sheikh J, Weller PF. Advances in diagnosis and treatment of eosinophilia. *Curr Opin Hematol* 2009;16:3-8.
12. Cools J, DeAngelo DJ, Gotlib J, Stover EH, Legare RD, Cortes J, *et al.* A tyrosine kinase created by fusion of the PDGFRA and FIP1L1 genes as a therapeutic target of imatinib in idiopathic hypereosinophilic syndrome. *N Engl J Med* 2003;348:1201-14.
13. Gleich GJ, Leiferman KM, Pardanani A, Tefferi A, Butterfield JH. Treatment of hypereosinophilic syndrome with imatinib mesylate. *Lancet* 2002;359:1577-8.
14. Griffin JH, Leung J, Bruner RJ, Caligiuri MA, Briesewitz R. Discovery of a fusion kinase in EOL-1 cells and idiopathic hypereosinophilic syndrome. *Proc Natl Acad Sci U S A* 2003;100:7830-5.
15. Pardanani A, Reeder T, Porrata LF, Li CY, Tazelaar HD, Baxter EJ, *et al.* Imatinib therapy for hypereosinophilic syndrome and other eosinophilic disorders. *Blood* 2003;101:3391-7.
16. Wilkins HJ, Crane MM, Copeland K, Williams WV. Hypereosinophilic syndrome: an update. *Am J Hematol* 2005;80:148-57.
17. Ballanger F, Barbarot S, Hamidou M. Syndromes hypereosinophiliques primitifs: actualites. *Ann Dermatol* 2006;133:1487-94.
18. Gleich GJ, Leiferman KM. The hypereosinophilic syndromes: still more heterogeneity. *Curr Opin Immunol* 2005;17:679-84.
19. Kazmierowski JA, Chusid MJ, Parrillo JE, Fauci AS, Wolff SM. Dermatologic manifestations of the hypereosinophilic syndrome. *Arch Dermatol* 1978;114:531-5.
20. Leiferman KM, O'Duffy JD, Perry HO, Greipp PR, Giuliani ER, Gleich GJ. Recurrent incapacitating mucosal ulcerations. A prodrome of the hypereosinophilic syndrome. *JAMA* 1982;247:1018-20.
21. Aractingi S, Janin A, Zini JM, Gauthier MS, Chauvenet L, Tobelem G, *et al.* Specific mucosal erosions in hypereosinophilic syndrome. Evidence for eosinophil protein deposition. *Arch Dermatol* 1996;132:535-41.
22. Barouky R, Bencharif L, Badet F, Salles G, Vital Durand D, Rousset H. Mucosal ulcerations revealing primitive hypereosinophilic syndrome. *Eur J Dermatol* 2003;13:207-8.
23. Ravoet M, Sibille C, Roufousse F, Duveillier H, Sotiriou C, Schandené L, *et al.* 6q- Is an early and persistent chromosomal aberration in CD3-CD4+ T-cell clones associated with the lymphocytic variant of hypereosinophilic syndrome. *Haematologica* 2005;90:753-65.
24. Gotlib J. Molecular classification and pathogenesis of eosinophilic disorders: 2005 update. *Acta Haematol* 2005;114:7-25.
25. Roufousse FE, Goldman M, Cogan E. Hypereosinophilic syndromes. *Orphanet J Rare Dis* 2007;11:2:37.
26. Ionescu MA, Murata H, Janin A. Oral mucosa lesions in hypereosinophilic syndrome – an update. *Oral Dis* 2008;14:115-22.
27. Klion AD, Noel P, Akin C, Law MA, Gilliland DG, Cools J, *et al.* Elevated serum tryptase levels identify a subset of patients with a myeloproliferative variant of idiopathic hypereosinophilic syndrome associated with tissue fibrosis, poor prognosis, and imatinib responsiveness. *Blood* 2003;101:4660-6.
28. Roufousse F, Cogan E, Goldman M. The hypereosinophilic syndrome revisited. *Annu Rev Med* 2003;54:169-84.
29. Hofmann SC, Technau K, Muller AMS, Lubbert M, Bruckner-Tuderman L. Bullous pemphigoid associated with hypereosinophilic syndrome: simultaneous response to imatinib. *J Am Acad Dermatol* 2007;56(5 Suppl):S68-72.
30. Letsinger JA, McCarty MA, Jorizzo JL. Complex aphthosis: a large case series with evaluation algorithm and therapeutic ladder from topicals to thalidomide. *J Am Acad Dermatol* 2005;52:500-8.
31. Desreumaux P, Janin A, Dubucquoi S, Copin MC, Torpier G, Capron A, *et al.* Synthesis of interleukin-5 by activated eosinophils in pa-

- tients with eosinophilic heart diseases. *Blood* 1993;82:1553-60.
32. Alter P, Maisch B. Endomyocardial fibrosis in Churg-Strauss syndrome assessed by cardiac magnetic resonance imaging. *Int J Cardiol* 2006;108:112-3.
  33. Mabilat-Pragnon C, Janin A, Michel L, Thomaidis A, Legrand Y, Soria C, *et al.* Urokinase localization and activity in isolated eosinophils. *Thromb Res* 1997;88:373-9.
  34. Daneshpouy M, Socie G, Lemann M, Rivet J, Gluckman E, Janin A. Activated eosinophils in upper gastrointestinal tract of patients with graft-versus-host disease. *Blood* 2002;99:3033-40.
  35. Janin A, Copin MC, Dubos JP, Rouland V, Delaporte E, Blanchet-Bardon C. Familial peeling skin syndrome with eosinophilia: clinical, histologic, and ultrastructural study of three cases. *Arch Pathol Lab Med* 1996;120:662-5.
  36. Colombel JF, Janin A, Torpier G. Activated eosinophils in coeliac disease. *Gut* 1990;31:583-4.
  37. Janin A, Socie G, Devergie A, Aractingi S, Esperou H, Vérola O, *et al.* Fasciitis in chronic graft-versus-host disease. A clinicopathologic study of 14 cases. *Ann Intern Med* 1994;120:993-8.
  38. Janin A, Torpier G, Capron M, Courtin P, Goselin B. Immunopathological study of eosinophils in eosinophilic granuloma of bone: evidence for release of three cationic proteins and subsequent uptake in macrophages. *Virchows Arch A Pathol Anat Histopathol* 1992;421:255-61.
  39. Janin A, Torpier G, Courtin P, Capron M, Prin L, Tonnel AB, *et al.* Segregation of eosinophil proteins in alveolar macrophage compartments in chronic eosinophilic pneumonia. *Thorax* 1993;48:57-62.
  40. Dubucquoi S, Janin A, Desreumaux P, Rigot JM, Copin MC, François M, *et al.* Evidence for eosinophil activation in eosinophilic cystitis. *Eur Urol* 1994;25:254-8.
  41. Jong EC, Mahmoud AA, Klebanoff SJ. Peroxidase mediated toxicity to schistosomula of *Schistosoma mansoni*. *J Immunol* 1981;126:468-71.
  42. Gosset P, Tillie-Leblond I, Janin A, Marquette CH, Copin MC, Wallaert B, *et al.* Expression of E-selectin, ICAM-1 and VCAM-1 on bronchial biopsies from allergic and non-allergic asthmatic patients. *Int Arch Allergy Immunol* 1995;106:69-77.
  43. Tsicopoulos A, Janin A, Akoum H, Lamblin C, Vorng H, Hamid Q, *et al.* Cytokine profile in minor salivary glands from patients with bronchial asthma. *J Allergy Clin Immunol* 2000;106:687-96.
  44. Desreumaux P, Bloget F, Seguy D, Capron M, Cortot A, Colombel JF, *et al.* Interleukin 3, granulocyte-macrophage colony-stimulating factor, and interleukin 5 in eosinophilic gastroenteritis. *Gastroenterology* 1996;110:768-74.
  45. Desreumaux P, Janin A, Delaporte E, Dubucquoi S, Piette F, Cortot A, *et al.* Parallel interleukin 5 synthesis by eosinophils in duodenal and skin lesions of a patient with dermatitis herpetiformis. *Gut* 1995;37:132-5.
  46. Daneshpouy M, Facon T, Jouet JP, Janin A. Acute flare-up of conjunctival graft-versus-host disease with eosinophil infiltration in a patient with chronic graft-versus-host disease. *Leuk Lymphoma* 2002;43:445-6.
  47. Tillie-Leblond I, Gosset P, Janin A, Salez F, Prin L, Tonnel AB. Increased interleukin-6 production during the acute phase of the syndrome of episodic angioedema and hypereosinophilia. *Clin Exp Allergy* 1998;28:491-6.
  48. Vandezande LM, Wallaert B, Desreumaux P, Tsicopoulos A, Lamblin C, Tonnel AB, Janin A. Interleukin-5 immunoreactivity and mRNA expression in gut mucosa from patients with food allergy. *Clin Exp Allergy* 1999;29:652-9.
  49. Desreumaux P, Janin A, Colombel JF, Prin L, Plumas J, Emilie D, *et al.* Interleukin 5 messenger RNA expression by eosinophils in the intestinal mucosa of patients with coeliac disease. *J Exp Med*. 1992;175:293-6.
  50. Dubucquoi S, Janin A, Klein O, Desreumaux P, Quandalle P, Cortot A, *et al.* Activated eosinophils and interleukin 5 expression in early recurrence of Crohn's disease. *Gut* 1995;37:242-6.
  51. Launay D, Delaporte E, Gillot JM, Janin A, Hachulla E. An unusual cause of vascular purpura: recurrent cutaneous eosinophilic necrotizing vasculitis. *Acta Derm Venereol* 2000;80:394-5.
  52. Leiferman KM. A current perspective on the role of eosinophils in dermatologic diseases. *J Am Acad Dermatol* 1991;24:1101-12.
  53. Weller PF. Eosinophils: structure and functions. *Curr Opin Immunol* 1994;6:85-90.

54. Janin A. Eosinophilic myocarditis and fibrosis. *Hum Pathol* 2005;36:592-3.
55. Sanderson CJ. Interleukin-5, eosinophils, and disease. *Blood* 1992;79:3101-9.
56. Mishra A, Hogan SP, Lee JJ, Foster PS, Rothenberg ME. Fundamental signals that regulate eosinophil homing to the gastrointestinal tract. *J Clin Invest* 1999;103:1719-27.
57. Rothenberg ME, Hogan SP. The eosinophil. *Annu Rev Immunol* 2006;24:147-74.
58. Wardlaw AJ. Molecular basis for selective eosinophil trafficking in asthma: a multistep paradigm. *J Allergy Clin Immunol* 1999;104:917-26.
59. Moqbel R, Levi-Schaffer F, Kay AB. Cytokine generation by eosinophils. *J Allergy Clin Immunol* 1994;94:1183-8.
60. Rothenberg ME. Eosinophilia. *N Engl J Med* 1998;338:1592-600.
61. Collins PD, Marleau S, Griffiths-Johnson DA, Jose PJ, Williams TJ. Cooperation between interleukin-5 and the chemokine eotaxin to induce eosinophil accumulation *in vivo*. *J Exp Med* 1995;182:1169-74.
62. Matthews AN, Friend DS, Zimmermann N, Sarafi MN, Luster AD, Pearlman E, *et al.* Eotaxin is required for the baseline level of tissue eosinophils. *Proc Natl Acad Sci U S A* 1998;95:6273-8.
63. Clutterbuck EJ, Sanderson CJ. Regulation of human eosinophil precursor production by cytokines: a comparison of recombinant human interleukin-1 (rhIL-1), rhIL-3, rhIL-5, rhIL-6, and rh granulocyte-macrophage colony-stimulating factor. *Blood* 1990;75:1774-9.
64. Rothenberg ME, MacLean JA, Pearlman E, Luster AD, Leder P. Targeted disruption of the chemokine eotaxin partially reduces antigen-induced tissue eosinophilia. *J Exp Med* 1997;185:785-90.
65. Zimmermann N, Hershey GK, Foster PS, Rothenberg ME. Chemokines in asthma: cooperative interaction between chemokines and IL-13. *J Allergy Clin Immunol* 2003;111:227-42.
66. Broide DH, Paine MM, Firestein GS. Eosinophils express interleukin 5 and granulocyte macrophage-colony stimulating factor mRNA at sites of allergic inflammation in asthmatics. *J Clin Invest* 1992;90:1414-24.
67. Lorentz A, Schwengberg S, Mierke C, Manns MP, Bischoff SC. Human intestinal mast cells produce IL-5 *in vitro* upon IgE receptor cross-linking and *in vivo* in the course of intestinal inflammatory disease. *Eur J Immunol* 1999;29:1496-503.
68. Phillips C, Coward WR, Pritchard DI, Hewitt CR. Basophils express a type 2 cytokine profile on exposure to proteases from helminths and house dust mites. *J Leukoc Biol* 2003;73:165-71.
69. Till S, Dickason R, Huston D, Humbert M, Robinson D, Larche M, *et al.* IL-5 secretion by allergen-stimulated CD4+ T cells in primary culture: relationship to expression of allergic disease. *J Allergy Clin Immunol* 1997;99:563-9.
70. Ionescu MA, Rivet J, Daneshpouy M, Briere J, Morel P, Janin A. *In situ* eosinophil activation in 26 primary cutaneous T-cell lymphomas with blood eosinophilia. *J Am Acad Dermatol* 2005;52:32-9.
71. Del Prete G, Tiri A, Maggi E, De Carli M, Macchia D, Parronchi P, *et al.* Defective *in vitro* production of gamma-interferon and tumor necrosis factor-alpha by circulating T cells from patients with the hyper-immunoglobulin E syndrome. *J Clin Invest* 1989;84:1830-5.
72. Genton C, Wang Y, Izui S, Malissen B, Delsol G, Fournié GJ, *et al.* The Th2 lymphoproliferation developing in LatY136F mutant mice triggers polyclonal B cell activation and systemic autoimmunity. *J Immunol* 2006;177:2285-93.
73. Kiehl P, Falkenberg K, Vogelbruch M, Kapp A. Tissue eosinophilia in acute and chronic atopic dermatitis: a morphometric approach using quantitative image analysis of immunostaining. *Br J Dermatol* 2001;145:720-9.
74. Kitano K, Ichikawa N, Shimodaira S, Ito T, Ishida F, Kiyosawa K. Eosinophilia associated with clonal T-cell proliferation. *Leuk Lymphoma* 1997;27:335-42.
75. Desreumaux P, Delaporte E, Colombel JF, Capron M, Cortot A, Janin A. Similar IL-5, IL-3, and GM-CSF syntheses by eosinophils in the jejunal mucosa of patients with celiac disease and dermatitis herpetiformis. *Clin Immunol Immunopathol* 1998;88:14-21.
76. Pazdrak K, Olszewska-Pazdrak B, Stafford S, Garofalo RP, Alam R. Lyn, Jak2, and Raf-1 kinases are critical for the antiapoptotic effect of interleukin 5, whereas only Raf-1 kinase is essential for eosinophil activation and degra-

- nulation. *J Exp Med* 1998;188:421-9.
77. Bain BJ. Hypereosinophilia. *Curr Opin Hematol* 2000;7:21-5.
78. Gotlib J, Cools J, Malone JM III, Schrier SL, Gilliland DG, Coutre SE. The FIP1L1-PDGFR alpha fusion tyrosine kinase in hypereosinophilic syndrome and chronic eosinophilic leukemia: implications for diagnosis, classification, and management. *Blood* 2004;103:2879-91.
79. Keung YK, Beaty M, Steward W, Jackle B, Pettinati M. Chronic myelocytic leukemia with eosinophilia, t(9;12)(q34;p13), and ETV6-ABL gene rearrangement: case report and review of the literature. *Cancer Genet Cytogenet* 2002;138:139-42.
80. Garrett JK, Jameson SC, Thomson B, Collins MH, Wagoner LE, Freese DK, *et al.* Antiinterleukin-5 (mepolizumab) therapy for hypereosinophilic syndromes. *J Allergy Clin Immunol* 2004;113:115-9.
81. Sefcick A, Sowter D, DasGupta E, Russell NH, Byrne JL. Alemtuzumab therapy for refractory idiopathic hypereosinophilic syndrome. *Br J Haematol* 2004;124:558-9.
82. Metzgeroth G, Walz C, Erben P, Popp H, Schmitt-Graeff A, Haferlach C, *et al.* Safety and efficacy of imatinib in chronic eosinophilic leukaemia and hypereosinophilic syndrome: a phase-II study. *Br J Haematol* 2008;143:707-15.



Luxury box for Pyramidon tablets. For free!; year 1934.  
(From the collection of Mr. Zlatko Puntijar)