

Influence of Gum Arabic Enriched with GABA Coating on Oxidative Damage of Walnut Kernels

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SUMMARY

Because of the higher content of unsaturated fatty acids (UNSFAs) and phenolics, walnut kernels are vulnerable to oxidative rancidity and browning due to unfavorable post-harvest handling procedures. This study investigates the impact of gum arabic coating enriched with γ -aminobutyric acid (GABA) on oxidative rancidity and browning of kernels during storage at 20 °C. The results showed that the walnut kernels coated with gum arabic (5 %) enriched with GABA (0.1 mM) exhibited lower oxidative rancidity and browning, manifested by lower peroxide value and malondialdehyde accumulation along with higher whiteness index. Moreover, kernels had higher UNSFA/SFA ratio as a response to lower lipoxygenase activity and H₂O₂ accumulation. The reduced oxidative browning in coated kernels was accompanied with lower polyphenol oxidase and higher phenylalanine ammonia-lyase activity leading to higher accumulation of phenolics and increased DPPH[•] scavenging capacity. Based on our findings, gum arabic coating (5 %) enriched with GABA (0.1 mM) may have a commercial potential for maintaining nutritional quality of walnut kernels.

Key words: DPPH scavenging capacity, γ -aminobutyric acid, gum arabic coating, postharvest nutritional quality, walnut kernel

INTRODUCTION

Owing to higher unsaturated fatty acid (UNSFAs) content, walnut kernels are nutritionally important in human diet. However, it makes walnut kernels vulnerable to oxidative rancidity under unfavorable postharvest handling (1). Oxidative rancidity is a result of UNSFA peroxidation by reactive oxygen species (ROS) accumulation or lipoxygenase (LOX) enzyme activity (2,3). Besides, oxidative rancidity is stimulated by a higher oxygen availability, higher temperatures and also by a long storage period. Peroxide and malondialdehyde (MDA) are the primary and secondary products of UNSFA peroxidation. Therefore, the peroxide value (PV) and MDA accumulation are often used as oxidative rancidity markers (2,3).

Under unfavorable postharvest handling procedures, in addition to oxidative rancidity, kernel browning occurs due to the oxidation of phenolics by polyphenol oxidase (PPO) (4). In walnuts, light kernel colour is a crucial sensory factor for the consumer preference. Oxidative rancidity and browning deteriorate the sensory and nutritional quality of walnut and cause their unpleasant off-flavour and shorten their shelf life, which is also unsafe to consumer health (5,6).

The kernel oxidative rancidity (due to high O₂ availability) and surface browning by UNSFA peroxidation via ROS/LOX and phenolic oxidation by PPO are responsible for the nutritional quality deterioration and for reducing the sensory quality (7). Reducing the O₂ availability by using edible coatings would be beneficial for attenuating oxidative rancidity and preventing browning of walnut kernels; resulting in safer kernels with high quality (6). The results obtained by Deng *et al.* (8) confirm that γ -aminobutyric acid (GABA), as an antioxidant, together with MDA inhibit the formation of reactive carbonyl intermediates during oxidative stress. The enrichment of edible coatings with powerful antioxidants such as GABA may help synergistically to preserve the kernel quality (7,9). Owing to the economic

value and nutritional importance of walnut kernels, great efforts have been made by researchers to attenuate oxidative rancidity, maintain sensory and nutritional quality, and extend their shelf life by applying edible coatings combined with antioxidants such as soy protein coating enriched with catechin (7) and chitosan enriched with green tea (10), which now have potential commercial applications.

In this study, the impact of gum arabic coating enriched with GABA on the oxidative rancidity and browning of walnut kernels during storage at 20 °C for 18 weeks were evaluated. Research emphasis was on mass fractions of fatty acids and total phenolics during storage. Kernel colour changes and enzyme (phenylalanine ammonia-lyase (PAL), PPO and LOX) activity were also included in the measured parameters. Peroxide value (PV) and malondialdehyde (MDA) accumulation in kernels were also studied.

MATERIALS AND METHODS

Walnut kernels and treatments

Walnut fruits (*Juglans regia* cv. Chandler) were harvested from fully mature walnut orchard from Karaj, Alborz province, Iran. After the harvest, the fruits were immediately carried to the postharvest physiology of horticultural crops laboratory at the University of Tehran, Karaj, Iran. Then, the green husk and the hard shell of walnut fruits were manually removed. In a preliminary experiment, unshelled nuts, without being dried, were coated with 0, 5, 10, 15 and 20 % (m/V) gum arabic alone or enriched with γ -aminobutyric acid (GABA) at a concentration of 0, 0.01, 0.1, 1 and 10 mM. Gum arabic solution was prepared according to Ali *et al.* (11) by dissolving 0, 5, 10, 15 and 20 g of gum arabic powder in 100 mL distilled water. The solutions were stirred at low heat (40 °C) for 60 min, then filtered to remove any undissolved impurities. The pH of the solutions was adjusted to 5.6 by 1 M NaOH.

GABA and gum arabic powder, food grade (kibble size: KB 120, i.e. <125 micron), was supplied by Labortecnica SA (Almeria, Spain). All other chemicals were purchased from Sigma-Aldrich, Merck (Madrid, Spain) and were of analytical grade. Different concentrations (0, 0.01, 0.1, 1 and 10 mM) of GABA were added to gum arabic solution according to Khaliq *et al.* (12). For coating treatments, healthy walnut kernels, macroscopically free from disorders and disease symptoms were placed in a mesh container and immersed in the coating solutions for 1 min. The coated walnut kernels were then dried for 3 h at room temperature and 100 g of kernel halves were kept at 20 °C for 6, 12 and 18 weeks (10).

In the treated kernels, peroxide value (PV) and malondialdehyde (MDA) content (as oxidative rancidity markers) increased and kernel surface whiteness index (WI), as oxidative browning marker, decreased during storage at 20 °C for 18 weeks (data not shown). Coating resulted in the lowest kernel oxidative rancidity and browning. Based on these results, the solutions of gum arabic 5 and 10 % with the addition of 0.1 and 1 mM GABA, respectively, were selected for further study. PV,

MDA accumulation and WI were evaluated every 6 weeks during storage at 20 °C. Phenylalanine ammonia-lyase (PAL) and polyphenol oxidase (PPO) enzyme activity, total phenolic accumulation, DPPH[•] scavenging capacity and LOX enzyme activity were also investigated.

Kernel surface whiteness

Kernel colour was measured on the upper part of the outer surface on each half by chromatometer (CR-400; Minolta, Tokyo, Japan) using CIE colour parameters L^* , a^* and b^* , and WI was calculated according to the following equation (13):

$$WI=100-[(100-L^*)^2+a^{*2}+b^{*2}]^{1/2} \quad /1/$$

Kernel PV and MDA content

Lipid extraction was measured according to Lee *et al.* (14). PV was measured by iodometric titration with 0.01 N Na₂S₂O₃ according to AOAC method 965.33 (15) using 1 g of lipids separated from walnut kernels and expressed in mmol peroxide per kg kernel oil. MDA content was measured by the thiobarbituric acid (TBA) method described by Nepote *et al.* (16) and expressed in nmol TBA per g dry mass.

Total phenolic content and DPPH[•] scavenging capacity

Total phenolic content was determined according to the Folin-Ciocalteu procedure (17). The absorbance of the samples was measured by UV/Vis spectrophotometer (PC-1650; Shimadzu Kyoto, Japan) and the content was expressed in mg gallic acid equivalents (GAE) per g dry mass (dm). Free radical DPPH[•] scavenging activity was traced according to Brand-Williams *et al.* (18). The reduction percentage of DPPH[•] was calculated according to the following equation:

$$\text{Inhibition of DPPH}^{\bullet} = [(A_{\text{control}} - A_{\text{sample}}) / A_{\text{control}}] \cdot 100 \quad /2/$$

where A_{control} is the absorbance of DPPH[•] solution without extract.

Fatty acid mass fraction

Fatty acids were quantified by forming fatty acid methyl esters according to Arena *et al.* (19), using gas chromatograph equipped with flame ionization detector (GC-FID model 6890 N; Agilent Technologies, Santa Clara, CA, USA). Fatty acid content was expressed in percentage as mass fraction in oil. The unsaturated/saturated fatty acid ratio was calculated by the formula:

$$\text{UNSFA/SFA} = (w(18:1) + w(18:2) + w(18:3)) / (w(16:0) + w(18:0)) \quad /3/$$

where according to C:D number 16:0 is palmitic, 18:0 stearic, 18:1 oleic, 18:2 linoleic and 18:3 linolenic acid.

PAL and PPO activities

PAL was assayed based on cinnamic acid production according to Nguyen *et al.* (20), and was expressed in μmol

cinnamic acid per mg protein per h. PPO was assayed by measuring the oxidation of catechol as substrate according to Nguyen *et al.* (20), and was expressed in U/mg protein.

LOX activity and H₂O₂ content

LOX activity was measured by linoleic acid as substrate according to Phetsirikoon *et al.* (21), and was expressed in U/mg protein. The H₂O₂ content was measured according to Patterson *et al.* (22) and expressed in μmol/g dm.

Statistical analysis

The experiment was conducted as split plot in time based on completely randomized design with three replications (100 g of kernels for each replication). Data analysis was performed using IBM SPSS software v. 21 (23). Differences were assessed by Tukey's test at p<0.05.

RESULTS AND DISCUSSION

Oxidative rancidity and browning

During storage at 20 °C for 18 weeks, we recorded a significant increase in PV and MDA accumulation and a decrease in WI of kernels (Table 1), indicating oxidative rancidity and browning incidence manifested in kernels by irreversible UNSFA peroxidation *via* ROS/LOX and phenolic oxidation by PPO. Kernels coated with gum arabic (5 %) enriched with GABA (0.1 mM) exhibited lower PV and MDA accumulation and higher WI than other treatments during storage at 20 °C for 18 weeks (p<0.01; Table 1). Coating with 5 % gum arabic

containing 0.1 mM GABA resulted in the lowest kernel oxidative rancidity and browning. Thus, it can be clearly indicated that gum arabic coating enriched with GABA could strongly protect kernel UNSFA peroxidation and phenolic oxidation, attenuating kernel oxidative rancidity and browning.

Fatty acid profile, LOX activity and H₂O₂ accumulation

As shown in Table 2, linoleic, linolenic and oleic acid contents decreased during storage at 20 °C for 18 weeks, while palmitic and stearic acid contents increased. As a consequence, unsaturated/saturated fatty acid (UNSFA/SFA) ratio decreased. Meanwhile, coated kernels with lower palmitic and stearic acid contents (p<0.01) and higher linoleic, linolenic and oleic acid contents (p<0.01) had higher UNSFA/SFA ratio during storage at 20 °C for 18 weeks (Table 2). Furthermore, kernels coated with 5 % gum arabic and 0.1 mM GABA had lower LOX activity (p<0.01) and H₂O₂ accumulation (p<0.01) during storage (Table 3). Oxidative rancidity of walnut kernels is a consequence of the peroxidation of UNSFA by ROS/LOX, and a decline in UNSFA/SFA ratio was concurrent with the higher PV and MDA accumulation, as oxidative rancidity markers. In addition to the oxidation of kernels, UNSFA and LOX can participate in O₂^{•-} accumulation (24). Therefore, overcoming the kernel oxidative rancidity is achievable by decreasing O₂ availability to prevent ROS production using edible coatings or by scavenging ROS radicals (7–10). Kang *et al.* (7) reported that the walnut kernels coated with soy protein enriched with antioxidant catechin exhibited significantly lower UNSFA oxidation with the lower PV and MDA accumulation during storage at 35 °C for 21 days, which was the result of the synergistic action of

Table 1. Oxidative rancidity and browning of walnut kernels coated with gum arabic enriched with γ-aminobutyric acid (GABA) during storage at 20 °C for 18 weeks

w(gum arabic)/%	c(GABA)/mM	t/week	Oxidative rancidity and browning marker		
			WI	PV/(mmol/kg)	b(MDA)/(nmol/g)
0	0	0	(54.3±0.3)	(0.7±0.1)	(8.3±1.3)
		6	(52.3±0.9) ^{abc}	(1.3±0.1) ^e	(11.3±1.2) ^d
		12	(48±0.6) ^d	(2.6±0.1) ^b	(14.8±1.2) ^b
		18	(41.7±0.9) ^e	(4.1±0.2) ^a	(16.1±0.5) ^a
0	1	6	(50.3±0.3) ^{cd}	(0.8±0.1) ^f	(10.3±1.1) ^e
		12	(48.7±0.4) ^d	(2.0±0.) ^d	(13.1±0.8) ^c
		18	(45±0.7) ^e	(2.8±0.1) ^b	(15.9±1.1) ^b
10	0	6	(53.7±0.4) ^{ab}	(0.8±0.01) ^f	(10.4±1.3) ^e
		12	(50.5±0.5) ^{bcd}	(2.1±0.2) ^c	(12.6±0.5) ^d
		18	(48±0.6) ^d	(2.9±0.2) ^b	(14.3±1.2) ^b
5	0.1	6	(54±0.6) ^a	(0.7±0.2) ^f	(9.2±1.1) ^f
		12	(52.3±0.9) ^{ab}	(1.8±0.2) ^d	(11.7±1.1) ^d
		18	(50±0.5) ^c	(2.3±0.1) ^c	(14.0±0.6) ^c
Significant		df			
Time		2	**	*	**
Treatment		3	**	**	**
T×T		6	**	**	**
CV		-	2.35	8.80	8.06

Data are presented as mean value±standard error of three replications. Different letters indicate significant differences at p=0.05. * and **significance at p=0.05 and 0.01, respectively. WI=whiteness index, PV=peroxide value per kernel oil mass, MDA=malondialdehyde expressed as thiobarbituric acid per dry mass

Table 2. Fatty acid mass fraction in walnut kernels coated with gum arabic enriched with γ -aminobutyric acid (GABA) during storage at 20 °C for 18 weeks

w(gum arabic)/%	c(GABA)/mM	t/week	w(fatty acid)/%					
			Palmitic	Stearic	Oleic	Linoleic	Linolenic	UNSAF/SFA
0	0	0	(8.0±0.4)	(2.8±0.6)	(22.9±0.5)	(52.3±0.5)	(13.0±0.7)	(8.7±0.1)
		6	(9.4±0.7) ^d	(3.8±0.1) ^e	(21.2±0.2) ^c	(51.5±1.2) ^b	(11.3±0.4) ^e	(6.4±0.1) ^e
		12	(9.8±0.1) ^c	(4.1±0.2) ^c	(19.5±0.4) ^f	(46.9±1.7) ^d	(10.5±0.6) ^f	(5.5±0.1) ^h
0	1	6	(9.4±0.2) ^d	(3.2±0.4) ^g	(21.4±0.9) ^c	(51.1±0.3) ^b	(12.1±0.3) ^{bc}	(6.7±0.1) ^d
		12	(9.7±0.4) ^c	(3.8±0.2) ^{de}	(19.9±0.4) ^e	(47.4±1.5) ^d	(11.3±0.9) ^e	(5.8±0.1) ^g
		18	(10.2±0.1) ^b	(4.3±0.3) ^b	(17.9±0.4) ^g	(42.3±0.5) ^g	(9.3±0.8) ^g	(4.8±0.1) ⁱ
10	0	6	(8.4±0.1) ^g	(3.2±0.5) ^g	(22.3±0.3) ^b	(52.8±1.2) ^a	(12.3±1.4) ^b	(7.5±0.1) ^b
		12	(9.1±0.1) ^e	(3.5±0.5) ^f	(21.4±0.8) ^c	(49.5±1.5) ^c	(11.7±1.1) ^{de}	(6.5±0.1) ^e
		18	(9.5±0.1) ^d	(3.9±0.4) ^{cd}	(20.3±1.1) ^d	(43.6±1.2) ^f	(10.5±1.1) ^f	(5.6±0.1) ⁱ
5	0.1	6	(8.1±0.1) ^h	(2.9±0.4) ^h	(22.8±0.3) ^a	(53.0±1.3) ^a	(12.9±0.6) ^a	(8.1±0.1) ^a
		12	(8.8±0.1) ^f	(3.2±0.2) ^g	(22.3±0.8) ^b	(51.3±1.3) ^b	(12.2±0.7) ^b	(7.2±0.2) ^c
		18	(9.1±0.2) ^e	(3.4±0.7) ^f	(21.4±0.4) ^c	(45.4±1.2) ^e	(11.7±0.5) ^{cd}	(6.2±0.1) ^f
Significant		df						
Time		2	**	**	**	**	**	**
Treatment		3	**	**	**	**	**	**
T×T		6	**	**	**	**	**	*
CV		-	1.21	1.64	1.44	1.96	1.35	1.09

Data are presented as mean value±standard error of three replications. Different letters indicate significant differences at $p=0.05$. * and **significance at $p=0.05$ and 0.01 , respectively. UNSFA/SFA=unsaturated/saturated fatty acid ratio

Table 3. Oxidative markers of walnut kernels coated with gum arabic enriched with γ -aminobutyric acid (GABA) during storage at 20 °C for 18 weeks

w(gum arabic)/%	c(GABA)/mM	t/week	Oxidative stress marker	
			LOX activity/(U/mg)	$b(\text{H}_2\text{O}_2)$ /($\mu\text{mol/g}$)
0	0	0	(10.3±0.3)	(14.3±0.3)
		6	(24±1.4) ^{defg}	(20.2±0.4) ^{cde}
		12	(36.1±1.4) ^{bc}	(24.7±0.4) ^b
		18	(50.6±1.3) ^a	(31.4±0.5) ^a
0	1	6	(20.7±0.5) ^{fgh}	(19.9±0.5) ^{cde}
		12	(31.9±1.0) ^{bcd}	(22.9±1.1) ^{bcd}
		18	(33.1±0.4) ^{bc}	(30.1±0.4) ^a
10	0	6	(17.0±0.9) ^{gh}	(17.4±0.4) ^{ef}
		12	(29.7±0.7) ^{bcde}	(25.5±1.5) ^b
		18	(37.5±1.1) ^b	(31.5±1.2) ^a
5	0.1	6	(12.9±1.2) ^h	(14.5±1.2) ^f
		12	(22.8±1.1) ^{efg}	(19.2±0.9) ^{def}
		18	(28.6±1.3) ^{cdef}	(23.3±1.2) ^{bc}
Significant		df		
Time		2	**	**
Treatment		3	**	**
T×T		6	*	*
CV		-	10.84	6.21

Data are presented as mean value±standard error of three replications. Different letters indicate significant differences at $p=0.05$. * and **significance at $p=0.05$ and 0.01 , respectively. LOX=lypoxigenase

O_2 barrier function of soy protein and antioxidant activity of catechin. The authors suggested that the edible soy protein coating enriched with catechin delayed UNSFA peroxidation of walnut kernels by protecting them from O_2 exposure during storage. Another study carried out by Vidrih *et al.* (25) showed that the unstable linolenic acid content decreased due to its oxidation during storage, while the other fatty acids remained

largely unaffected. Sabaghi *et al.* (10) reported that walnut kernels coated with chitosan enriched with antioxidant green tea exhibited a lower UNSFA peroxidation, confirmed by lower PV and MDA accumulation and higher sensory quality observable in lower kernel surface browning during storage at room temperature for 18 weeks, which was a consequence of the synergistic action of O_2 barrier function of chitosan and the

antioxidant activity of green tea. Christopoulos and Tsantili (3) reported that during storage at 20 °C for 9 months, walnut kernel UNSFA/SFA ratio decreased and the oleic, linoleic and linolenic acid peroxidation resulted in kernel rancidity assayed by the higher PV, as oxidative kernel rancidity marker. Additionally, they reported that walnut kernels stored at low temperature (1 °C) and packaged under N₂ or CO₂ exhibited lower PV, as oxidative kernel rancidity marker, which may be a consequence of the lower oleic, linoleic and linolenic acid peroxidation, resulting in higher kernel UNSFA/SFA ratio. Our results suggest that the debilitation of oxidative rancidity in walnut kernels coated with gum arabic (5 %) enriched with GABA (0.1 mM) may be caused by a lower LOX enzyme activity and lower H₂O₂ accumulation. Eventually, higher UNSFA/SFA ratio may be an outcome of the synergistic action of gum arabic, which lowers O₂ availability, with GABA, which scavenges the ROS (26). This suggests that the gum arabic coatings enriched with GABA delayed lipid oxidation of walnuts by protecting them from oxygen exposure during storage period.

Total phenolic accumulation, DPPH scavenging capacity and PAL/PPO activities

As shown in Table 4, walnut kernels coated with 5 % gum arabic enriched with GABA (0.1 mM) exhibited higher total phenolic accumulation ($p < 0.01$) and DPPH scavenging capacity ($p < 0.01$) during storage at 20 °C for 18 weeks. Moreover, kernels had lower PPO ($p < 0.01$) and higher phenylalanine ammonia-lyase (PAL) enzyme activity ($p < 0.01$).

In walnuts, light kernel colour is crucial for sensory quality attributes leading to consumer preference. According to

Christopoulos and Tsantili (27), phenolics are accumulated in walnut kernels by the activation of phenylpropanoid pathway through PAL enzyme activity, which is responsible for non-oxidative deamination of phenylalanine to *trans*-cinnamic acid. Walnut kernel phenolics have a high antioxidant capacity, which contributes to a higher DPPH or FRAP scavenging capacity. Phenolics are crucial for the human health and play a crucial role in attenuating oxidative UNSFA rancidity in the walnut kernels by obstructing ROS accumulation (28,29). However, under unfavourable postharvest handlings and due to membrane integrity damage caused by MDA accumulation, higher phenolic accumulation in walnut kernels may contribute to surface browning in kernels, as a result of the increased PPO (enzyme responsible for browning) activity, which leads to a lower consumer acceptability and reduced sensory quality (3,10,30). In line with oxidative rancidity, kernel surface browning as a consequence of phenolic oxidation by PPO results in a huge reduction in kernel antioxidant capacity (31). Fuentealba *et al.* (32) reported that the walnut kernel light colour was accompanied with higher phenolic accumulation resulting from lower phenolic oxidation by PPO activity and seemingly attributed higher DPPH scavenging capacity. Likewise, light colour kernels exhibit a higher arbutin accumulation and antioxidant activity, which is crucial for attenuating the browning incidence by hampering tyrosinase enzyme activity. Along with higher phenolic accumulation, light walnut colour was accompanied with higher linolenic acid accumulation, revealing the lessening of oxidative rancidity *via* antioxidant phenolics (32). Moreover, Wang *et al.* (4) reported that the green hull of fresh walnuts in the hull packed in modified atmosphere

Table 4. Phenolic metabolism of walnut kernels coated with gum arabic enriched with γ -aminobutyric acid (GABA) during storage at 20 °C for 18 weeks

w(gum arabic)/%	c(GABA)/mM	t/week	Phenolic metabolism			
			w(total phenolics as GAE)/(mg/g)	DPPH ⁺ scavenging capacity/%	PAL activity/ (μ mol/mg-h)	PPO activity/ (U/mg)
0	0	0	(54.4 \pm 0.9)	(65.5 \pm 0.9)	(62.4 \pm 1.1)	(9.1 \pm 0.8)
		6	(35.7 \pm 0.8) ^{bcd}	(52.1 \pm 0.2) ^{cd}	(36.8 \pm 0.8) ^d	(12.8 \pm 0.7) ^{ef}
		12	(26.9 \pm 0.6) ^{ef}	(42.1 \pm 0.3) ^f	(22.3 \pm 0.3) ^g	(28.1 \pm 1.3) ^c
		18	(19.4 \pm 1.5) ^f	(35.9 \pm 0.3) ^g	(17.9 \pm 0.2) ^h	(66.1 \pm 1.3) ^a
0	1	6	(41.0 \pm 1.9) ^b	(55.7 \pm 0.3) ^b	(41.9 \pm 0.3) ^c	(11.0 \pm 1.8) ^f
		12	(33.0 \pm 1.8) ^{cde}	(49.2 \pm 0.3) ^d	(36.1 \pm 0.4) ^d	(16.9 \pm 1.1) ^e
		18	(29.6 \pm 1.3) ^{de}	(45.9 \pm 0.6) ^e	(28.9 \pm 0.4) ^f	(37.6 \pm 0.2) ^b
10	0	6	(38.2 \pm 1.3) ^{bc}	(54.9 \pm 0.6) ^b	(45.9 \pm 0.4) ^b	(11.3 \pm 0.1) ^f
		12	(28.1 \pm 0.6) ^e	(50.4 \pm 0.9) ^{cd}	(38.2 \pm 0.2) ^d	(14.6 \pm 0.2) ^{ef}
		18	(20.3 \pm 0.9) ^f	(45.3 \pm 0.3) ^e	(30.7 \pm 0.4) ^{ef}	(31.9 \pm 0.1) ^c
5	0.1	6	(52.6 \pm 0.6) ^{ef}	(60.2 \pm 1.5) ^a	(57.4 \pm 1.6) ^a	(9.5 \pm 0.2) ^f
		12	(48.2 \pm 0.6) ^a	(55.6 \pm 0.4) ^b	(44.9 \pm 0.9) ^b	(11.4 \pm 0.1) ^f
		18	(39.1 \pm 0.6) ^{bc}	(53.1 \pm 0.6) ^{bc}	(32.1 \pm 0.3) ^e	(22.7 \pm 0.3) ^d
Significant		df				
Time		2	**	**	**	**
Treatment		3	**	**	**	**
TxT		6	ns	**	**	**
CV		-	7.39	2.15	2.92	7.52

Data are presented as mean value \pm standard error of three replications. Different letters indicate significant differences at $p = 0.05$. * and **significance at $p = 0.05$ and 0.01, respectively. GAE=gallic acid equivalents, PAL=phenylalanine ammonia-lyase, PPO=polyphenol oxidase

packaging during storage at 1 °C for 60 days exhibited higher phenolic and flavonoid accumulation, leading to higher FRAP scavenging capacity derived from higher PAL/PPO enzyme activity, which diminished the decay of fresh walnut in the hull and maintained kernel quality with lower PV (4). Christopoulos and Tsantili (31) reported that the walnut kernels stored at 20 °C for 9 months exhibited a higher surface browning, reflected by the lower L^* , h^o and WI, along with lower FRAP and DPPH' scavenging capacity due to phenolic oxidation. In comparison, walnut kernels stored at low temperature (1 °C) and packed under N₂ or CO₂ exhibited a lower kernel surface browning and higher FRAP and DPPH' scavenging capacity owing to the increased phenolic accumulation, as the consequence of reduced phenolic oxidation by the low temperature and/or low O₂ availability (30). Christopoulos and Tsantili (31) suggested that walnut kernel surface browning at a higher temperature and O₂ availability may be attributed to lower FRAP and DPPH' scavenging capacity and the lower phenolic accumulation caused by the higher phenolic oxidation. Based on the data from the present experiment, to overcome the oxidative browning in walnut kernels, coating with gum arabic (5 %) enriched with GABA (0.1 mM) would be an applicable method due to higher PAL/PPO enzyme activity concurrent with higher phenolic accumulation and higher DPPH' scavenging capacity, mainly due to the synergistic behaviour of gum arabic coating function through lowering O₂ availability with the ROS scavenging capacity of GABA. In addition to phenolics, GABA itself exhibits a strong ROS scavenging activity and may contribute to reduction of oxidative browning of walnut kernels (25).

CONCLUSIONS

In conclusion, the present study sheds light on the beneficial impacts of gum arabic coating enriched with γ -aminobutyric acid (GABA) on attenuating the oxidative rancidity and browning of walnut kernels stored at 20 °C for 18 weeks. The results of this study demonstrate that coating of nuts with 5 % gum arabic enriched with GABA (0.1 mM) could control kernel lipid oxidation and undesirable phenolic changes during storage at 20 °C, as compared to uncoated walnut kernels. The reduction of oxidative rancidity and browning of walnut kernels by gum arabic coating enriched with GABA may be the result of lower lipoxigenase activity along with lower H₂O₂ accumulation, giving rise to higher UN-SFA/SFA ratio, and of higher PAL/PPO enzyme activity, giving rise to higher phenolic accumulation and higher DPPH' scavenging capacity. Taken together, synergistic action of gum arabic coating, which lowers O₂ availability, and the antioxidant function of GABA may be responsible for the reduction of oxidative rancidity and browning of walnut kernels.

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REFERENCES

- Martínez ML, Labuckas DO, Lamarque AL, Maestri DM. Walnut (*Juglans regia* L.): Genetic resources, chemistry, by-products. *J Sci Food Agr*. 2010;90(12):1959–67. <https://doi.org/10.1002/jsfa.4059>
- Wang S, Ikediala JN, Tang J, Hansen JD, Mitcham E, Mao R, Swanson D. Radio frequency treatments to control codling moth in in-shell walnuts. *Postharvest Biol Technol*. 2001;22(1):29–38. [https://doi.org/10.1016/S0925-5214\(00\)00187-3](https://doi.org/10.1016/S0925-5214(00)00187-3)
- Christopoulos MV, Tsantili E. Oil composition in stored walnut cultivars – Quality and nutritional value. *Eur J Lipid Sci Technol*. 2015;117(3):338–48. <https://doi.org/10.1002/ejlt.201400082>
- Wang J, Li P, Gong B, Li S, Ma H. Phenol metabolism and preservation of fresh in-hull walnut stored in modified atmosphere packaging. *J Sci Food Agr*. 2017;97(15):5335–42. <https://doi.org/10.1002/jsfa.8422>
- Piccirillo P, Fasano P, Mita G, De Paolis A, Santino A. Exploring the role of lipoxigenases on walnut quality and shelf-life. *Acta Hort*. 2005;705:543–5. <https://doi.org/10.17660/ActaHortic.2005.705.79>
- Shahidi F, John JA. Oxidative rancidity in nuts. In: Harris LJ, editor. *Improving the safety and quality of nuts*. Cambridge, UK: Woodhead Publishing Ltd; 2013. pp. 198–229. <https://doi.org/10.1533/9780857097484.2.198>
- Kang HJ, Kim SJ, You YS, Lacroix M, Han J. Inhibitory effect of soy protein coating formulations on walnut (*Juglans regia* L.) kernels against lipid oxidation. *LWT – Food Sci Technol*. 2013;51(1):393–6. <https://doi.org/10.1016/j.lwt.2012.10.019>
- Deng Y, Xu L, Zeng X, Li Z, Qin B, He N. New perspective of GABA as an inhibitor of formation of advanced lipoxidation end-products: It's interaction with malondiadehyde. *J Biomed Nanotechnol*. 2010;6(4):318–24. <https://doi.org/10.1166/jbn.2010.1130>
- Aghdam MS, Naderi R, Jannatizadeh A, Sarcheshmeh MAA, Babalar M. Enhancement of postharvest chilling tolerance of anthurium cut flowers by γ -aminobutyric acid (GABA) treatments. *Sci Hort*. 2016;198:52–60. <https://doi.org/10.1016/j.scienta.2015.11.019>
- Sabaghi M, Maghsoudlou Y, Khomeiri M, Ziaifar AM. Active edible coating from chitosan incorporating green tea extract as an antioxidant and antifungal on fresh walnut kernel. *Postharvest Biol Technol*. 2015;110:224–8. <https://doi.org/10.1016/j.postharvbio.2015.08.025>

11. Ali A, Maqbool M, Alderson PG, Zahid N. Effect of gum arabic as an edible coating on antioxidant capacity of tomato (*Solanum lycopersicum* L.) fruit during storage. *Postharvest Biol Technol.* 2013;76:119–24.
<https://doi.org/10.1016/j.postharvbio.2012.09.011>
12. Khaliq G, Muda Mohamed MT, Ghazali HM, Ding P, Ali A. Influence of gum arabic coating enriched with calcium chloride on physiological, biochemical and quality responses of mango (*Mangifera indica* L.) fruit stored under low temperature stress. *Postharvest Biol Technol.* 2016;111:362–9.
<https://doi.org/10.1016/j.postharvbio.2015.09.029>
13. Boun HR, Huxsoll CC. Control of minimally processed carrot (*Daucus carota*) surface discoloration caused by abrasion peeling. *J Food Sci.* 1991;56(2):416–8.
<https://doi.org/10.1111/j.1365-2621.1991.tb05293.x>
14. Lee MS, Lee SH, Song KB. Effect of various natural antioxidants on the safflower oil. *Korean J Food Preserv.* 2004;11:126–9.
15. AOAC Official Method 965.33. Peroxide value of oils and fats. Official final action. Rockville, MD, USA: AOAC International; 1994.
16. Nepote V, Mestrallet MG, Grosso NR. Natural antioxidant effect from peanut skins in honey-roasted peanuts. *J Food Sci.* 2004;69(7):295–300.
<https://doi.org/10.1111/j.1365-2621.2004.tb13632.x>
17. Singleton VL, Orthofer R, Lamuela-Raventós RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods Enzymol.* 1999;299:152–78.
[https://doi.org/10.1016/S0076-6879\(99\)99017-1](https://doi.org/10.1016/S0076-6879(99)99017-1)
18. Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. *LWT – Food Sci Technol.* 1995;28(1):25–30.
[https://doi.org/10.1016/S0023-6438\(95\)80008-5](https://doi.org/10.1016/S0023-6438(95)80008-5)
19. Arena E, Campisi, S, Fallico B, Maccarone E. Distribution of fatty acids and phytosterols as a criterion to discriminate geographic origin of pistachio seeds. *Food Chem.* 2007;104(1):403–8.
<https://doi.org/10.1016/j.foodchem.2006.09.029>
20. Nguyen TBT, Ketsa S, van Doorn WG. Relationship between browning and the activities of polyphenoloxidase and phenylalanine ammonia lyase in banana peel during low temperature storage. *Postharvest Biol Technol.* 2003;30(2):187–93.
[https://doi.org/10.1016/S0925-5214\(03\)00103-0](https://doi.org/10.1016/S0925-5214(03)00103-0)
21. Phetsirikoon S, Ketsa S, van Doorn WG. Chilling injury in *Dendrobium* inflorescences is alleviated by 1-MCP treatment. *Postharvest Biol Technol.* 2012;67:144–53.
<https://doi.org/10.1016/j.postharvbio.2011.12.016>
22. Patterson BD, MacRae EA, Ferguson IB. Estimation of hydrogen peroxide in plant extracts using titanium(IV). *Anal Biochem.* 1984;139(2):487–92.
[https://doi.org/10.1016/0003-2697\(84\)90039-3](https://doi.org/10.1016/0003-2697(84)90039-3)
23. SPSS user's guide, v. 21, IBM Corporation, New York, NY, USA; 2012.
24. Apel K, Hirt H. Reactive oxygen species: Metabolism, oxidative stress, and signal transduction. *Annu Rev Plant Biol.* 2004;55:373–99.
<https://doi.org/10.1146/annurev.arplant.55.031903.141701>
25. Vidrih R, Hribar J, Solar A, Zlatić E. The influence of atmosphere on the oxidation of ground walnut during storage at 20 °C. *Food Technol Biotechnol.* 2012;50(4):454–60.
26. Liu C, Zhao L, Yu G. The dominant glutamic acid metabolic flux to produce γ -amino butyric acid over proline in *Nicotiana tabacum* leaves under water stress relates to its significant role in antioxidant activity. *J Integr Plant Biol.* 2011;53(8):608–18.
<https://doi.org/10.1111/j.1744-7909.2011.01049.x>
27. Christopoulos MV, Tsantili E. Participation of phenylalanine ammonia-lyase (PAL) in increased phenolic compounds in fresh cold stressed walnut (*Juglans regia* L.) kernels. *Postharvest Biol Technol.* 2015;104:17–25.
<https://doi.org/10.1016/j.postharvbio.2015.03.003>
28. Carvalho M, Ferreira PJ, Mendes VS, Silva R, Pereira JA, Jerónimo C, Silva BM. Human cancer cell antiproliferative and antioxidant activities of *Juglans regia* L. *Food Chem Toxicol.* 2010;48(1):441–7.
<https://doi.org/10.1016/j.fct.2009.10.043>
29. Shahidi F, Zhong Y. Novel antioxidants in food quality preservation and health promotion. *Eur J Lipid Sci Technol.* 2010;112(9):930–40.
<https://doi.org/10.1002/ejlt.201000044>
30. Manzocco L, Calligaris S, Mastrocola D, Nicoli MC, Lerici CR. Review of non-enzymatic browning and antioxidant capacity in processed foods. *Trends Food Sci Tech.* 2000;11:340–6.
[https://doi.org/10.1016/S0924-2244\(01\)00014-0](https://doi.org/10.1016/S0924-2244(01)00014-0)
31. Christopoulos MV, Tsantili E. Effects of temperature and packaging atmosphere on total antioxidants and colour of walnut (*Juglans regia* L.) kernels during storage. *Sci Hortic.* 2011;131:49–57.
<https://doi.org/10.1016/j.scienta.2011.09.026>
32. Fuentealba C, Hernández I, Saa S, Toledo L, Burdiles P, Chirinos R, *et al.* Colour and *in vitro* quality attributes of walnuts from different growing conditions correlate with key precursors of primary and secondary metabolism. *Food Chem.* 2017;232:664–72.
<https://doi.org/10.1016/j.foodchem.2017.04.029>