# Occurrence of *Listeria monocytogenes* in fish and fishery products

Rožman<sup>1</sup>, J., B. Njari<sup>2</sup>, L. Kozačinski<sup>2</sup>

Original scientific paper

### SUMMARY

Listeriosis is an infection that is caused by eating food contaminated by bacteria Listeria monocytogenes, which represents one of the major public health issues and mostly occurs during the transport of food worldwide. It is primarily associated with the consumption of finished products. This paper outlines our findings of bacteria L. monocytogenes in analysed samples of fresh fish (sea bass) and fishery products (smoked and marinated fish, gilt-head bream and sea bass). We also examined swabs taken from work surfaces and hands of fish-processing plant employees. Bacteria L. monocytogenes were not present in samples of fresh, smoked and marinated fish. However, bacteria L. inocua were present in samples of marinated and smoked fish. Although bacteria L. monocytogenes were not present in samples of swabs, bacteria L. inocua were present in swabs taken from work surfaces during handling, grading and processing of fish, i.e. in areas designated for evisceration and the control of fish processing, as well as on the hands of employees who operated the process for the filleting of fish.

Key words: L. monocytogenes, L. inocua, fish, fishery products, plant hygiene

# **INTRODUCTION**

The persistent issue of food safety in practice includes potential public health problems related to medical conditions in humans that are caused by consuming foods contaminated by bacteria. According to World Health Organization data, in addition to bacteria of the genera Salmonella, Listeria monocytogenes is considered the most significant public health problem. According to the European Food Safety Agency (EFSA) report, European Union Member States have in 2013 reported 1,763 cases of listeriosis and a total of 191 deaths, with 64 deaths recorded in France alone (Anon., 2015). Such infection can either manifest as an epidemic or sporadically. It is primarily associated with the consumption of "ready to eat" (RTE) products whose shelf life is extended at refrigerator temperatures. The occurrence of the infection is usually associated with dairy and meat products in both fish and minimally

processed fishery products (smoked fish). Accordingly, 17 EU Member States have reported findings of *L. monocytogenes* in fishery products in 2013. *L. monocytogenes* was found in 1.6 % of the total of 1,649 samples of various fishery products. Just as in previous years, *L. monocytogenes* was more often determined in RTE fish (usually smoked fish).

*L. monocytogenes* is a microorganism that is widely distributed in nature. It can be found in plant tissue, soil and water, as well as in infected animals. Infected animals represent a major route of transmission to humans. It is mostly transmitted to humans and animals by consuming contaminated food or feed. It grows best in cold conditions and not only tolerates drought and higher levels of salinity, but is able to survive mild heat treatment (Helwigh et al., 2012). *L. monocytogenes* can be present in different food products. Microbiological contamination of finished products is often associated with either the contamination of the product itself or

1 Jelena Rožman, student, University of Zagreb, Faculty of Agriculture

2 Ph. D. Bela Njari, Professor; Ph. D. Lidija Kozačinski, Professor; University of Zagreb, Faculty of Veterinary Medicine, Department of Hygiene, Technology and Food Safety, Heinzelova 55, Zagreb

Correspondence author: klidija@vef.hr

### Scientific and professional section

irregularities in production process. In the case of *L. monocytogenes*, the high-risk group comprises of fresh, unprocessed foodstuffs such as milk, meat, cream cheeses, as well as fish and fishery products, including frozen fish, cold and hot smoked fish, marinated fish, fermented fish and fish salads (Kozačinski et al., 2000; Dominguez et al., 2001; González-Rodríguez et al., 2002; Gombas et al., 2003; Popović and Đurđević-Milošević, 2008; Papadopoulos et al., 2010. Tocmo et al., 2014; Kuzmanović et al., 2011).

Fishery products can be easily polluted (contaminated) by *L. monocytogenes* during production. Raw fish can be an important source of pollution on equipment and in facilities that will result in the contamination of products (Miettinen and Wirtanen, 2005). Both evisceration and cleaning of fish before retail can cause the spread of bacteria in the production facility that will, in turn, result in cross-contamination of fish, equipment, employees and the environment in general (Papadopoulos et al., 2010).

This paper will therefore present the results of examination for the presence of *L. monocytogenes* that was conducted on samples of fresh fish (sea bass) and fishery products (smoked sea bass and gilt-head bream). Since these bacteria are ubiquitous, we shall analyse the results of their findings in fish processing facilities and try to compare the incidence of reported bacteria findings in fresh fish and nature to the incidence of reported bacteria findings in fishery products. This paper shall also present a part of results obtained within the framework of the project FP7 Selection and Improving of Fit - For - Purpose Sampling Procedures for Specific Foods and Risks – BASELINE Grant Agreement number 222738; 2009 – 2013 (Faculty of Veterinary Medicine Team Leader Ph. D. Lidija Kozačinski, Professor).

## MATERIALS AND METHODS

This paper outlines our findings of bacteria *L. mo-nocytogenes* in analysed samples of fresh fish and fishery products. We examined a total of 21 samples, namely nine samples of fresh fish (sea bass), six samples of marinated fish (three samples of marinated sea bream and three samples of marinated sea bass) and six samples of smoked fish (three samples of smoked sea bream and three samples of smoked sea bass). Our study has also investigated a part of the results obtained by bacteriological examination of fish, namely sea bass (n = 39) obtained within the framework of the project BASELINE, and the results of the evaluation of microbiological purity of fish processing facilities related to findings of *L. monocytogenes* (n = 40).

Provided samples of fish and fishery products were

delivered to the Department of Hygiene, Technology and Food Safety after due production process, at the moment they were ready to be distributed on the market. Samples were delivered in a portable freezer without damaging the packaging. Fresh fish (sea bass) weighed about 300 g and was packaged in modified atmosphere. Marinated sea bass and gilt-head bream were packaged in containers; each weighing 100 g. Marinated fish was filleted. Smoked sea bass and gilt-head bream fillets were also packaged in packs weighing about 100 g each.

Activities within the framework of the project BA-SELINE included two deliveries of fresh sea bass crates. Samples were collected after fishing and after primary handling (evisceration) in fish processing facility, prior to packaging for retail. In the second sampling, samples of sea bass were collected on the fish market. During the collection of fish samples in processing facilities, we also took swabs from work surfaces and hands of fish-processing plant employees. Both fish samples and swabs were examined only for the presence of *L. monocytogenes*.

In accordance with the standard HRN EN ISO 11290-2:1999 (Microbiology of food and animal feeding stuffs - Horizontal method for the detection and enumeration of *Listeria monocytogenes* - Part 1: Enumeration method) (HRN EN ISO 11290-1:1999 + HRN EN ISO 11290-1:1999/A1:2008), all samples of fish and products were screened for the presence of *L. monocytogenes* per 25 g. *L. monocytogenes* was identified using a commercially available API Listeria test kit, as prescribed by the manufacturer (Biomerieux).

# **RESULTS AND DISCUSSION**

The results of bacteriological examination of fish and products, as well as the results of *L. monocytogenes* findings among analysed samples of fish and products are presented in Table 1. *L. monocytogenes* was not identified in any of the examined samples of fish and fishery products. However, during the biochemical identification, bacteria *L. inocua* were confirmed in all analysed samples of fishery products (n = 12).

**Table 1:** Results of examination for the presence of L. monocytogenes in fish and fishery products

Sample	n	L. monocytogenes / 25 g	Note
Fresh sea bass	9	Neg.	-
Maaimated gilt-head	3	Neg.	<i>Listeria inocua</i> / 25 g = Pos.
Marinated sea bass	3	Neg.	<i>Listeria inocua</i> / 25 g = Pos.
<b>Sneeke</b> d gilt-head	3	Neg.	<i>Listeria inocua</i> / 25 g = Pos.
Smoked sea bass	3	Neg.	<i>Listeria inocua</i> / 25 g = Pos.

#### Scientific and professional section

**Table 2:** Rezultati pretrage svježe ribe (brancin) na prisutnost bakterije

 L. monocytogenes

Uzork	1	
Pogon prerade ribe	Maloprodaja	L. monocytogenes/25 g
8	31	neg.

Table 2 presents the results of microbiological analysis of fresh cultivated fish (sea bass) samples collected in dispatch centres and retail facilities. Bacteria *L. monocytogenes* were not identified in any of the examined samples.

*L. monocytogenes* was not identified in any of the examined swabs taken from work surfaces (Table 3) and hands of employees. However, *L. inocua* was identified in three instances of both samplings, namely in swabs taken from work surfaces designated for handling, gra-

**Table 3:** Results of microbiological examination of work surfaces and hands

 of employees in fish processing facilities

Sampling site		Sampling 1			Sampling 2		
		Lm	Li	n	Lm	Li	
Work surface - evisceration Work surface - fish handling		Neg.	Neg.	2	Neg.	Pos.	
		Neg.	Pos.	2	Neg.	Neg.	
Work surface - fish grading	2	Neg.	Pos.	2	Neg.	Neg.	
Work surface - fish filleting	2	Neg.	Pos.	2	Neg.	Neg.	
Work surface - fish salting	2	Neg.	Neg.	2	Neg.	Neg.	
Work surface - control of fish processing Hands of workers - evisceration Hands of workers - filleting Hands of workers - grading Hands of workers - control		Neg.	Neg.	2	Neg.	Pos.	
		Neg.	Neg.	2	Neg.	Neg.	
		Neg.	Neg.	2	Neg.	Pos.	
		Neg.	Neg.	2	Neg.	Neg.	
		Neg.	Neg.	2	Neg.	Neg.	
Total	20	0	3	20	0	3	

Lm = Listeria monocytogenes; Li = Listeria inocua

ding and processing of fish, i.e. areas designated for evisceration, control of fish processing, and hands of employees who operated the process for the filleting of sea bass in second sampling.

Research results presented in Table 1 and 2 demonstrate that bacteria *L. monocytogenes* were not identified in analysed samples of fresh fish that were sampled in fish processing facility prior to placement on the market, or in fish samples collected on the fish market, or in samples of smoked and marinated fish. Although other authors identified *L. monocytogenes* in fresh fish and fishery products - smoked and marinated fish (Jeyasekaran et al., 1996; Dominguez et al., 2001; González-Rodríguez et al., 2002; Papadopoulos et al., 2010; Dimitijevic et al., 2011; Tocmo et al., 2014; Lakicevic et al., 2015), samples in our research tested negative. However, bacteria *L. inocua* were present in samples of marinated and smoked fish (Table 1). Obtained results reflect the research conducted by other authors that identified bacteria L. inocua in fishery products (Jeyasekaran et al., 1996; Dominguez et al., 2001). Moreover, these bacteria are often present along with L. monocytogenes and have very similar atypical haemolytic strains (Petran and Swanson, 1993; Johnson et al., 2004; Moreno et al., 2012). Findings of L. inocua in our research reflect the research conducted by Kuzmanović et al. (2011) that identified L. inocua in 8.51% of examined fish samples. Our research results (Table 1) confirmed the occurrence of subsequent contamination in fish processing facilities that can according to Papandopoulos et al. (2010) be considered a by-product of fish processing (evisceration, removal of gills, scaling, filleting). Above all, such findings reveal the need for additional caution, primarily because the transfer of L. inocua in production facilities resembles the transfer of L. monocytogenes. Since these bacteria are ubiquitous, they can survive in extreme conditions such as high pH environments, both high and low temperatures, as well as high levels of salinity (Moreno et al., 2012). Although they are very similar to L. monocytogenes, they are not considered a pathogenic bacterial species. Further research on the source and manner of bacterial contamination in fish processing facilities is needed. Upon considering the research conducted by Vitt et al. (2008) and Sabanadesan et al. (2000) on the effects of smoke, i.e. type of liquid smoke preparations and the duration of smoking, we concluded that additional attention should also be paid to parameters of processing fish fillets with smoke.

L. monocytogenes was not identified in samples of the examined swabs taken from work surfaces and hands of employees (n = 40) (Table 3). Bacteria L. inocua were identified in three instances of both samplings conducted on work surfaces during handling, grading and processing of fish, i.e. in areas designated for evisceration and the control of fish processing, as well as on the hands of employees who operated the process for the filleting of sea bass (15 % of examined swabs). If we compare the occurrence of L. inocua on work surfaces in fish processing facilities (Table 3) to conclusions reached by Papandopoulos et al. (2010), we can consider the contamination of marinated and smoked fish by L. inocua a result of the occurrence of such bacteria in the environment. Jørgensen et al. (1998) reported that all examined samples of smoked fish in processing facilities involved in their study tested positive for L. monocytogenes. The fact that L. monocytogenes has not been identified in either fish or products involved in our study does not eliminate a possibility of subsequent contamination in any latter production stage

pertaining to handling, salting, filleting or smoking of fish. Furthermore, Miettinen and Wirtanen (2005) claim that raw fish can be an important source of pollution on equipment and machines, which will result in the contamination of products. In addition, Lakicevic et al. (2015) found that 2.3 % of swabs obtained from production facility samples tested positive for L. monocytogenes. Unlike our study, the study conducted by Dimitrijevic et al. (2011) identified *L. monocytogenes* in 7 out of 99 examined swabs (7.0 %) obtained from work surfaces and the hands of employees in fish processing facilities. Authors have identified *L. monocytogenes* in 3 out of 66 examined swabs (4.5 %) in the second production facility.

Findings of any bacterial species and, in particular, L. monocytogenes, as well as findings of L. inocua (Table 3) during handling, control of fish processing and on hands of employees therefore indicate a strong need for the monitoring and control of hygiene in due facilities. According to the provisions of the Ordinance on the Frequency of Controls and Standards for Microbiological Purity in Facilities under Sanitary Supervision (Official Gazette No. 137/09), assessments of microbiological purity of the facility for the production of food must be undertaken. The number of required samples depends on the type and purpose of such facility, and microbiological purity is assessed based on the number of aerobic mesophilic bacteria and enterobacteria revealed in results of due bacteriological examinations. The results of our study confirm that cleaning, washing and disinfection should be increased, while bacteriological examinations of not only finished products but also swabs that were obtained within the framework of internal control activities should be introduced, all in line with due recommendations for the reduction of Listeria bacteria findings in production facilities (Gram, 2004). Since Listeria bacteria are ubiquitous, they can survive in different environmental conditions and can hide in brine, cutting machines, areas designated for filleting of fish, work surfaces, floors, etc. We therefore emphasize authors' notes that point out that in order to eliminate bacteria that come in contact with food from production facilities and work surfaces, implemented programmes of good production and hygienic practices must be based on the specific sampling of due environments and obtained evidence of bacteria found in facilities (Rorvik et al., 1995; Autio et al., 1999; Bagge Ravn et al., 2003; Dimitrijevic et al., 2011).

## CONCLUSION

The results of bacteriological examination revealed that samples of fresh fish (sea bass), smoked (gilt-he-

ad bream and sea bass) and marinated fish (gilt-head bream and sea bass) did not test positive for L. monocytogenes. According to the provisions of the regulations on microbiological criteria for foodstuffs (Regulation No. 2073/2005), examined samples of fish and fishery products can be considered satisfactory and microbiologically safe. However, bacteria L. inocua were present in samples of fishery products. Furthermore, L. inocua was also identified 6 sampled swabs obtained from fish processing facilities. The results of microbiological analysis of swabs collected from fish processing facilities suggest the violation of hygiene standards and good hygienic practices in such facilities. Recommendations aiming at reducing the number of Listeria occurrences indubitably include extensive cleaning, washing and disinfection of facilities, as well as effective implementation of bacteriological examinations intended for not only finished products but also swabs obtained within the framework of internal control activities.

**Acknowledgements:** This paper also presented a part of results obtained within the framework of the project FP7 Selection and Improving of Fit - for - Purpose Sampling Procedures for Specific Foods and Risks – BASELINE Grant Agreement number 222738; 2009 – 2013.

# REFERENCES

**Anonimno (2015):** Scientific report of EFSA and ECDC. The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2013. EFSA Journal 13(1):3991

Autio T., S. Hielm, M. Miettinen, A.-M Sjoberg, K. Aarnisalo, J. Bjorkroth, T. Mattila-Sandholm, H. Korkeala (1999): Sources of Listeria monocytogenes contamination in a cold-smoked rainbow trout processing plant detected by pulsed-field gel electrophoresis typing. Appl. Environ. Microbiol. 65, 150–155

**Bagge-Ravn D., K. Gardshodn, L. Gram, B. Fonnesbech Vogel (2003):** Comparison of fog and foam sanitizing procedures in a salmon smokehouse with respect to the general hygienic level and survival of Listeria monocytogenes. J. Food Prot. 66, 592–598

Dimitrijević, M., R. C. Anderson, N. Karabasil, N. Pavličević, S. Jovanović, J. Nadaljeković trajlović, V. Teodorović, M. Marković, S. Dojčinović (2011): environmental prevalence and persistence of Listeria monocytogenes in cold-smoke trout processing plants. Acta Veterinaria (Beograd) 61,4, 429-442.

**Dominguez, C., I. Gomez, J. Zumalacarregui (2001):** Prevalence and Contamination Levels of Listeria monocytogenes in Smoked Fish and Pâté Sold in Spain. J. Food Protection 12, 2075-2077(3)

Gombas, D.E., Y. Chen, R.S. Clavero, V.N. Scott (2003): Survey of Listeria monocytogenes in Ready-to-Eat Foods. J. Food Prot. 66, 570–577

González-Rodríguez M.N., J.J. Sanz, J. A. Santos, A. Otero. M.L. García-

#### Scientific and professional section

**López (2002):** Foodborne pathogenic bacteria in prepackaged fresh retail portions of farmed rainbow trout and salmon stored at 3°C. Int J Food Microbiol 76:135–141

Gram, L. H.H. Huss (2000): Fresh and processed fish and shellfish. In the Microbiological Safety and Quality of Food. B. M. Lund, A. C. Baird – Parker and G. W. Gould, editors. Chapman and Hall., London, U. K., pp 472-506.

Jeyasekaran, G., I. Karunasagar, I. Karunasagar (1996): Incidence of Listeria spp. in tropical fish. Int. J. Food Microbiol. 31, 1–3, 333–340

Johnson, J., K. Jinneman, G. Stelma, B.G. Smith, D. Lye, J. Messer, J. Ulaszek, L. Evsen, S. Gendel, R.W. Bennet, B. Swaminathan, J. Pruckler, A. Steigerwalt, S. Kathariou, S. Yildirim, D. Volokhov, A. Rasooly, V. Chizhikov, M. Wiedmann, E. Fortes. R.E. Duvall, A.D. Hitchins (2004): Natural atypical Listeria innocua strains with Listeria monocytogenes pathogenicity island 1 genes. Appl Env Microbiol 70, 4256–4266.

Jørgensen L.V. H.H. Huss (1998): Prevalence and growth of Listeria monocytogenes in naturally contaminated seafood. Int. J. Food Microbiol. 42, 127–131

Kozačinski Lidija, M. Hadžiosmanović, B. Mioković, B. Njari, Darinka Pranjić i Željka Cvrtila (2000): The meaning of Listeria monocytogenes in veterinary-sanitary inspection. Vet. arhiv, 70 (suppl.), S7-S13.

Kuzmanović, J., R. Ašanin, M. Baltić, D. Mišić. M. Dimitrijević, M. Stojanović, N. Ašanin I. Kovačević (2011): Prisustvo Listerija vrsta u uzorcima riba, proizvoda od ribe i morskih plodova. Acta veterinaria 61, 2-3, 193-203, doi:10.2298/AVB1103193K

Lakicevic, B., T. Baltic, V. Jankovic, B. Velebita, M. Dimitrijevic, N. Parunovic, D. Milicevic (2015): Occurrence of Listeria monocytogenes in a Serbian salmon and seafood processing line during 2013. Procedia Food Science 5, 156 – 159

Miettinen H., G. Wirtanen (2005): Prevalence and location of Listeria monocytogenes in farmed rainbow trout. Int J Food Microbiol 104, 135–143 Moreno, L. Z., R. Paixao, D.D. Gobbi, D. C. Raimundo, T.P. Ferreira, E. Hofer, M.H. Matte, A.M. Moreno (2012): Characterization of atypical Listeria innocua isolated from swine slaughterhouses and meat markets. Res Microbiol 163, 268–271.

Papadopoulos T., A. Abrahim, D. Sergelidis, I. Kirkoudis, K. Bitchava (2010): Prevalence of Listeria spp. in freshwater fish (Oncorhynchus mykiss and Carassius gibelio) and the environment of fish markets in Northern Greece. J Hell Vet Med Soc 61:15–22

Petran, R.L., K. M. J. Swanson (1993): Simultaneous growth of Listeria monocytogenes and Listeria innocua. J. Food Prot. 56, 616-618.

Popović G., D. Đurđević-Milošević (2008): Prisustvo bakterija Listeria monocytogenes u namirnicama i prateći rizik za zdravlje potrošača, Zbornik naučnih radova Instituta PKB Agroekonomik, vol. 14, 151-159.

Rorvik L., D. Caugant, M. Yndestad (1995): Contamination pattern of Listeria monocytogenes and other Listeria spp. in a salmon slaughterhouse and smoked salmon processing plant. Int. J. Food Microbiol. 25, 19–27

Sabanadesan, S., A.M. Lammerding, M.W. Griffiths (2000): Survival of Listeria innocua in Salmon following Cold-Smoke Application. J. Food Prot. 6, 715-720

Tocmo R., K. Krizman, W.J. Khoo, L.K. Phua, M. Kim, H.G. Yuk (2014): Listeria monocytogenes in vacuum-packed smoked fish products: occurrence, routes of contamination, and potential intervention measures. Compr Rev Food Sci F13:172–189

Vitt, S.M., B.H. Himelbloom, C.A. Crapo (2007): Inhibition of Listeria innocua and L. monocytogenes in a laboratory medium and cold-smoked salmon containing liquid smoke. J. Food Safety 21, 111-125

Received: 10.3.2016.

Accepted: 31.3.2016.

