Synthesis and antiplasmodial evaluation of novel mefloquine-based fumardiamides

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The paper is focused on the synthesis and screening of the antiplasmodial activity of novel fumardiamides **5–10** with the mefloquine pharmacophore and a Michael acceptor motif. Multi-step reactions leading to the title compounds included two amide bond formations. The first amide bond was achieved by the reaction of (*E*)-ethyl 4-chloro-4-oxobut-2-enoate (**1**) and N^1-(2,8-bis(trifluoromethyl)quinolin-4-yl) butane-1,4-diamine (**2**). The obtained ester **3** was hydrolyzed and gave acid **4**, which then reacted with the selected halogenanilines in the presence of HATU/DIEA and formed products **5–10**. Title compounds showed marked, dose dependent activity *in vitro* against hepatic stages of *Plasmodium berghei*. IC_{50} values of the most active compounds **5**, **7** and **9** bearing 3-fluoro, 3-chloro and 3-trifluoromethyl substituents were 3.04–4.16 μmol L^{-1}, respectively. On the other hand, the compounds exerted only weak activity against the erythrocytic stages of two *P. falciparum* strains (*Pf*3D7 and *Pf*Dd2) *in vitro*, with the exception of compound **5** (IC_{50} = 2.9 μmol L^{-1}).

Mefloquine (MQ) was launched more than 30 years ago for the prophylaxis and treatment of malaria caused by the multidrug-resistant *Plasmodium falciparum*. Like other quinoline antimalarials, mefloquine binds to the toxic ferriprotoporphyrin IX and prevents its detoxification (1). In addition, the proposed mefloquine targets include ribosomes, phosphatidylinositol, volume-regulated anion channels and endocytosis. The main disadvantages of mefloquine include its photochemical instability and cardiovascular, hepatic and/or neuropsychiatric side effects (2). Also, resistance to mefloquine, connected with increased expression of P-glycoprotein *Pf*MDR1 in the digestive vacuole, has developed (3).

To overcome mefloquine drawbacks, numerous mefloquine derivatives have been prepared, not only as novel antimalarials (4), but also as potential anticancer agents (5, 6).

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The activity of mefloquine and mefloquine derivatives against *Echinococcus multilocularis* has also been reported (7). More recently, mefloquine has received considerable attention as an antimycobacterial agent (8, 9). It is active against *Mycobacterium avium* and *M. tuberculosis* strains resistant to macrolides, quinolones and rifamycins and shows a synergistic effect with ethambutol and moxifloxacin. Antimycobacterial activity of mefloquine derivatives has also been extensively reported (10–12).

In the last ten years, our research group has designed and prepared more than one hundred primaquine and chloroquine derivatives with urea, bis-urea, acylsemicarbazide, semicarbazide or amide functionalities (13–24). Biological evaluation assays revealed that many of these compounds showed potential as antiproliferative, antioxidative, antitubercular, antiplasmodial or biofilm eradication agents. Further, we have prepared several SAHAquines based on suberoylanilide hydroxamic acid (SAHA) and primaquine motifs, which showed antiplasmodial activity against erythrocytic stages of two *P. falciparum* strains and against *P. berghei* hepatic stages (22).

In our continued pursuit of novel, biologically active quinoline derivatives, we now present the synthesis and evaluation of the antiplasmodial activity of novel fumardiamides in which one amide bond was achieved with N^1-(2,8-bis(trifluoromethyl)quinolin-4-yl)butane-1,4-diamine and the other amide was achieved with the corresponding halogenaniline. Our compounds can be considered mefloquine derivatives, since they bear the same 2,8-bis(trifluoromethyl)quinoline motif as mefloquine. The carbonyl group conjugated with the carbon-carbon double bond of the central part of molecules also classifies these compounds as Michael acceptors, able to react with biological targets bearing thiol groups. We envisaged that combining the mefloquine pharmacophore with Michael acceptor and halogenaniline moieties could result in the development of novel antimalarial hits.

<center>EXPERIMENTAL</center>

General

Melting points were determined on an SMP3 apparatus (Barloworld Scientific, UK) in open capillaries and are uncorrected. A CEM Discover microwave reactor was used for microwave reactions (CEM GmbH, Germany). IR spectra were recorded on a Spectrum One (Perkin-Elmer, UK) and UV-Vis spectra on a Lambda 20 double beam spectrophotometer (Perkin-Elmer, UK). NMR ^{1}H and ^{13}C spectra were recorded at 25 °C on an NMR Avance 600 spectrometer (Bruker, Germany) at 300.13 or 600.13 and 75.47 or 150.9 MHz for ^{1}H and ^{13}C nuclei, resp. Chemical shifts (δ) are reported in parts per million (ppm) relative to tetramethylsilane in the ^{1}H and the dimethyl sulfoxide (DMSO) residual peak as a reference in the ^{13}C spectra (39.51 ppm). Coupling constants (*J*) are reported in Hz. Mass spectra were collected on an HPLC-MS/MS instrument (HPLC, Agilent Technologies 1200 Series; MS, Agilent Technologies 6410 Triple Quad) using electrospray ionization in positive mode (Agilent Technologies, USA). Elemental analyses were performed on a CHNS LECO analyzer (LECO Corporation, USA). All compounds were routinely checked by TLC with Merck silica gel 60F-254 glass plates using dichloromethane/methanol 9.5:0.5, 9:1, 8.5:1.5 and cyclohexane/ethyl acetate/methanol 1:1:0.5 as solvent systems. Spots were visualized by short-wave UV light and iodine vapor. Column chromatography was performed on silica gel 0.063–0.200 mm.

All chemicals and solvents were of analytical grade and were purchased from commercial sources. 4-Chloro-2,8-bis(trifluoromethyl)quinoline, butane-1,4-diamine, (*E*)-4-ethoxy--4-oxobut-2-enoic acid (mono-ethyl fumarate), 3-fluoroaniline, 4-fluoroaniline, 3-chloroaniline, 4-chloroaniline, 3-trifluoromethylaniline, 4-trifluoromethylaniline and triethylamine (TEA) were purchased from Merck GmbH (Germany), and (1-[bis(dimethylamino) methylene]-1*H*-1,2,3-triazolo[4,5-*b*]pyridinium 3-oxid hexafluorophosphate (HATU) and *N,N*-diisopropylethylamine (DIEA) were purchased from Alfa Aesar (USA). Anhydrous solvents were dried and redistilled prior to use.

Syntheses

(E)-*ethyl 4-chloro-4-oxobut-2-enoate (Mono-ethyl fumarate chloride) (***1***)*. – A solution of 0.288 g mono-ethyl fumarate (2 mmol) in 7 mL thionyl chloride was kept overnight and evaporated under reduced pressure. The residue was triturated several times with anhydrous dichloromethane and the solvent was evaporated again. The crude product (0.315 g, 97 %) was used in further reaction without purification.

N^1-*[2,8-bis(trifluoromethyl)quinolin-4-yl]butane-1,4-diamine (***2***)*. – A mixture of 0.599 g (0.02 mol) 4-chloro-2,8-bis(trifluoromethyl)quinoline and 1.763 g 1,4-diaminobutane (0.02 mol) was stirred under microwave irradiation (300 W) at 95 °C. After 0.5 h, the reaction mixture was diluted with dichloromethane, extracted with 5 % NaOH (4 × 40 mL) and washed with water (2 × 40 mL). The organic layer was dried over anhydrous sodium sulfate, filtrated and evaporated under reduced pressure. White solid **2** was obtained (0.632 g, 90 %) and used in further reaction without purification.

*Ethyl (*E*)-4-[(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)amino]-4-oxobut-2-enoate (***3***)*. – A solution of 0.527 g (1.5 mmol) of amine **2** and 0.152 g (1.5 mmol) TEA in 10 mL of anhydrous dichloromethane was added dropwise to chloride **1** dissolved in 10 mL anhydrous dichloromethane. The reaction mixture was stirred for 1 h at room temperature and extracted 3 times with brine. The organic layer was dried over sodium sulfate, filtered and evaporated under reduced pressure. After purification by column chromatography (mobile phase dichloromethane/methanol 9.5:0.5) and crystallization from ether, 0.473 g (66 %) of pale yellow solid **3** was obtained.

*(*E*)-4-[(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)amino]-4-oxobut-2-enoic acid (***4***)*. – A solution of 0.126 g (3 mmol) lithium hydroxide monohydrate in 10 mL water was added to a solution of 0.286 g (0.6 mmol) ester **3** in 10 mL methanol. The reaction mixture was stirred for 3 h at room temperature. Methanol was evaporated under reduced pressure and the aqueous residue was acidified with 10 %-HCl to pH 1. The precipitated product was filtered and washed with water until neutral pH was reached. 0.261 g (97 %) of white solid **4** was obtained.

*Fumardiamides ***5–10***: General procedure*. – A solution of 0.121 g (0.27 mmol) of compound **4**, 0.068 g (0.54 mmol) DIEA and 0.103 g (0.27 mmol) HATU in 1 mL of *N,N*-dimethylformamide was stirred at room temperature. After 10 min, 0.297 mmol of the corresponding halogenaniline was added. The reaction mixture was stirred 2–24 h at room temperature, evaporated under reduced pressure, dissolved in 8 mL ethyl acetate and extracted 3 times with water. The organic layer was dried over sodium sulfate, filtered and evaporated un-

der reduced pressure. The crude products were purified by column chromatography (mobile phase dichloromethane/methanol 9.5:0.5) and crystallized from ether or petroleum ether.

(2E)-N'-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-N-(3-fluorophenyl)but-2-enediamide (5). – Reaction of 0.121 g acid **4** and 0.033 g (0.297 mmol) 3-fluoroaniline; after purification, 0.063 g (43 %) of white solid **5** was obtained.

(2E)-N'-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-N-(4-fluorophenyl)but-2-enediamide (6). – Reaction of 0.121 g acid **4** and 0.033 g (0.297 mmol) 4-fluoroaniline; after purification, 0.056 g (38 %) of white solid **6** was obtained.

(2E)-N'-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-N-(3-chlorophenyl)but-2-enediamide (7). – Reaction of 0.121 g acid **4** and 0.038 g (0.297 mmol) 3-chloroaniline; after purification, 0.072 g (48 %) of white solid **7** was obtained.

(2E)-N'-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-N-(4-chlorophenyl)but-2-enediamide (8). – Reaction of 0.121 g acid **4** and 0.038 g (0.297 mmol) 4-chloroaniline; after purification, 0.045 g (31 %) of white solid **8** was obtained.

(2E)-N'-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-N-[3-(trifluoromethyl)phenyl]but-2-enediamide (9). – Reaction of 0.121 g acid **4** and 0.048 g (0.297 mmol) 3-trifluoromethylaniline; after purification, 0.056 g (35 %) of white solid **9** was obtained.

(2E)-N'-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-N-[4-(trifluoromethyl)phenyl]but-2-enediamide (10). – Reaction of 0.121 g acid **4** and 0.048 g (0.297 mmol) 4-trifluoromethylaniline; after purification, 0.037 g (23 %) of white solid **10** was obtained.

Biological evaluation

In vitro activity against P. berghei *hepatic stages*. – Activity of the synthesized compounds against *P. berghei* infection in a human hepatoma cell line was assessed employing a luminescence-based method, as previously described (25). Briefly, hepatic infection was determined by measuring the luminescence intensity of lysates of Huh7 cells, a human hepatoma cell line, infected with firefly luciferase-expressing *P. berghei* sporozoites (*Pb*-Luc). Huh7 cells ($1.0 × 10^4$ per well) were seeded in 96-well plates on the day before infection. One hour prior to infection, the medium was replaced by medium containing the appropriate drug concentration. Addition of $1.0 × 10^4$ *Pb*-Luc sporozoites was followed by centrifugation at 1800×*g* for 5 min and the parasite infection load was measured 48 h after parasite addition with a bioluminescence assay (Biotium, USA) using a multi-plate reader Infinite M200 (Tecan, Switzerland). The effect of different treatments on the viability of HuH7 cells was assessed using the CellTiter-Blue assay (Promega, USA) according to the manufacturer's protocol. Nonlinear regression analysis was employed to fit the normalized results of the dose-response curves, and IC_{50} values were determined using GraphPad Prism V 5.0.

In vitro drug sensitivity against P. falciparum *erythrocytic stages*. – *In vitro* activity of tested compounds was evaluated against asexual erythrocytic stages of two *P. falciparum*

laboratory strains, 3D7 (chloroquine sensitive) and Dd2 (chloroquine resistant). Antiplasmodial activity of fumardiamides **5–10** was assessed by histidine-rich protein 2 (HRP2) ELISA, as described before (26, 27). As controls, mefloquine hydrochloride (Sigma, USA) and chloroquine diphosphate (Sigma) were used. In brief, 96-well plates were pre-coated with the respective compounds or control drugs at a threefold dilution before highly synchronized ring stage parasites were added into complete culture medium at a hematocrit of 1.5 % and parasitaemia of 0.05 %. After three days of incubation at 37 °C, 5 % CO_2 and 5 % oxygen, plates were freeze thawed three times until analyzed by HRP2-ELISA. All compounds were evaluated in duplicate in at least two independent experiments. The 50 % inhibitory concentration (IC_{50}) was determined by analyzing the nonlinear regression of log concentration-response curves using the drc-package v0.9.0 of R v3.2.2. (28).

Cytotoxicity assay. – Cytotoxicity of novel compounds against the human liver cancer cell line Hep G2 (Leibniz Institute DSMZ – German Collection of Microorganisms and Cell Cultures, Germany) was evaluated using the neutral red assay (29). In brief, human cells were seeded to a 96-well plate in culture medium; on the following day, a serial dilution of the respective compound was added. After one day incubation, cytotoxicity was assessed by addition of Neutral Red and subsequent lysis of cells and measuring absorbance at λ = 570 nm in a plate reader. Inhibitory concentrations were calculated.

RESULTS AND DISCUSSION

Novel fumaric acid diamides **5–10**, bearing 2,8-bis(trifluoromethyl)quinoline and 3- or 4-substituted fluoro-, chloro- or trifluoromethylaniline moieties, were prepared by the simple reaction procedures summarized in Scheme 1. The first precursor, (*E*)-ethyl 4-chloro-4-oxobut-2-enoate) (**1**), was prepared from mono-ethyl fumarate and thionyl chloride, while the second, N^1-(2,8-bis(trifluoromethyl)quinolin-4-yl)butane-1,4-diamine (**2**), was prepared from 4-chloro-2,8-bis(trifluoromethyl)quinoline and 1,4-diaminobutane under microwave irradiation. Compounds **1** and **2** reacted under mild reaction conditions to give product **3**. In this reaction step, TEA was used as a hydrochloride acceptor. Basic hydrolysis
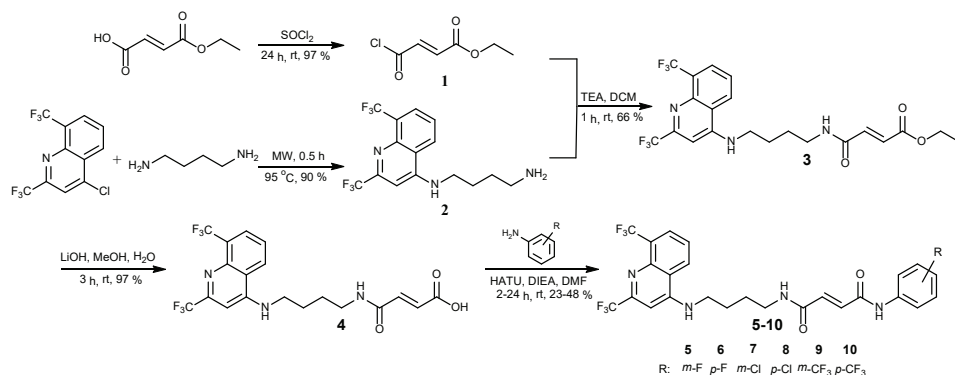
Scheme 1

of **3** afforded the corresponding acid **4**, which reacted with selected halogenanilines and formed the title fumardiamides **5**–**10**. HATU was used as a coupling reagent and DIEA as a base (known as Hünig's base). The common precursor **4** enables relatively easy preparation of diverse target compounds.

Purification of compounds was carried out using crystallization methods and/or column chromatography. Yields for the first two reaction steps were high, but rather low for the last one, 23–48 %.

Structures of the new compounds were confirmed by ^{1}H, ^{13}C NMR and IR spectra and additionally by elemental analysis. Chemical shifts, multiplicities and coupling constants confirmed the proposed structures. Analytical and spectral data of the target compounds **5**–**10** and their precursors are given in Table I. Atom numbering used in the current study is indicated in Fig. 1.
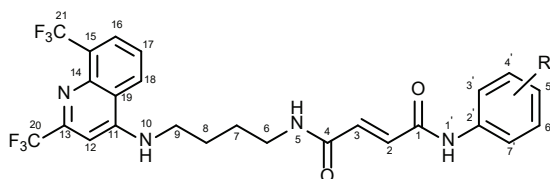

Fig. 1. Atom numbering of compounds **5**–**10**.

In ^{1}H NMR spectra, NH-1′ signals were visible as sharp singlets between δ 10.78 and 10.45, NH-5 signals as triplets at 8.55–8.47 ppm, while NH-10 signals were mixed with aromatic H-atoms. All NH signals were D_2O exchangeable. In ^{13}C NMR spectra, the CF_3 group at position 21 had a signal between δ 129.41 and 116.72 ppm and between 127.11 and 116.23 ppm at position C-20. Two additional CF_3 groups in compounds **9** and **10** appeared at similar ppm values (δ 130.08-129.00 and 128.72-127.75 ppm). All CF_3 signals were quartets with high coupling constants (277.70-272.73 Hz). The carbon atoms bearing trifluoromethyl groups, *i.e.*, atoms C-15, C-13 and C-4′ or C-5′, in compounds **9** and **10** showed quartets with markedly lower coupling constants (33.22-27.30 Hz), while fluorosubstituted carbons 4′ and 5′ in compounds **5** and **6** appeared as doublets at δ 163.71-160.51 and 159.03-157.44 ppm. Signals of the analogous chlorosubstituted carbons in compounds **7** and **8** were downshifted at δ 133.12 and 127.38 ppm. In all compounds, carbonyl C-atoms C-1 and C-4 appeared between δ 163.41 and 162.16 ppm. IR spectra revealed the presence of two sets of amide I and amide II signals between 1652 and 1512 cm^{-1}.

Chemical structure of the new compounds was also supported by mass spectrometry analysis. Molecular ion peaks corresponding to the expected molecular masses were obtained for all compounds. *m/z* data and the expected relative molecular masses are given in Table I, while all IR, NMR and mass spectra are available as Supplementary material.

We have also applied Chemicalize.org (30) and SwissADME programs (31) to compute physicochemical descriptors as well as to predict ADME parameters, pharmacokinetic properties, drug-like nature and medicinal chemistry friendliness of our fumardiamides. A set of physicochemical parameters, *i.e.*, number of atoms, molecular mass (MW), partition coefficient (log *P*), number of H-bond donors (HBD), number of H-bond acceptors (HBA), molar refractivity (MR) and topological polar surface area (TPSA) are given in Table

Table I. Analytical and spectral data of compounds 5–10

Cmpd.	M.p. (°C)	IR (KBr) v_{max} (cm^{-1})	^{1}H NMR (DMSO-d_6, δ ppm, J/Hz)	^{13}C NMR (DMSO-d_6, δ ppm, J/Hz)	MS (m/z)	Molecular formula (M_r)	CHN analysis calcd. found (%)		
3	174–175	3358, 3088, 2962, 2868, 2362, 1712, 1676, 1652, 1596, 1558, 1502, 1444, 1368, 1316, 1228, 1166, 1114, 1046, 1020, 964, 936, 908, 864, 818, 768, 718, 686, 618, 546	8.66 (d, 1H, 16, J = 8.4), 8.55 (t, 1H, 5, J = 5.4), 8.16 (d, 1H, 18, J = 72), 7.98 (t, 1H, 17, J = 5.2), 7.69 (t, 1H, 10, J = 7.9), 7.00 (d, 1H, 3, J = 15.5), 6.89 (s, 1H, 12), 6.57 (d, 1H, 2, J = 15.5), 4.19 (q, 1H, 1', J = 7.1), 3.47-3.36 (m, 1H, 9), 3.26 (q, 1H, 6, J = 5.8), 1.77-1.67 (m, 2H, 7), 1.64-1.55 (m, 2H, 8), 1.33-1.17 (t, 3H, 2')	164.95 (4), 162.71 (1), 152.23 (14), 147.94-147.73 (q, 13, J = 32.13), 143.78 (11), 137.46 (18), 130.17-121.27 (q, 21, J = 272.02), 128.91-128.71 (q, 16, J = 5.93), 128.07 (2), 126.86 (3), 126.76-125.88 (q, 15, J = 27.30), 126.57-117.64 (q, 20, J = 274.05), 124.31 (17), 119.31 (19), 94.08 (12), 60.50 (1'), 42.12 (9), 38.40 (6), 26.34 (7), 24.92 (8), 13.87 (2')	478.1 [MH]$^+$	C$_{21}$H$_{21}$F$_6$N$_3$O$_3$ (477.15)	52.83 52.79	4.43 4.47	8.80 8.82
4	212–215	3432, 3392, 2948, 2872, 3686, 2562, 1712, 1674, 1598, 1536, 1442, 1326, 1294, 1182, 1112, 1048, 974, 930, 820, 762, 722, 346, 622, 552	12.86 (s, 1H, 1'), 8.65 (d, 1H, 16, J = 8.5), 8.51 (t, 1H, 5, J = 5.5), 8.15 (d, 1H, 18, J = 7.2), 7.99 (t, 1H, 17, J = 5.2), 7.68 (t, 1H, 10, J = 7.9), 7.05-6.79 (d+s, 2H, 3, 12), 6.52 (d, 1H, 2, J = 15.5), 3.49-3.19 (m, 4H, 9, 6), 1.78-1.52 (m, 4H, 7, 8)	166.50 (4), 163.08 (1), 152.32 (14), 148.53-147.30 (q, 13, J = 31.03), 143.86 (11), 137.05 (18), 129.45 (2), 129.37-118.69 (q, 21, J = 271.78), 129.12-128.88 (q, 16, J = 5.55), 127.04-117.82 (q, 20, J = 272.64), 127.00 (3), 126.92-125.80 (q, 15, J = 28.22), 119.38 (19), 94.19 (12), 42.20 (9), 38.45 (6), 26.50 (7), 25.01 (8)	450.0 [MH]$^+$	C$_{19}$H$_{17}$F$_6$N$_3$O$_3$ (449.12)	50.79 50.66	3.81 3.79	9.35 9.40
5	230–231	3472, 3298, 3146, 3072, 2948, 2878, 2362, 1652, 1645, 1598, 1544, 1494, 1442, 1312, 1192, 1120, 1048, 976, 950, 864, 820, 760, 718, 668, 620, 596, 564, 516	10.64 (s, 1H, 1'), 8.66 (d, 1H, 16, J = 8.3), 8.53 (t, 1H, 5, J = 5.7), 8.15 (d, 1H, 18, J = 7.2), 7.99 (t, 1H, 17, J = 5.2), 7.68 (dd, 2H, 10, 3', J = 9.6, 5.2), 7.43-7.33 (m, 2H, 6, 7'), 7.04 (dd, 2H, 2, 3, J = 20.23, 3.87), 6.95-6.87 (m, 2H, 12, 5'), 3.43 (dd, 2H, 9, J = 12.1, 6.4), 3.26 (dd, 2H, 6, J = 12.3, 6.4), 1.79-1.67 (m, 2H, 7), 1.66-1.55 (m, 2H, 8)	163.71-160.51 (d, 4', J = 246.5), 163.34 (4), 162.60 (1), 152.32 (14), 148.56-147.25 (q, 13, J = 33.22), 143.85 (11), 140.56-140.41 (d, 2', J = 11.07), 134.37 (18), 132.35 (2), 130.58-130.45 (d, 6', J = 8.41), 129.41-118.63 (q, 21, J = 275.38), 129, 15-128.78 (q, 16, J = 7.43), 127.11-116.23 (q, 20, J = 277.70), 127.00 (3), 126.89-126.51 (q, 15, J = 27.30), 124.46 (17), 119.37 (19), 115.16 (7'), 110.72-109.9 (q, 3', J = 20.74), 106.32-105.97 (d, 5', J = 26.64), 94.23 (12), 42.22 (9), 38.51 (6), 26.54 (8), 25.06 (7)	565.1 [M+Na]$^+$	C$_{25}$H$_{21}$F$_7$N$_4$O$_2$ (542.45)	55.35 55.27	3.90 3.99	10.33 10.51

No.	m.p. (°C)	IR	^{1}H NMR	^{13}C NMR	MS	Formula (M_r)	C	H	N
6	249–252	3458, 3284, 3150, 3078, 2948, 2874, 1638, 1596, 1546, 1512, 1438, 1408, 1312, 1216, 1124, 1046, 986, 950, 836, 760, 718, 686, 666, 618, 554, 514	10.45 (s, 1H, 1'), 8.64 (d, 1H, 16, J = 8.3), 8.47 (t, 1H, 5, J = 5.5), 8.14 (d, 1H, 18, J =7.2), 7.95 (t, 1H, 17, J = 5.2), 7.80-7.63 (m, 3H, 10, 3', 7'), 7.19-7.13 (m, 2H 4', 6'), 6.99 (dd, 2H, 2, 3, J = 34.03, 18.25), 6.87 (s, 1H, 12), 3.41 (q, 2H, 9, J = 6.8), 3.25 (q, 2H, 6, J = 7.0), 1.75-1.68 (m, 2H, 7), 1.64-1.55 (m, 2H, 8)	163.41 (4), 162.16 (1), 159.03-157.44 (d, 5', J = 240.38), 152.28 (14), 148.41-147.03 (q, 13, J =33.21), 143.81 (11), 135.18 (2'), 133.90 (18), 132.53 (2), 128.96-128.81 (q, 16, J = 5.77), 128.44-119.48 (q, 21, J = 274.97), 126.92 (3), 126.85-125.82 (q, 15, J = 32.4), 126.07-117.14 (q, 20, J = 272.73), 124.38 (17), 121.08-121.03 (d, 3', 7', J =7.71), 119.34 (19), 115.43-115.28 (d, 4', 6', J = 21.3), 94.14 (12), 42.19 (9), 38.44 (6), 26.50 (8), 25.03 (7)	543.0 [MH]$^+$	$C_{25}H_{21}F_7N_4O_2$ (542.45)	55.35 55.15	3.90 3.74	10.33 10.29
7	235–237	3484, 3302, 3184, 3122, 3068, 2946, 2882, 1640, 1636, 1598, 1544, 1478, 1438, 1408, 1388, 1312, 1218, 1190, 1122, 1050, 1028, 986, 950, 912, 888, 864, 834, 762, 720, 662, 564, 516	10.59 (s, 1H, 1'), 8.65 (d, 1H, 16, J = 8.5), 8.52 (t, 1H, 5, J = 5.6), 8.14 (d, 1H, 18, J =7.2), 7.97 (t, 1H, 17, J = 5.3), 7.90 (s, 1H, 3'), 7.67 (t, 1H, 10, J = 7.9), 7.50 (d, 1H, 7', J = 8.3), 7.36 (t, 6', J = 7.9, 6', J = 8.1), 7.15 (dd, 1H, 5', J = 7.9, 1.2), 7.01 (dd, 2H, 2, 3, J = 26.90, 9.90), 6.88 (s, 1H, 12), 3.42 (q, 2H, 9, J = 6.7), 3.26 (q, 2H, 6, J =6.7), 1.77-1.68 (m, 2H, 7), 1.65-1.56 (m, 2H, 8)	163.33 (4), 162.60 (1), 152.30 (14), 148.43-147.35 (q, 13, J = 32.90), 143.83 (11), 140.19 (2'), 134.39 (18), 133.12 (4'), 132.27 (2), 130.51 (6'), 128.99-128.84 (q, 16, J = 5.36), 128.44-119.53 (q, 21, J = 272.82), 126.94 (3), 126.84-125.87 (q, 15, J = 29.88), 126.13-117.15 (q, 20, J = 275.42), 124.42 (17), 123.50 (5'), 119.36 (19), 118.79 (7'), 117.76 (3'), 94.18 (12), 42.21 (9), 38.49 (6), 26.50 (7), 25.04 (8)	559.1 [MH]$^+$	$C_{25}H_{21}ClF_6N_4O_2$ (558.90)	53.72 53.54	3.79 3.98	10.02 9.88
8	267–268	3470, 3294, 3186, 3120, 3070, 2934, 2874, 1640, 1636, 1598, 1544, 1492, 1438, 1400, 1312, 1220, 1166, 1126, 1048, 1012, 980, 882, 762, 718, 668, 620, 564, 508	10.56 (s, 1H, 1'), 8.66 (d, 1H, 16, J = 5.5), 8.52 (t, 1H, 5, J = 5.5), 8.16 (d, 1H, 18, J =7.2), 7.99 (t, 1H, 17, J = 4.9), 7.69 (dd, 3H, 10, 3', 7', J = 11.9, 8.63), 7.40 (dd, 2H, 4', 6', J = 8.8), 7.01 (dd, 2H, 2, 3, J = 23.75, 5.29), 6.89 (s, 1H, 12), 3.43 (dd, 2H, 9, J =12.0, 6.4), 3.26 (dd, 2H, 6, J = 12.3, 6.2), 1.80-1.67 (m, 2H, 7), 1.67-1.54 (m, 2H, 8)	163.38 (4), 162.41 (1), 152.32 (14), 148.55-147.26 (q, 13, J = 32.72), 143.85 (11), 137.76 (2'), 134.18 (18), 132.44 (2), 129.41-118.62 (q, 21, J = 273.33), 129.14-128.82 (q, 16, J = 5.34), 128.76 (3', 7'), 127.38 (5'), 127.06-116.27 (q, 20, J = 274.67), 126.99 (3), 126.91-125.79 (q, 15, J = 28.75), 124.45 (17), 120.86 (4', 6'), 119.37 (19), 94.22 (12), 42.22 (9), 38.50 (6), 26.54 (7), 25.06 (8)	559.0 [MH]$^+$	$C_{25}H_{21}ClF_6N_4O_2$ (558.90)	53.72 53.59	3.79 3.55	10.02 9.92

No.	mp (°C)	IR	¹H NMR	¹³C NMR	MS	Formula (M)	C	H	N
9	230–233	3460, 3284, 3164, 3094, 29420, 2876, 2362, 1650, 1645, 1598, 1558, 1494, 1446, 1312, 1172, 1120, 1070, 974, 950, 888, 800, 760, 720, 698, 662, 618, 594, 518	10.76 (s, 1H, 1'), 8.66 (d, 1H, 16, J = 8.3), 8.56 (t, 1H, 5, J = 5.6), 8.21-8.121 (m, 2H, 18, 3'), 8.00 (t, 1H, 17, J = 5.3), 7.84 (d, 1H, 10, J = 8.4), 7.68 (t, 1H, 7', J = 79), 7.59 (t, 1H, 6, J = 8.0), 7.45 (d, 1H, 5', J = 7.8), 7.03 (dd, 2H, 2, 3, J = 23.74, 5.72), 6.90 (s, 1H, 12), 3.43 (dd, 2H, 9, J = 12.3, 6.6), 3.27 (dd, 2H, 6, J = 12.6, 6.5), 1.76-1.69 (m, 2H, 7), 1.66-57 (m, 2H, 8)	163.30 (4), 162.80 (1), 152.32 (14), 148.51-147.32 (q, 13, J = 33.07), 143.86 (11), 139.56 (2'), 134.53 (18), 132.24 (2), 131.70-121.80 (q, 8', J = 276.06), 130.14 (6), 130.08-129.00 (q, 4', J = 29.26), 129.07-128.86 (q, 16, J = 5.53), 127.00 (3), 126.86-125.86 (q, 15, J = 27.99), 126.64-116.72 (q, 21, J = 276.06), 126.26-116.36 (q, 20, J = 276.06), 124.46 (17), 122.92 (7'), 120.28-120.02 (q, 5', J = 2.08), 119.38 (19), 115.48-115.24 (q, 3', J = 4.15), 94.22 (12), 42.22 (9), 38.24 (6), 26.54 (7), 25.07 (8)	593.1 [MH]⁺	$C_{26}H_{21}F_9N_4O_2$ (592.46)	52.71 / 52.84	3.57 / 3.81	9.46 / 9.68
10	271–273	3486, 3302, 3130, 3080, 2962, 2938, 2876, 1650, 1645, 1600, 1540, 1438, 1412, 1328, 1220, 1188, 1156, 1110, 1068, 1018, 974, 950, 920, 838, 760, 718, 680, 620, 594, 564, 510	10.78 (s, 1H, 1'), 8.66 (d, 1H, 16, J = 8.3), 8.55 (t, 1H, 5, J = 5.6), 8.15 (d, 1H, 18, J = 7.2), 7.99 (t, 1H, 17, J = 5.3), 7.88 (d, 2H, 4', 6', J = 8.5), 7.69 (dd, 3H, 10, 3, 7', J = 16.6, 8.3), 7.05 (dd, 2H, 2, 3, J = 24.88, 8.14), 6.89 (s, 1H, 12), 3.43 (dd, 2H, 9, J = 12.3, 6.6), 3.27 (dd, 2H, 6, J = 12.7, 6.7), 1.78-1.68 (m, 2H, 7), 1.66-1.56 (m, 2H, 8)	163.30 (4), 162.86 (1), 152.32 (14), 148.49-147.30 (q, 13, J = 32.96), 143.86 (11), 142.34 (2'), 134.67 (18), 132.24 (2), 129.19-119.45 (q, 8', J = 271.11), 129.09-128.84 (q, 16, J = 5.77), 128.94-119.11 (q, 21, J = 273.93), 128.72-127.75 (q, 5', J = 27.00), 127.00 (3), 126.64-116.70 (q, 20, J = 276.74), 126.30-126.02 (q, 4', 6', J = 3.01), 124.44 (17), 124.33-123.19 (q, 15, J = 31.41), 119.38 (19), 119.30 (3', 7'), 94.25 (12), 42.22 (9), 38.52 (6), 26.53 (7), 25.06 (8)	593.1 [MH]⁺	$C_{26}H_{21}F_9N_4O_2$ (592.46)	52.71 / 52.99	3.57 / 3.86	9.46 / 9.35

Table II. Calculated physiochemical descriptors of diamides 5–10 (30)

Cmpd.	Molecular formula	Number of atoms	M_r	log P	HBD	HBA	MR (cm^3 mol^{-1})	TPSA (Å^2)
5	$C_{25}H_{21}F_7N_4O_2$	59	542.458	5.10	3	4	129.11	83.12
6	$C_{25}H_{21}F_7N_4O_2$	59	542.458	5.10	3	4	129.11	83.12
7	$C_{25}H_{21}ClF_6N_4O_2$	59	558.910	5.57	3	4	133.70	83.12
8	$C_{25}H_{21}ClF_6N_4O_2$	59	558.910	5.57	3	4	133.70	83.12
9	$C_{26}H_{21}F_9N_4O_2$	62	592.466	5.84	3	4	134.87	83.12
10	$C_{26}H_{21}F_9N_4O_2$	62	592.466	5.84	3	4	134.87	83.12

M_r – relative molecular mass; log P – partition coefficient; HBD – H-bond donor; HBA – H-bond acceptor; MR – molecular refractivity; TPSA – topological polar surface area

II (30). All compounds showed minimum aberration in molecular masses and log P, while the other four parameters are fully in agreement with Lipinski's and Gelovani's rules. SwissADME bioavailability radars for the most active compounds **5**, **7** and **9** are given in Fig. 2 (31). Radars enable a first glance at the drug-likeness of the molecules. The pink area represents the optimal range for each property: lipophilicity (LIPO) (XLOGP3 between −0.7 and +5.0), molecular mass (SIZE) (between 150 and 500 g mol^{-1}), polarity (POLAR) (TPSA between 20 and 130 Å^2), solubility (INSOLU) (log S not higher than 6), saturation (INSATU) (fraction of carbons in the sp^3 hybridization not less than 0.25), and flexibility (FLEX) (no more than 9 rotatable bonds). All three compounds are too flexible and are thus predicted as not orally bioavailable.

Biological evaluation

To evaluate the antiplasmodial potential of the title fumardiamides, *in vitro* assays against two strains of *P. falciparum* erythrocytic stages, *Pf*3D7 and *Pf*Dd2, and against *P. berghei* hepatic stages were performed. Our results showed that only compound **5** displayed marked activity against *P. falciparum* blood stages, with an IC_{50} of 2.9 μmol L^{-1} (Table III). On the other hand, all the compounds evaluated in this work showed promising antiplasmodial *in vitro* activity against *P. berghei*. Fumardiamides were tested at 1 and 10 μmol L^{-1}, and DMSO was used as a negative control (Fig. 3). The most active compounds (**5**, **7** and **9**) had very similar IC_{50} values, ranging between 3.04 and 4.16 μmol L^{-1} (Fig. 4), indicating that the type of halogen (fluorine or chlorine) had no influence on the compound activity against the parasite's hepatic stages, but had a slight impact on cytotoxicity. Among the three most active compounds, 3-fluoro derivative **5** showed the best activity/toxicity ratio (Table IV). Replacement of the halogen atom with CF$_3$ group caused slight perturbations of electron density of the aromatic scaffold, but this change also had a negligible effect on the activity. Comparison of the structural isomers **5** and **6**, **7** and **8**, and **9** and **10** revealed that *meta*-substituted derivatives were slightly more active than the analogous *para*-substituted derivatives. Cytotoxicity studies performed on the human liver cancer cell line (HepG2) cells revealed low cytotoxicity of fumardiamides **5–10**. These studies
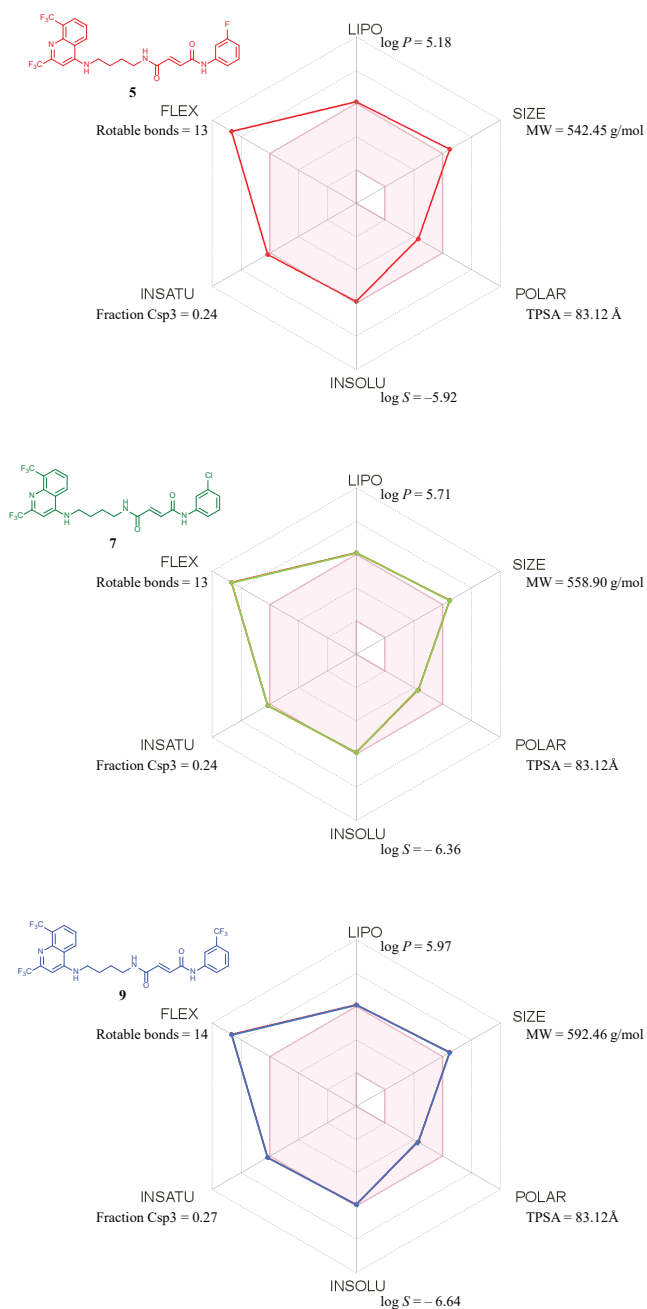
Fig. 2. Bioavailability radars for the most active compounds **5**, **7** and **9**. The pink area represents the optimal range for each property (lipophilicity, size, polarity, solubility, saturation and flexibility).
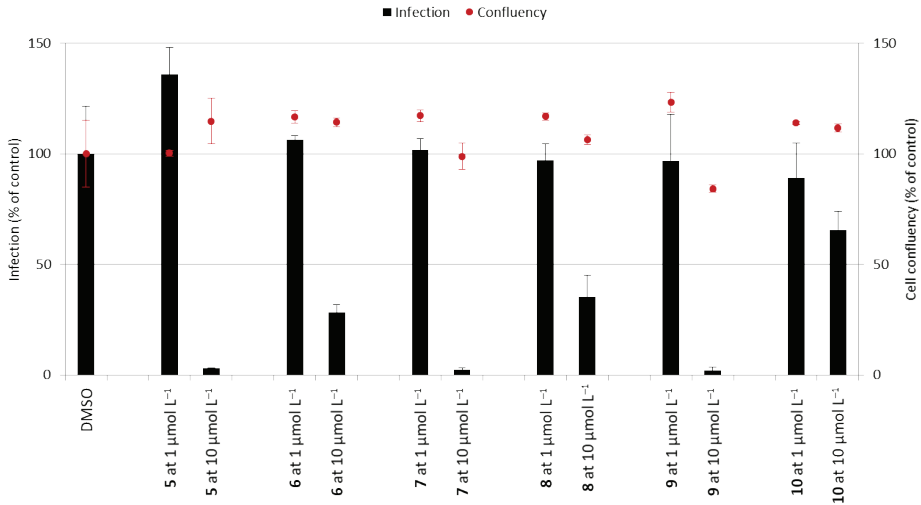
Fig. 3. Activity of compounds **5–10** against *P. berghei* liver-stages at 1 and 10 µmol L^{-1} concentrations. Anti-infective activity (infection scale, bars) is shown. Results represent mean ± SD, *n* = 1.
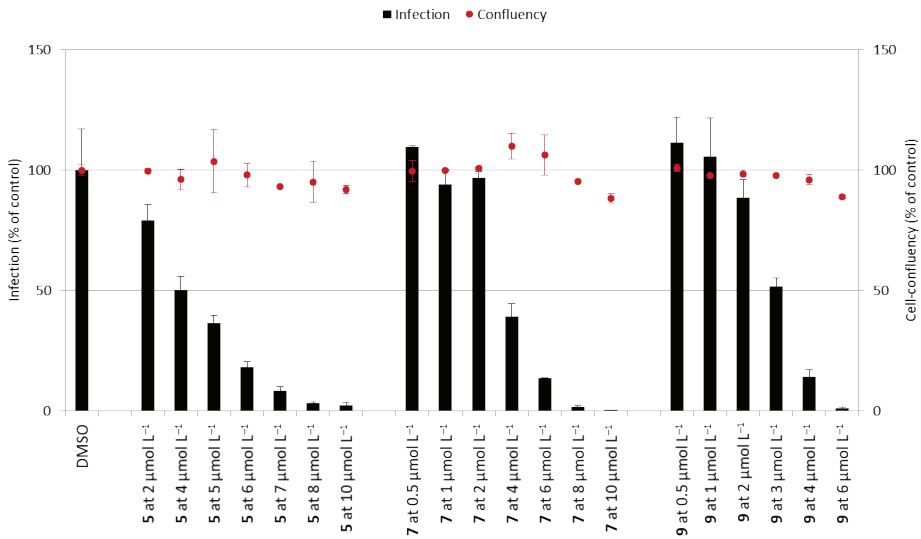

Fig. 4. *IC$_{50}$* of selected MQ-derivatives against *P. berghei* hepatic stages. *IC$_{50}$* = 4.16 (**5**), 3.64 (**7**) and 3.04 (**9**) µmol L^{-1}. Results represent mean ± SD, *n* = 1.

are only preliminary. To get a better insight into the toxicity of prepared compounds, more cell lines should be tested and additional cytotoxicity tests should be performed.

Table III. IC$_{50}$ *values of compounds* **5–10** *against the erythrocytic stage of two* P. falciparum *strains*
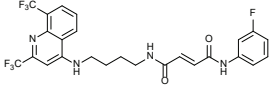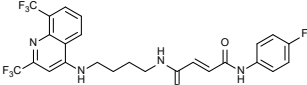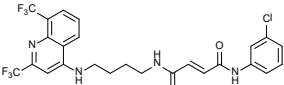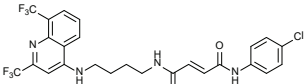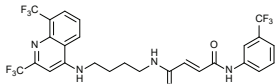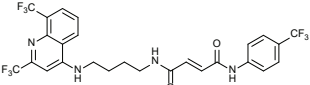
Cmpd.	Chemical structure	IC$_{50}$ (μmol L^{-1})	
		*Pf*3D7	*Pf*Dd2
5		2.9	15.2
6		> 13.0	> 13.0
7		18.5	> 13.0
8		> 13.0	> 13.0
9		> 13.0	16.9
10		> 13.0	> 13.0
Mefloquine		6.5 × 10^{-3}	6.6 × 10^{-3}
Chloroquine		3.7 × 10^{-3}	0.2
Primaquine		2.0	1.8

Table IV. Cytotoxicity screening of compounds **5–10** *towards the human liver cancer cell line (Hep G2)*

Cmpd.	IC$_{50}$ (μmol L^{-1})
5	64.1 ± 54.2
6	> 100
7	10.0 ± 3.6
8	> 100
9	13.0 ± 2.6
10	> 100
Primaquine	148.0 ± 3.5

CONCLUSIONS

Novel fumardiamides **5–10** with the mefloquine pharmacophore and Michael acceptor motif were designed and synthesized through multi-step reactions. Three compounds, namely (2*E*)-*N'*-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-*N*-(3-fluorophenyl) but-2-enediamide (**5**), (2*E*)-*N'*-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl]amino}butyl)-*N*-(3-chlorophenyl)but-2-enediamide (**7**) and (2*E*)-*N'*-(4-{[2,8-bis(trifluoromethyl)quinolin-4-yl] amino}butyl)-*N*-[3-(trifluoromethyl)phenyl]but-2-enediamide (**9**), provided fairly promising results in the *in vitro* bioassay against *P. berghei* liver stages, while compound **5** was also active against the blood stages of two *P. falciparum* strains. These compounds could serve as starting points for further optimization and development of more potent agents.

Supplementary material is available upon request.

Abbreviations, acronyms, symbols. – CQ – chloroquine; DIEA – *N,N*-diisopropylethylamine; HATU – 1-[bis(dimethylamino)methylene]-1*H*-1,2,3-triazolo[4,5-*b*]pyridinium 3-oxid hexafluorophosphate; HBA – H-bond acceptor; HBD – H-bond donor; Hep G2 – human liver cancer cell line; MQ – mefloquine; PQ – primaquine; TEA – trimethylamine; TPSA – topological polar surface area.

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