

TAXONOMIC SURVEY OF BENTHIC DIATOMS IN NEUM BAY, SOUTHEASTERN ADRIATIC

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The objective of this work was to contribute to the knowledge of microphytobenthos in the Adriatic Sea, by studying for the first time the benthic diatom communities in Neum Bay in Bosnia and Herzegovina. For this purpose, the benthic diatoms naturally growing on stones and macroalgae were identified at one station in the small semi-enclosed oligotrophic Neum Bay during 2010 and 2011. A total of 24 samples were collected at two different depths, 0.5 m and 8 m, and analysed with light and scanning electron microscopy. Altogether, 425 pennate and 58 centric taxa (species and infraspecific taxa) belonging to 60 families and 115 genera were noted. Genera with the greatest number of taxa were: *Mastogloia* (46 taxa), *Navicula* (36), *Diploneis* (35), *Nitzschia* (34), *Amphora* (31), *Cocconeis* (27), *Achnanthes* (14), *Halimphora* (12), *Lyrella* (11), and *Surirella* and *Licmophora* (10 each). *Amphora bigibba* var. *interrupta* and *Cocconeis scutellum* were the most frequent taxa, being present in 87.5% of the samples. In total, 142 taxa were found only once (sporadic taxa). Although benthic diatom richness is high in the bay, taxa are apparently not distributed evenly temporally. Consistent quantitative and qualitative data are still needed for a better determination of the seasonal and spatial changes of the epilithic assemblages in the region.

Keywords: Bacillariophyta, shallow oligotrophic bay, species identification, biodiversity, Bosnia and Herzegovina.

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Cilj rada bio je doprinijeti poznавању mikrofitobentosa u Jadranskom moru istražujući po prvi put bentoske dijatomejske zajednice u Neumskom zaljevu u Bosni i Hercegovini. Bentoske dijatomeje su određene na jednoj postaji u poluzatvorenom oligotrofnom Neumskom zaljevu 2010. i 2011. Sakupljeno je 24 uzorka na dubinama 0,5 i 8 m, analizirani su s pomoću svjetlosnog i skenirajućeg elektronskog mikroskopa. U uzorcima je utvrđeno 425 penata i 58 centričnih svojti (vrsta i nižih taksonomskih kategorija) dijatomeja unutar 60 porodica i 115 rodova. Najveći broj svojti nađen je unutar sljedećih robova: *Mastogloia* (46 svojti), *Navicula* (36), *Diploneis* (35), *Nitzschia* (34), *Amphora* (31), *Cocconeis* (27), *Achnanthes* (14), *Halimphora* (12), *Lyrella* (11) te *Surirella* i *Licmophora* (svaki po 10 svojti). Najčešće svojte su *Amphora bigibba* var. *interrupta* i *Cocconeis scutellum* s 87,5% učestalosti u ukupnom broju uzorka. Utvrđene su 142 sporadične svojte (utvrđene u samo jednom uzorku). Iako je broj svojti relativno visok, nije utvrđena pravilnost u njihovoј vremenskoj raspodjeli. Radi preciznijeg utvrđivanja sezonskih i prostornih promjena strukture epilitskih dijatomejskih zajednica, potrebni su detaljniji kvalitativni i kvantitativni podaci.

Ključne riječi: Bacillariophyta, plitki oligotrofni zaljev, taksonomska istraživanja, bogatstvo svojti, Bosna i Hecegovina

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INTRODUCTION

Benthic diatoms are colonial or unicellular diatoms, free living or attaching to the substratum by gelatinous extrusion. They are mostly associated with substrata throughout their life cycles. Some of them move actively in sediments and sometimes resuspend in the water column. Benthic diatoms are the most abundant and diversified part of marine microphytobenthos playing an important role in marine ecosystems (FALKOWSKI *et al.*, 2004). These organisms are used as water quality indicators as well as in palaeoecological reconstructions due to their ecophysiological features (CIBIC & BLASUTTO, 2011; STEVENSON & PAN, 1999). In addition, knowledge of the structure of the benthic diatom community and the autoecology of individual taxa is a unique source of information in the study of the dynamics of the marine microphytobenthos.

In the Mediterranean Sea, ÇOLAK SABANCI & KORAY (2010), ÇOLAK SABANCI (2012) and ÁLVAREZ-BLANCO & BLANCO (2014) recently investigated the structure and taxonomic composition of benthic diatom communities. In the Adriatic Sea, the diatom taxa from natural sediment samples and artificial substrata of the Gulf of Trieste (BARTOLE *et al.*, 1991-94; SDRIGOTTI *et al.*, 1999; MUNDA, 2005), the Venice lagoon (TOLOMIO & ANDREOLI, 1989; TOLOMIO *et al.*, 1999; FACCA *et al.*, 2002; TOLOMIO *et al.*, 2002; FACCA & SFRISO, 2007), the northwestern Adriatic coast (TOTTI, 2003; TOTTI *et al.*, 2007; FRANZO *et al.*, 2015, and references therein) were reported. Several studies on epibiotic diatom communities on marine hydroids in the Ligurian Sea have also been reported (ROMAGNOLI *et al.*, 2007, 2014, and references therein). Finally, a checklist on microphytobenthos of the Italian seas, including benthic diatom taxa, was compiled by CIBIC & FACCA (2010).

On the other hand, to the best of our knowledge, the structure and taxonomic composition of benthic diatoms have been only rarely taken into consideration in the eastern Adriatic Sea (BURIĆ *et al.*, 2004; MIHO & WITKOWSKI, 2005; CAPUT *et al.*, 2008; LEVKOV *et al.*, 2010; CAR *et al.*, 2012; NENADOVIĆ *et al.*, 2015; MEJANDŽIĆ *et al.*, 2015). Diatoms were investigated synoptically and mostly related to estuaries or coastal wetlands.

The objective of this work was to contribute to the knowledge of microphytobenthos in the Adriatic Sea, by studying for the first time the benthic diatom communities in Neum Bay in Bosnia and Herzegovina.

MATERIAL AND METHODS

Study area

Bosnia and Herzegovina has a total sea area of ca. 8 km² with only 21.2 kilometers of coastline (Fig. 1). The coast is low and rocky, built up of karstified carbonates and represents only 0.26% of the total length of the Adriatic coastline (BLAKE & TOPALOVIĆ, 1996; PIKEJ & JURAČIĆ, 2013). In fact, the sea area of the country is represented by the small, semi-enclosed Neum Bay, located between mainland and the 7 km-long Klek Peninsula (DUPČIĆ-RADIĆ *et al.*, 2013).

Neum Bay is part of the larger Mali Ston Bay, which is enclosed on the seaward side by the 62 km long Pelješac peninsula. It expands to the northwest and connects with the Neretva River channel which is linked with the open sea. Due to its ecological and economic importance, this area with a centuries-long mariculture tradition was in 1983 proclaimed the Mali Ston Bay and Malo More Special Marine Reserve, including Neum Bay. The most important factors that affect the ecological conditions in Mali Ston Bay,

primarily salinity and nutrient enrichment, are the inflow of freshwater from the Neretva River in the outer part of the bay and from underwater springs ("vruljas") in the inner part (JASPRICA, 1989; JASPRICA *et al.*, 1997; JASPRICA & CARIĆ, 2001; ČALIĆ *et al.*, 2013).

The average and maximum water depth in Neum Bay are 17 and 27 m, respectively. The bottom structure ranges from sandy sediments in the central part of the bay to rocks where the system communicates with the coastline. The town of Neum (population of 3,236 estimated 2013) is the only coastal settlement in Bosnia and Herzegovina and an important tourism destination for the country, which means that the area is under significant anthropogenic pressure during summer.

Additional details on the area are given by JASPRICA *et al.* (2012) and ČALIĆ *et al.* (2013).

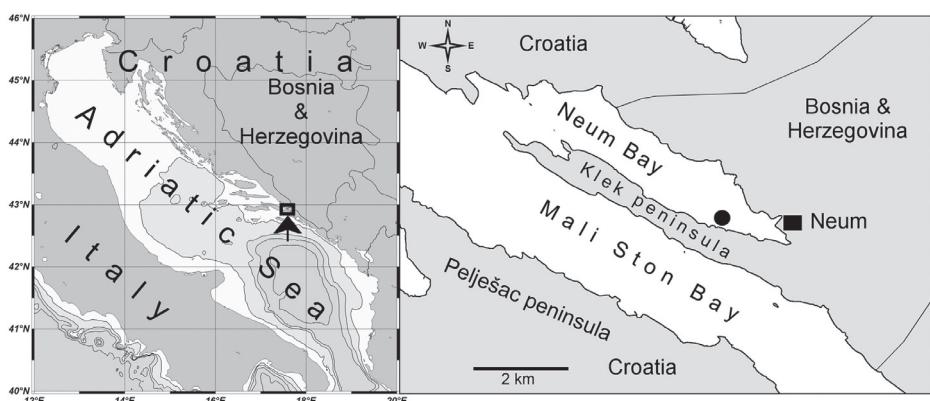


Fig. 1. Sampling location (●) in Neum Bay, Bosnia and Herzegovina.

Sampling and analyses

The investigated station (WGS84 42°54'58.7" N; 17°36'17.6" E) was located in the southern part of Neum Bay (Fig. 1). Samples were collected from a depth of 0.5 m in March, May and July of 2010 and at monthly intervals from January to December 2011 (see Tab. 1 for sampling design). In addition, samples were also collected from a depth of 8 m from April to December 2011. Altogether, 24 samples were analysed.

Temperature was measured with an inverted thermometer. Water samples for salinity were taken by 5-L Niskin bottles and determined by argentometric titration (GRASSHOFF *et al.*, 1983).

Tab. 1. Sampling design of benthic diatom survey conducted in the Neum Bay, Bosnia and Herzegovina (● - sampled; ⊗ - not sampled) in 2010 and 2011.

The sampling points were located at the distance of 1 m for the shallow samples and 25 m for the deeper samples from the coastline due to the small bathymetric gradients. The diatom samples were obtained by scraping of the submersed stones and collecting attached macroalgae [mostly *Cystoseira* spp., *Codium vermilara* (Olivi) Delle Chiaje, *Ceramium* spp., *Dictyota dichotoma* (Hudson) J.V.Lamouroux]. To identify epilithic diatoms, stones of 15–20 cm in diameter were collected. Deeper stones were collected by scuba diving. Stones were collected as randomly as possible amongst those that are not completely covered by filamentous algae and on which the diatom biofilm was visible. Stones were put into a plastic bag of 1 L in which 200 mL of distilled water was added. The upper parts of the stones were rubbed with a toothbrush and the mixture was decanted into 250 mL polyethylene bottles (WINTER & DUTHIE, 2000). The samples were preserved with a final concentration of 4% formaldehyde.

In the laboratory, the material collected was cleaned of organic material for light (LM) and scanning electron microscopy (SEM) observations. Material was treated with sulfuric acid-potassium permanganate, decolorized with oxalic acid, and acid residues were removed by repeated decantation. The cleaned material was pipetted onto ethanol-cleaned cover-slips and left to air-dry, before mounting in Canada balsam when they were ready for light microscopic examination as permanent slides (HASLE & FRYXELL, 1970). In order to register all diatom taxa, the entire surface area of each permanent slide has been surveyed. Whenever possible, diatoms were identified to species rank and counted at 1000 x magnification by phase-contrast optics with a Microstar binocular microscope (AO Scientific Instruments), 100 x PlanApo oil immersion objective. Slides have been deposited in the diatom collection of the Institute for Marine and Coastal Research, University of Dubrovnik, Dubrovnik, Croatia.

Ultrastructural analysis was performed using SEM. A drop of the cleaned sample was air-dried overnight on aluminium stubs and coated with gold-palladium or osmium. SEM observations were made at the Warsaw University of Technology, Faculty of Materials Science and Engineering, using a Hitachi S-3500, SU-70 and SEM/ STEM S-5500.

Identifications were made following PERAGALLO & PERAGALLO (1897–1908), HENDEY (1964), RICARD (1974, 1975, 1977), POULIN *et al.* (1984, 1990), BÉRARD-THERRIAULT *et al.* (1986, 1987), HARTLEY (1986), SNOEIJS (1993, 1999), SNOEIJS & POTAPOVA (1995), SNOEIJS & KASPEROVICIENÉ (1996), SNOEIJS & BALASHLOVA (1998), HARTLEY *et al.* (1996), and WITKOWSKI *et al.* (2000). Nomenclature and data on general environments of taxa follow AlgaeBase (GUIRY & GUIRY, 2017), and only partially ÁLVAREZ-BLANCO & BLANCO (2014).

RESULTS

Environmental conditions

In 2011, surface water temperature varied between 10.0 and 27.1°C, and maximum was recorded in September, while the minimum was noted in January (Tab. 2). The salinity ranged between 32.0 in April and 38.4 in December. Warm summer months were characterized by lower salinity, while in the cooler months of winter and spring salinity was mostly related to the precipitation regime and the extremely dynamic water flow from the underwater karstic springs (“vruljas”) that feed the bay.

Tab. 2. Monthly distribution of the temperature and salinity on the surface (0.5 m depth) in Neum Bay in 2010 and 2011.

Years	2010			2011											
	3	5	7	1	2	3	4	5	6	7	8	9	10	11	12
Months	3	5	7	1	2	3	4	5	6	7	8	9	10	11	12
Temperature (°C)	10.4	15.9	25.2	10.0	10.9	10.2	15.9	15.5	22.5	24.9	26	27.1	23.1	.	15.7
Salinity	36.7	37.2	36.4	34.2	33.5	36.6	32.0	36.9	34.9	35.6	34.7	35.2	.	.	38.4

Taxonomic composition of the benthic diatom community

During this study, a total of 483 specific and infraspecific diatom taxa were identified in Neum Bay (Tab. 3). The list of taxa included 425 pennate and 58 centric diatoms belonging to all three recognized diatom classes: Coscinodiscophyceae, Mediophyceae and Bacillariophyceae (MEDLIN & KACZMarska, 2004), and 60 families (Tab. 3). Among them, two taxa were included in Bacillariophyta *incertae sedis* (*sensu* GUIRY & GUIRY, 2017). Ten families were included in Coscinodiscophyceae, 11 in Mediophyceae and 39 in Bacillariophyceae.

Families with the greatest number of genera were: Naviculaceae (8 genera), Bacillariaceae (6), Achnanthidiaceae (6), Suriellaceae (6), Fragilariaceae (6), Eupodiscaceae (6) and Plagiogrammaceae (4). Additionally, the highest number of species and infraspecific taxa belonged to Naviculaceae (56).

Altogether, 115 genera were found in all samples. Thirteen genera were included in Coscinodiscophyceae, 17 in Mediophyceae and 87 in Bacillariophyceae. Genera with the greatest number of taxa were: *Mastogloia* (46 taxa), *Navicula* (36), *Diploneis* (35), *Nitzschia* (34), *Amphora* (31), *Cocconeis* (27), *Achnanthes* (14), *Halamphora* (12), *Lyrella* (11) and *Suriella* and *Licmophora* (10 each). In total, 24 genera had two taxa each, while 60 were composed of one taxon only. Regarding the habitat type (*sensu* GUIRY & GUIRY, 2017), the greatest number of diatom taxa (152) have been characterized as exclusively marine, 15 are brackish and 29 exclusively freshwater diatoms, while one (*Mastogloia exigua*) has been characterized as exclusively a soil diatom. Among truly marine diatoms, three *Amphora* taxa (*Amphora bigibba* var. *interrupta*, *A. hyalina* and *A. laevissima*) showed high frequency of appearance and were found in more than 50% of the samples.

However, *Amphora bigibba* var. *interrupta* and *Cocconeis scutellum* were the most frequent taxa, being present in 87.5% of the samples. Taxa with high frequencies (79.1–83.3%) were: *Achnanthes brevipes* (83.3%), *Toxarium undulatum* (79.1%) and *Licmophora remulus* (79.1%), while others (centric diatoms *Actinocyclus subtilis*, *Biddulphia biddulphiana*; and pennates *Grammatophora oceanica*, *Halamphora coffeiformis*, *Brebissonia lanceolata*, *Coronia decorus*, *Synedra fulgens*, *Trachyneis aspera* and *Suriella fastuosa*, Fig. 2) were found from 62.5% to 66.7% of the samples. Altogether, 34 taxa were found in at least 50% or more of the samples and could be characterised as taxa with higher frequency of appearance. In total, 142 taxa were found only once (sporadic taxa) during the investigated period (Tab. 3).

Some taxa appeared only in particular seasons and/or depths, but the determination of the relationship between their appearance and abundances, and the ecological conditions in the bay falls outside of the scope of the present paper.

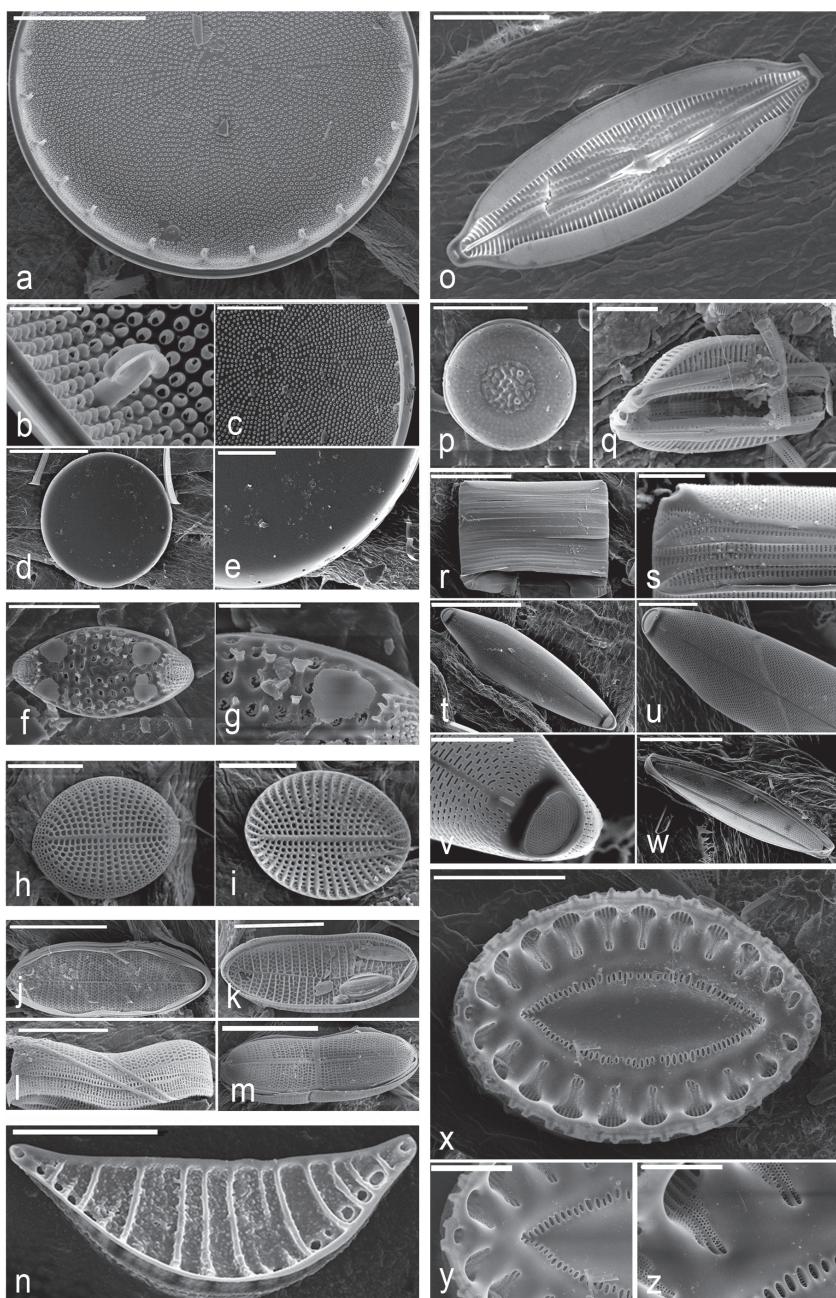


Fig. 2. Scanning electron microscope (SEM) micrographs of benthic diatoms in the Neum Bay: a-e) *Actinocyclus subtilis* (W.Gregory) Ralfs; f-g) *Dimerogramma minus* (W.Gregory) Ralfs; h-i) *Cocconeis scutellum* Ehrenberg; j-m) *Achnanthes brevipes* C.Agardh; n) *Rhopalodia pacifica* Krammer; o) *Mastogloia ignorata* Hustedt; p) *Cyclotella meneghiniana* Kützing; q) *Halamphora coffeiformis* (C.Agardh) Levkov; r-w) *Striatella unipunctata* (Lyngbye) C.Agardh; x-z) *Surirella fastuosa* (Ehrenberg) Ehrenberg. Scale bars = 40 µm (d), 30 µm (j, m, r, t), 20 µm (a, k, l, w, x), 10 µm (c, e, h, i, n, o, u, y), 5 µm (f, p, q, s, v, z), 2 µm (b, g).

Tab. 3. Species and infraspecific taxa of benthic diatoms in Neum Bay in 2010 and 2011, including data on their family-level affiliations, general environment (GE; S – soil taxa, M – marine, B – brackish, F – freshwater, *sensu* Gurny & Gurny, 2017), seasonal distribution in shallow (0.5 m) and deeper (8.0 m) samples, absolute (n) and percentage (%) frequency of appearance.

Taxon	Family	GE						2010						2011						
		S	M	B	F	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	
<i>Bacillaria socialis</i> (Gregory) Ralfs	Bacillariaceae	.	+	+	.	.	+	.	+	.	+	.	+	.	8	33.33
<i>Bacteriastrom</i> sp.1	Chaelocerataceae	.	+	.	+	.	.	+	.	.	+	.	+	.	+	.	+	.	2	8.33
<i>Berkeleyeracaea</i>	Berkelyaceae	.	+	.	+	.	.	+	.	.	+	.	+	.	+	.	+	.	3	12.50
E.J. Cox																				
<i>Bidulphia alternans</i> (Bailey) Van Heurck	Bidulphiaceae	.	+	.	+	.	.	+	.	.	+	.	+	.	+	.	+	.	1	4.17
<i>Bidulphia bidulphioides</i> (J.E.Smith) Boyer	Bidulphiaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	16	66.67
<i>Bidulphia tenuifolia</i> (J.W.Bailey) Roper	Bidulphiaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	4	16.67
<i>Brachystira aponina</i> Kützing	Brachystiraceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Brachystira estonianum</i> Witkowski; Lange-Bentalot & Merzelin	Brachystiraceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	2	8.33
<i>Brekissonia lanceolata</i> (C.Agardh)	Cymbellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	15	62.50
R.K.Mahoney & Reimer																				
<i>Caloneis ablutens</i>	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	2	8.33
<i>Caloneis bicincta</i> (Grunow) Boyer	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	9	37.50
<i>Caloneis clavicularis</i> Zverkov & Metzelin	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Caloneis egenii</i> (A.Schmidt) Cleve	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Caloneis elongata</i> (Grunow) Boyer	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Caloneis excentrica</i> (Grunow) Boyer	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	5	20.83
<i>Caloneis liber</i> (W.Smith) Cleve	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	8	33.33
<i>Caloneis liber</i> var. <i>linearis</i> Cleve	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	10	41.67
<i>Caloneis</i> sp.1	Caloniaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	6	25.00
<i>Canthyladiscus clypeus</i> (Ehrenberg) Ehrenberg ex Kitzing	Surrellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Canthyladiscus echinus</i> Ehrenberg ex Kützing	Surrellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Canthyladiscus fastigiatus</i> Ehrenberg	Surrellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Canthyladiscus immunitatus</i> R.Ross & Aldrin	Surrellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	2	8.33
<i>Canthyladiscus punctulatoides</i> M.Ricard	Surrellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	3	12.50
<i>Canthyladiscus</i> sp.1	Surrellaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	3	12.50
<i>Cannistrum latum</i> (E.Cox) G.Reid	Pleurosigmataceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	2	8.33
<i>Cadambus gallinii</i> (Bory) D.M.Williams & Round	Lichenophoraceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	9	37.50
<i>Centanopsis closterium</i> Ehrenberg	Fragilariae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	2	8.33
<i>Clinacospherina notabilis</i> Ehrenberg	Clinacosphaeriaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Cocconeosis orthoneoides</i> (Hustedt) Wikowski	Cocconeidae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Achnanthidiaceae</i>	Achnanthidiaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Cocconeosis apicalis</i> (Greville) A.W.F.Schmidt	Achnanthidiaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Cocconeosis hyalina</i> Negrelli ex Kitzing	Achnanthidiaceae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Cocconeosis californica</i> Grunow	Cocconeidae	.	+	.	+	.	+	+	.	+	+	.	+	.	+	.	+	.	1	4.17

Tab. 3. continued

Tab. 3. continued

Tab. 3. continued

Tab. 3. continued

Tab. 3. continued

Taxon	Family	GE						2010						2011									
		S			M			March			April			May			June			July			
		S	M	B	F	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
<i>Plagiotropis lepidophora</i> (W.Gregory) Kuntze	Plagiotropidaceae	.	+	+	+	.	+	.	.	+	.	+	.	+	.	+	.	13	54.17
<i>Plagiotropis tarefts</i> T.B.B.Paddock	Plagiotropidaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	12	50.00
<i>Planohlidium delcatulatum</i> (Kützing) Round & Bublikaytova	Achanthidiaceae	.	+	+	+	+	+	.	+	+	.	+	.	+	.	+	.	5	20.83
<i>Planohlidium quaternensis</i> (Grunow) Witkowski, Lange-Bertalot & Metzeltin	Achanthidiaceae	.	+	+	+	+	+	.	+	+	.	+	.	+	.	+	.	10	41.67
<i>Platesia salinaria</i> (Grunow) Lange-Bertalot	Naviculaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Pleurosigna angulatum</i> (J.T.Quekett)	Pleurosignataceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	4	16.67
W.Smith	Pleurosignataceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	7	29.17
<i>Pleurosigna formosan</i> W.Smith	Pleurosignataceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	7	29.17
<i>Pleurosigna itam</i> Ricard	Pleurosignataceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	13	54.17
<i>Pleurosigna</i> sp.1	Pleurosignataceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	4	16.67
<i>Pleurostria lacris</i> (Ehrenberg) Compère	Eupodiscaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	0.00	
<i>Podoctis adriatica</i> (Kützing) Rallis	Fragilariaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Podostis</i> sp.1	Hydrodictyaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	3	12.50
<i>Prestatumnis proricta</i> Grunow	Staurodiaceae	.	+	+	+	.	+	.	+	+	.	+	.	+	.	+	.	0.00	
<i>Protokelia chlonekyi</i> (M.H.Giffen) Round & Basson	Rhopalodiaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Protokelia</i> sp.1	Rhopalodiaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	2	8.33
<i>Protomelissia atlantica</i> R.A.Gibson	Protoraphidaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	7	29.17
<i>Psammadictyon panduriforme</i> (W.Gregory) D.G.Mann	Bacillariaceae	.	+	+	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
<i>Psammadictyon radum</i> (Chodat)	Bacillariaceae	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
D.G.Mann	Psammodesmus nitidus	(W.Gregory) Round	.	+	+	.	+	.	+	+	.	+	.	+	.	+	.	6	25.00
D.G.Mann	<i>Pseudostaurastropsis saccogelarum</i> (Wikowski) E.A.Morales & D.C.Mann	.	+	+	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
Rhizobanina minima	Rhizobanina minima	Kützing	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	10	41.67
Rhizosolenia stellifera	Rhizosolenia stellifera	T.Brightwell	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	2	8.33
Rhizosolenia abreviata	(C.A.Gardil) Lange-Bertalot	.	+	+	+	.	+	.	+	+	.	+	.	+	.	+	.	0.00	
Rhizosolenia linearis	Rhizosolenia linearis	Ostrop	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	2	8.33
Rhizosolenia marinaria	Rhizosolenia marinaria	(Kützing) M.Schmidt	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	3	12.50
Rhizopoda acuminata	Rhizopoda acuminata	Krammer	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	3	12.50
Rhizopoda musculus	Rhizopoda musculus	(Kützing) Otto Müller	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	2	27.50
Rhizopoda sternbergii	Rhizopoda sternbergii	Krammer	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	9	37.50
Rhizopoda pacifica	Rhizopoda pacifica	Krammer	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17
Rhizopoda diadema	Rhizopoda diadema	Krammer	.	+	+	.	.	.	+	.	+	.	+	+	.	+	.	+	.	+	.	1	4.17

Tab. 3. continued

DISCUSSION

In the present study, small forms belonging to some of the most taxonomically complex genera (e.g. *Amphora*, *Navicula* and *Nitzschia*), characterized by a considerable morphological variability, were recognized (SULLIVAN & CURRIN, 2000). A similar diatom community has been reported for different hard substrates from the Adriatic Basin (MUNDA, 2005; TOTTI *et al.*, 2007) and for coastal rocks of the Mediterranean (cf. ÁLVAREZ-BLANCO & BLANCO, 2014). Regarding their ecology, the genus *Navicula* includes taxa with a very wide ecological range (KRAMMER & LANGE-BERTALOT, 1991), while *Amphora* and some *Achnanthes* taxa are mostly typical of nutrient-poor regions (AGATZ *et al.*, 1999). The difficulty in their identification was a major problem in this study. As a matter of fact, more than 40 taxa in Tab. 3 are not fully determined to species level. Additionally, it has been pointed out that some taxa may exhibit more than one growth form, reflecting the different strategies of spatial utilization (cf. ROMAGNOLI *et al.*, 2014, and references therein) adopted for the competitive advantage they provide. In this study, although the samples contained a mixture of forms growing on the stones and some epiphytic forms, we refer to this collective community as epilithon (*sensu* ÇOLAK SABANCI, 2012).

The presence of taxa associated with fresh to brackish waters may be correlated with the precipitation regime and the dynamic water flow from the underground karstic springs ("vruljas") that feed the bay. These taxa were well adapted to this changing environment and these results also support the idea that some species could have different responses to the environmental conditions, resulting in a better or worst adaptation to them (UNDERWOOD *et al.*, 1998; RIBEIRO *et al.*, 2003; MIHO & WITKOWSKI, 2005).

In our study, the genus *Mastogloia* was the richest in taxa number, and the most frequent taxa were *Amphora bigibba* var. *interrupta*, *Cocconeis scutellum*, *Achnanthes brevipes*, *Toxarium undulatum* and *Licmophora remulus*. *Mastogloia* species, one of the largest diatom genera (PENNESI *et al.*, 2011, and references therein), can be found within different biotopes (ÇOLAK SABANCI, 2013). In our study, the most frequent *Mastogloia* was *M. cyclops* which has been characterized as good indicator of coastal zones (WACHNICKA *et al.*, 2010). Some taxa (e.g., *C. scutellum*, *Halamphora coffeiformis*) were found on different substrata and do not seem to have a preference either for geographic region or for the substrate type (ROMAGNOLI *et al.*, 2014). By contrast, *C. scutellum* is considered as typical epiphytic taxon (ULANOVA & SNOEIJIS, 2006).

It is difficult to determine the possible seasonal fluctuations in the assemblages based on qualitative data. Many taxa such as *Achnanthes brevipes*, *Actinocyclus subtilis*, *Grammatophora oceanica* and *Cocconeis scutellum* did not show any kind of marked seasonality. This was also found for several *Cocconeis* taxa (*C. molesta*, *C. placentula*, *C. scutellum*) in the Venice Lagoon (TOLOMIO & ANDREOLI, 1989). Additionally, ÇOLAK SABANCI (2012) in the study based on the seasonal sampling, showed that some of the above mentioned taxa did not have any apparent preference for a given season in the Aegean lagoon in Turkey. In our case, a range of factors can naturally be expected to affect diatom development in the bay, and this research will be investigated in depth in the future.

In our study, the number of taxa was higher than found in floristic surveys conducted in other, similar habitats, in a broad sense and over a wide geographic area in Adriatic or Mediterranean (e.g., MIHO & WITKOWSKI, 2005; ÇOLAK SABANCI, 2011, 2012). Other authors obtained quite different results due to differences in the size of the studied areas, environmental conditions and seasons. In addition, special attention must be paid to the use of different cleaning and processing treatments of benthic marine diatom samples

from Mediterranean oligotrophic areas (VERMEULEN *et al.*, 2012); marine diatoms are likely to be very sensitive to common treatments used in studies of freshwater populations because of the lower availability of silicic acid and of the interference of salinity in silica fixation (CONLEY *et al.*, 1989).

To summarize, this study represents the first floristic list focused on benthic diatoms for Bosnia and Herzegovina. Our analysis shows that, although benthic diatom richness is high in the bay, taxa are apparently not distributed evenly temporally. This is most likely due to the discrete design of the surveys, as many of the rare taxa may occur commonly in unexplored localities of the Adriatic Basin. Also, consistent quantitative and qualitative data are still needed for a better determination of the seasonal and spatial changes of the epilithic assemblages in the region.

From a purely scientific standpoint, it will be intriguing to quantify their contribution to the flow of energy and cycling of material in the bay. More practically, these data will be useful for a rational management of this important regional resource. Namely, the bay is part of a complex ecosystem of the wider area of the Mali Ston Bay (Croatia), which is an important part of the region's natural heritage and they must in fact be protected according to international obligations. Thus, sustainable land management must ensure protection of the centuries-old cultivation of shellfish *Ostrea edulis* in whole of Mali Ston Bay (VILIČIĆ, 2017).

AUTHOR CONTRIBUTIONS

N.J. designed the study and wrote the manuscript, D.H. conducted the field sampling, prepared the samples for counting and analysed the samples using the light microscope, A.C. performed scanning electron microscopy analysis. All authors commented on the manuscript.

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