



Functional neuroanatomy of nociception and pain

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Abstract

Pain is a complex sensory state based on the integration of a variety of nociceptive inputs processed centrally through many parallel and overlapping neural systems. The traditional anatomical concept implies that nociceptive information is dominantly used to generate and regulate perception of pain through one major sensory pathway. It becomes recognized that experiencing the affective component of the pain is at least as important as perception. Also, nociceptive information is strongly influencing brain centers for regulating homeostasis. So, understanding neuroanatomical organization of central processing of nociceptive information is of great clinical importance. There is an attempt to simplify this complex set of interacting networks to a core set of brain regions or a generalizable pain signature. Herewith we wish to give a short overview of recent advances by presenting principles about neuroanatomical organization for processing various aspects of nociceptive inputs.

GENERAL NEUROANATOMICAL PRINCIPLES IN NOCICEPTIVE PROCESSING

Pain is the most distinctive of all the sensory modalities (1) but can be simply defined as the subjective experience associated with actual or potential tissue damage (2–5). It serves an important protective function and warns to avoid or treat injury. The perception of pain is subjective and can vary greatly among individuals. Moreover, in the same individual an identical sensory stimulus can elicit quite distinct conscious responses under different conditions. This includes also psychological conditions, such as fear or anxiety that can significantly influence the experience of pain. So, more than most sensory modalities, the perception of pain is influenced by emotional state and environmental contingency, is dependent on experience, and varies so markedly from person to person (6–16), and consequently remains notoriously difficult to treat.

Noxious stimuli, including tissue injury, activate nociceptors (from the Latin, *noceo* = to injure, hurt) which are present in peripheral structures and transmit information to the CNS: from the body to the spinal cord dorsal grey column, from the skin of the head to the spinal and principal trigeminal nucleus, and from the neck mucosa to the lower 2/3rd of solitary tract nucleus (17–19). To generate perception of pain the information should continue ultimately to the cerebral cortex. From anatomical point of view it should be noted that nociception refers to the process through which information about peripheral stimuli is transmitted by primary afferent nociceptors to the spinal cord, brainstem, thalamus, and subcortical structures. For the experience of pain, activity of thalamocortical networks that process the information conveyed by pathways of nociception is needed (20–26) (Figure 1).

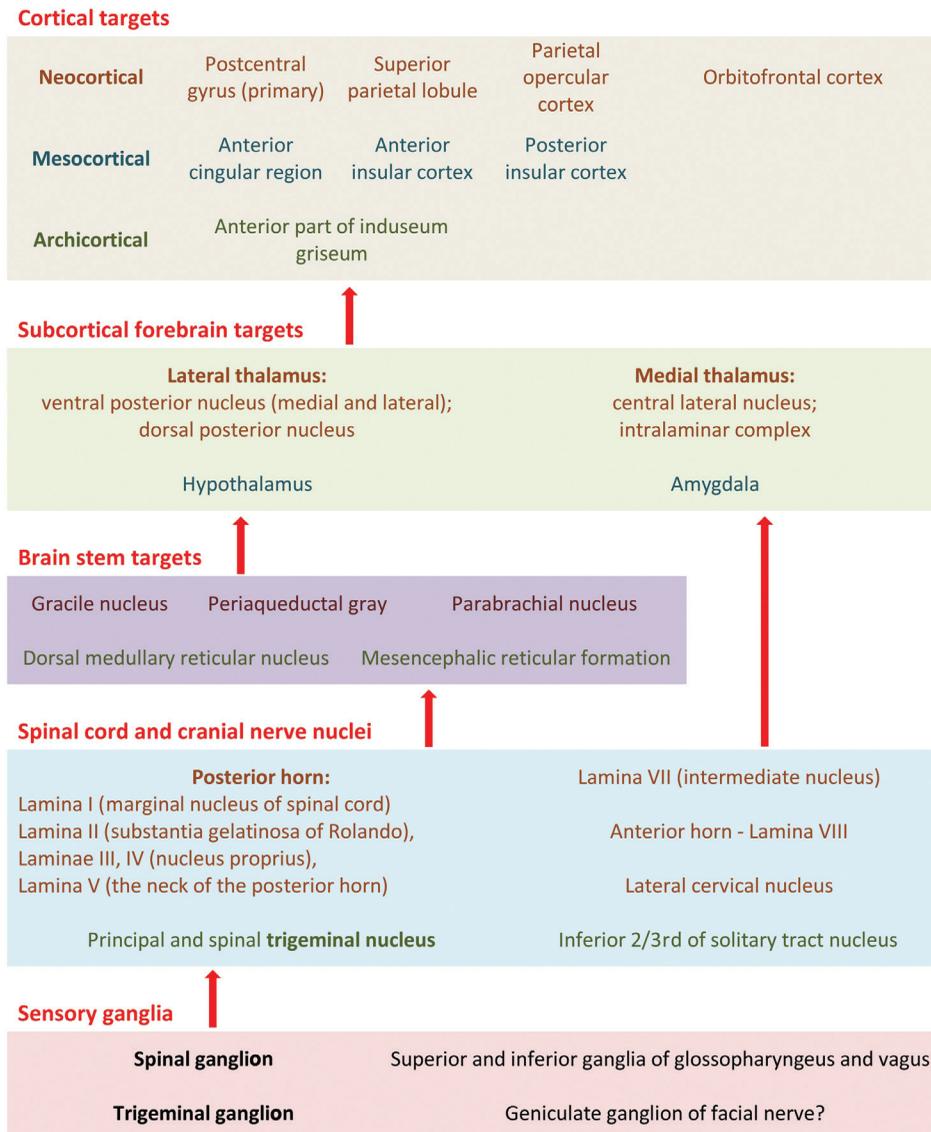


Figure 1. Structures containing neurons processing nociception and pain.

Pain sensation is not the direct expression of a sensory event. It has strong urgent and primitive quality with affective and emotional components (7; 26–32), and has also influence on homeostatic functioning (33; 34). By classical neuroanatomical description pain processing is dominantly related to spino- and trigemino-thalamic pathway (17–19), still this pathway is evolutionary newest in conducting nociceptive information (35). Pain is actually a complex sensory state that reflects the integration of many sensory signals and is the product of elaborate brain processing. Particularly complex processing is needed for experiencing the affective component of the pain and for the influencing on homeostasis. Numerous parallel and overlapping neural systems processing nociceptive information allow such a complex impact on the brain functioning (22; 36–40). Among the ascending pathways

arising from the spinal cord and its trigeminal homologue are the spinothalamic and spinoreticulothalamic tracts, as well as the spinoparabrachial-amygdala pathway, which provides more direct access to limbic emotional circuits in the brain. There is an attempt to simplify this complex set of interacting networks to a core set of brain regions or a generalizable pain signature. Such approaches identify the following areas as key to experiencing pain: the thalamus, the posterior and anterior insula, the postcentral gyrus, the anterior cingulate cortex, and the periaqueductal gray matter. Functional specificity of this complex pattern is still unresolved.

Altogether, understanding neuroanatomical organization of central processing of nociceptive information is of great clinical importance (41–45). Herewith we wish to give a short overview of recent advances in under-

TABLE 1

Various classes of nociceptors associate with specific function in the detection of distinct pain sub-modalities.

Modality	Submodality	Nociceptors/fibers
Nociceptive Pain (somatic or visceral)	sharp cutting pain	mechanical nociceptors A δ fibers
	dull burning pain	polymodal nociceptors C fibers
	deep aching pain from viscera	visceral nociceptors A δ , C fibers
	burning pain	thermal-mechanical (hot) A δ fibers
	freezing pain	thermal-mechanical (cold) C fibers
Non-Nociceptive Pain (neurophatic or symphatetic)	complex regional pain syndrome	no nociceptors
	post-herpetic neuralgia	
	phantom limb pain	
	anesthesia dolorosa	

standing neuroanatomical organization for various aspects of nociceptive processing and to relate neural structures and pathways to various submodalities of nociceptive processing.

MOLECULAR ANATOMY OF PERIPHERAL NOCICEPTIVE PROCESSING

Nociceptors, structures that respond selectively to stimuli that can damage tissue are widely distributed in the skin and deep tissues and include thermal, mechanical, and polymodal sensory receptors (46-49). They respond directly to mechanical and thermal stimuli, and indirectly to other stimuli by means of chemicals released from cells in the traumatized tissue. Fourth class, so called “silent” nociceptors are found in the viscera. The nociceptors are free nerve endings of the peripheral axonal branches originating from primary sensory neurons. The cell bodies of nociceptors that convey information from the body are located in the dorsal root ganglia. Trigeminal ganglia and superior ganglia of glossopharyngeus and vagus contain somatic neurons processing nociceptive stimuli from skin, cranial periost and dura, the mucosa of eye, nasal sinuses, oral and nasal cavity and possibly part of tympanic tube and cavity. For the rest of tympanic tube and cavity, and the whole mucosa of pharynx and larynx primary visceral neurons are located in the inferior ganglia of glossopharyngeus and vagus (17). The central branches of these primary neurons enter the central nervous system through spinal or cranial nerves where they make synaptic connections with a complex array of neurons that play different roles in nociceptive processing and pain. Trigeminal, glossopharyngeal and vagal nerve convey somatic information into the trigeminal nuclei, and visceral information into the nucleus of the solitary

tract. The spinal nerve conveys nociceptive information from the body and viscera and terminates on neurons in the dorsal horn of the spinal cord. Afferent fibers innervating viscera project to the CNS through autonomic nerves; some spinal afferents travel along hypogastric, lumbar colonic and splanchnic nerves to terminate in thoracolumbar regions as part of sympathetic innervation, while vagal and pelvic afferents respectively terminate in the brainstem and lumbosacral cord and contribute to parasympathetic innervation. Thoracic branches of vagus also contain afferent fibers, but the nature to conduct pain information from thoracic and abdominal viscera is not clarified.

Based on the myelination of their afferent fibers nociceptors in the skin, muscle, joints, and visceral receptors fall into two broad classes (2; 50-52). The first class are small diameter, thin myelinated A δ fibers that innervate nociceptors producing short-latency pain described as sharp and pricking. The majority are mechanoreceptors because they are activated by sharp objects that penetrate, squeeze, or pinch the skin. In addition, many of these A δ fibers also respond to noxious heat that can burn the skin. Electrophysiological studies have subdivided A δ nociceptors into two main classes: type I and type II. Type I respond to both mechanical and chemical stimuli, but have relatively high heat thresholds (>50°C). These fibers mediate the first pain provoked by pinprick and other intense mechanical stimuli. Type II A δ nociceptors have a much lower heat threshold, but a very high mechanical threshold; these fibers mediate the “first” acute pain response to noxious heat. The second class are nociceptors innervated by small diameter, unmyelinated C fibers producing diffusely distributed and poorly tolerated dull, burning pain. Under this heterogeneous class the most common type are polymodal nociceptors activated by a variety of noxious mechanical, thermal, and chemical stimuli, such as

pinch or puncture, noxious heat and cold, and irritant chemicals applied to the skin. Electrical stimulation of these fibers in humans evokes prolonged sensations of burning pain. Of particular interest are the heat responsive, but mechanically insensitive unmyelinated afferents (so-called silent nociceptors) that develop sensitivity when the chemical milieu of inflammation alters their properties (53).

Visceral fibers can serve sensory function and evoke conscious sensations and also afferent function and regulate autonomic flow. Visceral sensory afferents are almost exclusively thinly myelinated A δ -fibres and unmyelinated C-fibres. However, the distinction between nociceptive afferents and non-nociceptive afferents in the viscera is not clear. Nociceptors in the viscera that produce sensations of intense pain are activated by distension or swelling. Silent nociceptors found in the viscera are not normally activated by noxious stimulation. However, after tissue insult these nociceptors “wake up” in response to endogenous chemical mediators associated with tissue injury. Silent nociceptors are typically associated with increased spontaneous activity and responsiveness to noxious and even innocuous stimulus intensities. Their activation is thought to contribute to the development of secondary hyperalgesia and central sensitization, two prominent pain syndromes.

The membrane of the nociceptor contains protein complexes forming receptors and channels that transduce thermal, mechanical, or chemical energy of noxious stimuli into electrical impulses, which are propagated along the peripheral and central axon of the nociceptor into the central nervous system. Biochemical and molecular analysis of the nociceptor has identified multiple nociceptor subclasses, each expressing a distinctive repertoire of membrane ion channels, receptors and intracellular signaling proteins that are attractive targets for the development of drugs for therapeutic intervention in clinical pain conditions (54-56). This includes voltage-gated sodium, potassium and calcium channels, leak channels, and ligand-gated channels such as acid-sensing ion channels and transient receptor potential (TRP) channels (57). The role of transient receptor potential ion channels in thermal sensation was originally discovered by analyses of natural substances such as capsaicin and menthol that produce burning or cooling sensations when applied to the skin or injected subcutaneously. Capsaicin, the active ingredient in chili peppers, has been used extensively to activate nociceptive afferents that mediate sensations of burning pain (58). The varieties of TRP channels in nociceptors underlie the perception of a wide range of temperatures from extreme cold to intense heat. Some classes of TRP receptors are activated by cold temperatures and inactivated by warming (TRPM8 and TRPA1 receptors) while some types of TRP receptors are activated by warm or hot temperatures and inactivated by cooling (TRPV3, TRPV1, TRPV2 and TRPV4 receptors) (59-61). A key

role in the perception of pain in humans plays the voltage-gated sodium channel NaV1.7, found exclusively in the periphery and its mutation can lead to the inability to experience pain (62). An ionotropic purinergic receptor, PTX3, that is activated by adenosine triphosphate (ATP) released from peripheral cells after tissue damage is also expressed by nociceptors (63-65). In addition, nociceptors express members of the Mas-related G protein-coupled receptor (Mrg) family, which are activated by peptide ligands. They serve to sensitize nociceptors to other chemicals released in their local environment (66-68).

Neuroanatomical and molecular characterization of nociceptors demonstrated their heterogeneity; various functionally and molecularly heterogeneous classes of nociceptors associate with specific function in the detection of distinct pain submodalities (69).

MECHANISMS MODULATING NOCICEPTIVE PROCESSING

The proper function of the nociceptive system enables and enforces protective behavioral responses such as withdrawal or avoidance to acutely painful stimuli. In case of an injury the vulnerability of the affected tissue is going to increase (70-75). The nociceptive system adapts to this enhanced vulnerability by locally lowering the nociceptive thresholds and by facilitation of nocifensive responses. The behavioral correlates of these adaptations are allodynia (condition in which non-noxious events are perceived as noxious) and hyperalgesia (mildly noxious events are perceived as highly noxious). However, hyperalgesia and allodynia may persist long after the initial cause for pain or may occur due to dysfunction of parts of the peripheral or central nervous system and become maladaptive rather than protective.

Two types of hyperalgesia, primary and secondary, are associated with different mechanisms. Primary hyperalgesia develops at the site of tissue injury associated with an increased sensitivity of the peripheral nerve fibers involved in pain. Secondary hyperalgesia develops in uninjured tissue surrounding the site of injury. This form of hyperalgesia is not caused by sensitization of nociceptive nerve endings but due to changes in the excitability of neurons in the central nervous system, including the spinal cord and supra-spinal sites in the brain. Recent evidence suggests, however, that altered processing in the central nervous system is equally important in the induction of primary hyperalgesia (70).

Numerous mediators in the peripheral and central nervous systems contribute to the processes of sensitization. The sensitization of nociceptors is triggered by chemical mediators released from distinct cell types that accumulate at the site of tissue injury and act together to decrease the threshold of nociceptor activation, such as histamine, anandamide, acetylcholine, serotonin, norepinephrine,

prostaglandin, bradykinin, substance P and calcitonin gene-related peptide. Central sensitization is a considerably more complicated process that can result from increased release of excitatory neurotransmitter (e.g., glutamate, substance P) and/or enhanced synaptic efficacy. These changes relate to several cellular mechanisms (a) presynaptic changes, (b) postsynaptic changes, (c) interneuron changes, (d) changes in descending modulation, and (e) immune/microglial mechanisms (76; 77).

The understanding of pain mechanisms and pain control has focused on the properties of primary afferent and dorsal horn nociceptive neurons and ascending pathways. However, there is an active regulation of sensory transmission at the level of the dorsal horn by descending projections arising from a number of brain sites (78-89). Descending control plays a critical role in determining the experience of pain and can be facilitatory as well as inhibitory. The balance between inhibition and facilitation is dynamic, and can be altered in different behavioral, emotional and pathological states. For example, intense stress and fear are associated with decreased responsiveness to noxious stimuli that reflects a shift towards descending inhibition. By contrast, inflammation and nerve injury, sickness, and chronic opioid administration are associated with hyperalgesia that in part reflects a shift towards descending facilitation. There is much evidence to suggest that descending facilitation of spinal nociception is a major contributor to central sensitization and the development of secondary hyperalgesia.

Descending control arises from a number of supraspinal sites, including the midline periaqueductal gray-rostral ventromedial medulla (PAG-RVM) system, and the more lateral and caudal dorsal reticular nucleus (DRt) and ventrolateral medulla (VLM). Serotonergic neurons settled in the raphe nuclei and neorepinephrine neurons settled in the locus coeruleus play the major role in descending control. Inhibitory control from the PAG-RVM system preferentially suppresses nociceptive inputs mediated by C-fibers, preserving sensory-discriminative information conveyed by more rapidly conducting A-fibers. Analysis of the circuitry within the RVM reveals that the neural basis for bidirectional control from the midline system is two populations of neurons, ON-cells and OFF-cells that are differentially recruited by higher structures important in fear, illness and psychological stress to enhance or inhibit pain.

FUNCTIONAL NEUROANATOMY OF CENTRAL NOCICEPTIVE PROCESSING

The pain-related circuitry extends over both the cognitive and the emotional-motivational domains of the brain. It includes the spinothalamic component of the anterolateral fasciculus, the trigeminothalamic tract and pathways that connect the spinal cord and the sensory nuclei

of the trigeminal nerve with the brain stem, the hypothalamus and the basal forebrain (17).

The central branches of primary neurons enter the central nervous system and make synaptic connections with a complex array of neurons. This triggers the release of neurotransmitters such as glutamate and substance P, which activate second-order neurons that project to the brain. The spinal gray dorsal horn-column (and its equivalent, the caudal part of trigeminal nerve nucleus) constitutes one of the main relay stations for primary afferents of the dorsal roots.

Besides nociceptive-specific neurons, the spinal dorsal horn (especially deeper laminae) also contains wide dynamic range (WDR) neurons that respond to both innocuous and noxious stimuli. After entering the spinal cord (through lateral division of peripheral nerve) small myelinated A δ and unmyelinated C fibers terminate in the dorsal horn. The spinal gray matter in the dorsal horn is divided into six layers of cells (90-93). Nociception specific neurons in the lamina I respond selectively to noxious inputs from A δ or C fibers and project to higher brain structures. Other classes of lamina I responded to both innocuous and noxious mechanical stimulation and thus are termed wide-dynamic-range neurons. Neurons in laminae II and III are interneurons that receive inputs from A δ and C fibers, and make excitatory or inhibitory connections to neurons in lamina I, IV, and V that project to higher brain centers. Wide-dynamic-range neurons in lamina V typically respond to a wide variety of noxious stimuli and project to the brain stem and thalamus. Neurons in lamina V also receive input from nociceptors in visceral tissues.

Many neurons located in laminae VII and VIII, building the intermediate grey and deep part of ventral gray column, have complex response properties to noxious stimuli because the inputs from nociceptors to these neurons are conveyed through many intervening synapses. Whereas most dorsal horn neurons receive unilateral input, neurons in lamina VII often respond to stimulation of both side of the body and therefore contribute to the diffuse quality of many pain conditions. Visceral C fibers terminate ipsilaterally in laminae I, II, V, and X and also in lamina V and X of the contralateral gray matter.

Finally, fibers from lamina I and the nucleus of the solitary tract convey information about impending and actual tissue damage, and a wide range of visceral stimuli; the responses of neurons in lamina V correlate closely with reports of pain intensity, while the lamina VII plays a significant role in emotional responses to sensory stimuli.

Five major ascending pathways contribute to the central processing of nociceptive information: the spinothalamic, spinoreticular, spinomesencephalic, cervicothalamic, and spinohypothalamic tracts.

From neurons in laminae I, V, and VII, the main targets of the small-diameter fibers with sensory information destined for conscious perception, originates the spinothalamic tract that is the principal pathway transmitting noxious, thermal, and visceral information to the thalamus and cerebral cortex (94). The axons of most neurons in lamina I cross the midline, just ventral to the central canal, and ascend in the contralateral lateral spinothalamic tract located in the lateral funiculus. Axons of lamina V neurons cross the spinal cord and ascend in the contralateral ventral spinothalamic tract. Axons from the caudal part of spinal trigeminal nucleus form tractus trigeminothalamicus lateralis that crosses the midline and joins the spinothalamic tract. As a result of the decussation of spinothalamic fibers in the spinal cord, noxious information from each dermatome is transmitted contralaterally in the anterolateral column. The importance in pain processing through the spinothalamic tract is demonstrated by experimental and clinical evidence. Electrical stimulation is sufficient to elicit the sensation of pain and anterolateral cordotomy results in a marked reduction in pain sensation on the side of the body contralateral to that of the lesion. As mentioned, information carried by small-diameter sensory fibers also reaches the cerebral cortex through several polysynaptic routes other than the spinothalamic tract (38-40; 95). Although many of these pathways originate from neurons in lamina I and V, they arise from a different group of neurons and project to brain stem nuclei which in turn project to the thalamus and to other sites, such as the hypothalamus and amygdala. Neurons that project to sites other than the thalamus are involved in homeostatic control, by regulating endocrine release and autonomic activity. The spinoreticular tract ascends in the anterolateral quadrant of the spinal cord and terminates in both the reticular formation and the thalamus. It contains the axons of projection neurons in laminae VII and VIII and do not cross the midline. The spinomesencephalic (spinoparabrachial) tract projects in the anterolateral quadrant of the spinal cord to the mesencephalic reticular formation and periaqueductal gray matter. It contains the axons of projection neurons in laminae I and V. Information transmitted along this tract are thought to contribute to the affective component of pain. They also course through the dorsal part of the lateral funiculus and project to the parabrachial nucleus. Neurons of the parabrachial nucleus project to the amygdala, a key nucleus of the limbic system that regulates emotional states. The spinohypothalamic tract projects to hypothalamic nuclei that serve as autonomic control centers involved in the regulation of the neuroendocrine and cardiovascular responses that accompany pain syndromes. It contains the axons of neurons in laminae I, V, and VIII. The cervicothalamic tract runs in the lateral white matter of the upper two cervical segments of the spinal cord and contains the axons of neurons of the lateral cervical nucleus, which receives input from neurons in laminae III and IV of the dorsal horn. Most axo-

ns in the cervicothalamic tract cross the midline and ascend in the medial lemniscus of the brain stem, terminating in midbrain nuclei and in the ventroposterior nuclei of the thalamus. The signals from nociceptors in the pelvic and abdominal viscera are relayed through other neurons in laminae III and IV of the sacral and midthoracic spinal cord which project axon through the most medial part of the white dorsal column terminating in the gracile nucleus.

The thalamus represents a key integrative structure for the processing of pain and thus contains several relay nuclei that participate in the central processing of nociceptive information.

The lateral nuclear group comprises the ventroposterior medial nucleus, the ventroposterior lateral nucleus, and the dorsal posterior nucleus that receive inputs from nociception-specific and wide-dynamic-range neurons in laminae I and V of the dorsal horn through the spinothalamic tract. The lateral thalamus is thought to be concerned with the processing of information about the precise location of an injury, information usually conveyed to consciousness as acute pain. The medial nuclear group of the thalamus comprises the central lateral nucleus of the thalamus and the intralaminar complex and receives major input from neurons in laminae VII and VIII of the spinal cord. Many neurons in the medial thalamus respond optimally to noxious stimuli and project widely to the basal ganglia and different cortical areas. The pathway to the medial thalamus was the first spinothalamic projection evident in the evolution of mammals and is therefore known as the paleospinothalamic tract. It is also sometimes referred to as the spinoreticulothalamic tract because it includes indirect connections through the reticular formation of the brain stem. The projection from the lateral thalamus to the ventroposterior nuclei is most developed in primates, and is termed the neospinothalamic tract.

Regarding cortical processing it should be emphasized that pain is a complex perception that involves many areas. Spatial, temporal and intensity aspects of pain perception are processed in the primary and secondary somatosensory cortex (S1 and S2, respectively). The anterior cingulate cortex and insular cortex are involved in processing emotional states associated with pain, while the dorsal posterior insula contributes to the autonomic component of pain responses. Moreover, the prefrontal cortex with corresponding mediodorsal thalamic nucleus and premotor areas in connection with cerebellum are also commonly activated by painful stimulation.

FUNCTIONAL NEUROANATOMY OF PAIN SUBMODALITIES

The sense of pain includes a response to external events that damage or harm the body and therefore belongs to

exteroceptive sensation. Although the sense of the function of the major organ systems of the body and its internal state (interoception) do not become conscious sensation, abnormal function in major organ systems resulting from disease or trauma can evoke conscious sensations of pain.

When pain is experienced it can be acute, persistent or chronic (76). Persistent pain characterizes many clinical conditions and is usually the major reason why patients seek medical attention. In contrast, chronic pain does not serve a useful biological function; it only makes patients miserable (7; 31; 32; 96-101). Persistent pain can be subdivided into two broad classes, nociceptive and neuropathic (Table 1). Nociceptive pains result from the direct activation of nociceptors in the skin or soft tissue in response to tissue injury and usually arise from accompanying inflammation. Sprains and strains produce mild forms of nociceptive pain, whereas arthritis or a tumor that invades soft tissue produces a much more severe nociceptive pain. Pain may also result from injury to sensory fibers or from damage to the central nervous system. These types of pain are designated as non-nociceptive or neuropathic pain (77; 102-105). Neuropathic pains include the syndromes of reflex sympathetic dystrophy, also called complex regional pain syndrome, and post-herpetic neuralgia, the severe pain experienced by patients after a bout of shingles. Other neuropathic pains include phantom limb pain (106-109), the pain that occurs after limb amputation. In some instances pain can even occur without a peripheral stimulus, a phenomenon termed anesthesia dolorosa. This syndrome can be triggered following attempts to block chronic pain, for example after therapeutic transection of sensory afferent fibers in the dorsal roots.

Sharp, pricking pain is produced by nociceptors innervated by A δ fibers and activated by sharp objects that penetrate, squeeze, or pinch the skin (mechanical nociceptors). Dull, burning pain that is diffusely localized and poorly tolerated is produced by nociceptors innervated by C fiber (polymodal nociceptors). When a stone hits your elbow, you initially feel a sharp, fast, momentary pain, called first pain and conveyed by A δ fibers followed by a more prolonged aching and sometimes burning pain ("second pain") which is transmitted by the C fibers.

There is a clear demarcation between the perception of innocuous warmth and noxious heat. This enables us to avoid temperatures capable of causing tissue damage. If skin temperature changes slowly, we are unaware of changes in the range 31° to 36°C. Below 31°C the sensation progresses from cool to cold and finally, beginning at 10° to 15°C, to pain. Above 36°C the sensation progresses from warm to hot and then, beginning at 45°C, to pain. The freezing pain is produced by cold nociceptors innervated by C fibers, while the burning pain is produced by heat nociceptors innervated by A δ fibers. Thermal stimuli activate specific classes of TRP ion channels in the membrane (60). Two classes of TRP receptors are

activated by cold temperatures and inactivated by warming; TRPM8 receptors respond to temperatures below 25°C (such temperatures are perceived as cool or cold), while TRPA1 receptors have thresholds below 17°C (this range is described as cold or frigid). Four types of TRP receptors are activated by warm or hot temperatures and inactivated by cooling; TRPV3 receptors are expressed in warm type fibers and they respond to warming of the skin above 35°C (generate sensations ranging from warm to hot). TRPV1 and TRPV2 receptors are expressed in heat nociceptors and respond to temperatures exceeding 45°C (mediate sensations of burning pain). TRPV4 receptors are activated by temperatures above 27°C and respond to normal skin temperatures.

Visceral pain (deep aching pain) is sensations of intense, diffuse, often poorly localized pain. Diffuse nature and difficulty in locating visceral pain is due to a low density of visceral sensory innervation and extensive spinal distribution of visceral C fibers. Visceral pain is often associated with marked autonomic phenomena: nausea, gastrointestinal disturbances, pallor, profuse sweating, and changes in blood pressure, heart rate and body temperature. In addition, visceral pain frequently produces referred pain, a condition in which pain from injury to a visceral tissue is perceived as originating from a region of the body surface. Referred pain is sharper, better localized and less likely to be accompanied by autonomic signs, and therefore difficult to differentiate from pain of somatic origin. Convergence of visceral and somatic afferent fibers may account for referred pain phenomenon. Nociceptive afferent fibers from the viscera and fibers from specific areas of the skin converge on the same projection neurons in the dorsal horn (lamina V neurons). Thus, a signal from this neuron does not inform higher brain centers about the source of the input and the brain incorrectly attributes the pain to the skin. Another anatomical explanation for instances of referred pain is that the axons of nociceptive sensory neurons branch in the periphery, innervating both skin and visceral targets (110; 111).

There is one unpleasant sensation showing considerable apparent overlap in the neuronal cells and circuits that transmit pain, the itch. It is characterized by a strong innate urge to scratch and is confined to the skin, the ocular conjunctiva, and the mucosa. Itch was considered to be related to pain, because painful stimuli such as scratching inhibits while some analgesics (e.g. opioids) can cause itch. In addition, some itch-causing compounds (pruritogens) induce inflammatory pain, and under certain circumstances some pain-causing compounds (allogens) induce itch. Finally, there is a broad overlap between relevant mediator systems in pain and itch. Parallels between pain and itch processing are even more evident in the pattern of central sensitization. Touch- and pin prick-induced pain (allodynia and punctate hyperalgesia) correlate to touch- and pin prick induced itch (alloknesis and punctate hyperknesis). However, recent discoveries about

molecular and cellular basis of itch cover diverse aspects of itch sensation, from the identification of new receptors to the characterization of spinal cord itch circuits. These studies demonstrate that itch sensory signals are clearly demarcated from input of other somatosensory modalities (pain, touch, temperature) that is achieved by the expression of dedicated receptors and transmitters in a select population of sensory neurons which detect pruritogens. Also, itch specificity is maintained in a spinal cord circuit by the utilization of specific neurotransmitters and cognate receptors to convey input along a distinct cellular pathway (112-116).

Recent advances in molecular and anatomical processing of pain: clinical implications

Impressive insights into the molecular mechanisms of peripheral pain transduction has been recently developed opening the way for more effective pain therapies. Human genetics and molecular biology have revealed specific channels expressed selectively by nociceptive sensory neurons, such as TRP and Na channels, what has led to the development of many small-molecule channel antagonists some of which may prove to be effective as selective peripheral analgesics. The use of transcutaneous and dorsal-column electrical stimulation in the control of certain types of peripheral pain was encouraged by the finding that the balance of activity in small- and large-diameter sensory fibers modulates the perception of pain. Also, an observation that stimulation of specific sites in the brain stem produces deep analgesia has encouraged efforts to control pain by activating endogenous modulatory systems. In certain clinical conditions intrathecal and epidural administration of opiates induces a potent analgesia. However, understanding of the organization of central pain circuits under normal and pathological conditions remains relatively incomplete and thereby for most central pain syndromes there are still no effective pain therapies. It is expected that the future progress in pain therapy will depend on the research finding about brain circuits that transmit nociceptive signals (2; 117-120).

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REFERENCES

- WOOLF C J 2010 What is this thing called pain? *J Clin Invest* 120: 3742-3744
- BASBAUM A, JESSELL T 2012 Pain. In: Kandel E R, Schwartz J H, Jessell T M (eds) *Principles of Neural Science*. McGraw-Hill, New York, p 1709
- HOLDCROFT A, JAGGAR S 2006 *Core Topics in Pain*. Cambridge University Press, Cambridge.
- MCMAHON S, KOLTZENBURG M, TRACEY I, TURK D C 2013 *Wall and Melzack's Textbook of Pain*. Elsevier.
- MILLAN M J 1999 The induction of pain: an integrative review. *Prog Neurobiol* 57: 1-164
- CORBETTA, HUSEBO B S, ACHTERBERG W P, AARSLAND D, ERDAL A, FLO E 2014 The importance of pain management in older people with dementia. *Br Med Bull* 111: 139-148
- BUSHNELL M C, CEKO M, LOW L A 2013 Cognitive and emotional control of pain and its disruption in chronic pain. *Nat Rev Neurosci* 14: 502-511
- HASHMI J A, DAVIS K D 2014 Deconstructing sex differences in pain sensitivity. *Pain* 155: 10-13
- MOGIL J S, BAILEY A L 2010 Sex and gender differences in pain and analgesia. *Prog Brain Res* 186: 141-157
- PALLER C J, CAMPBELL C M, EDWARDS R R, DOBS A S 2009 Sex-based differences in pain perception and treatment. *Pain Med* 10: 289-299
- WIECH K, PLONER M, TRACEY I 2008 Neurocognitive aspects of pain perception. *Trends CognSci* 12: 306-313
- LEKNES S, TRACEY I 2008 A common neurobiology for pain and pleasure. *Nat Rev Neurosci* 9: 314-320
- HADJISTAVROPOULOS T, HERR K, PRKACHIN K M, CRAIG K D, GIBSON S J, LUKAS A, SMITH J H 2014 Pain assessment in elderly adults with dementia. *Lancet Neurol* 13: 1216-1227
- ATLAS L Y, WAGER T D 2012 How expectations shape pain. *Neurosci Lett* 520: 140-148
- KATONA C, PEVELER R, DOWRICK C, WESSELY S, FEINMANN C, GASK L, LLOYD H, WILLIAMS A C, WAGER E 2005 Pain symptoms in depression: definition and clinical significance. *Clin Med* 5: 390-395
- LAVIN R, PARK J 2014 A characterization of pain in racially and ethnically diverse older adults: a review of the literature. *J Appl Gerontol* 33: 258-290
- NIEUWENHUYNS R, VOOGD J, VAN HUIJZEN C 2007 *The Human Central Nervous System*. Springer-Verlag, Berlin Heidelberg, New York.
- USUNOFF K G, POPRATILOFF A, SCHMITT O, WREE A 2006 Functional neuroanatomy of pain. *Adv Anat Embryol Cell Biol* 184: 1-115
- CRAIG A D 2003 Pain mechanisms: labeled lines versus convergence in central processing. *Annu Rev Neurosci* 26: 1-30
- NEUGEBAUER V, GALHARDO V, MAIONE S, MACKAY S C 2009 Forebrain pain mechanisms. *Brain Res Rev* 60: 226-242
- GARCIA-LARREA L 2012 The posterior insular-opercular region and the search of a primary cortex for pain. *Neurophysiol Clin* 42: 299-313
- WOO C W, KOBAN L, KROSS E, LINDQUIST M A, BANICH MT, RUZIC L, ANDREWS-HANNA J R, WAGER T D 2014 Separate neural representations for physical pain and social rejection. *Nat Commun* 5: 5380
- VIERCK C J, WHITSEL B L, FAVOROV O V, BROWN A W, TOMMERDAHL M 2013 Role of primary somatosensory cortex in the coding of pain. *Pain* 154: 334-344
- WILLIS W D JR. 2007 The somatosensory system, with emphasis on structures important for pain. *Brain Res Rev* 55: 297-313
- WOOLF C J 2011 Central sensitization: implications for the diagnosis and treatment of pain. *Pain* 152: S2-S15
- VISWANATHAN A, HARSH V, PEREIRA E A, AZIZ T Z 2013 Cingulotomy for medically refractory cancer pain. *Neurosurg Focus* 35: E1
- FLATEN M A 2014 Pain-related negative emotions and placebo analgesia. *Handb Exp Pharmacol* 225: 81-96
- ELMAN I, BORSOOK D, VOLKOW N D 2013 Pain and suicidality: insights from reward and addiction neuroscience. *Prog Neurobiol* 109: 1-27

29. SHACKMAN A J, SALOMONS T V, SLAGTER H A, FOX A S, WINTER J J, DAVIDSON R J 2011 The integration of negative affect, pain and cognitive control in the cingulate cortex. *Nat Rev Neurosci* 12: 154-167
30. SANDKUHLER J, LEE J 2013 How to erase memory traces of pain and fear. *Trends Neurosci* 36: 343-352
31. ZHUO M 2014 Long-term potentiation in the anterior cingulate cortex and chronic pain. *Philos Trans R Soc Lond B Biol Sci* 369: 20130146
32. TOYODA H, LI X Y, WU L J, ZHAO M G, DESCALZI G, CHEN T, KOGA K, ZHUO M 2011 Interplay of amygdala and cingulate plasticity in emotional fear. *Neural Plast* 2011: 813749
33. CRAIG A D 2003 A new view of pain as a homeostatic emotion. *Trends Neurosci* 26: 303-307
34. LUMB B M 2002 Inescapable and escapable pain is represented in distinct hypothalamic-midbrain circuits: specific roles for Adelta- and C-nociceptors. *Exp Physiol* 87: 281-286
35. BONAVITA V, DE SIMONE R 2011 Pain as an evolutionary necessity. *Neurol Sci* 32 Suppl 1: S61-66
36. GARCIA-LARREA L, PEYRON R 2013 Pain matrices and neuropathic pain matrices: a review. *Pain* 154 (Suppl 1): S29-43
37. WAGER T D, ATLAS L Y, LINDQUIST M A, ROY M, WOO C W, KROSS E 2013 An fMRI-based neurologic signature of physical pain. *N Engl J Med* 368: 1388-1397
38. WOO C W, ROY M, BUHLE J T, WAGER T D 2015 Distinct brain systems mediate the effects of nociceptive input and self-regulation on pain. *PLoS Biol* 13: e1002036
39. WILLIS W D, WESTLUND K N 1997 Neuroanatomy of the pain system and of the pathways that modulate pain. *J Clin Neurophysiol* 14: 2-31
40. ALMEIDA T F, ROIZENBLATT S, TUFIK S 2004 Afferent pain pathways: a neuroanatomical review. *Brain Res* 1000: 40-56
41. ZERNIKOW B, WAGER J, BREHMER H, HIRSCHFELD G, MAIER C 2015 Invasive treatments for complex regional pain syndrome in children and adolescents: a scoping review. *Anesthesiology* 122: 699-707
42. SCHWEINHARDT P, BUSHNELL M C 2010 Pain imaging in health and disease—how far have we come? *J Clin Invest* 120: 3788-3797
43. WAXMAN S G, MERKIES I S, GERRITS M M, DIB-HAJJ S D, LAURIA G, COX J J, WOOD J N, WOODS C G, DRENTH J P, FABER C G 2014 Sodium channel genes in pain-related disorders: phenotype-genotype associations and recommendations for clinical use. *Lancet Neurol* 13: 1152-1160
44. FRIEDRICHS DORF S J, POSTIERA, EULL D, WEIDNER C, FOSTER L, GILBERT M, CAMPBELL F 2015 Pain Outcomes in a US Children's Hospital: A Prospective Cross-Sectional Survey. *Hosp Pediatr* 5: 18-26
45. GILRON I, DICKENSON A H 2014 Emerging drugs for neuropathic pain. *Expert Opin Emerg Drugs* 19: 329-341
46. BASBAUM A I, BAUTISTA D M, SCHERRER G, JULIUS D 2009 Cellular and molecular mechanisms of pain. *Cell* 139: 267-284
47. MENDEL L M, ALBERS K M, DAVIS B M 1999 Neurotrophins, nociceptors, and pain. *Microsc Res Tech* 45: 252-261
48. DUBIN A E, PATAPOUTIAN A 2010 Nociceptors: the sensors of the pain pathway. *J Clin Invest* 120: 3760-3772
49. GREENSPAN J D 1997 Nociceptors and the peripheral nervous system's role in pain. *J Hand Ther* 10: 78-85
50. RAJA S N, MEYER R A, CAMPBELL J N 1988 Peripheral mechanisms of somatic pain. *Anesthesiology* 68: 571-590
51. LEWIN G R, MOSHOURAB R 2004 Mechanosensation and pain. *J Neurobiol* 61: 30-44
52. WOOLF C J, MA Q 2007 Nociceptors—noxious stimulus detectors. *Neuron* 55: 353-364
53. SCHMIDT R, SCHMELZ M, FORSTER C, RINGKAMP M, TOREBJORK E, HANDWERKER H 1995 Novel classes of responsive and unresponsive C nociceptors in human skin. *J Neurosci* 15: 333-341
54. FINE P G, ROSENFELD M J 2013 The endocannabinoid system, cannabinoids, and pain. *Rambam Maimonides Med J* 4: e0022
55. JULIUS D, BASBAUM A I 2001 Molecular mechanisms of nociception. *Nature* 413: 203-210
56. PATAPOUTIAN A, TATE S, WOOLF C J 2009 Transient receptor potential channels: targeting pain at the source. *Nat Rev Drug Discov* 8: 55-68
57. WAXMAN S G, ZAMPONI G W 2014 Regulating excitability of peripheral afferents: emerging ion channel targets. *Nat Neurosci* 17: 153-163
58. O'NEILL J, BROCK C, OLESEN A E, ANDRESEN T, NILSSON M, DICKENSON A H 2012 Unravelling the mystery of capsaicin: a tool to understand and treat pain. *Pharmacol Rev* 64: 939-971
59. WILLIS W D JR. 2009 The role of TRPV1 receptors in pain evoked by noxious thermal and chemical stimuli. *Exp Brain Res* 196: 5-11
60. JULIUS D 2013 TRP channels and pain. *Annu Rev Cell Dev Biol* 29: 355-384
61. WANG H, WOOLF C J 2005 Pain TRPs. *Neuron* 46: 9-12
62. COX J J, REIMANN F, NICHOLAS A K, THORNTON G, ROBERTS E, SPRINGELL K, KARBANI G, JAFRI H, MANNAN J, RAASHID Y, AL-GAZALI L, HAMAMY H, VALENTE E M, GORMAN S, WILLIAMS R, MCHALE D P, WOOD J N, GRIBBLE F M, WOODS C G 2006 An SCN9A channelopathy causes congenital inability to experience pain. *Nature* 444: 894-898
63. BURNSTOCK G 2013 Purinergic mechanisms and pain—an update. *Eur J Pharmacol* 716: 24-40
64. TSUDA M, TOZAKI-SAITOH H, INOUE K 2010 Pain and purinergic signaling. *Brain Res Rev* 63: 222-232
65. BURNSTOCK G 2009 Purinergic receptors and pain. *Curr Pharm Des* 15: 1717-1735
66. OKUDA-ASHITAKA E, ITO S 2015 Pain regulation by nocistatin-targeting molecules: G protein-coupled-receptor and nocistatin-interacting protein. *Vitam Horm* 97: 147-165
67. HU S S, HO Y C, CHIOU L C 2014 No more pain upon Gq-protein-coupled receptor activation: role of endocannabinoids. *Eur J Neurosci* 39: 467-484
68. PAN H L, WU Z Z, ZHOU H Y, CHEN S R, ZHANG H M, LI D P 2008 Modulation of pain transmission by G-protein-coupled receptors. *Pharmacol Ther* 117: 141-161
69. SNIDER W D, MCMAHON S B 1998 Tackling pain at the source: new ideas about nociceptors. *Neuron* 20: 629-632
70. SANDKUHLER J 2009 Models and mechanisms of hyperalgesia and allodynia. *Physiol Rev* 89: 707-758
71. JENSEN T S, FINNERUP N B 2014 Allodynia and hyperalgesia in neuropathic pain: clinical manifestations and mechanisms. *Lancet Neurol* 13: 924-935
72. RAFFA R B, PERGOLIZZI J V JR. 2013 Opioid-induced hyperalgesia: is it clinically relevant for the treatment of pain patients? *Pain Manag Nurs* 14: e67-83
73. HOLTMAN J R JR., JELLISH W S 2012 Opioid-induced hyperalgesia and burn pain. *J Burn Care Res* 33: 692-701
74. HUANG J, ZHANG X, MCNAUGHTON P A 2006 Inflammatory pain: the cellular basis of heat hyperalgesia. *Curr Neuropharmacol* 4: 197-206

75. HADDAD J J 2007 On the enigma of pain and hyperalgesia: A molecular perspective. *Biochem Biophys Res Commun* 353: 217-224
76. BASBAUM A I 1999 Distinct neurochemical features of acute and persistent pain. *Proc Natl Acad Sci U S A* 96: 7739-7743
77. CAMPBELL J N, MEYER R A 2006 Mechanisms of neuropathic pain. *Neuron* 52: 77-92
78. MILLAN M J 2002 Descending control of pain. *Prog Neurobiol* 66: 355-474
79. LEE M C, WANIGASEKERA V, TRACEY I 2014 Imaging opioid analgesia in the human brain and its potential relevance for understanding opioid use in chronic pain. *Neuropharmacology* 84: 123-130
80. PARSADANIANTZ S M, RIVAT C, ROSTENE W, REAUX-LE GOAZIGO A 2015 Opioid and chemokine receptor crosstalk: a promising target for pain therapy? *Nat Rev Neurosci* 16: 69-78
81. HEINRICHER M M, TAVARES I, LEITH J L, LUMB B M 2009 Descending control of nociception: Specificity, recruitment and plasticity. *Brain Res Rev* 60: 214-225
82. FIELDS H 2004 State-dependent opioid control of pain. *Nat Rev Neurosci* 5: 565-575
83. FIELDS H L 2000 Pain modulation: expectation, opioid analgesia and virtual pain. *Prog Brain Res* 122: 245-253
84. PERTOVAARA A 2013 The noradrenergic pain regulation system: a potential target for pain therapy. *Eur J Pharmacol* 716: 2-7
85. PERTOVAARA A 2006 Noradrenergic pain modulation. *Prog Neurobiol* 80: 53-83
86. HAGELBERG N, JAASKELAINEN S K, MARTIKAINEN I K, MANSIKKA H, FORSELL H, SCHEININ H, HIETALA J, PERTOVAARA A 2004 Striatal dopamine D2 receptors in modulation of pain in humans: a review. *Eur J Pharmacol* 500: 187-192
87. MASON P 1999 Central mechanisms of pain modulation. *Curr Opin Neurobiol* 9: 436-441
88. STEIN C, CLARK J D, OH U, VASKO M R, WILCOX G L, OVERLAND A C, VANDERAH T W, SPENCER R H 2009 Peripheral mechanisms of pain and analgesia. *Brain Res Rev* 60: 90-113
89. OSSIPOV M H, MORIMURA K, PORRECA F 2014 Descending pain modulation and chronification of pain. *Curr Opin Support Palliat Care* 8: 143-151
90. CRAIG A D 2000 The functional anatomy of lamina I and its role in post-stroke central pain. *Prog Brain Res* 129: 137-151
91. BRAZ J, SOLORZANO C, WANG X, BASBAUM A I 2014 Transmitting pain and itch messages: a contemporary view of the spinal cord circuits that generate gate control. *Neuron* 82: 522-536
92. TODD A J 2010 Neuronal circuitry for pain processing in the dorsal horn. *Nat Rev Neurosci* 11: 823-836
93. KUMAR S, RASTOGI S, MAHENDRA P, BANSAL M, CHANDRA L 2013 Pain in trigeminal neuralgia: neurophysiology and measurement: a comprehensive review. *J Med Life* 6: 383-388
94. RALSTON H J, 3RD 2005 Pain and the primate thalamus. *Prog Brain Res* 149: 1-10
95. LIMA D, ALMEIDA A 2002 The medullary dorsal reticular nucleus as a pronociceptive centre of the pain control system. *Prog Neurobiol* 66: 81-108
96. APKARIAN A V, BALIKI M N, GEHA P Y 2009 Towards a theory of chronic pain. *Prog Neurobiol* 87: 81-97
97. DENK F, MCMAHON S B, TRACEY I 2014 Pain vulnerability: a neurobiological perspective. *Nat Neurosci* 17: 192-200
98. BORSOOK D, ERPELDING N, BECERRA L 2013 Losses and gains: chronic pain and altered brain morphology. *Expert Rev Neurother* 13: 1221-1234
99. ZHUO M 2015 Targeting injury-related synaptic plasticity for the treatment of chronic pain. *Curr Pharm Des* 21: 914-919
100. XU B, DESCALZI G, YE H R, ZHUO M, WANG Y W 2012 Translational investigation and treatment of neuropathic pain. *Mol Pain* 8: 15
101. MAY A 2008 Chronic pain may change the structure of the brain. *Pain* 137: 7-15
102. TRUINI A, GARCIA-LARREA L, CRUCCU G 2013 Reappraising neuropathic pain in humans—how symptoms help disclose mechanisms. *Nat Rev Neurol* 9: 572-582
103. VON HEHN C A, BARON R, WOOLF C J 2012 Deconstructing the neuropathic pain phenotype to reveal neural mechanisms. *Neuron* 73: 638-652
104. TREEDE R D, JENSEN T S, CAMPBELL J N, CRUCCU G, DOSTROVSKY J O, GRIFFIN J W, HANSSON P, HUGHES R, NURMIKKO T, SERRA J 2008 Neuropathic pain: redefinition and a grading system for clinical and research purposes. *Neurology* 70: 1630-1635
105. MARCHAND F, JONES N G, MCMAHON S B 2009 Future treatment strategies for neuropathic pain. *Handb Exp Pharmacol*: 589-615
106. FOELL J, BEKRATER-BODMANN R, FLOR H, COLE J 2011 Phantom limb pain after lower limb trauma: origins and treatments. *Int J Low Extrem Wounds* 10: 224-235
107. FLOR H, NIKOLAJSEN L, STAEHELIN JENSEN T 2006 Phantom limb pain: a case of maladaptive CNS plasticity? *Nat Rev Neurosci* 7: 873-881
108. MCCORMICK Z, CHANG-CHIEN G, MARSHALL B, HUANG M, HARDEN R N 2014 Phantom limb pain: a systematic neuroanatomical-based review of pharmacologic treatment. *Pain Med* 15: 292-305
109. ZHUO M 2012 Cortical depression and potentiation: basic mechanisms for phantom pain. *Exp Neurobiol* 21: 129-135
110. SIKANDAR S, DICKENSON A H 2012 Visceral pain: the ins and outs, the ups and downs. *Curr Opin Support Palliat Care* 6: 17-26
111. CHRISTIANSON J A, BIELEFELDT K, ALTIER C, CENAC N, DAVIS B M, GEBHART G F, HIGH K W, KOLLARIK M, RANDICH A, UNDEM B, VERGNOLLE N 2009 Development, plasticity and modulation of visceral afferents. *Brain Res Rev* 60: 171-186
112. DAVIDSON S, MOSER H, GIESLER G 2014 Ascending Pathways for Itch.
113. IKOMA A 2013 Updated neurophysiology of itch. *Biol Pharm Bull* 36: 1235-1240
114. IKOMA A, STEINHOFF M, STANDER S, YOSIPOVITCH G, SCHMELZ M 2006 The neurobiology of itch. *Nat Rev Neurosci* 7: 535-547
115. SCHMELZ M 2010 Itch and pain. *Neurosci Biobehav Rev* 34: 171-176
116. LAMOTTE R H, DONG X, RINGKAMP M 2014 Sensory neurons and circuits mediating itch. *Nat Rev Neurosci* 15: 19-31
117. TRACEY I 2011 Can neuroimaging studies identify pain endophenotypes in humans? *Nat Rev Neurol* 7: 173-181
118. PERKINS J R, LEES J, ANTUNES-MARTINS A, DIBOUN I, MCMAHON S B, BENNETT D L, ORENGO C 2013 Pain-Networks: a web-based resource for the visualisation of pain-related genes in the context of their network associations. *Pain* 154: 2586 e2581-2512
119. SAAB C Y 2012 Pain-related changes in the brain: diagnostic and therapeutic potentials. *Trends Neurosci* 35: 629-637
120. MAY A 2011 Structural brain imaging: a window into chronic pain. *Neuroscientist* 17: 209-220