CCA-1622

YU ISSN 0011-1643 UDC 547.92 Original Scientific Paper

Steroidal Analogue of Deoxyvernolepin. Synthesis of δ -Lactone Key Intermediate

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Received February 7, 1985

Sesquiterpene lactones of elemanolide type vernolepin 1 and deoxyvernolepin 2 exhibit a pronounced antitumor activity against the Walker intramuscular carcinosarcoma 256 and CCRF — CEM human lymphoblastic leukemia cells in culture and were therefore, subjects of many partial and total syntheses. The synthesis of the steroidal analogue of deoxyvernolepin

The synthesis of the steroidal analogue of deoxyvernolepin 3, i.e., its key intermediate, δ -lactone 28-hydroxy-2-oxa-5 α ,10 α -cholest-6-en-3-one 4 starting from cholesterol is the subject of this report.

Known reactions, as well as modified ones, were used in the synthesis. A new reaction for the preparation of dimethyl acetals with pyridinium chlorochromate and methanol was also developed. It was found that the acetalization reaction using pyridinium chlorochromate and methanol could be performed without affecting an acid-sensitive functional groups such as the oxirane ring.

During the last 30 years more than 800 sesquiterpene lactones have been isolated and identified from various plant species of *Compositae* and *Umbelliferae*, as well as from *Lauraceae*, *Burserceae*, *Magnoliaceae* and *Hepaticeae*.^{1,2}

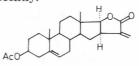
It was found that the pronounced actineoplastic (antitumor) activity in vitro, seldom in vivo, of sesquiterpene lactones is due to the α -ethylene- γ and/or δ -lactone ring and the stereochemistry of epoxide, hydroxy and other functional groups in their molecule.²⁻⁷

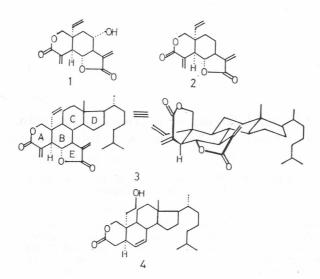
Bis- α -methylene sesquiterpene lactones of elemanolide type vernolepin $1^{3,5^{-10}}$ and deoxyvernolepin 2^{11-13} (Figure 1) exhibit a pronounced antitumor activity against the Walker intramuscular carcinosarcoma 256⁸ and CCRF — CEM human lymphoblastic leukemia cells in culture¹² and were, therefore, subjects of many partial and total syntheses.^{11,13-22}

These facts encouraged us to synthesize the steroidal analogue of deoxyvernolepin 3, i.e., its δ -lactone key intermediate, 28-hydroxy-2-oxa--5 α ,10 α -cholest-6-en-3-one 4, starting from cholesterol*.

* Part of Ph. D. thesis of B. Š.

* The steroidal $\gamma\text{-lactone 5}$ which exhibits antitumor activity against HeLa cells has been synthesized recently.^23





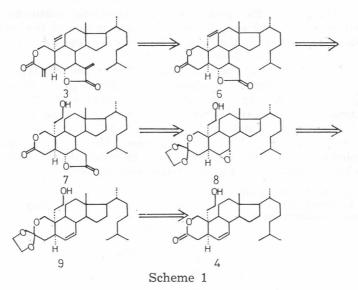
Retrosynthetic analysis of steroidal dilactone 3 (Scheme 1) points to the need for the following transformations of the cholesterol framework:

— simultaneous introduction of exocyclic double bonds into $\alpha\text{-position}$ of the $\gamma\text{-}$ and $\delta\text{-lactone rings,}^{24}$

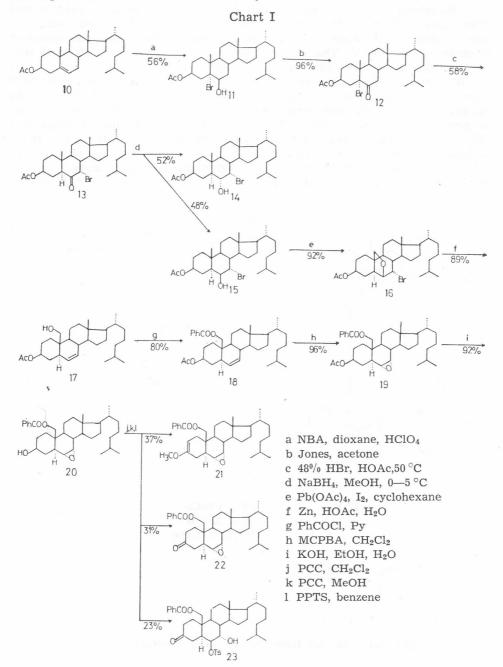
— synthesis of the γ -lactone ring (ring E) by the opening of the 6α , 7α -epoxide ring with a suitable nucleophile,

— synthesis of the δ -lactone ring (ring A) by ozonolysis of Δ^2 -double bond and subsequent acification of the obtained product and

— conversion of primary alcohol into terminal double bond (the vinyl group) *via* the corresponding arylselenyl derivative.



The synthesis should envisage an intermediate with cis-5 α ,10 α -fused A and B rings because the nucleophilic attack on the 6 α ,7 α -epoxide ring of the cholestane (5 α ,10 β -) derivative yields the inappropriate trans-6 β ,7 α -diaxial product.^{28,29} It was expected that the regiospecific β -attack of the nucleophile (dilithio acetate,^{16,18,19} t-butyl dilithioacetoacetate²⁰ or some other



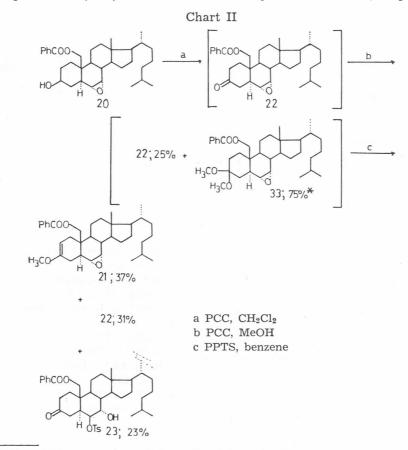
appropriate nucleophile) on the C-7 carbon atom of compound 8 should give $trans-\gamma$ -lactone upon acidification.

In this report, beside the δ -lactone 4, the synthesis of 2-oxa-5 α ,10 α --cholest-6,19-dien-3-one 29 via the corresponding selenide 28 is described (Chart III).

Cholesteryl acetate 10 was treated with HOBr in dioxane solution and bromohydrin 11 was obtained in $56^{0}/_{0}$ yield³⁰ (Chart I). Bromohydrin 11 was oxidized with Jones reagent in acetone yielding the corresponding bromoketone 12 (960/₀). The latter compound was further transformed into its 7 α -isomer with 480/₀ HBr in acetic acid³¹ in 580/₀ yield.

Reduction of bromoketone 13 with sodium borohydride in methanol³² gave 7α -bromo- 5α -cholestan- 3β , 6β -diol 3-monoacetate 15 (48%), which was further cyclized with lead tetraacetate and iodine in cyclohexane solution to give 6β ,19-epoxide 16 (92%).^{30,33-36} The 6β ,19-epoxide ring cleavage was effected with zinc in acetic acid and water (89%) and the obtained primary alcohol 17 protected by esterification with benzoyl chloride in pyridine (80%).

The epoxidation of 18 with *m*-chloroperoxybenzoic acid gave only the $6\alpha,7\alpha$ -epoxide 19 (96%) which was further hydrolized into $6\alpha,7\alpha$ -epoxy-5\alpha-



* The yield was estimated from the ¹H NMR spectrum.

-cholestan-3β,19-diol 19-monobenzoate 20 under mild conditions (0.1 M KOH, H_2O) in $92^{0}/_{0}$ yield.

Synthesis of enol ether 21 was accomplished via the intermediate dimethyl acetal 33 (Chart II).

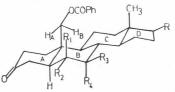
The intermediate compound 33 was prepared from alcohol 20, which was oxidized with pyridinium chlorochromate (PCC) into the corresponding ketone 22. The latter compound was dissolved in hot methanol together with »black gum«³⁷ and 3,3-dimethyl acetal 33 was obtained in 75% yield*, according to ¹H NMR spectrum (not isolated).

In order to establish the reaction mechanism of this facile route for the preparation of dimethyl acetals³⁹ from alcohols or ketones, we treated the previously prepared 6α , 7α -epoxy-19-benzoyloxy- 5α -cholestan-3-one 22 with boiling methanol in the presence of pyridinium ion (originated from PyH+Cl⁻ or pyridinium p-toluenesulphonate (PPTS)), but without success. However, when the same ketone was treated with methanol and PCC as catalyst, dimethyl acetal 33 was obtained in almost the same yield as in the case when the intermediate ketone was not isolated. Thus we presume that the Cr⁴⁺ ion takes part in this reaction**.

This new method for the preparation of dimethyl acetals directly from alcohols, without isolation of intermediate ketones was presented in our previous paper.38

The mixture of 22 and 33, which could not be separated on a silica gel column, was treated with PPTS in boiling benzene^{13,39} to give the expected enol ether 21 (37%), ketone 22 (31%) and 7a-hydroxy-6β-p-toluenesulphonyloxy-19-benzoyloxy-5a-cholestan-3-one 23 (23%), a product of nucleophilic attack of tosyloxy anion on the C-6 carbon atom of 6α , 7α -epoxide ring (Chart II).

Inspection of Dreiding models and the ¹H NMR data of 22, 23 and 34 (Figure 2) point to chair conformation of the B ring of p-toluenesulphonate 23 in spite of the 10 β -position of the bulky —CH₂OCOPh group.



34 $R_1 = R_2 = R_3 = R_4 = H$ 22 $R_1 = R_3 = H$ $R_2 + R_4 = 0$ 23 $R_1 = OTs$

> $R_2 = R_3 = H$ $R_4 = OH$ $R = C_8 H_{17}$

2H-19 δ 4.72, s⁴⁰ H-6 δ 2.84, d, $J_{6.7} = 4$ Hz H-7 δ 3.16, dd, $J_{7.6} = 4$ Hz, $J_{7.8} = 2.5$ Hz δ 4.49 and δ 4.71, AB syst., J = 12 Hz 2H-19 δ 4.36, dd, $J_{6.5} = 3.75$ Hz, $J_{6.7} = 2$ Hz H-6 H-7 δ 3.91, m, W_{1/2} = 7.5 Hz 2H-19 δ 4.53 and δ 4.63, AB syst., J = 13 Hz

Figure 2

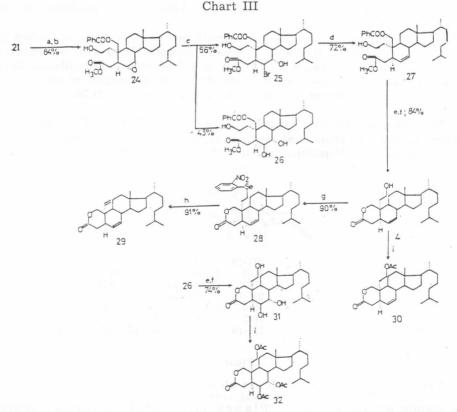
* See experimental part. It is not clear what are the reduction products of the Cr^{6+} ion in these reactions; several authors proposed that Cr^{4+} might be one. For further information see. G. Piancatelli and M. D'Aruia, Synthesis, (1982) 245 and references therein.
 ** Analogous results were obtained with 5α-cholest-3-one, 6β,19-epoxy-5α-

⁻cholestan-3-one and cholest-5-en-3-one.

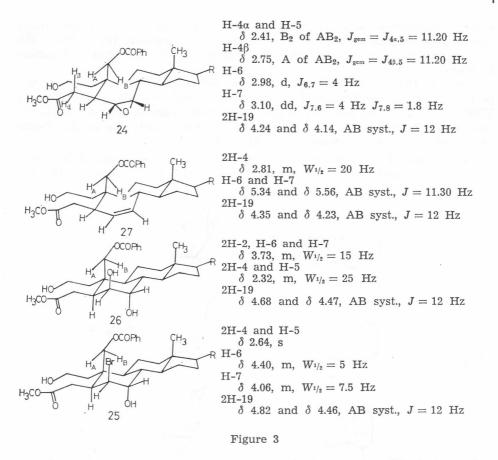
Coupling constants of $J_{6,5} = 3.75$ Hz and $J_{6,7} = 2$ Hz are in agreement with the proposed conformation.

Methylene 2H-19 protons of keto-benzoate 34 appear as a singlet at δ 4.72⁴⁰ as a consequence of the diamagnetic anisotropic effect of the 3-keto group (chemical shift of methylene R—CH₂OCOPh protons in δ 4.20⁴¹).^{42a-f} However, one of the H-19 protons of 6 α ,7 α -epoxy-19-benzoyloxy-5 α -cholestan-3-one 22 is shifted to upper field. Thus the chemical shifts of δ 4.71 for H_A-19 and δ 4.49 for H_B-19 are assigned. Both methylene protons of tosylate 23 are shifted to the upper field and their chemical shifts are H_A-19 δ 4.63 and H_B-9 δ 4.53.

Ozonolysis of the enol ether 21 at -78 °C in dichloromethane-methanol solution and subsequent reduction with sodium borohydride of intermediate hydroperoxide gave methyl-6 α ,7 α -epoxy-2-hydroxy-19-benzoyloxy-2,3-seco-5 α -cholestan-3-oate 24 (84%); Chart III). The 6 α ,7 α -epoxide ring was treated with 48% hydrobromic acid in dioxane to give bromohydrin 25 (56%) and triol 26 (43%). Surprisingly the chair conformation was established for compound 25 on the basis of the ¹H NMR data (Figure 3).



a O₃, CH₂Cl₂, MeOH, -78 °C; b NaBH₄, -20 °C, HOAc; c $48^{0/0}$ HBr, dioksan, H₂O; d Zn, HOAc, H₂O; e KOH, MeOH, H₂O; f conc. HCl; g $o-O_2NC_6H_4SeCN$, THF, $n-Bu_3P$; h $30^{0/0}$ H₂O₂, THF; i Ac₂O, Py.



Shold the B ring of 25 exist in one of the two possible boat conformations (bot unfavourable because of strong 1,2-diequatorial interaction of the 6 β -bromine atom and 7 α -hydroxy group), the H-7 proton will appear as dd with $J \approx 10-12$ Hz, or as a singlet. However, $W_{1/2} = 7.5$ Hz was found for the H-7 proton, which together with a great downfield shift of both H-19 protons* (Table I) confirms the proposed conformation of the B ring.

Finally, the δ -lactone 4 was prepared from bromohydrin 25 by reductive introduction of the Δ^6 -double bond with zinc in acetic acid and water (72%).

TABLE	Ι
	_

		$\mathbf{H}_{\mathtt{A}}$	H_{B}	anteng Ri
24	2H-19	δ 4.24	δ 4.14	$\Delta v_{AB} = 8.0$ Hz
27	2H-19	δ 4.35	δ 4.23	$\Delta v_{AB} = 9.6$ Hz
26	2H-19	δ 4.68	δ 4.47	$\Delta v_{\rm AB} = 16.8$ Hz
25	2H-19	δ 4.82	δ 4.46	$\Delta v_{AB} = 28.8 \text{ Hz}$

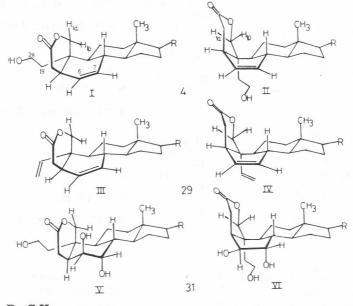
* Characteristic influence of axial 6β-substituents.42e,42f

alkaline hydrolysis and subsequent acidification with concentrated hydrochloric acid without isolation of the intermediate compound $(84^{0}/_{0};$ Chart III).

An analogous reaction scheme was applied to the synthesis of δ -lactone 31 from triol 26.

Introduction of the vinyl group to the 10α -position of the lactone 4 was performed with o-nitrophenyl selenocyanate and tri-*n*-butylphosphine in THF^{26,27} and the obtained yellow 28-o-nitrophenylseleno-2-oxa-5 α ,10 α -cholest-6-en-3-one 28 (90%) was treated with 30% hydrogen peroxide to give 2-oxa-5 α ,10 α -cholest-6,19-dien-3-one 29 in 91% yield.

Inspection of Dreiding models suggested that the synthesized δ -lactones 4, 28, 29, 31, and acetates 30 and 32 may exist *i.e.* in one of the two



 $R = C_8 H_{17}$

Figure 4

conformations^{*} shown in Figure 4. Analysis of the ¹H NMR data of the obtained lactones (Table II) indicates that conformations I, III and V of 4, 29 and 31, respectively, can be found in CDCl₃ or acetone- d_6 solutions. Distorted chair conformation of the δ -lactone ring is proposed on the basis of the great downfield shift of H-1 β protons (δ 4.37, δ 4.39 and δ 4.89) caused by the diamagnetic anisotropic effect of the carbonyl group of the lactone ring on H-1 β proton.

The great downfield shift of the H-1 β proton of δ -lactone 31 (δ 4.89) is indicative of an additional effect of the axial 6 β -hydroxy group.

The half-chair conformation of the B ring of 4, 28 and 29 is proposed on the basis of $J_{6,7} = 10$ Hz, $J_{6,5} = 1.2$ Hz and $J_{6,8} = 1$ Hz. In the case of the boat conformation of the B ring (structures II and IV; Figure 4) the coupling constant $J_{6,5} \approx 8$ —10 Hz should be found.

* Twisted congormations could also be taken into consideration.

	3.83 B of AB, J=12	3.84 B of AB, J=12	3.86, dd $J_{gem} = 11.25, J = 2.5$	4.01, dd J_{gem} =11.25, J =2.5	3.84 B of AB, J=12	
Η-1β	4.37 A of AB, J=12	4.39 A of AB, J=12	$rac{4.89}{J_{ m gem}} = 11.25$	$4.76 J_{gem} = 11.25$	4.41 A of AB, J=12	4.39, s
H-4	2.80, m, W _{1/2} =25	2.78, m, W _{1/2} =25	2.81, m, $W_{1/2} = 26.25$	2.86, m, W _{1/2} =41.25	2.86, m, $W_{1/2}$ =41.25 2.83, m, $W_{1/2}$ =28.75	
H-5	2.29, m, W1/1=31.25		2.32, m, $W^{1/2}$ =31.252.36, m, $W^{1/4}$ =15	2.31, m, $W_{1/2} = 10$	2.33, m, $W_{1/2}$ =26.25	2.56, m, $W_{1/2} = 50$
9-Н	$5.43, \text{ ddd}, J_{6,7}=10$ $J_{6,5}=1.2, J_{6,8}=1$	$5.43, \text{ddd}, J_{6.7}=10$ $J_{6.5}=1.2, J_{6.8}=1$		5.05, t, $J=3.25$	5.48, ddd, $J_{6,7} = 10$ $J_{6,5} = 1.2, J_{6,8} = 1$	5.48, ddd, $J_{6,7} = 10$ $J_{6,5} = 1.2, J_{6,8} = 1$
7-H	5.73, d, $J_{7,6} = 10$	5.73, d, $J_{7,6} = 10$	$3.74, m, W_{1/2} = 12.5$	4.86, m, $W_{1/2} = 5.5$	5.82, d, $J_{7,6} = 10$	5.66, d, $J_{7,6} = 10$
H-18	0.71, S	0.73, s	0.71, S	0.69, s	0.74, s	0.71, s
H-19	1.73, t, <i>J</i> =7.5	1.83, t, <i>J</i> =7.5	I	1	1	$J_{19,28h} = 15$ $J_{19,28h} = 10$
H-26 and H-27	0.86, d, J=6	0.89, d, $J=6$	0.86, d, J=6	0.86, d, J=6	0.86, d, J=6	0.88, d, J=6
H-28	3.80, t, J=7.5	4.14, t, $J=7.5$	3.64, t, J=7.5	4.18, t $J=7.5$	4.18, m, W ^{1/2} =28.75	1
-OCOCH3	1	2.07	I	2.06, 2.08, 2.11	-	1
$H-28_A$	1	1	1	Ι	I	5.43, dd
						$J_{28A,19} = 15$ $J_{28A,28B} = 2.5$
H-28 _B	1	1	I	1	1	5.14, dd $J_{82B,19} = 10$ $J_{28B,28A} = 2.5$
^a Chemical ^b CDCl ₃ ^e acetone-d ₆	^b Chemical shifts expressed in terms of ppm (δ) values and coupling constants (J) in Hz. ^b CDCl ₃ ^c acetone-d ₆	n terms of ppm	(d) values and co	upling constants (.	7) in Hz.	

STEROIDAL ANALOGUE OF DEOXYVERNOLEPIN

9

EXPERIMENTAL

Melting points were taken on the Boetius PHMK apparatus and were not corrected. IR spectra were recorded (KBr pellets or film) on a Perkin-Elmer spectrophotometer, model 457. ¹H NMR spectra were recorded on a Varian FT-80A spectrometer in CDCl₃ and acetone- d_6 using tetramethylsilane as internal standard. Chemical shifts were expressed in terms of ppm (δ) values and coupling constants (J) in Hz. ¹³C NMR spectra were recorded on a Varian FT-80A spectrometer at 20 MHz in CDCl₃ and acetone- d_6 .

5α -Bromo- 5α -cholestan- 3β , 6β -diol 3-monoacetate 11

Bromohydrin 11 was synthesized from cholesterol according to the known procedure of Kalvoda *et al.*³⁰ (38 g, $56^{0}/_{0}$ m. p. 172—174 °C).

5α -Bromo- 3β -acetoxy- 5α -cholestan-6-one 12

Bromohydrin 11 was dissolved in acetone and oxidized with Jones reagent. After the usual work-up procedure, bromoketone 12 (36.5 g, $96^{0}/_{0}$; m. p. 158—160 °C (EtOH)) was obtained.

7α -Bromo- 3β -acetoxy- 5α -cholectan-6-one 13

Isomerization of 5α -bromo-6-ketone 12 into 7α -bromo isomer 13 was performed with $48^{0}/_{0}$ hydrobromic acid in acetic acid according to Heilbron *et. al.*³¹ (20.8 g, $58^{0}/_{0}$; m. p. 142—145 ° (EtOH)).

7α -Bromo-5- α -cholestan- 3β , 6β -diol 3-monoacetate 15

 7α -Bromoketone 13 was reduced with sodium borohydride in methanol solution at 0—5 °C. After work-up and separation on a silica gel column bromohydrin 15 was obtained (14 g, 48%; m. p. 179—180 °C (EtOH)) together with its epimer, 7α -bromo- 5α -cholestan- 3β , 6α -diol 3-monoacetate 14 (15 g, 52%; m. p. 162—163 °C (EtOH)).

7α -Bromo-6 β , 19-epoxy-5 α -cholestan-3 β -ol 3-acetate 16

A suspension of lead tetraacetate (76 g; 0.171 mole) and calcium carbonate (34 g) in cyclohexane (1666 ml) were stirred at reflux for 20 min. Upon addition of bromohydrin 15 (15 g; 0.029 mole) and iodine (16.5 g; 0.065 mole) the reaction mixture was irradiated with a Wolfram lamp (500 W) for 40 minutes. The cooled reaction mixture was filtered through celite, the filtrate was diluted with ether and washed with $10^{0/6}$ sodium tiosulphate and water and dried over anhydrous magnesium sulphate. Magnesium sulphate was filtered off, the filtrate evaporated *in vacuo* and the residue crystallized from methanol and ether to give 7 α -bromo- -6β ,19-epoxy- 5α -cholestan- 3β -ol 3-acetate 16, 13.7 g (92⁰/₀), m. p. 126—128 °C (MeOH//ether), lit.³³ m. p. 129—130 °C; $[\alpha]_{p}^{20} = -62.30^{\circ}$ (c = 1.00, chl.); IR (KBr): ν 1740, 1245, 1235, 1197, 1038, 1022, 970, 962, 936, 922, 905 cm⁻¹; ¹H NMR (CDCl₃): δ 0.75 (3H, s, H-18), 0.88 (6H, d, J = 6 Hz, H-26 and H-27), 0.93 (3H, d, J = 6 Hz, H-21), 2.05 (3H, s, $-\text{OCOCH}_3$), 2.56 (1H, dd, $J_{8.7} = 4$ Hz, $J_{8.9} = 12$ Hz, H-8), 3.69 and 3.87 (2H, AB system, J = 8.8 Hz, H-19), 4.09 (2H, m, $W_{1/2} = 17.5$ Hz, H-6 and H-7), 4.73 (1H, m, $W_{1/2} = 25$ Hz, H-3).

5α -Cholest-6-en- 3β , 19-diol 3-monoacetate 17

Powdered zinc (91 g) was added to a solution of 6β ,19-epoxide 16 (13.6 g; 0.026 mole) in glacial acetic acid (456 ml) and water (20 ml) at 45—50 °C while the reaction mixture was vigorously stirred. After 40 min the reaction mixture was colled to room temperature, inorganic material filtered off and the filtrate evaporated *in vacuo*. The obtained oily residue was dissolved in the mixture of ether and chloroform (5:1) and washed with a saturated solution of sodium bicarbonate and water. The dried solution was evaporated *in vacuo* and the residue crystallized from a mixture of methanol and chloroform to give 5a-cholest-6-en- 3β ,19-diol 3-monoacetate 17, 10.4 g (89%), m. p. 147—149 °C (MeOH/CHCl₃), lit.³³ m. p. 102—103 °C; $[\alpha]_{\rm p}^{15} = -94.94^{\circ}$ (c = 1.12, chl); IR (KBr): v 3520, 3040, 1650, 1265, 1055, 1045, 1029.

995, 985 cm⁻¹ ¹H NMR (CDCl₃): δ 0.73 (3H, s, H-18), 0.87 (6H, d, J = 6 Hz, H-26 and H-27), 0.90 (3H, d, J = 6 Hz, H-21), 2.03 (3H, s, $-\text{OCOCH}_3$), 3.85 (2H, s, H-19), 4.75 (1H, m, $W_{1/2} = 30$ Hz, H-3), 5.24 and 5.51 (2H, AB system, J = 10 Hz, H-16 and H-7).

Anal. $C_{29}H_{48}O_3$ (444.72) calc'd: C 78.33; H 10.88% found: C 78.01; H 10.74%.

5α -Cholest-6-en- 3β , 19-diol 3-acetate 19-benzoate 18

Monoacetate 17 (10.4 g; 0.023 mole), dissolved in pyridine (48 ml), was treated with benzoyl chloride (10.4 ml; 0.09 mole) at room temperature. After the usual work-up the obtained oily residue crystallized from methanol to give 5*a*-cholest-6-en-3 β ,19-diol 3-acetate 19-benzoate 18, 10.1 g (80%), m. p. 126—128 °C (MeOH); [α]p¹⁷ = -58.15° (c = 0.92, chl.); IR (KBr): ν 3030, 1745, 1730, 1650, 1620, 1600, 1275, 1240, 1180, 1115, 1073, 1055, 1030, 715 cm⁻¹; ¹H NMR (CDCl₃): δ 0.65 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.93 (3H, d, J = 6 Hz, H-21), 2.05 (3H, s, -OCOCH₃), 4.46 and 4.68 (2H, AB system, J = 12 Hz, H-19), 4.80 (1H, m, W_{1/2} = 30 Hz, H-3), 5.28 and 5.56 (2H, AB system, J = 10 Hz, H-6 and H-7), 7.28— -8.05 (5H, arom.).

Anal. C₃₆H₅₂O₄ (548.81) calc'd.: C 78.79; H 9.55⁰/₀ found: C 79.04; H 9.57⁰/_e.

$6\alpha.7\alpha$ -Epoxy-5 α -cholestan-3 β ,19-diol 3-acetate 19-benzoate 19

10.1 g (0.018 mole) of 18, dissolved in dichloromethane (163 ml), was treated with *m*-chloroperoxybenzoic acid (4.2 g; 0.024 mole). The reaction mixture was left overnight, washed with a saturated solution of sodium bicarbonate and water. The dried organic solution was evaporated in vacuo and the obtained residue crystallized from methanol to give 6a,7a-epoxy-5a-cholestan- 3β ,19-diol 3-acetate 19-benzoate 19, 10 g (96%), m. p. 110—112 °C (MeOH); $[a]_{p}^{16} = -18.24^{\circ}$ (c = 1.17, chl.); IR (KBr): v 3050, 1730, 1595, 1582, 1262, 1245, 1180, 1105, 1074, 1050, 1025, 725 cm⁻¹; ¹H NMR (CDCl₃): δ 0.64 (3H, s, H-18), 0.87 (6H, d, J = 6 Hz, H-26 and H-27), 0.91 (3H, d, J = 6 Hz, H-21), 2.05 (3H, s, —OCOCH₃), 2.83 (1H, d, $J_{6,7} = 4$ Hz, $H_{7,8} = 2.5$ Hz, H-7), 4.37 and 4.64 (2H, AB system, J = 12 Hz, H-19), 4.76 (1H, m, $W_{1/2} = 30$ Hz, H-3), 7.28—8.10 (5H, arom.).

Anal. C₃₆H₅₂H₅ (564.81) calc'd.: C 76.55; H 9.28% found: C 76.90; H 9.33%.

6α , 7α -Epoxy- 5α -cholestan- 3β , 19-diol 19-monobenzoate 20

A solution of potassium hydroxide in water (1 g KOH in 2.7 ml H₂O) was added to the stirred solution of 19 (10 g; 0.018 mole) in ethanol (259 ml) at room temperature. After 90 min, the reaction mixture was poured into cold water and acidified with diluted hydrochloric acid. The precipitate was filtered off, washed well with water and crystallized from methanol to give 6α , 7α -epoxy- 5α -cholestan- -3β ,19-diol 19-monobenzoate 20, 8.5 g (92%), m. p. 107–109 °C (MeOH); $[\alpha]_{p}^{18} = -13.09^{\circ}$ (c = 1.12, chl.); IR (KBr): v 3350, 1730, 1610, 1595, 1180, 1115, 1075, 1032, 719 cm⁻¹; ¹H NMR (CDCl₃): δ 0.63 (3H, s, H-18), 0.85 (6H, d, J = 6 Hz, H-26 and H-27), 0.88 (3H, d, J = 6 Hz, H-21), 2.83 (1H, d, $J_{6,7} = 4$ Hz, H-6), 3.13 (1H, dd, $J_{7.6} = 4$ Hz, $J_{7.8} = 2.5$ Hz, H-7), 3.70 (1H, m, W₁/₂ = 18 Hz, H-3), 4.36 and 4.60 (2H, AB system, J = 12 Hz, H-19), 7.28–8.08 (5H, arom.).

Anal. $C_{34}H_{50}O_4$ (522.81) calc'd.: C 78.12; H 9.64% found: C 77.84; H 9.45%

3-Methoxy-6a,7a-epoxy-5a-cholest-2-en-19-ol 19-benzoate 21

To a solution of alcohol 20 (8.5 g; 0.016 mole) in dichloromethane (223 ml) pyridinium chlorochromate (12.6 g; 0.058 mole) was added in small portions. The reaction was mechanically stirred for four hours at room temperature. After the oxidation was finished (TLC), the supernatant was poured off and the precipitate washed with dry ether (3×100 ml). The combined solutions were left at room temperature for 45 min, the precipitate was filtered off and the filtrate evaporated

in vacuo. The obtained brown-reddish oil was dissolved in methanol (100 ml) and solution refluxed for 20 minutes. The resulting green-coloured the solution was evaporated in vacuo and the oily residue eluted with benzene through a short silica gel column. The obtained colourless oil was dissolved in dry benzene (223 ml), pyridinium p-toluenesulphonate was added (0.95 g; 0.004 mole) and the reaction mixture refluxed for 30 minutes. The cooled reaction mixture was twice washed with water, dried over anhydrous magnesium sulphate and evaporated in vacuo. After separation on a silica gel column enol ether 21, ketone 22 and p-toluene-sulphonate 23 were isolated. 3-methoxy-6a,7a-epoxy-5a-cholest-2-en-19-ol 19-benzoate 21, 3.2 g (37%), oil; $[a]_{p}^{18} = +24.44^{\circ}$ (c = 0.71, chl.); IR (KBr): v 3055, 1718, 1665, 1600, 1582, 1270, 1213, 1180, 1110, 1070, 1028, 712 cm⁻¹; ¹H NMR (CDCl₃): δ 0.61 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.90 (3H, s, J = 6 Hz, H-21), 2.95 (2H, d, $J_{6.7} = 4$ Hz, H-6), 3.13 (1H, dd, $J_{7.6} = 4$ Hz, $J_{7.8} = 2.5$ Hz, H-7), 3.50 (3H, s, -OCH₃), 4.30 and 4.40 (2H, AB system, J = 12 Hz, H-19), 4.58 (1H, d, $J_{2.1z} = 3.5$ Hz, H-2), 7.28–8.08 (5H, arom.).

Anal. C₃₅H₅₀O₄ (534.81) calc'd.: C 78.61; H 9.42⁰/₀ found: C 78.01; H 9.69⁰/₀.

6a,7a-epoxy-19-benzoyloxy-5a-cholestan-3-one 22, 2.7 g (31%), oil; $[a]_{b}^{23} = +14.71^{\circ}$ (c = 0.64, chl.); IR (film): v 3055, 1717, 1600, 1585, 1270, 1110, 1100, 1072, 1028, 715 cm⁻¹; ¹H NMR (CDCl₃): δ 0.69 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.88 (3H, d, J = 6 Hz, H-21), 2.84 (1H, d, $J_{6.7} = 4$ Hz, H-6), 3.16 (1H, dd, $J_{7.6} = 4$ Hz, $J_{7.8} = 2.5$ Hz, H-7), 4.49 and 4.71 (2H, AB system, J = 12 Hz, H-19), 7.27—8.08 (5H, arom.).

7α-hydroxy-6β-p-toluenesulphonyloxy-19-benzoyloxy-5α-cholestan-3-one 23, 2.5 g (23%), oil; $[a]_{\rm b}^{20.5} = +11.48^{\circ}$ (c = 0.82, chl.); IR (film): v 3500, 3060, 1715, 1600, 1585, 1271, 1191, 1179, 1112, 1099, 1072, 1023, 938, 715, 677 cm⁻¹; ¹H NMR (CDCl₃): δ 0.69 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.89 (3H, d, J = 6 Hz, H-21), 2.45 (3H, s, CH₃/TsO-), 3.91 (1H, m, $W_{1/2} = 7.5$ Hz, H-7), 4.36 (1H, dd, $J_{6.5} = 3.75$ Hz, $J_{6.7} = 2$ Hz, H-6), 4.53 and 4.63 (2H, AB system, J = 13 Hz, H-19), 7.33-8.04 (9H, arom.).

Methyl-6a,7a-epoxy-2-hydroxy-19-benzoyloxy-2,3-seco-5a--cholestan-3-oate 24

Through a solution of enol ether 21 (0.05 g; 9×10^{-5} mole) in dichloromethane (4.1 ml) and methanol (0.1 ml) ozone (~ 5%) was bubbled for 5 min at -78 °C. After the reaction was over (TLC), ozone was replaced by an argon atmosphere, the reaction mixture warmed to -20 °C and methanol (1 ml) added, followed by sodium borohydride (0.014 g; 3.70×10^{-4} mole). After the reduction was completed, the reaction mixture was acidified with glacial acetic acid (-20 °C $\rightarrow 0$ °C). The organic layer was washed with water, dried over anhydrous magnesium sulphate and evaporated *in vacuo* to give, *methyl-6a,7a-epoxy-2-hydroxy-19-benzoyloxy-2,3-seco-5a-cholestan-3-oate 24*, 0.044 g (84%), oil; $[a]_{\rm b}^{19} = -27.66^{\circ}$ (c = 0.53, chl.); IR (KBr): ν 3440, 3055, 1735, 1718, 1660, 1582, 1270, 1177, 1110, 1070, 1028, 712 cm⁻¹; ¹H NMR (CDCl₃): δ 0.66 (3H, s, H-18), 0.88 (6H, d, J = 6 Hz, H-26 and H-27), 0.91 (3H, d, J = 6 Hz, H-21), 2.41 (2H, B₂ of AB₂ system, $J_{\rm gem} = J_{4\pi,5} = 11.2$ Hz, H-4 α and H-5), 2.75 (1H, A of AB₂ system, $J_{\rm gem} = J_{4\pi,5} = 11.2$ Hz, H-4 β), 2.98 (1H, d, $J_{6,7} = 4$ Hz, $H_{7,8} = 1.8$ Hz, H-7), 3.69 (2H, t, J = 6 Hz, H-2), 3.72 (3H, s, -COCH₃), 4.14 and 4.24 (2H, AB system, J = 12 Hz, H-19), 7.29–8.06 (5H, arom.).

$Methyl-6\beta-bromo-2,7\alpha-dihydroxy-19-benzoyloxy-2,3-seco-5\alpha--cholestan-3-oate\ 25$

A solution of epoxide 24 (1.60 g; 2.8×10^{-3} mole) in dioxane (57 ml) and water (4.73 ml) was treated with $48^{0/0}$ hydrobromic acid (4.70 ml) and stirred for 45 min at room temperature. The reaction mixture was diluted with water (20 ml), extracted with ether (3 × 40 ml) and the etheral solution washed with a saturated sodium bicarbonate solution and water. The dried solution was evaporated *in* vacuo and after separation on a silica gel column bromohydrin 25 and triol 26 were obtained, methyl-6\beta-bromo-2,7a-dihydroxy-19-benzoyloxy-2,3-seco-5a-cholestan-3-

-oate 25, 1.02 g (56%), m. p. 149—151 °C (chl./hexane); $[a]_{D}^{20} = +11.07^{\circ}$ (c = 0.85, chl.); IR (KBr): ν 3435, 3055, 1722, 1710, 1600, 1585, 1275, 1176, 1027, 712 cm⁻¹; ¹H NMR (CDCl₃): δ 0.75 (3H, s, H-18), 0.89 (6H, d, J = 6 Hz, H-26 and H-27), 0.93 (3H, d, J = 6 Hz, H-21), 2.64 (3H, s, H-4 and H-5), 3.71 (3H, s, —COOCH₃), 3.75 (2H, m, W_{1/2} = 15 Hz, H-2), 4.06 (1H, m, W_{1/2} = 7.5 Hz, H-7), 4.40 (1H, m, W_{1/2} = 5 Hz, H-6), 4.46 and 4.82 (2H, AB system, J = 12 Hz, H-19), 7.29—8.19 (5H, arom.).

Anal. C₃₅H₅₃BrO₆ (649.70) calc'd.: C 64.70; H 8.22⁰/₀ found: C 64.80; H 7.88⁰/₀

methyl-2,6 β ,7 α -trihydroxy-19-benzyloxy-2,3-seco-5 α -cholestan-3-oate 26, 0.71 g (43%), m. p. 177.5—179.5 °C (chl./hexane); $[\alpha]_{\rm p}^{19} = 0^{\circ}$ (c = 0.67, chl.); IR (KBr): ν 3450, 3330, 3245, 3055, 1732, 1690, 1600, 1582, 1277, 1240, 1180, 1168, 1029, 712 cm⁻¹; ¹H NMR (CDCl₃): δ 0.68 (3H, s, H-18), 0.87 (6H, d, J = 12 Hz, H-26 and H-27), 2.32 (3H, m, $W_{1/2} = 25$ Hz, H-4 and H-5), 3.67 (3H, s, —COOCH₃), 3.73 (4H, m, $W_{1/2} = 15$ Hz, H-2, H-6 and H-7), 4.47 and 4.68 (2H, AB system, J = 12 Hz, H-19), 7.28—8.11 (5H, arom.).

Anal. C₃₅H₅₄O₇ (586.81) calc'd.: C 71.64; H 9.28% found: C 71.87; H 9.10%.

Methyl-2-hydroxy-2,3-seco-5a-cholest-6-en-3-oate 27

Powdered zinc (5.41 g) was added to a solution of bromohydrin 25 (1.02 g; 1.6×10^{-3} mole) in glacial acetic acid (27 ml) and water (1.2 ml) at 45–50 °C while the reaction mixture was vigorously stirred. After 40 min the reaction mixture was cooled to room temperature, the inorganic material filtered off and the filtrate evaporated in vacuo. The obtained oily residue was dissolved in the mixture of ether and chloroform (5:1) and washed with saturated sodium bicarbonate solution and water. The dried solution was evaporated in vacuo and the residue eluted through a short silica gel column to give methyl-2-hydroxy-2,3-seco-5a-cholest-6-en-3-oate 27, 0.62 g (72%), oil; $[a]_{p^{23}} = -68.05^{\circ}$ (c = 1.15, chl.); IR (film): ν 3500, 3055, 3015, 1735, 1720, 1640, 1600, 1585, 1272, 1178, 1112, 1022, 978, 712 cm⁻¹; ¹H NMR (CDCl₃): δ 0.66 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.89 (3H, d, J = 6 Hz, H-21), 2.81 (2H, m, $W_{1/2} = 20$ Hz, H-4), 3.66 (3H, s, -COOCH₃), 3.71 (2H, m, $W_{1/2} = 15$ Hz, H-2), 4.23 and 4.35 (2H, AB system, J = 12 Hz, H-19), 5.34 and 5.56 (2H, AB system, J = 11.3 Hz, H-6 and H-7), 7.28–8.08 (5H, arom.).

28-Hydroxy-2-oxa-5a,10a-cholest-6-en-3-one 4

A solution of potassium hydroxide (0.2 g KOH in 2 ml H₂O) was added to a suspension of 27 (0.54 g; 9.8×10^{-4} mole) in methanol (3 ml) and the reaction mixture was left on a steam bath for 15 min and acidified with concentrated hydrochloric acid. The obtained suspension was extracted with ether and chloroform (5:1), the separated organic solution washed with saturated sodium bicarbonate solution and water, dried over anhydrous magnesium sulphate and evaporated *in vacuo*. Crystallization from chloroform and hexane gave 28-hydroxy-2-oxa-5a,10a-cholest-6-en-3-one 4, 0.34 g (84%), m. p. 102—103.5 °C (chl./hexane); $[a]_p^{18} = -78.16^\circ$ (c = 0.77, chl.); IR (KBr): ν 3440, 3010, 1735, 1650, 1060 cm⁻¹; ¹H NMR (CDCl₃): δ 0.71 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.92 (3H, d, J = 6 Hz, H-21), 1.73 (2H, t, J = 7.5 Hz, H-19), 2.29 (1H, m, $W_{1/2} = 25$ Hz, H-5), 2.80 (2H, m, $W_{1/2} = 31.25$ Hz, H-4), 3.80 (2H, d, J = 7.5 Hz, H-28), 3.83 and 4.37 (2H, AB system, J = 12 Hz, H-1), 5.43 (1H, ddd, $J_{6.7} = 10$ Hz, $J_{6.5} = 1.2$ Hz, $J_{6.8} = 1$ Hz, H-6), 5.73 (1H, d, $J_{7.6} = 10$ Hz, H-7); ¹³C NMR (CDCl₃): δ 172.9 (C-3), 132.0 and 129.0 (C-6 and C-7), 71.5 (C-1), 58.4 (C-28), 36.1 (C-22), 35.7 (C-20), 28.1 (C-16), 28.0 (C-24), 23.8 (C-23), 22.5 and 22.6 (C-26 and C-27), 18.7 (C-21), 11.9 (C-18).

> Anal. C₂₇H₄₄O₃ (416.64) calc'd: C 77.84; H 10.64⁰/₀ found: C 78.10; H 10.54⁰/₀.

Acetylation of 4 with acetic anhydride in pyridine gave 28-acetoxy-2--oxa-5a,10a-cholest-6-en-3-one 30, oil; IR (film): ν 3010, 1755, 1737, 1660, 1232, 1065 cm⁻¹; ¹H NMR (CDCl₃): δ 0.73 (3H, s, H-18), 0.89 (6H, d, J = 6 Hz, H-26

and H-27), 0.93 (3H, d, J = 6 Hz, H-21), 1.83 (2H, t, J = 7.5 Hz, H-19), 2.07 (3H, s, -OCOCH₃), 2.32 (1H, m, $W_{1/2} = 25$ Hz, H-5), 2.78 (2H, m, $W_{1/2} = 31.25$ Hz, H-4), 3.84 and 4.39 (2H, AB system, J = 12 Hz, H-1), 4.14 (2H, t, J = 7.5 Hz, H-28), 5.43 (1H, ddd, $J_{6.7} = 10$ Hz, $J_{6.5} = 1.2$ Hz, $J_{6.8} = 1$ Hz, H-6), 5.73 (1H, d, $J_{7.6} = 10$ Hz, H-7).

6β ,7 α ,28-Trihydroxy-2-oxa-5 α ,10 α -cholestan-3-one 31

δ-Lactone 31 was obtained in the same manner from triol 26 as 4 from 27. 6β ,7a,28-trihydroxy-2-oxa-5a,10a-cholestan-3-one 31, 0.37 g, (74%), m. p. 193—195 °C (EtOAc); [a]_p¹⁹⁻⁵ = +17.35° (c = 0.70, chl.); IR (KBr): ν 3410, 1705, 1225, 1067, 1050, 1021, 986 cm⁻¹; ¹H NMR (acetone-d₆): δ 0.71 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, 2.81 (2H, m, $W_{1/2} = 26.85$ Hz, H-4), 3.64 (2H, d, J = 7.5 Hz, H-28), 3.74 (2H, d, $W_{1/2} = 12.5$ Hz, H-6 and H-7), 3.86 (1H, dd, $J_{gem} = 11.25$ Hz, H-28), 3.74 (2H, d, 4.89 (1H, d, $J_{gem} = 11.25$ Hz, H-1β); ¹³C NMR (aceton-d₆): δ 171.2 (C-3), 76.2 and 75.7 (C-6 and C-7), 71.4 (C-1), 57.2 (C-28), 36.7 (C-22), 35.9 (C-20), 24.1 (C-23), 23.1 and 22.8 (C-26 and C-27), 19.1 (C-21), 12.2 (C-18).

> Anal. C₂₇H₄₆O₅ (450.66) calc'd: C 71.96; H 10.29% found: C 72.46; H 10.53%.

Acetylation of 31 with acetic acid anhydride in pyridine gave $6\beta_{,7\alpha,28}$ -triacetoxy-2-oxa-5a,10a-cholestan-3-one 32, oil; IR (KBr): ν 1755, 1735, 1265, 1234, 1210, 1060, 1032, 970 cm⁻¹; ¹H NMR (CDCl₃): δ 0.69 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz, H-26 and H-27), 0.91 (3H, d, J = 6 Hz, H-21), 2.06 (3H, m, $-\text{OCOCH}_3$), 2.11 (3H, s, $-\text{OCOCH}_3$), 2.31 (1H, m, $W_{1/2}$ = Hz, H-5), 2.86 (2H, m, $W_{1/2}$ = 41.25 Hz, H-4), 4.01 (1H, dd, J_{gem} = 11.25 Hz, J = 2.5 Hz, H-1a), 4.18 (2H, t, J = 7.5 Hz, H-28), 4.76 (1H, d, J_{gem} = 11.25 Hz, H-1 β), 4.86 (1H, m, $W_{1/2}$ = 5.5 Hz, H-7), 5.05 (1H, t, J = 3.25 Hz, H-6).

28-o-nitrophenylseleno-2-oxa-5a,10a-cholest-6-en-3-one 28

δ-Lactone 4 (0.15 g, 3.6×10^{-4} mole) dissolved in dry tetrahydrofurane (2 ml) was treated with o-nitrophenyl selenocyanate (0.13 g; 5.7×10^{-4} mole)⁴³ and trin-butylphosphine. When the reaction was over (determined by disappearance of the deep-red colour) the solution was evaporated in vacuo and the obtained oil was eluted through a short silica gel column. Crystallization from hexane afforded 28-o-nitrophenylseleno-2-oxa-5a,10a-cholest-6-en-3-one 28, 0.18 g (90%), m. p. 105—106 °C (hexane); $[\alpha]_p^{20} = -15.55^\circ$ (c = 0.86, acetone); IR KBr): ν 3010, 1750, 1720, 1690, 1582, 1560, 1503, 1335, 1297, 847, 782, 730 cm⁻¹; H NMR (CDCl₃): δ 0.74 (3H, s, H-18), 0.86 (6H, d, J = 6 Hz H-26 and H-27), 0.93 (3H, d, J = 6 Hz, H-21), 2.33 (1H, m, $W_{1/2} = 26.25$ Hz, H-5), 2.83 (4H, m, $W_{1/2} = 28.75$ Hz, H-4 and H-28), 3.84 and 4.41 (2H, AB system, J = 12 Hz, H-1), 5.48 (1H, ddd, $J_{6.7} = 10$ Hz, $J_{6.5} = 1.2$ Hz, $J_{6.8} = 1$ Hz, H-6), 5.82 (1H, d, $J_{7.6} = 10$ Hz, H-7).

Anal. C₃₃H₄₇O₄NSe (600.70) calc'd.: C 65.98; H 7.89; N 2.33⁰/₀ found: C 66.62; H 7.73; N 3.61⁰/₀.

2-Oxa-5a,10a-cholest-6,19-dien-3-one 29

A solution of 28 (0.15 g; 2.5×10^{-4} mole) in tetrahydrofurane (2 ml) was cooled to 0 °C and treated with 30% hydrogen peroxide (1 ml). The reaction mixture was left overnight at room temperature, extracted with ethyl acetate and washed with saturated sodium bicarbonate solution and water. The dried solution was evaporated *in vacuo* and the resulting oil crystallized from methanol to give 2-oxa-5a,10a-cholest-6,19-dien-3-one 29, 0.091 g (91%), m. p. 84-85 °C (MeOH); $[a]_{p}^{25} = -87.70^{\circ}$ (c = 0.61, chl.); IR (KBr): *v* 3070, 3010, 1750, 1725, 1712, 1690, 1627, 1280, 1205, 1181, 1147, 1052, 1031, 1007, 920, 729, 700 cm⁻¹; ¹H NMR (CDCl₃): δ 0.71 (3H, s, H-18), 0.88 (6H, d, J = 6 Hz, H-26 and H-27), 0.91 (3H, d, J = 6 Hz, H-21), 2.56 (3H, m, W_{1/2} = 50 Hz, H-4 and H-5), 4.39 (2H, s, H-1), 5.14 (1H, dd, $J_{28_{h},19}=10$ Hz, $J_{28_{h},28_{h}}=2.5$ Hz, H-28), 5.43 (1H, dd, $J_{28_{h},19}=15$ Hz, $J_{28_{h},28_{h}}=2.5$ Hz, H-28), 5.43 (1H, dd, $J_{28_{h}}=1$ Hz, H-6), 5.66 (1H, d, $J_{-7}=10$ Hz, H-7), 5.68 (1H, dd, $J_{19,28_{h}}=15$ Hz, $J_{19,28_{h}}=10$ Hz, H-7).

Anal. C₂₇H₄₂O₂ (398.63) calc'd: C 18.35; H 10.62⁰/₀ found: C 81.53; H 10.23⁰/₀. Acknowledgement. — The authors are grateful to the Serbian Academy of Sciences and Arts and to the Serbian Research Fund for the financial support.

REFERENCES

- 1. W. Herz, Izrael J. Chem. 16 (1977) 32.
- 2. E. R. Rodriguez, G. H. N. Towers, and J. C. Mitchell, Phytochemistry 15 (1976) 1573.
- 3. S. M. Kupchan, D. C. Fessler, M. A. Eakin, and T. J. Giacobbe, Science 168 (1970) 376.
- 4. E. E. van Tamelen and S. R. Bach, J. Amer. Chem. Soc. 77 (1955) 4683.
- 5. S. M. Kupchan, T. J. Giacobbe, and I. S. Krull, Tetrahedron Lett. (1970) 2859.
- S. M. Kupchan, T. J. Giacobbe, I. S. Krull, A. M. Thomas, M. A. Eakin, and D. C. Fessler, J. Org. Chem. 35 (1970) 3539.
- 7. R. L. Hanson, H. A. Lardy, and S. M. Kupchan, Science 168 (1970) 378.
- S. M. Kupchan, R. J. Hemingway, D. Werner, A. Karim, A. T. McPhail, and G. A. Sim, J. Amer. Chem. Soc. 90 (1968) 3596.
- 9. G. M. Laekeman, J. Martens, H. Bult, A. Vlietinck, and A. G. Herman, J. Nat. Prod. 46 (1983) 161.
- 10. L. Sequeira, R. J. Hemingway, and S. M. Kupchan, Science 161. (1968) 790.
- 11. P. A. Grieco, J. A. Noguez, and Y. Masaki, J. Org. Chem. 42 (1977) 495.
- 12. P. A. Grieco, J. A. Noguez, Y. Masaki, K. Hiroi, and M. Nishizawa, J. Med. Chem. 20 (1977) 71.
- 13. Y. Fujimoto, H. Miura, T. Shimizu, and T. Tatsuno, Tetrahedron Lett. (1980) 3409.
- 14. P. A. Grieco, M. Nishizawa, S. D. Burke, and N. Marinović, J. Amer. Chem. Soc. 98 (1976) 1612.
- P. A. Grieco, M. Nishizawa, T. Oguri, S. D. Burke, and N. Marinović, J. Amer. Chem. Soc. 99 (1977) 5773.
- S. Danishefsky, T. Kitahara, P. F. Schuda, and S. J. Entheredge, J. Amer. Chem. Soc. 98 (1976) 3028.
- 17. S. Danishefsky, T. Kitahara, R. McKee, and P. F. Schuda, J. Amer. Chem. Soc. 98 (1976) 6715.
- S. Danishefsky, P. F. Schuda, T. Kitahara, and S. J. Entheredge, J. Amer. Chem. Soc. 99 (1977) 6066.
- 19. F. Zutterman, H. De Wilde, P. De Clercq, and M. Vandewalle, Tetrahedron 35 (1979) 2389.
- G. R. Kieczykowski, M. L. Quesada, and R.H. Schlessinger, J. Amer. Chem. Soc. 102 (1980) 789.
- 21. H. Iio, M. Isobe, T. Kawai, and T. Goto, Tetrahedron 35 (1979) 941.
- 22. H. Iio, M. Iosobe, T. Kawai, and T. Goto, J. Amer. Chem. Soc. 101 (1979) 6076.
- M. Kocor, M. M. Kabat, J. Wicha, and W. Peczynska-Czoch, Steroids 41 (1983) 55.
- 24. For a review of the methods for the synthesis of α-methylene-γ- and δ-lactones see P. A. Grieco, Synthesis (1975) 67, A. W. Murray and R. G. Reid, J. Chem. Soc. Chem. Communn. (1984) 132 and references 17, 18 and 25.
- 25. J. Schreiber, M. Haag, N. Nishimoto, and A. Eschenmoser, Angew. Chem. 10 (1971) 355; Angew. Chem. Int. Ed. Engl. 10 (1971) 330.
- 26. P. A. Grieco, S. Gilman, and M. Nishizawa, J. Org. Chem. 41 (1976) 1485.
- 27. D. L. J. Clive, Tetrahedron 34 (1978) 1049.

- 28. C. Djerassi, Steroid Reactions, Holden Day, Inc., San Francisco 1963 p 593.
- 29. D. N. Kirk and M. P. Hartshorn, Steroid Reaction Mechanisms, Elsevier Publishing Comp., 1968 p. 113.
- 30. J. Kalvoda, K. Heusler, H. Ueberwasser, G. Anner, and A. Wettstein, Helv. Chim. Acta 46 (1963) 1361.
- 31. I. M. Heilbron, E. R. H. Jones, and F. S. Spring, J. Chem. Soc. (1937) 801.
- 32. D. R. James and C. W. Shopee, J. Chem. Soc. (1954) 4224.
- 33. P. Kočovsky, L. Kohout, and V. Černý, Coll. Czech. Chem. Communn. 45 (1980) 559.
- 34. J. Fried and J. A. Edwards, Organic Reactions in Steroid Chemistry, Van Nostrand Reinhold Company, Vol. II, 1972 pp. 158, 431.
- 35. a. W. Carruthers, Some Modern Methods of Organic Syntheses, Cambridge University Press, 1971 p 274.
 - b. Modern Synthetic Reactions, 2nd Ed., W. A. Benjamin, Inc., Menlo Park, 1972 p 358.
- 36. a. M. Lj. Mihailović and Ž. Čeković, Synthesis (1970) 209.
 b. Selective Organic Transformations, Vol II, M. Lj. Mihailović and R. E. Partch, B. S. Thyagarjan (Ed.), Wiley-Interscience, New York, London 1975.
- 37. E. J. Corey and N. Suggs, Tetrahedron Lett. (1975) 2647.
- 38. I. Aljančić-Šolaja, M. Bralović, B. Šolaja, and M. Stefanović, Bull. Soc. Chim. Beograd, 48 (1983) 299.
- 39. F. A. J. Meskens, Synthesis (1981) 501.
- 40. Y. Watanabe, Y. Mizuhara, and M. Shiota, Canad. J. Chem. 47 (1969) 1495.
- R. M. Silverstein, C. G. Bassler, and T.C. Morrin, Spectrometric Identification of Organic Compounds, 3rd Ed., John Wiley and Sons Inc., 1974.
- 42. a. J. R. Dyer, Applications of Absorption Spectroscopy of Organic Compounds, Prentice-Hall, Inc., 1965.
 - b. F. Scheinmann, An Introduction of Spectroscopic Methods for the Identification of Organic Compounds, Vol. I, Pergamon Press Ltd., Headington Hall, Oxford 1965.
 - c. E. Pretsch, T. Clerc, J. Siebl, and W. Simon, Tabelen für Strukturaufklärung organischer Verbindungen mit spektroskopischen Metoden, Springer-Verlag, Berlin, Heidelberg 1976.
 - d. R. J. Abraham and P. Loftus, Proton and Carbon-13 NMR Spectroscopy; an Integrated Approach, Heyeden and Son Ltd., London 1980.
 - e. Struktur und Absorptionsspektroskopie der Steroide und Alkaloide, Georg Thieme Verlag, Stuttgart 1975.

f. N. S. Bhacca and D. H. Williams, Applications of NMR Spectroscopy in Organic Chemsitry. Ilustrations from the Steroid Field, Holden-Day, Inc., San Francisco 1964.

43. Sublimed, yellow, o-nitrophenyl selenocyanate, m.p. 144 °C, was used in this reaction. For further informations see H. Bauer, Chem. Ber. 46 (1913) 92;
K. B. Sharpless, M. W. Young, and D. Lauer, Tetrahedron Lett. (1973) 1979;
K. B. Sharpless and M. W. Young, J. Org. Chem 40 (1975) 947 and references 26 and 27.

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IZVOD

Steroidni analozi deoksivernolepina. Sinteza δ -laktonskog ključnog intermedijera

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Seskviterpenski laktoni elemanolidnog tipa vernolepin 1 i deoksivernolepin 2 pokazuju izraženu antitumorsku aktivnost protiv Walker-ovog intramuskularnog karcinosarkoma 256 i CCRF — CEM humanih limfoblastičnih ćelija leukemije u kulturi i stoga su bili predmet mnogih parcijalnih i totalnih sinteza.

Zbog značajnog aktineoplastičnog (antitumorskog) dejstva seksviterpenskih laktona sa α -metilenskom funkcijom na γ - i δ -laktonskom prstenu sintetizovan je polazeći od holesterola, steroidni analog deoksivernolepina 3, odnosno ključni intermedijer u njegovoj sintezi 28-hidroksi-2-oksa-5 α ,10 α -holest-6-en-3-on(4).

U sintezi su primenjene poznate reakcije, neke su modifikovane, a upotrebljena je i nova reakcija za dobivanje dimetil-acetala pomoću metanola i piridinijum-hlorohromata. Ovim reagensom mogu se reakcije acetalizacije vršiti u prisustvu osetljivih funkcionalnih grupa kao što je epoksidna grupa.