



# Hedgehog signaling and cross-talk therapeutic potential

## SONJA LEVANAT

Laboratory for Hereditary Cancer  
Division of Molecular Medicine  
Ruđer Bošković Institute, Zagreb, Croatia  
E-mail: sonja.levanat@irb.hr

## INTRODUCTION

Normal and tumor cells use many pathways for survival, proliferation and communication with environment. Many defects during lifetime occur and consequently many other sets of alternative pathways switch on, in a fighting for survival, repair and reproduction. In such dynamic processes depending on a moment, environment, conditions and events, cells learn how to survive. To find effective tools to attack particular sets of those pathways in particular cells is main strategic road to treat and fight with cancer.

Accumulating wide lines of experimental evidence revealed that aberrant activation of Hedgehog–Gli (Hh–Gli) pathway and pathways involving receptor tyrosine kinases (RTK), such as the EGF signaling, frequently occur during cancer initiation and progression, and these tumorigenic cascades may cooperate through multiple signaling cross-talks to the malignant transformation of cells, treatment resistance and disease relapse.

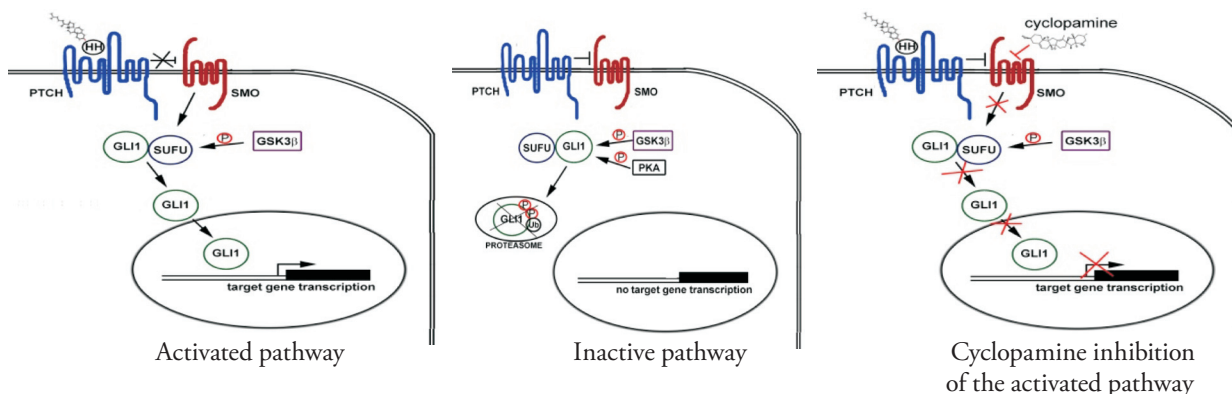
In this context, the most relevant issue for clinical application is: How to attack molecular mechanisms and specific downstream signaling elements that may contribute to the cooperative or synergistic interactions of the Hh–Gli and RTK signaling pathways, including EGFR, in cancer and metastasis-initiating cells?

Moreover, it is of great therapeutic interest to define drug resistance-associated molecules, including ABC transporters modulated through the inhibition of Hh and/or EGFR pathways, that could be targeted for reversing the chemoresistance of cancer and metastasis-initiating cells.

In view of the promising results from preclinical studies, targeting the Hh cascade seems to represent a therapeutic strategy of great clinical potential.

## Misregulation of molecular signaling pathways

Misregulation of molecular signaling pathways that control fundamental cellular processes such as growth and cell death has been directly associated with a variety of inherited and sporadic diseases. Targeting such pathways, as is the Hedgehog (Hh) signaling pathway represents a promising new paradigm for drug discovery. Cyclopamine, plant-derived steroidal alkaloid, was the first discovered inhibitor for this pathway, shown to bind to the heptahelical transmembrane part of Smo, inhibiting its activity.



**Figure 1.** Schematic presentation of main steps in Hh-Gli signaling pathway. Pathway is activated when the ligand Hh binds to 12-transmembrane receptor Ptch (Activated Pathway), or it is inactive when the ligand is not present (Inactive pathway); and the activated pathway can be blocked by cyclopamine inhibition (Cyclopamine inhibition of the activated pathway). The interactions of the components of the Hh-Gli pathway occur in the primary cilia of cells (10). Functional Hh protein is generated in a two-step process that involves autocatalytic cleavage of a precursor molecule to release a cholesterol-modified N-terminal signaling domain, followed by addition of palmitate to the N terminus. This protein is then secreted from the membranes of the producing cells and initiates the Hh signaling cascade upon binding to the Patched (Ptch) (11). In the absence of the Hh ligand, the Ptch receptor inhibits the activity of the downstream co-receptor Smoothed (Smo), which in general topology resembles G-protein-coupled receptors (GPCRs). It is proposed that Ptch effects this inhibition by excluding the ciliary localization of Smo (12). Binding of Hh protein to Ptch causes Smo, stored in intracellular vesicles, to move to the cilium and activate signal transduction. Active Smo then signals via a cytosolic complex of proteins including Suppressor of Fused (SuFu), and the cascade culminates by triggering activation of the glioma (Gli) family of transcription factors and their translocation to the nucleus. This activation results in the expression of specific genes that promote cell proliferation and differentiation.

After that, small molecule Hh-Gli regulators have become very rapidly expanding field. They come in varieties depending on their source (synthetic versus natural products), as well as on the locus of action: those that inhibit SMO versus those that act downstream of SMO, including those that may block GLI function directly. Blocking antibodies, peptides and small RNA inhibitors are more recent aspects of new targeting tools.

### The Hedgehog-Gli (Hh-Gli) signaling pathway

The Hh-Gli signaling pathway is a developmental pathway, which is often found aberrantly active in various tumors. The Hh pathway is a key regulator of patterning, growth, and cell migration during embryonic development (1, 2), and inhibition of the Hh pathway at this stage has been shown to cause severe birth defects such as cyclopia (3). In adult organisms, the Hh pathway contributes to homeostasis and regeneration of certain tissues such as skin and bone, it is active almost exclusively in somatic stem cells, but aberrant activation of the Hh pathway has been linked to tumorigenesis in various and severe types of cancers (4). The fact that it is frequently activated in cancer stem cells makes it an interesting target for future therapies.

Hedgehog (Hh) signaling pathway was first discovered in *Drosophila* in early 1980s (5). The pathway's name originates from the observations that mutations in the

gene encoding the secreted protein, one of the key regulators of the pathway in fruit flies, give rise to an unusual spiky-haired phenotype. In mammals, the proteins are Sonic hedgehog (Shh), named after the popular video game hero, Indian hedgehog (Ihh) and Desert hedgehog (Dhh), the latter two named after existing species of living hedgehogs. Sonic hedgehog (Shh) is the most widely characterized of the three vertebrate Hedgehog homologs, and is essential for proper embryonic development.

The pathway activation begins when the secreted Shh protein binds to its receptor, Patched (Ptch1), a twelve transmembrane protein, resulting in the de-repression of Smoothed (Smo) a seven transmembrane protein, that has a function of co-receptor. This triggers a cascade of events in the cytoplasm leading to activation of the transcription zinc finger factors Gli and transcription of their target genes. Several components of the Hh-Gli pathway (PTCH, GLI1, GLI2 and HHIP) are Gli transcriptional targets that induce positive or negative feedback (6). The Gli proteins are regulated by the Suppressor of Fused (SuFu), Protein Kinase A (PKA), Glycogen Synthase Kinase 3 $\beta$  (GSK3 $\beta$ ) and Casein Kinase 1 (CK1). GLI targets mediate various cellular responses, notably enhanced cell proliferation and survival by upregulating D-type cyclins and antiapoptotic proteins (7, 8, 9).

Many studies have shown that the activity of GLI proteins can be additionally modified by integration of distinct signals, such as the MEK/extracellular signal-regu-

**TABLE 1**

Small molecule Hh-Gli pathway inhibitors and indicative targets in clinical trials (some of the data from Mas and Ruiz i Altaba 2010-(16)).

Compound	Target	Cancer type	Status
GDCO449	SMO	medulloblastoma, glioblastoma, BCC Colorectal cancer, stomach, ovarian, pancreatic	phase II
GDCO449 +gemcitabine	SMO+DNA Replication	metastatic pancreatic	"
BM-833923	SMO	BCC, BCNS, small cell lung	phase I
BM-833923	SMO	small cell lung	"
+carboplatin	DNA alkylation	small cell lung	"
+etoposide	Topo II	small cell lung	"
BM-833923	SMO		
+cisplatin	Topo II	metastatic gastric and esophageal	"
BM-833923	SMO	multiple myeloma	

lated kinase (ERK) and phosphoinositide-3 kinase (PI3K)/AKT pathway, and they have been described as non-canonical Hh-Gli activators in cancer.

However, signaling events immediately downstream of Smo are still not clearly understood. Accumulating evidence from several groups indicates an important but not yet fully defined step: mammalian Smo is during signaling translocated to primary cilia. This was found in most vertebrate cells (10).

The pathway is a highly coordinated and orchestrated network, linking events from ligand binding on the membrane, toward events in cytoplasm and transcription factors Gli. Therefore, it deserved the name Hh-Gli signaling pathway, today in predominant use.

### Hh-Gli pathway inhibition

The first small-molecule inhibitor of the Hh-Gli pathway, natural product alkaloid cyclopamine, achieves inhibition by direct binding to the seven-transmembrane alpha-helical bundle of the Smoothed, the co-receptor Smo (13). The majority of Hh-Gli pathway inhibitors target Smo, and this has led to the identification and development of many other Smo antagonists and derivatives of cyclopamine. Since a number of cancer cells have been found insensitive to Smo inhibition, because of the mostly acquired resistance to Smo antagonists through mutations in SMO that prevent binding of the antagonist (14, 15), there was a need to target downstream effectors. Today there is a long list of small molecule inhibitors of the Hh-Gli pathway, acting from the level of attacking the ligand (any of Hh varieties), or from the Smo level, or acting on downstream targets in cytoplasm. Some of them are promising and are in clinical trials (table 1). Also, there is a long list of many new potential antagonists and agonists of the pathway, some of them listed in table 2.

**TABLE 2**

Some of known Hh-Gli antagonists and agonists.

Antagonists	Target	References
cyclopamine	Smo	17, 18, 19
KAAD-cyclopamine	Smo	20
Robotnikinin	Shh	21
SANT1,2,3,4	Smo	22
SANT74, SANT75	Smo	23
Cur-61414	Smo	24
GANT58	Gli	25
GANT61	Gli	25
Hh-Ag	Smo	26
SAG	Smo	22

Even though GLI1 is a transcription factor and thus a priori a bad target, it is a rather unusual factor with multiple lives in different cellular compartments (25). GLI1, and the other GLIs, are exquisitely regulated at different levels, including phosphorylation, acylation, sequestration and degradation (27–31). Each of these steps, as well as the partners that physically interact with the Gli proteins, provides possible sites for small molecule action. Therefore, Gli1 is not only a valid target but so far it is also the only reliable and general marker of a cell's response to Hh signaling. Measuring GLI1 levels in relevant human cells is thus a requisite (32).

### Inhibitors of the pathway

Because of its accessibility on the membrane and its importance in regulation of the pathway, SMO has been the primary focus in the development of small-molecule inhibitors of the Hh-Gli pathway. GDC-0449 (vismodegib; Genentech) is an orally administered agent that selectively suppresses SMO activity and was the first SMO

inhibitor to progress to clinical trials. It has produced promising antitumor responses in patients with advanced basal cell carcinoma and medulloblastoma (33, 34), but resistance has been reported (15, 35). The resistance to SMO inhibitors highlights the therapeutic need to target downstream effectors. So, the small molecule GANT61 was identified as a specific inhibitor of GLI1 and GLI2. It suppresses the DNA-binding capacity of GLIs and inhibits GLI-mediated transcription. GANT61 reduces proliferation and induces apoptosis in a GLI-specific fashion in prostate cancer (25), colon carcinoma (36, 37), oral squamous cell carcinoma (38), pancreatic cancer (39), neuroblastoma (40), and chronic lymphocytic leukemia (41). However, today it is generally recognized that this inhibitor is not really specific inhibitor for Gli, and unfortunately the Hh-Gli pathway has no specific inhibitors created yet.

### MicroRNA regulation

MicroRNAs (miRNA), small RNA molecules which bind to regulatory elements in the mRNA molecules and control their stability, are crucial post-transcriptional regulators of gene expression, cell differentiation and proliferation. They are involved in normal cell development and in development of various types of tumors. The role of miRNA in regulation of Hh-Gli signaling pathway has been suggested using screening approaches and bioinformatics, and a direct link between these two mechanisms has been investigated in various cancers. Downregulation or even misregulation of specific miRNAs allows high levels of Hh-dependent gene expression leading to tumor cell proliferation, sustaining cancer development (42). Specific miRNAs involved in the regulation of the Hh signalling (miR-125b, miR-324-5p and miR-326), downregulated in medulloblastoma, target the activator components of the pathway, Smo and Gli1, thereby suppressing tumor cell growth. This was the first discovered mechanism of regulation of Hh signaling through miRNA-mediated control of Smo and Gli1 and of involvement of miRNA-mediated control of the Hh pathway in malignancy. The concept is still under research, particularly for severe types of cancer (43), and may have promising implications for miRNA based therapies (44).

### Link between developmental biology and cancer

First discoveries related to the Hh-Gli pathway and human disorders were made on a range of PTCH1 alteration profiles, including genetic mutation, LOH, and promoter hypermethylation, and the two-hit theory was explored to dissect all possible genetic and epigenetic mechanisms (45–49).

At this level, the key player in the pathway is PTCH1. Inactivation of PTCH1 allows hedgehog ligand-independent activation of SMO, causing a downstream activation

of the pathway that may lead to neoplastic growth. Mutations in the PTCH (PTCH1) gene are the underlying cause of nevoid basal cell carcinoma syndrome (NBCCS) or Gorlin syndrome. And thanks to this syndrome, tumor suppressor PTCH was cloned, its role in development and cancer was unraveled. Cancers driven by mutations within the Hh signaling, mostly BCC and tumors described within Gorlin syndrome (50, 51), had not been in wider focus, until aberrant activation of the pathway and its inhibitors (i.e. the natural alkaloid, cyclopamine) were described. After that, various studies through *in vitro* and *in vivo* models explored and attempted to explain mechanisms of ligand-dependent, ligand-independent, autocrine, canonical and non-canonical Hh-Gli pathway activation in multiple tumors.

### MECHANISMS OF HH-GLI SIGNALING PATHWAY DEREGLATION THAT MAY LEAD TO CANCER DEVELOPMENT

The first and widely described, ligand-independent mechanism, usually involves mutations in which loss of PTCH1 or its functionality leads to loss of suppression, whereas SMO mutations create a constitutively active form of the protein (20, 52, 53, 54).

But it was also shown that PTCH1 function can be lost through methylation (48, 55, 56).

Amplifications or mutations of genes downstream of Ptc contribute to activation of the Hh-Gli pathway, e.g. high amplification of Gli1 was reported in glioblastomas (6).

Another under widely explored, ligand-dependent mechanism, can be achieved through ligand hyperproduction or by downstream activation processes.

Ligand-dependent Hh-Gli signaling has been reported in different stages of carcinogenesis in different tumors: pancreatic cancer, lung cancer, esophageal cancer, prostate cancer, breast cancer, gastric cancer, colon cancer, ovarian cancer and hepatocellular cancer (58–66), suggesting that Hh-Gli signaling has significant role in carcinogenesis of these tumors.

In support of these findings, transgenic mice with pancreatic-specific expression of SHH or GLI2 develop pancreatic tumors (59, 67). In some other tumors (gastric, prostate cancer) Hh signaling activation is associated with cancer progression, and consistent with these findings, inhibition of Hh signaling in prostate and gastric cancer cells reduces cell invasiveness (63, 68). Also, it was published that Hh signaling is required for development and progression of melanoma, gliomas, breast cancer, ovarian cancer, leukemia and B-cell lymphomas (69, 70).

In addition, the modes of Hedgehog signaling in cancer development may be variable. Activated Hh-Gli signaling can act in an autocrine or paracrine manner. In the

autocrine manner Hh is produced by the cancer cells themselves. In the paracrine manner (various studies in pancreatic, lung, esophageal cancer) stromal tumor cells are included in receiving signals. Even more, it was demonstrated that tumor-infiltrating monocytes or macrophages secrete ligand Shh, that activates Hh-Gli pathway in cancer cells (71, 72, 73). Shh or Ihh ligands secreted by the tumor cells activate Hh signaling in the stromal cells (74). It is also evident from studies of Dierks et al 2009 (74) and Zhao et al 2009 (75) that Hh signaling is required for maintenance of cancer stem cell population.

In our research we have observed hyperproduction of the Shh ligand by tumor cells in ovarian cancer, that lead to cell proliferation, as an example of the autocrine activation (76). These results indicate that in ovarian tumors pathogenesis through SHH gene expression differs in borderline tumors and carcinoma. Also, it was shown by others that the Hh-Gli signalling pathway plays an important role in ovarian tumorigenesis as well as in the activation of cell proliferation, thus could be as molecular target of new treatment strategies for ovarian carcinoma (66).

On the other hand, in breast cancer we observed a cross-talk between Hh-Gli signaling (Shh ligand) and estrogen receptors creating an autoregulatory loop (77, 78). Furthermore, in colon cancer we observed hyperactivation of the regulatory kinase GSK3 $\beta$  that leads to overproduction of activator form of Gli3 and to the pathway hyperactivation (79). This suggests a major role for the interplay of GSK3 and Gli3 in the regulation of this pathway in colon cancer (publication in preparation).

Such examples from recent research of our group and many others document various ways of Hh-Gli signaling activation in many types of cancer, indicating different tumors have different modes of interaction with the pathway. Therefore, this pathway might indeed be a suitable target for cancer therapy.

## CANCER THERAPY TARGETING HH SIGNALING

Today it is generally recognized that Hh-Gli signalling pathway is activated in various types of cancer and at various levels, and contributes to cancer proliferation, progression and invasiveness, so this pathway is anticipated to provide a new avenue for cancer therapy.

There are probably more than hundred compounds disclosed to have inhibitory effects on Hh signalling. Some are under clinical trials. Hh-signaling inhibitors are mainly targeting three sites in Hh-Gli pathway: Ligand Hh (by neutralizing antibodies, Robotnikinin), Smo protein (cyclopamine and its derivatives) and Gli inhibitors. Several Smo inhibitors have been proposed as potential candidates for cancer therapy either as a single agent or in combination regimens with conventional chemotherapy. Most pathway inhibitors can be divided into three groups:

natural products (cyclopamine), novel synthetic compounds and Hh-signaling modulators.

Cyclopamine is Hh-Gli pathway inhibitor on the level of direct interaction with Smo (3). Some derivatives of cyclopamine differing in solubility (IPI-926) or in structure (GDC-0499, LDE225, BMS-833923), or inhibitors of the transformation of inactive Smo into active Smo (SANT 74-75), and more others have been developed, and some are in clinical use (80, 81, 82). Most drug development programs and recent clinical trials are focused on Smo inhibitors.

However, it was also shown that on the level of Gli-mediated transcription, which constitutes the final step in the pathway, some tumors could be selectively inhibited (GANT58 and GANT 61) (25). Another recently identified Gli inhibitor, Gli-antagonist, is arsenic trioxide (ATO), which FDA approved as a drug for the treatment of acute promyelocytic leukemia. ATO binds directly to Gli1 inhibiting its transcriptional activity and suppressing tumor growth *in vitro* and *in vivo* (83, 84).

Rapid advancement in the discovery of novel Hh signaling inhibitors has provided many opportunities for developing novel cancer therapeutic strategies. It is not surprising to learn that several major challenges still exist to prevent the use of Hh signaling inhibitors in clinics. These challenges include a lack of basic understanding of the molecular mechanisms by which Hh signaling mediates carcinogenesis; no clear criteria to identify the right tumors for therapeutic application; only a few reliable, physiologically relevant, and reproducible mouse models for cancer metastases to test and optimize drug dosages in order to minimize side effects; and a lack of clear strategies to mitigate drug resistance. Over the last years, research in this area has greatly improved. It is anticipated that additional novel therapeutic strategies will be developed for cancer clinical trials using Hh signaling inhibitors in the next years.

## INTERACTIONS BETWEEN HH-GLI SIGNALING AND OTHER PATHWAYS OR CROSS-TALK

We may assume that pathways that enable particular cell to survive are interacting among themselves, and that in many cases Hh-Gli signaling pathway is involved.

Examples include regulation of SHH expression by Ras, NF $\kappa$ B and ER $\alpha$ , as well as regulation of Ihh by Msx2 (67, 85). Also, expression of Gli1 is regulated by TGF $\beta$ , Ras and Jun oncoprotein (86, 87). Furthermore, the interaction between PKC and Hh signaling varies depending on PKC isoforms and cell types; although PKC alpha is shown to activate Hh signaling, PKC delta inhibits it (88).

Particularly interesting are the interactions with another developmental pathway, Wnt pathway, which is also often active in some cancers, like colon cancer. Hh and Wnt signaling can form a positive or negative feedback loop depending on tissue content. In gastric cancer Hh signaling can exert negative effects on Wnt signaling through elevated expression of Wnt inhibitor sFRP-1 (89). But in Hh-mediated skin carcinogenesis Wnt signaling is required (90), mostly through beta-catenin expression (91).

Many studies have shown p53 pathway collaboration with Hh pathway in skin carcinogenesis. In melanoma it was shown that p53 negatively regulates Gli1 expression through MDM2 (92); this feedback regulatory loop is required for maintaining stem cell number and cancer cell number.

Synergistic effects with Hh-Gli signaling was shown for some growth factors (IGF-I, VEGF, PDGF alpha, EGF) and their receptors, affecting MEK/ERK/JUN pathway (93). This raises the question whether targeting Hh signaling with inhibitors of the pathway is also a good target for growth factor pathways, and could such strategy make contribution to better treatment of cancer (inhibitors of Hh-Gli pathway and EGF/EGFR, IGF inhibitors etc).

The importance of the Hh-Gli signaling pathway investigations related to its role in cross-talk is underlined by the estimates that the pathway may be active in one third of all cancers. Better understanding of the modes of Hh-Gli pathway regulation and tumor response, as well as of interactions of the pathway with other signaling pathways, has an obvious potential for development of better therapies that would be based on combined effects of the Hh-Gli and other pathways inhibitors.

## REFERENCES

1. INGHAM P W 2001 Hedgehog signaling in animal development: paradigms and principles. *Genes Dev* 15(23): 3059–87
2. INGHAM P W, NAKANO Y, SEGER C 2011 Mechanisms and functions of Hedgehog signalling across the metazoa. *Nat Rev Genet* 12(6): 393–406
3. CHEN J K, TAIPALE J, COOPER M K, BEACHY P A 2002 Inhibition of Hedgehog signaling by direct binding of cyclopamine to Smoothened. *Genes Dev* 16(21): 2743–8
4. BRISCOE J, THÉRON P P 2013 The mechanisms of Hedgehog signalling and its roles in development and disease. *Nat Rev Mol Cell Biol* 14(7): 418–31
5. NÜSSLEIN-VOLHARD C, WIESCHAUS E 1980 Mutations affecting segment number and polarity in *Drosophila*. *Nature* 287(5785): 795–801
6. RUIZ I ALTABA A, MAS C, STECCA B 2007 The Gli code: an information nexus regulating cell fate, stemness and cancer. *Trends Cell Biol* 17: 438–47
7. DUMAN-SCHEEL M, WENG L, XIN S, DU W 2002 Hedgehog regulates cell growth and proliferation by inducing Cyclin D and Cyclin E. *Nature* 417: 299–304
8. BIGELOW R L H, CHARI N S, UND-EN A B, SPURGERS K B, LEE S, ROOP D R *et al.* 2004 Transcriptional regulation of bcl-2 mediated by the sonic hedgehog signaling pathway through gli-1. *J Biol Chem* 279: 1197–205
9. REGL G, KASPER M, SCHNIDAR H, EICHBERGER T, NEILL G W, PHILPOTT M P 2004 *et al.* Activation of the BCL2 promoter in response to Hedgehog/Gli Signal Transduction Is Predominantly Mediated by GLI2. *Cancer Res* 64: 7724–31
10. CORBIT K C, AANSTAD P, SINGLA V, NORMAN A R, STAINIER D Y R, REITER J F 2005 Vertebrate Smoothened functions at the primary cilium. *Nature* 437(7061): 1018–21
11. JOHNSON R L, MILENKOVIC L, SCOTT M P 2000 In Vivo Functions of the Patched Protein: Requirement of the C Terminus for Target Gene Inactivation but Not Hedgehog Sequestration. *Mol Cell* 6(2): 467–78
12. CHEN Y, YUE S, XIE L, PU X, JIN T, CHENG S Y 2011 Dual phosphorylation of Suppressor of Fused (SuFu) by PKA and GSK-3beta regulates its stability and localization in the primary cilium. *J Biol Chem* 286(15): 13502–13511
13. CHEN J K K, BEACHY P A 2002 Inhibition of hedgehog signaling by direct binding of cyclopamine to smoothened. *Genes Dev* 16: 2743–2748
14. METCALFE C, DE SAUVAGE F J 2011 Hedgehog fights back: mechanisms of acquired resistance against Smoothened antagonists. *Cancer Res* 71(15): 5057–61
15. YAUCH R L, DIJKGRAAF G J P, ALICKE B, JANUARIO T, AHN C P, HOLCOMB T 2009 *et al.* Smoothened mutation confers resistance to a Hedgehog pathway inhibitor in medulloblastoma. *Science* 326: 572–4
16. MAS C, RUIZ I ALTABA A 2010 Small molecule modulation of HH-Gli signalling: current leads, trials and tribulations. *Biochem Pharmacol* 80: 712–723
17. COOPER M K, PORTER J A, YOUNG K E, BEACHY P A 1998 Teratogen-mediated inhibition of target tissue response to Shh signaling. *Science* 280(5369): 1603
18. BINNS W, JAMES L F, KEELER R F, BALLS L D 1968 Effects of teratogenic agents in range plants. *Cancer Res* 28: 2323–6
19. INCARDONA J P, GAFFIELD W, KAPUR R P, ROELINK H 1998 The teratogenic veratrum alkaloid cyclopamine inhibits sonic hedgehog signal transduction. *Development* 125: 3553–62
20. TAIPALE CHEN J K, COOPER M K, WANG B, MANN R K, MILENKOVIC L *et al.* 2000 Effects of oncogenic mutations in Smoothened and Patched can be reversed by cyclopamine. *Nature* 406: 1005–9
21. STANTON B Z, PENG L F, MALOOF N, NAKAI K, WANG X, DUFFNER J L *et al.* 2009 A small molecule that binds Hedgehog and blocks its signalling in human cells. *Nat Chem Biol* 5: 154–6
22. CHEN J K, TAIPALE J, YOUNG K E, MAITI T, BEACHY P A 2002 Small molecule modulation of Smoothened activity. *Proc Natl Acad Sci U S A* 99(22): 14071–6
23. YANG H, XIANG J, WANG N, ZHAO Y, HYMAN J, LI S *et al.* 2009 Converse conformational control of smoothened activity by structurally related small molecules. *J Biol Chem* 284: 20876–84

24. WILLIAMS J A, GUICHERIT O M, ZAHARIAN BI, XU Y, CHAI I, WICHTERLE H *et al.* 2003 Identification of a small molecule inhibitor of hedgehog signalling pathway : effects on basal cell carcinoma-like lesions. *Proc Natl Acad Sci USA* 100: 4616–21
25. LAUTH M, BERGSTROM A, SHIMOKAWA T, TOFTGARD R 2007 Inhibition of Gli-mediated transcription and tumor cell growth by small-molecule antagonists. *Proc Natl Acad Sci USA* 104: 8455–60
26. FRANK-KAMENETSKY M, ZHANG X M, BOTTEGA S, GUICHERIT O, WICHTERLE H, DUDEK H *et al.* 2002 Small-molecule modulators of Hedgehog signaling: identification and characterization of Smoothened agonists and antagonists. *J Biol Chem* 277: 10
27. STECCA B, RUIZ I ALTABA A 2009 A GLI1-p53 inhibitory loop controls neural stem cell and tumour cell numbers. *EMBO J* 28: 663–76
28. DI MARCOTULLIO L, FERRETTI E, GRECO A, DE SMAELE E, PO A, SICO M A *et al.* 2006 Numb is a suppressor of Hedgehog signalling and targets Gli1 for Itch-dependent ubiquitination. *Nat Cell Biol* 8(12): 1415–23
29. RIOBÓN A, LUK K, AIX, HAINES G M, EMERSON C P 2006 Phosphoinositide 3-kinase and Akt are essential for Sonic Hedgehog signaling. *Proc Natl Acad Sci U S A* 103(12): 4505
30. CANETTIERI G, DI MARCOTULLIOL, GRECO A, CONI S, ANTONUCCIL, INFANTE P *et al.* 2010 Histone deacetylase and Cullin 3-REN(KCTD11) ubiquitin ligase interplay regulates Hedgehog signaling through Gli acetylation. *Nat Cell Biol* 12: 132–42
31. CHAMOUN Z, MANN R K, NELLEN D, VON KESSLER D P, BELLOTTO M, BEACHY P A *et al.* 2001 Skinny hedgehog, an acyltransferase required for palmitoylation and activity of the hedgehog signal. *Science* 293: 2080–4
32. DAHMANE N, LEE J, ROBINS P, HELLER P, RUIZ I ALTABA A 1997 Activation of the transcription factor Gli1 and the Sonic hedgehog signaling pathway in skin tumours. *Nature* 389: 876–81
33. RUDIN C M, HANN C L, LATERRA J, YAUCH R L, CALLAHAN C A, FU L *et al.* 2009 Treatment of medulloblastoma with Hedgehog pathway inhibitor GDC-0449. *N Engl J Med* 361: 1173–8
34. VON HOFF D D, LORUSSO P M, RUDIN C M, REDDY J C, YAUCH R L, TIBES R *et al.* 2009 Inhibition of the Hedgehog pathway in advanced basal-cell carcinoma. *N Engl J Med* 361: 1164–72
35. DIJKGRAAF G J P, ALICKE B, WEINMANN L, JANUARIO T, WEST K, MODRUSAN Z *et al.* 2011 Small molecule inhibition of GDC-0449 refractory smoothened mutants and downstream mechanisms of drug resistance. *Cancer Res* 71: 435–44
36. MAZUMDAR T, DEVECCHIO J, SHI T, JONES J, AGYEMAN A, HOUGHTON J A 2011 Hedgehog signaling drives cellular survival in human colon carcinoma cells. *Cancer Res* 71: 1092–102
37. MAZUMDAR T, DEVECCHIO J, AGYEMAN A, SHI T, HOUGHTON J A 2011 Blocking Hedgehog survival signaling at the level of the GLI genes induces DNA damage and extensive cell death in human colon carcinoma cells. *Cancer Res* 71: 5904–14
38. YAN M, WANG L, ZUO H, ZHANG Z, CHEN W, MAO L *et al.* 2011 HH/GLI signalling as a new therapeutic target for patients with oral squamous cell carcinoma. *Oral Oncol* 47: 504–9
39. FU J, RODOVA M, ROY S K, SHARMA J, SINGH K P, SRIVASTAVA R K *et al.* 2013 GANT-61 inhibits pancreatic cancer stem cell growth in vitro and in NOD/SCID/IL2R gamma null mice xenograft. *Cancer Lett* 330: 22–32
40. WICKSTROM M, DYBERG C, SHIMOKAWA T, MILOSEVIC J, BARYAWNO N, FUSKEVA G O M *et al.* 2013 Targeting the hedgehog signal transduction pathway at the level of GLI inhibits neuroblastoma cell growth in vitro and in vivo. *Int J Cancer* 132: 1516–24
41. DESCH P, ASSLABER D, KERN D, SCHNIDAR H, MANGELBERGER D, ALINGER B *et al.* 2010 Inhibition of GLI, but not Smoothened, induces apoptosis in chronic lymphocytic leukemia cells. *Oncogene* 29: 4885–95
42. FERRETTI E, DE SMAELE E, MIELE E, LANEVE P, PO A, PELLONI M, PAGANELLI, DI MARCOTULLIO L, CAFFARELLI E, SCREPANTI I, BOZZONI I, GULINO A 2008 Concerted microRNA control of Hedgehog signalling in cerebellar neuronal progenitor and tumour cells. *EMBO Journal* 27: 2616–2627
43. GU W, SHOU J, GU S, SUN B, CHE X 2014 Identifying Hedgehog signaling specific MicroRNAs in glioblastoma. *Int J Med Sci* 11(5): 488/493
44. GONZÁLEZ-GUGEL E, VILLA-MORALES M, SANTOS J, BUENO M J, MALUMBRES M, RODRÍGUEZ-PINILLA S M, PIRIS M A, FERNÁNDEZ-PIQUERAS J 2013 Down-regulation of specific miRNAs enhances the expression of the gene Smoothened and contributes to T-cell lymphoblastic lymphoma development. *Carcinogenesis* doi:10.1093/carcin/bgs404, 2013.
45. HAHN H, WICKING C, ZAPHIROPOULOUS P G, GAILANI M R, SHANLEY S, CHIDAMBARAM A, VORECHOVSKY I, HOLMBERG E, UNDEN A B, GILLIES S, NEGUS K, SMYTH I *et al.* 1996 Mutations of the human homologue of *Drosophila patched* in the nevoid basal cell carcinoma syndrome. *Cell* 85: 841–851
46. SHIMKETS R, GAILANI M, SIU V, YANG-FENG T, PRESSMAN C, LEVANAT S, GOLDSTEIN A, DEAN M, BALE A E 1996 Molecular analysis of chromosome 9q deletions in two Gorlin syndrome patients. *Am J Hum Genet* 59: 417–422
47. LEVANAT S, GORLIN R J, FALLET S, JOHNSON D R, FANTASIA J E, BALE A E 1996 A two-hit model for developmental defects in Gorlin syndrome. *Nature genetics* 12: 85–87
48. CRETNIK M, MUSANI V, ORESKOVIC S, LEOVIC D, LEVANAT S 2007 The Patched gene is epigenetically regulated in ovarian dermoids and fibromas, but not in basocellular carcinomas. *Int J Mol Med* 19: 875–883
49. CAR D, SABOL M, MUSANI V, OZRETIC P, LEVANAT S 2010 Epigenetic regulation of the Hedgehog-Gli signaling pathway in cancer. *Period Biol* 112(4): 419–423
50. GAILANI M R, STAHL-BACKDAHL M, LEFFELL D J, GLYNN M, ZAPHIROPOULOS P G, PRESSMAN C, UNDEN A B, DEAN M, BRASH D E, BALE A E, TOFTGARD R 1996 The role of the human homologue of *drosophila patched* in sporadic basal cell carcinomas. *Nat Genet* 14: 78–81
51. MUSANI V, CRETNIK M, SITUM M, BASTA-JUZBASIC A, LEVANAT S 2009 Gorlin Syndrome patient with large deletion in 9q22.32-q22.33 detected by Quantitative Multiplex Fluorescent PCR. *Dermatology* 219(2): 111–118
52. LEVANAT S, KONČAR MUBRIN M, CRNIĆ I, ŠITUM M, BASTA-JUZBAŠIĆ A 2000 Variable expression of Gorlin syndrome may reflect complexity of signalling pathway. *Pflugers Archiv – European Journal of Physiology* 439: R31–R33

53. LAM C W, XIE J, TO K F, NG H K, LEE K C, YUEN N W, LIM P L, CHAN L Y, TONG S F, MCCORMICK F 1999 A frequent activated smoothened mutation in sporadic basal cell carcinomas. *Oncogene* 18(3): 833–836
54. KALLASSY M, TOFTGARD R, UEDA M, NAKAZAWA K, VORECHOVSKY I, YAMASAKI H, NAKAZAWA H 1997 Patched (ptch)-associated preferential expression of smoothened (smoh) in human basal cell carcinoma of the skin. *Cancer Res* 57(21): 4731–4735
55. ECHE I, PETRY F, ROSENBERGER A, TAUBER S, MO NKE-MEYER S, HESS I, DULLIN C, KIMMINA S, PIRNGRUBER J, JOHNSEN S A, UHMANN A, NITZKI F, WOJNOWSKI L, SCHULZ-SCHAEFFER W, WITT O, HAHN H 2009 Antitumor Effects of a Combined 5-Aza-2 Deoxycytidine and Valproic Acid Treatment on Rhabdomyosarcoma and Medulloblastoma in Ptch Mutant Mice. *Cancer Res* 69(3): 887–895
56. PENG L, HU J, LI S, WANG Z, XIA B, JIANG B, LI B, ZHANG Y, WANG J, WANG X 2013 Aberrant methylation of the PTCH1 gene promoter region in aberrant crypt foci. *Int J Cancer* 132: E18–E25
57. STECCA B, RUIZ I ALTABA A 2005 Brain as a paradigm of organ growth: Hedgehog-Gli signaling in neural stem cells and brain tumors. *J Neurobiol* 64(4): 476–90
58. KUBO M, NAKAMURA M, TASAKI A, YAMANAKAN, NAKASHIMA H, NOMURA M, KUROKI S, KATANO M 2004 Hedgehog signaling is a new therapeutic target for patients with breast cancer. *Cancer Res* 64: 6071–4
59. THAYER S P, DI MAGLIANO M P, HEISER P W, NIELSEN C M, ROBERTS D J, LAUWERS G Y, QI Y P, GYSIN S, FERNÁNDEZ-DEL CASTILLO C, YAJNIK V, ANTONIU B, MCMAHON M, WARSHAW A L, HEBROK M 2003 Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. *Nature* 425: 851–6
60. WATKINS D N, BERMAN D M, BURKHOLDER S G, WANG B, BEACHY P A, BAYLIN S B 2003 Hedgehog signalling within airway epithelial progenitors and in small-cell lung cancer. *Nature* 422(6929): 313–7
61. BERMAN D M, KARHADKAR S S, MAITRA A, MONTES DE OCA R, GERSTENBLITH M R, BRIGGS K *et al.* 2003 Widespread requirement for Hedgehog ligand stimulation in growth of digestive tract tumours. *Nature* 23;425(6960): 846–51
62. FAN L, PEPICELLI C V, DIBBLE C C, CATBAGAN W, ZARYCKI J L, LACIAK R 2004 *et al.* Hedgehog Signaling Promotes Prostate Xenograft Tumor Growth. *Endocrinology* 1;145(8): 3961–70.
63. MA X, CHEN K, HUANG S, ZHANG X, ADEGBOYEGA P A, EVERS B M *et al.* 2005 Frequent activation of the hedgehog pathway in advanced gastric adenocarcinomas. *Carcinogenesis* 26: 1698–1705
64. QUALTROUGH D, BUDA A, GAFFIELD W, WILLIAMS A C, PARASKEVA C 2004 Hedgehog signalling in colorectal tumour cells: induction of apoptosis with cyclopamine treatment. *Int J Cancer* 20;110(6): 831–7
65. CHENG W T, XU K, TIAN D Y, ZHANG Z G, LIU L J, CHEN Y 2009 Role of Hedgehog signaling pathway in proliferation and invasiveness of hepatocellular carcinoma cells. *Int J Oncol* 34: 829–36
66. CHEN X, HORIUCHI A, KIKUCHI N, OSADA R, YOSHIDA J, SHIOZAWA T, KONISHI I. 2007 Hedgehog signal pathway is activated in ovarian carcinomas, correlating with cell proliferation: Its inhibition leads to growth suppression and apoptosis. *Cancer Sci* 98: 68–76
67. PASCA DI MAGLIANO M, SEKINE S, ERMILOV A, FERRIS J, DLUGOSZ A A 2006 Hebrok M Hedgehog /Ras interactions regulate early stages of pancreatic cancer. *Genes Dev* 20: 3161–3173
68. KARHADKAR S S, BOVA G S, ABDALLAH N, DHARA S, GARDNER D, MAITRA A *et al.* 2004 Hedgehog signaling in prostate regeneration, neoplasia and metastasis. *Nature* 431: 707–712
69. STECCA B, MAS C, CLEMENT V, ZBINDEN M, CORREA R, PIGUET V, BEERMANN F, RUIZ I ALTABA A 2007 Melanomas require HEDGEHOG-GLI signaling regulated by interactions between GLI1 and the RAS-MEK/AKT pathways. *Proc Natl Acad Sci U S A* 3;104(14): 5895–900
70. KASPER M, JAKS V, FIASCHI M, TOFTGÅRD R 2009 Hedgehog signalling in breast cancer. *Carcinogenesis* 30(6): 903–11
71. TIAN H, CALLAHAN C A, DUPREE K J, DARBONNE W C, AHN C P, SCALES S J *et al.* 2009 Hedgehog signaling is restricted to the stromal compartment during pancreatic carcinogenesis. *Proc Natl Acad Sci* 106(11): 4254–9
72. YAUCH R L, GOULD S E, SCALES S J, TANG T, TIAN H, AHN C P, MARSHALL D, FU L, JANUARIO T, KALLOP D, NANNINI-PEPE M, KOTKOW K, MARSTERS J C, RUBIN L L, DE SAUVAGE F J A 2008 paracrine requirement for hedgehog signalling in cancer. *Nature, Sep 18; 455(7211): 406–10*
73. YAMASAKI A, KAMEDA C, XU R, TANAKA H, TASAKA T, CHIKAZAWA N, SUZUKI H, MORISAKI T, KUBO M, ONISHI H, TANAKA M, KATANO M 2010 Nuclear factor kappaB-activated monocytes contribute to pancreatic cancer progression through the production of Shh. *Cancer Immunol Immunother* 59(5): 675–86
74. DIERKS C, BEIGI R, GUO G R, ZIRLIK K, STEGERT M R, MANLEY P *et al.* 2008 Expansion of Bcr-Abl positive leukemic stem cells is dependent on Hedgehog pathway activation. *Cancer Cell* 14: 238–249
75. ZHAO C, CHEN A, JAMIESON C H, FERESHTEH M, ABRAHAMSSON A, BLUM J *et al.* 2009 Hedgehog signaling is essential for maintenance of cancer stem cells in myeloid leukaemia. *Nature* 458: 776–779
76. MAURAC I. Interaction of the Hh-Gli signaling pathway with BRCA1 and BRCA2 genes in ovarian cancer, doctoral thesis 2011 University of Zagreb, Croatia.
77. SABOL M, TRNSKI D, UZAREVIC Z, OZRETIC P, MUSANI V, RAFAJ M, CINDRIC M, LEVANAT S 2014 Combination of cyclopamine and tamoxifen promotes survival and migration of mcf-7 breast cancer cells – interaction of hedgehog-gli and estrogen receptor signaling pathways. *PLoS One* 9(12):e114510. doi: 10.1371/journal.pone.0114510
78. UZAREVIC Z 2011 The Hh-Gli signaling pathway activity in estrogen dependent (MCF-7) and estrogen independent (SkBr-3) breast cancer cell lines, doctoral thesis 2011, University Josip Juraj Strossmayer Osijek, Ruđer Bošković Institute, University of Dubrovnik, University Postgraduate Interdisciplinary Doctoral study Molecular biosciences, Croatia.
79. GOJEVIC A 2011 Expression of Gli isoforms in sporadic colon cancer, doctoral thesis 2011 University of Zagreb, Croatia.
80. LOW J A 2010 de Sauvage FJ. Clinical experience with Hedgehog pathway inhibitors. *J Clin Oncol* 28: 225–60
81. TREMBLAY M R, LESCARBEAU A, GROGAN M J 2009 Discovery of a potent and orally active hedgehog pathway antagonist (IPI-926). *J Med Chem* 52: 4400–18



82. TREMBLAY M R, NESLER M, WEATHERHEAD R, CASTRO A C 2009 Recent patents for Hedgehog pathway inhibitors for the treatment of malignancy. *Expert Opin Ther Pat* 19: 1039–56
83. KIM J, LEE J J, KIM J, GARDNER D, BEACHY P A 2010 Arsenic antagonizes the Hedgehog pathway by preventing ciliary accumulation and reducing stability of the Gli2 transcriptional effector. *Proc Natl Acad Sci USA* 107: 13432–37
84. BEAUCHAMP E M, RINGER L, BULUT G, SAJWAN K P, HALL M D, LEE Y C *et al.* 2011 Arsenic trioxide inhibits human cancer cell growth and tumor development in mice by blocking Hedgehog/Gli pathway. *J Clin Invest* 121: 148–60
85. KOGA K, NAKAMURA M, NAKASHIMA H, AKIYOSHI T, KUBO M, SATO N, KUROKI S, NOMURA M, TANAKA M, KATANO M. 2008 Novel link between estrogen receptor alpha and hedgehog pathway in breast cancer. *Anticancer Res* 28(2A): 731–40
86. JI Z, MEI F C, XIE J, CHENG X 2007 Oncogenic Kras suppresses GLI1 degradation and activates hedgehog signaling pathway in pancreatic cancer cells. *J Biol Chem* 282: 14048–14055
87. LANER-PLAMBERGERS, KASERA, PAULISCHTA M, HAUSER-KRONBERGER C, EICHBERGER T, FRISCHAUF A M 2009 Cooperation between GLI1 and JUN enhances transcription of JUN and selected GLI1 target genes. *Oncogene* 28: 1639–1651
88. CAI Q, LI J, GAO T, XIE J, EVERS B M 2009 Protein kinase Cdelta negatively regulates hedgehog signaling by inhibition of Gli1 activity. *J Biol Chem* 284: 2150–2158
89. YANAI K, NAKAMURA M, AKIYOSHI T, NAGAI S, WADA J, KOGA K *et al.* 2008 Crosstalk of hedgehog and Wnt pathways in gastric cancer. *Cancer Lett* 263: 145–156
90. YANG S H, ANDL T, GRACHTCHOUK V, WANG A, LIU J, SYU L J *et al.* 2008 Pathological responses to oncogenic Hedgehog signaling in skin are dependent on canonical Wnt/beta3-catenin signaling. *Nature Genet* 40: 1130–1135
91. MUSANI V, GORRY P, BASTA-JUZBASIC A, STIPIC T, MIKLIC P, LEVANAT S 2006 Mutation in exon 7 of PTCH deregulates SHH/PTCH/SMO signaling: possible linkage to WNT. *Int J Mol Med* 17: 755–759
92. STECCA B, RUIZ I, ALTABA A 2009 Gli1-p53 inhibitory loop controls neural stem cell and tumour cell numbers. *EMBO J* 28: 663–676
93. SCHNIDAR H, EBERL M, KLINGLER S, MANGELBERGER D, KASPER M, HAUSER-KRONBERGER C *et al.* 2009 Epidermal growth factor receptor signaling synergizes with Hedgehog-Gli in oncogenic transformation via activation of the MEK/ERK/JUN pathway. *Cancer Res* 69: 1284–1292.