

Oxidative stress under general intravenous and inhalation anaesthesia

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Oxidative stress occurs when reactive oxygen species (ROS) production overwhelms cell protection by antioxidants. This review is focused on general anaesthesia-induced oxidative stress because it increases the rate of complications and delays recovery after surgery. It is important to know what effects of anaesthetics to expect in terms of oxidative stress, particularly in surgical procedures with high ROS production, because their either additive or antagonistic effect may be pivotal for the outcome of surgery. *In vitro* and animal studies on this topic are numerous but show large variability. There are not many human studies and what we know has been learned from different surgical procedures measuring different endpoints in blood samples taken mostly before and after surgery. In these studies most intravenous anaesthetics have antioxidative properties, while volatile anaesthetics temporarily increase oxidative stress in longer surgical procedures.

KEY WORDS: glutathione; malondialdehyde; reactive oxygen species; superoxide dismutase; TBARS

Abbreviations

CPB – cardiopulmonary bypass; F₂-IsoPs – F₂-isoprostanes; CAT – catalase; GPx – glutathione peroxidase; GSH – glutathione; GSHR – glutathione reductase; GST – glutathione transferase; HNE – 4-hydroxynonenal; LOO[•] – lipidperoxyl radicals; MDA – malondialdehyde; 8-OHdG – 8-hydroxydeoxyguanosine; OSI – oxidative stress index; PUFA – polyunsaturated fatty acids; ROS – reactive oxygen species; SIRS – systematic inflammatory response syndrome; SOD – superoxide dismutase; TAC – total antioxidative capacity; TAS – total antioxidative status; TBARS – thiobarbituric acid reacting substances; TOS – total oxidative status

Oxidative stress is the imbalance of reactive oxygen species (ROS) production and antioxidative cell defence, which disrupts redox signalling and control and causes molecular damage (1). It is considered to be involved in the process of ageing, inflammations, cancers, degenerative diseases (2), and exposure to xenobiotics and drugs, such as anaesthetics (3). Anaesthetics-induced oxidative stress may affect lipids, proteins, and DNA. Among them, the most susceptible to oxidation are lipids (4). ROS interact with polyunsaturated fatty acids (PUFA) to form highly reactive lipoperoxides. Oxidative attack on lipids may result in the production of aldehydes such as 4-hydroxynonenal

(HNE) and malondialdehyde (MDA) or prostaglandin-like compounds called F₂-isoprostanes (F₂-IsoPs) (5). Anaesthetics-related oxidative stress is most often measured as plasma MDA levels or thiobarbituric acid reacting substances (TBARS) (6).

Oxidative damage to proteins results in their functional impairment or loss and in increased susceptibility to proteases (7, 8). Biomarkers of oxidative stress of proteins are carbonyl groups introduced into amino acid side chains, but they are not measured frequently in studies on anaesthetics-induced oxidative stress.

DNA damage is established by upregulation of 8-hydroxydeoxyguanosine (8-OHdG) and migration of broken DNA measured either by single-cell gel electrophoresis (comet assay) or more specific comet assays such as formamidopyridine DNA *N*-glycolase (Fpg) or enzyme-modified and human 8-oxoguanine DNA glycosylase (hOGG1) comet assay. In human studies DNA damage is measured in peripheral blood lymphocytes. In animal studies measurements may involve other organs as well.

Oxidative stress can be prevented or decreased by natural cell antioxidant defence, which transforms highly reactive oxygen species to less reactive compounds. Among numerous antioxidants the most frequently measured are glutathione (GSH), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione reductase (GSHR), and glutathione transferase (GST).

Other biomarkers of oxidative stress include total antioxidant status (TAS), total oxidative status (TOS), and

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oxidative stress index (OSI). OSI is the ratio between TOS and TAS and indicates the level of oxidative stress.

Surgery patients undergo procedures that can cause surgical trauma, inflammation, and ischaemia-reperfusion injury, all of which cause oxidative stress (9–11). To avoid further tissue injury caused by anaesthetics, it is very important to choose the ones that would minimise oxidative stress. This review seeks to summarise the effects of the most common intravenous and inhalation (volatile) anaesthetics in terms of oxidative stress and related genotoxic potential. We relied on preclinical (*in vitro* and animal) and available clinical data. Even though they are often inconclusive and even controversial, because the experiments greatly vary in cell cultures, animal species, concentrations, times of exposure, and different end-points, our goal was to find common grounds that could provide some informed advice to anaesthetists and surgeons.

INTRAVENOUS ANAESTHETICS

Propofol

Propofol (2,6-diisopropylphenol) is a widely used anaesthetic with many favourable properties. It reduces intracranial pressure and prevents inflammation and convulsions (12). Its pharmacokinetics in adults has been aptly described by Kanto and Gepts (13) and in children by Rigby-Jones and Sneyd (14). Children have higher clearance than adults and need higher induction and maintenance doses to achieve the same propofol blood concentration.

As propofol contains a phenolic OH-group, *in vitro* studies suggest that it has a high potential to prevent lipid peroxidation (15) by scavenging lipidperoxyl radicals (LOO^\cdot) and could replace α -tocopherol in that respect (16). It also seems to strengthen the GSH system in rat cells by inhibiting GPx and increasing GSHR and GST activity (17).

Genotoxicity studies seem to agree that propofol does not damage DNA. In primary rat astroglial cells it has been reported to protect DNA from lesions by scavenging peroxynitrite (18). In patients who underwent ear surgery propofol did not cause DNA damage in peripheral blood lymphocytes (19, 20). Similar was reported for 21 patients who underwent open-heart surgery and showed no increase in chromosomal aberrations of peripheral blood lymphocytes (21).

In rat studies *in vivo*, propofol has been reported to protect from kidney (22), heart (23), and liver ischaemia/reperfusion injury (24), decrease lipid peroxidation in the spinal cord after injury (25), attenuate postoperative signs of organ injury after liver transplantation (26, 27), protect from obstructive jaundice (28), hepatocyte necrosis, and infiltration of inflammatory cells, and restore liver architecture after halothane-induced intoxication (29).

In dogs anaesthetised with propofol, however, findings are less straightforward regarding total antioxidant capacity (TAC). One study reported unchanged TAC up to 48 h after gas evacuation for pneumoperitoneum (30), and another study a significant decrease in TAS with unchanged TOS at the end of anaesthesia for laparotomy and gastrotomy, which resulted with significant increase in OSI (TOS and TAS ratio) (31). A study on rats did not give consistent results regarding plasma TBARS, GSH/GSSG, SOD, CAT, and GSHR (32).

Findings in humans are generally favourable. One study (33) showed no significant plasma MDA changes in 29 liver donors anaesthetised with propofol before and after surgery. Another study (19) in 15 patients who underwent minor surgery and were receiving propofol anaesthesia for two hours showed increased plasma TAS and tocopherol concentrations, which confirmed its antioxidative properties. One clinical trial (34) comparing 50 patients under propofol anaesthesia and 50 patients on ketamine reported lower blood lipid peroxidation, GPx, and SOD activity in the first group. In 10 children with acyanotic heart disease, plasma MDA, GSH, lactate, and pyruvate concentrations remained unchanged after surgery (35). One study with 12 patients who underwent laparoscopic cholecystectomy (36) showed a significant drop in plasma MDA 1 min before desufflation and 20 min after desufflation when compared with concentrations before insufflation. In contrast, another study (37) reported higher plasma MDA concentrations in patients (N=17) undergoing partial hepatectomy after 60 min of propofol anaesthesia. These returned to preoperative values 24 h after surgery. A comparative study in patients with radical oesophagectomy showed significantly lower incidence of severe postoperative complications with propofol (7 of 92) than with sevoflurane anaesthesia (18 of 94) ($P=0.03$) (38).

Because of its antioxidative properties, propofol has, in fact, been proposed for application other than anaesthetic and sedative. In cultured neonatal rat cardiomyocytes it reduced doxorubicin-induced stress, and the authors of the study suggested that it could be used as adjuvant in doxorubicin therapy (39). It was also suggested for the treatment of glioblastoma, as it triggered apoptosis in human brain glioblastoma multiforme cells by decreasing Bcl-2 and increasing Bax and caspase activity (40). It also showed beneficial effects in critically ill patients with systemic inflammatory response and sepsis (41).

Thiopental

Thiopental or penthiobarbital (2-thio-5-ethyl-5-secpentylbarbituric acid) is a rapid-onset short-acting general anaesthetic. Even though it has largely been replaced by propofol, it is still used for rapid induction of anaesthesia in emergency cases. Research of its antioxidative and immunomodulating properties in human neutrophils showed a significant drop in O_2^- , H_2O_2 , and OH levels in the

presence of clinically relevant thiopental plasma concentrations (30 µg/L) (42). Thiopental also showed anti-inflammatory properties by significantly reducing chemotaxis and phagocytosis (43).

Animal studies of thiopental effects on oxidative stress report uneven findings. In rats with spinal cord contusion injury it showed beneficial effects, because spinal cord MDA was significantly lower than in control rats not treated with thiopental (25). In another study (44), in contrast, rats showed higher MDA and lower GSH, GPx, and GSHR levels in brain, heart, and bronchial tissues. In dogs with surgical trauma thiopental significantly increased plasma TOS, while TAS remained unchanged, which resulted with increased OSI (31).

In humans, the study with patients who underwent laparoscopic cholecystectomy (36) showed no changes in plasma MDA throughout the whole procedure, which suggests antioxidative effects of thiopental.

Ketamine

Ketamine (C₁₃H₁₆ClNO) is an *N*-methyl-D-aspartate (NMDA) receptor antagonist (45).

In vitro, ketamine was reported to produce toxic effects only at levels much higher than those corresponding to anaesthetic concentrations (46, 47). When they did correspond to plasma concentrations in anaesthesia (30 µg/mL), human neutrophil ROS (O₂⁻, H₂O₂, and OH) production did not differ from controls (43).

In rats, ketamine showed protective effects against oxidative stress. In fact, ketamine anaesthesia resulted in significantly lower kidney and liver tissue MDA than propofol, thiopental, and fentanyl (28, 48). In another study (44), the authors attributed significantly higher MDA and lower GSHR and GPx levels to ketamine-triggered boost of adrenaline, which is known to induce oxidative stress (49). In mice, ketamine also turned out to mitigate (secondary) brain injury which occurs when ROS production overwhelms the antioxidant system (50). It lowered brain MDA content in these mice and increased GPx, SOD, and NRF2 activity. These effects were accompanied by a significant reduction of brain water content and improved brain function (grip test) score.

Reports of ketamine effects in humans are positive. In adult and paediatric patients with cardiopulmonary bypass (CPB) ketamine mitigated the systemic inflammatory response syndrome (SIRS) by reducing proinflammatory IL-6 and IL-8, and/or increasing anti-inflammatory IL-10 (51–53).

Etomidate

Etomidate (R-1-(1-ethylphenyl)imidazole-5-ethyl ester) is a short-acting intravenous anaesthetic, whose R (+) enantiomer has a much higher hypnotic activity than the S (-) enantiomer (54). It has been in clinical use since 1972 thanks to high therapeutic index and minimal effects on

blood pressure and breathing. However, its use as anaesthetic has been limited since reports of serious adrenocortical suppression and myoclonus were published (55). It is no longer used to induce anaesthesia in critically ill patients, because adrenal suppression decreases cortisol release and thus weakens anti-inflammatory response. This means longer stays at intensive care units and higher mortality rate (56).

In vitro, etomidate does not affect TBARS production and the glutathione defence system (GPx, GSHR, GST) (17).

Similar was observed in animal studies. It does not seem to affect MDA levels or change SOD activity (57) in rat brain, while it significantly decreased MDA and increased GSH levels in the spinal cord after injury (58).

In human studies, its effect on plasma MDA was compared with that of thiopental and propofol. In patients who underwent laparoscopic cholecystectomy it fared worse than either (36), as it increased MDA levels before and after desufflation compared to other anaesthetics. In another study with 60 patients who underwent tibial fracture surgery (59) etomidate did not affect SOD activity, and patients receiving it had shorter hospitalisation time because of fewer postoperative complications such as numbness, lower limb pain, and coldness than patients on propofol.

INHALATIONAL (VOLATILE) ANAESTHETICS

Sevoflurane

Sevoflurane [fluoromethyl-2,2,2trifluoro-1-(trifluoromethyl) ethyl ether] has been used for general anaesthesia since the 1990s. It has a lower blood-gas partition coefficient than other volatile anaesthetics, which allows rapid induction of anaesthesia and quick reawakening. Because it does not cause irritation, acts as bronchodilator, and barely has any smell, it is the anaesthetic of choice in children (60), especially those with respiratory infections, airway hyper-reactivity, and seasonal allergies (61).

Genotoxicity studies point to none to low DNA damage. In one study (62), alkaline comet assay showed no sevoflurane genotoxicity in human peripheral lymphocytes, but in another (63) DNA damage was found in kidney homogenates of rats exposed for two hours over three consecutive days. Excessive exposure in this experiment may explain why the comet tail length and intensity increased gradually until the end of experiment (24 h). In peripheral lymphocytes of patients who underwent minimally invasive surgery that lasted two hours, the assay revealed no DNA damage (20), and no damage was reported in peripheral lymphocytes of children under sevoflurane anaesthesia for about one hour (64). In patients with lower abdominal surgery DNA damage was short-lived and tail

parameters started to return to normal on the third day, and repair was complete five days after anaesthesia (65).

In animal studies, findings of its effects on oxidative stress parameters vary, however. One study (66) reported higher GST and SOD activity and TBARS level in the liver of young and adult female rats after sevoflurane anaesthesia. In adult male rats it also increased MDA in the lungs, but decreased it in the kidney and brain (67). It also significantly reduced liver MDA levels in rats with ischaemia/reperfusion injury (57). Compared to isoflurane in another study (68), it produced significantly lower TBARS levels.

Several human studies suggest that sevoflurane does not increase oxidative stress, especially in minor surgeries (3), and where it does (as in major surgeries such as orthopaedic, cholecystectomy, hysterectomy, and alike) oxidative stress parameters return to normal 24–48 h after exposure has stopped. In patients who underwent laparoscopic surgery it did not increase plasma MDA and protein carbonyls (69) nor did it affect MDA, GSH, lactate, and pyruvate levels in peripheral blood of ten acyanotic children who underwent elective cardiac surgery (35) or GPx activity and TOS in 15–50-year-old patients who underwent laparoscopic cholecystectomy (70), while their TAS significantly increased. In a study of 12–36 months old children undergoing hypospadias repair surgery (71) sevoflurane increased SOD, GPx, and caspase-3 mRNA levels two hours after surgery, but these returned to normal three days later. The authors conclude that sevoflurane temporarily increases oxidative stress in children.

Desflurane

Desflurane (1,2,2,2-tetrafluoroethyl difluoromethyl ether) has the blood-gas partition coefficient lower than any other anaesthetic, which grants rapid recovery from general anaesthesia. It is contraindicated in children because of its pungent smell, which makes children hold their breath, cough, and salivate profusely (61).

Animal studies report controversial findings. One study comparing the effects of desflurane on oxidative stress in pig peripheral blood and bronchoalveolar lavage (BAL) fluid with those of sevoflurane and propofol (72) reported significantly higher MDA levels and lower GPx activity. Another, comparing it with sevoflurane, reported no brain accumulation of IL-6 and TNF- α or cognitive impairment in six days old mice (73).

Clinical studies also report inconclusive findings. In mothers with elective Caesarean section serum TOS and OSI were significantly lower after surgery than before it (74), but compared to sevoflurane, these parameters were significantly higher, especially in umbilical artery blood, which also had higher LOOH. In patients with laparoscopic cholecystectomy (70) it increased total oxidant capacity, but did not affect TAS and GPx activity. Similar were the findings reported for Turkish patients who underwent unspecified elective surgery (75): while serum GPx and

erythrocyte SOD activity did not change, MDA levels soared, and α -tocopherol plummeted in the first hour after surgery, but both returned to baseline values 12 h later. This transitory increase in MDA and protein carbonyl content was also reported six hours after desflurane anaesthesia (given together either with fentanyl or N₂O), which started to return to normal 24 h later (69).

As for its genotoxic potential, one study reports that desflurane does not cause DNA damage if used for minor surgeries for at least 90 minutes (76), while another study (77) revealed significantly higher 8-OHdG concentrations in plasma samples in addition higher comet tail length and intensity in BAL samples taken after anaesthesia, which points to oxidative stress as the mechanism of genotoxicity (77).

Isoflurane

Isoflurane (2-chloro-2-(difluoromethoxy)1,1,1-trifluoro-ethane) has been used as anaesthetic since the 1980s (3, 20) thanks to particularly (s)low metabolism and solubility, which results in very short induction and recovery time.

Isoflurane has antioxidative properties *in vitro* (3, 78), but *in vivo* it induces oxidative stress by increasing lipid peroxidation (68). In an animal experimental model simulating liver transplantation isoflurane induced significantly higher TBARS levels than sevoflurane.

Clinical studies evidence some oxidative stress. In one, isoflurane increased plasma MDA in patients undergoing donor hepatectomy, but it did not affect TOS, TAC, and SOD activity (33). Plasma SOD and GPx activities were also not affected in another study (79).

Genotoxicity/mutagenicity studies generally suggest no effects (77, 80), especially not after a minor surgery (19, 20). However, after a major lower abdominal surgery, one study (65) reported significantly increased comet assay parameters in peripheral blood lymphocytes two hours after the beginning of isoflurane anaesthesia. These parameters returned to normal three days later.

Nitrous oxide

Having the lowest potency among inhalational anaesthetics, nitrous oxide (N₂O) is often combined with more powerful ones (81) or used as anxiolytic. As it does not cause addiction but causes brief (several minutes long) bursts of euphoria, dissociation, and excitement, it is not prohibited by law, and people have also been using it for recreational purposes ever since 1775 (laughing gas parties) (82).

Even so, its use as anaesthetic has been in decline over postoperative nausea and vomiting, which are more pronounced if used for more than one hour (83). However, the European Society of Anaesthesiology believes that its benefits outweigh the adverse effects and find no real arguments to abandon it.

N₂O is not mutagenic in bacterial and mammalian cell systems (84). It is teratogenic to embryos of pregnant animals exposed to high doses of N₂O for 24 h during organogenesis but is not carcinogenic (76, 84). However, one study in nurses showed a positive correlation between the level of exposure to N₂O and oxidative DNA damage measured with the Fpg-modified comet assay (85). These nurses also had significantly higher lymphocyte ROS and plasma and urine TBARS levels and lower GPx activity than unexposed healthcare workers.

It is known that N₂O irreversibly inactivates methionine synthase (86), which leads to increased concentrations of homocysteine in plasma (87), which, in turn, inhibits the expression of antioxidant enzymes, including GPx. This effect can be counteracted by oral vitamin therapy (folate 2.5 mg, B₆ 25 mg and B₁₂ 500 mg) for one week before surgery (88). In addition, N₂O causes endothelial dysfunction in patients with cardiovascular disease undergoing non-cardiac surgery, which, in turn, increases the risk of postoperative cardiac events and mortality (87). One clinical study in patients anaesthetised with desflurane with or without N₂O (76) found no difference in lipid peroxidation, protein carbonyls, DNA damage, and ferric-reducing antioxidant power, which indicates that N₂O does not increase oxidative stress.

Volatile anaesthetic developmental neurotoxicity

One specific aspect of concern with anaesthetics – and particularly with sevoflurane, which is the most frequently used anaesthetic in paediatric surgery – is their neurotoxic potential. Considering that anaesthetics are lipophilic and readily pass the placental barrier, they may cause brain injury in foetuses and children. Judging by animal model studies (reviewed in ref. 89), exposure to sevoflurane during uterine quiescence can induce neuroinflammation, neuroapoptosis, and eventually memory impairment. Cognitive impairment was also reported in young mice repeatedly exposed to sevoflurane, which caused an increase in proinflammatory proteins IL-6 and TNF- α (73). IL-6 and Nrf2 were also increased in rats exposed to sevoflurane for five hours (90). Another review article (91) of preclinical intervention studies in neonatal mice suggests that volatile anaesthetics can lead to oxidative stress-related acute neurotoxicity and learning and memory deficits later in life. The risks of attention deficit, hyperactivity disorder, disabilities in language acquisition, and disabilities in abstract reasoning after prolonged anaesthesia have also been highlighted in a review article by Olsen and Brambrink (92). However, a randomised multicentre controlled trial following up 722 children for two and five years (93) after their one-hour general anaesthesia reported no significant neurocognitive or behavioural deficits.

CONCLUSIONS

Research of oxidative/antioxidative effects of anaesthetics has produced ambiguous findings so far, which stem from differences in exposure related to specific surgical procedures, length of anaesthesia, and timing of blood and tissue sampling. Even so, intravenous anaesthetics generally have antioxidant properties and short exposure to volatile anaesthetics does not seem to increase oxidative stress, while the effects of longer exposure are transient. Transient anaesthetics-induced oxidative stress may be the most relevant for the outcome of surgery with procedures that cause oxidative stress themselves. In such cases anaesthetics with antioxidative properties are drugs of choice.

REFERENCES

1. Jones DP. Redefining oxidative stress. *Antioxid Redox Signal* 2006;8:1865–79. doi: 10.1089/ars.2006.8.1865
2. Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D, Abete P. Oxidative stress, aging, and diseases. *Clin Interv Aging* 2018;13:757–72. doi: 10.2147/CIA.S158513
3. Lee Y-M, Song C, Yeum K-J. Impact of volatile anesthetics on oxidative stress and inflammation. *Biomed Res Int* 2015;2015:242709. doi: 10.1155/2015/242709
4. Pisoschi AM, Pop A. The role of antioxidants in the chemistry of oxidative stress: A review. *Eur J Med Chem* 2015;97:55–74. doi: 10.1016/j.ejmech.2015.04.040
5. Milne GL, Yin H, Brooks JD, Sanchez S, Roberts LJ, Morrow JD. Quantification of F₂-isoprostanes in biological fluids and tissues as a measure of oxidant stress. *Methods Enzymol* 2007;433:113–26. doi: 10.1016/S0076-6879(07)33006-1
6. Del Rio D, Stewart AJ, Pellegrini N. A review of recent studies on malondialdehyde as toxic molecule and biological marker of oxidative stress. *Nutr Metab Cardiovasc Dis* 2005;15:316–28. doi: 10.1016/j.numecd.2005.05.003
7. Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative stress and antioxidant defense. *World Allergy Organ J* 2012;5:9–19. doi: 10.1097/WOX.0b013e3182439613
8. Davies MJ. Singlet oxygen-mediated damage to proteins and its consequences. *Biochem Biophys Res Commun* 2003;305:761–70. doi: 10.1016/S0006-291X(03)00817-9
9. Ozdogan M, Devay AO, Gurer A, Ersoy E, Devay SD, Kulacoglu H, Gundogdu H. Plasma total anti-oxidant capacity correlates inversely with the extent of acute appendicitis: A case control study. *World J Emerg Surg* 2006;1:6. doi: 10.1186/1749-7922-1-6
10. Baysal Z, Togrul T, Aksoy N, Cengiz M, Çelik H, Boleken ME, Kaya M, Yavuz G. Evaluation of total oxidative and antioxidative status in pediatric patients undergoing laparoscopic surgery. *J Pediatr Surg* 2009;44:1367–70. doi: 10.1016/j.jpedsurg.2008.11.031
11. Glantzounis GK, Tselepis AD, Tambaki AP, Trikalinos TA, Manataki AD, Galaris DA, Tsimoyiannis EC, Kappas AM. Laparoscopic surgery-induced changes in oxidative stress markers in human plasma. *Surg Endosc* 2001;15:1315–9. doi: 10.1007/s00464-001-0034-2

12. Marik PE. Propofol: An immunomodulating agent. *Pharmacotherapy* 2005;25:28S–33S. doi: 10.1592/phco.2005.25.5_Part_2.28S
13. Kanto J, Gepts E. Pharmacokinetic implications for the clinical use of propofol. *Clin Pharmacokinet* 1989;17:308–26. doi: 10.2165/00003088-198917050-00002
14. Rigby-Jones AE, Sneyd JR. Propofol and children – What we know and what we do not know. *Paediatr Anaesth* 2011;21:247–54. doi: 10.1111/j.1460-9592.2010.03454.x
15. Boisset S, Steghens JP, Favetta P, Terreux R, Guitton J. Relative antioxidant capacities of propofol and its main metabolites. *Arch Toxicol* 2004;78:635–42. doi: 10.1007/s00204-004-0585-9
16. Aarts L, van der Hee R, Dekker I, de Jong J, Langemeijer H, Bast A. The widely used anesthetic agent propofol can replace α -tocopherol as an antioxidant. *FEBS Lett* 1995;357:83–5. doi: 10.1016/0014-5793(94)01337-Z
17. De La Cruz JP, Seden G, Carmona JA, de la Cuesta FS. The *in vitro* effects of propofol on tissular oxidative stress in the rat. *Anesth Analg* 1998;87:1141–6. doi: 10.1213/00000539-199811000-00031
18. Acquaviva R, Campisi A, Murabito P, Raciti G, Avola R, Mangiameli S, Musumeci I, Barcelona ML, Vanella A, Volti GL. Propofol attenuates peroxynitrite-mediated DNA damage and apoptosis in cultured astrocytes: An alternative protective mechanism. *Anesthesiology* 2004;101:1363–71. doi: 10.1097/0000542-200412000-00017
19. Braz MG, Braz LG, Freire CMM, Lucio LMC, Braz JRC, Tang G, Salvadori DMF, Yeum KJ, Amornytin S. Isoflurane and propofol contribute to increasing the antioxidant status of patients during minor elective surgery a randomized clinical study. *Medicine (Baltimore)* 2015;94(31):e1266. doi: 10.1097/MD.0000000000001266
20. Braz MG, Braz LG, Barbosa BS, Giacobino J, Orosz JEB, Salvadori DMF, Braz JRC. DNA damage in patients who underwent minimally invasive surgery under inhalation or intravenous anesthesia. *Mutat Res* 2011;726:251–4. doi: 10.1016/j.mrgentox.2011.09.007
21. Karahalil B, Yağar S, Bahadır G, Durak P, Şardaş S. Diazepam and propofol used as anesthetics during open-heart surgery do not cause chromosomal aberrations in peripheral blood lymphocytes. *Mutat Res* 2005;581:181–6. doi: 10.1016/j.mrgentox.2004.10.021
22. Li Y, Zhong D, Lei L, Jia Y, Zhou H, Yang B. Propofol prevents renal ischemia-reperfusion injury via inhibiting the oxidative stress pathways. *Cell Physiol Biochem* 2015;37:14–26. doi: 10.1159/000430329
23. Xie L-J, Zhao S, Zhang J-X, Li L. Propofol protects hearts from ischemia-reperfusion injury through interfering with mitochondria-dependent apoptotic pathway. *Chinese J Pharmacol Toxicol* 2007;21:247–54.
24. Bellanti F, Mirabella L, Mitarotonda D, Blonda M, Tamborra R, Cinnella G, Fersini A, Ambrosi A, Dambrosio M, Vendemiale G, Serviddio G. Propofol but not sevoflurane prevents mitochondrial dysfunction and oxidative stress by limiting HIF-1 α activation in hepatic ischemia/reperfusion injury. *Free Radic Biol Med* 2016;96:323–33. doi: 10.1016/j.freeradbiomed.2016.05.002
25. Kaptanoglu E, Sen S, Beskonakli E, Surucu HS, Tuncel M, Kilinc K, Taskin Y. Antioxidant actions and early ultrastructural findings of thiopental and propofol in experimental spinal cord injury. *J Neurosurg Anesthesiol* 2002;14:114–22. doi: 10.1097/00008506-200204000-00005
26. Yuan D, Su G, Liu Y, Chi X, Feng J, Zhu Q, Cai J, Luo G, Hei Z. Propofol attenuated liver transplantation-induced acute lung injury via connexin43 gap junction inhibition. *J Transl Med* 2016;14:194. doi: 10.1186/s12967-016-0954-1
27. Yao W, Han X, Zhang Y, Guan J, Ge M, Chen C, Wu S, Chen J, Luo G, Huang P, Hei Z. Intravenous anesthetic protects hepatocyte from reactive oxygen species-induced cellular apoptosis during liver transplantation *in vivo*. *Oxid Med Cell Longev* 2018;2018:4780615. doi: 10.1155/2018/4780615
28. Yildiz H, Coskuner I, Bulbuloglu E, Silay E, Kurutas EB, Dogan Z, Kantarceken B, Oksuz H, Senoglu N, Yuzbasioglu MF, Cetinkaya A, Ciralik H. The protective effects of ketamine and propofol in obstructive jaundice: an experimental study. *Bratisl Med J* 2012;113:139–44. doi: 10.4149/BLL_2012_034
29. Brasil LJ, San-Miguel B, Kretzmann NA, Amaral JLG Do, Zettler CG, Marroni N, González-Gallego J, Tuñón MJ. Halothane induces oxidative stress and NF- κ B activation in rat liver: Protective effect of propofol. *Toxicology* 2006;227:53–61. doi: 10.1016/j.tox.2006.07.013
30. Alipour F, Emami MR, Mohri M. Endocrine and oxidative stress characteristics in different anesthetic methods during pneumoperitoneum in dogs. *Comp Clin Pathol* 2018;27:1667–73. doi: 10.1007/s00580-018-2792-4
31. Lee JY. Oxidative stress due to anesthesia and surgical trauma and comparison of the effects of propofol and thiopental in dogs. *J Vet Med Sci* 2012;74:663–5. doi: 10.1292/jvms.11-0221
32. Félix LM, Correia F, Pinto PA, Campos SP, Fernandes T, Videira R, Oliveira MM, Peixoto FP, Antunes LM. Propofol affinity to mitochondrial membranes does not alter mitochondrial function. *Eur J Pharmacol* 2017;803:48–56. doi: 10.1016/j.ejphar.2017.03.044
33. Ucar M, Özgül U, Polat A, Toprak HI, Erdogan MA, Aydoğan MS, Durmus M, Ersoy MO. Comparison of antioxidant effects of isoflurane and propofol in patients undergoing donor hepatectomy. *Transplant Proc* 2015;47:469–72. doi: 10.1016/j.transproceed.2014.11.043
34. Khoshraftar E, Ranjbar A, Kharkhane B, Heidary ST, Gharebaghi Z, Zadkhosh N. Antioxidative effects of propofol vs. Ketamin in individuals undergoing surgery. *Arch Iran Med* 2014;17:486–9. doi: 0141707/AIM.008
35. Dumaresq DMH, de Vasconcelos RC, Guimarães SB, Cavalcante SL, Garcia JHP, de Vasconcelos PRL. Metabolic and oxidative effects of sevoflurane and propofol in children undergoing surgery for congenital heart disease. *Acta Cir Bras* 2011;26:66–71. doi: 10.1590/S0102-86502011000700014
36. Yagmurdur H, Cakan T, Bayrak A, Arslan M, Baltaci B, Inan N, Kilinc K. The effects of etomidate, thiopental, and propofol in induction on hypoperfusion-reperfusion phenomenon during laparoscopic cholecystectomy. *Acta Anaesthesiol Scand* 2004;48:772–7. doi: 10.1111/j.0001-5172.2004.00417.x
37. Laviolle B, Basquin C, Aguillon D, Compagnon P, Morel I, Turmel V, Seguin P, Boudjema K, Bellissant E, Mallédant Y. Effect of an anesthesia with propofol compared with desflurane on free radical production and liver function after

- partial hepatectomy. *Fundam Clin Pharmacol* 2012;26:735–42. doi: 10.1111/j.1472-8206.2011.00958.x
38. Tsuchiya M, Shiimoto K, Mizutani K, Fujioka K, Suehiro K, Yamada T, Sato EF, Nishikawa K. Reduction of oxidative stress a key for enhanced postoperative recovery with fewer complications in esophageal surgery patients: Randomized control trial to investigate therapeutic impact of anesthesia management and usefulness of simple blood test for pre. *Medicine (Baltimore)* 2018;97(47):e12845. doi: 10.1097/MD.00000000000012845
39. Lai HC, Yeh YC, Wang LC, Ting CT, Lee WL, Lee HW, Wang KY, Wu A, Su CS, Liu TJ. Propofol ameliorates doxorubicin-induced oxidative stress and cellular apoptosis in rat cardiomyocytes. *Toxicol Appl Pharmacol* 2011;257:437–48. doi: 10.1016/j.taap.2011.10.001
40. Hsu SS, Jan CR, Liang WZ. Evaluation of cytotoxicity of propofol and its related mechanism in glioblastoma cells and astrocytes. *Environ Toxicol* 2017;32:2440–54. doi: 10.1002/tox.22458
41. Baronica R, Tonković D, Bačić G, Karadjole T, Šuran J, Bačić Baronica K, editors. *Croatian International Symposium on Intensive Care Medicine / Simpozij intenzivne medicine s međunarodnim sudjelovanjem*. *Neurol Croat* 2013;62(Suppl 2):1–170.
42. Wittmann S, Daniels S, Ittner KP, Fröhlich D. Thiopentone and methohexitone enantiomers do not act stereoselectively on the oxidative response in human neutrophils *in vitro*. *Pharmacology* 2004;72:12–9. doi: 10.1159/000078627
43. Nishina K, Akamatsu H, Mikawa K, Shiga M, Maekawa N, Obara H, Niwa Y. The inhibitory effects of thiopental, midazolam, and ketamine on human neutrophil functions. *Anesth Analg* 1998;86:159–65. doi: 10.1097/00000539-199801000-00032
44. Ahiskalioglu EO, Aydin P, Ahiskalioglu A, Suleyman B, Kuyrukluylidiz U, Kurt N, Altuner D, Coskun R, Suleyman H. The effects of ketamine and thiopental used alone or in combination on the brain, heart, and bronchial tissues of rats. *Arch Med Sci* 2018;14:645–54. doi: 10.5114/aoms.2016.59508
45. Dinis-Oliveira RJ. Metabolism and metabolomics of ketamine: a toxicological approach. *Forensic Sci Res* 2017;2:2–10. doi: 10.1080/20961790.2017.1285219
46. Bai X, Yan Y, Canfield S, Muravyeva MY, Kikuchi C, Zaja I, Corbett JA, Bosnjak ZJ. Ketamine enhances human neural stem cell proliferation and induces neuronal apoptosis via reactive oxygen species-mediated mitochondrial pathway. *Anesth Analg* 2013;116:869–80. doi: 10.1213/ANE.0b013e3182860fc9
47. Ito H, Uchida T, Makita K. Ketamine causes mitochondrial dysfunction in human induced pluripotent stem cell: Derived neurons. *PLoS One* 2015;10(5):e0128445. doi: 10.1371/journal.pone.0128445
48. Hatipoglu S, Yildiz H, Bulbuloglu E, Coskuner I, Kurutas EB, Hatipoglu F, Ciralik H, Berhuni MS. Protective effects of intravenous anesthetics on kidney tissue in obstructive jaundice. *World J Gastroenterol* 2014;20:3320–6. doi: 10.3748/wjg.v20.i12.3320
49. Parvin R, Akhter N. Protective effect of tomato against adrenaline-induced myocardial infarction in rats. *Bangladesh Med Res Counc Bull* 2008;34:104–8. doi: 10.3329/bmrcb.v34i3.1974
50. Liang J, Wu S, Xie W, He H. Ketamine ameliorates oxidative stress-induced apoptosis in experimental traumatic brain injury via the Nrf2 pathway. *Drug Des Devel Ther* 2018;12:845–53. doi: 10.2147/DDDT.S160046
51. Welters ID, Feurer M-K, Preiss V, Miller M, Scholz S, Kwapisz M, Mogk M, Neuhuser C. Continuous S-(+)-ketamine administration during elective coronary artery bypass graft surgery attenuates pro-inflammatory cytokine response during and after cardiopulmonary bypass. *Br J Anaesth* 2011;106:172–9. doi: 10.1093/bja/aeq341
52. Dale O, Somogyi AA, Li Y, Sullivan T, Shavit Y. Does intraoperative ketamine attenuate inflammatory reactivity following surgery? A systematic review and meta-analysis. *Anesth Analg* 2012;115:934–43. doi: 10.1213/ANE.0b013e3182662e30
53. Durandy Y. Minimizing systemic inflammation during cardiopulmonary bypass in the pediatric population. *Artif Organs* 2014;38:11–8. doi: 10.1111/aor.12195
54. Forman SA. Clinical and molecular pharmacology of etomidate. *Anesthesiology* 2011;114:695–707. doi: 10.1097/ALN.0b013e3181ff72b5
55. Raines DE. The Pharmacology of Etomidate and Etomidate Derivatives. *Int Anesthesiol Clin* 2015;53:63–75. doi:10.1097/AIA.0000000000000050.
56. Allen C, Washington S. The role of etomidate as an anaesthetic induction agent for critically ill patients. *Br J Hosp Med* 2016;77:282–6. doi: 10.12968/hmed.2016.77.5.282
57. Li X, Lu F, Li W, Xu J, Sun XJ, Qin LZ, Zhang QL, Yao Y, Yu QK, Liang XL. Underlying mechanisms of memory deficits induced by etomidate anesthesia in aged rat model: Critical role of immediate early genes. *Chin Med J (Engl)* 2016;129:48–53. doi: 10.4103/0366-6999.172570
58. Cayli SR, Ates O, Karadag N, Altinoz E, Yucel N, Yologlu S, Kocak A, Cakir CO. Neuroprotective effect of etomidate on functional recovery in experimental spinal cord injury. *Int J Dev Neurosci* 2006;24:233–9. doi: 10.1016/j.ijdevneu.2006.04.003
59. Li R, Fan L, Ma F, Cao Y, Gao J, Liu H, Li Y. Effect of etomidate on the oxidative stress response and levels of inflammatory factors from ischemia-reperfusion injury after tibial fracture surgery. *Exp Ther Med* 2017;13:971–5. doi: 10.3892/etm.2017.4037
60. Lv X, Yan J, Jiang H. Inhaled anesthetic sevoflurane: neurotoxicity or neuroprotection in the developing brain. *Int J Clin Exp Med* 2017;10:9930–8.
61. Kaye AD, Fox CJ, Padnos IW, Ehrhardt KP, Diaz JH, Cornett EM, Chandler D, Sen S, Patil S. Pharmacologic considerations of anesthetic agents in pediatric patients: A comprehensive review. *Anesthesiol Clin* 2017;35(2):e73–e94. doi: 10.1016/j.anclin.2017.01.012
62. Szyfter K, Szulc R, Mikstacki A, Stachecki I, Rydzanicz M, Jałoszyński P. Genotoxicity of inhalation anaesthetics: DNA lesions generated by sevoflurane *in vitro* and *in vivo*. *J Appl Genet* 2004;45:369–74. PMID: 15306730
63. Brozović G, Oršolić N, Rozgaj R, Knežević F, Knežević AH, Maričić M, Kršnik D, Benković V. Sevoflurane and isoflurane genotoxicity in kidney cells of mice. *Arh Hig Rada Toksikol* 2017;68:228–35. doi: 10.1515/aiht-2017-68-2941
64. Krause T, Scholz J, Jansen L, Boettcher H, Koch C, Wappler F, Schulte am Esch J. Sevoflurane anaesthesia does not induce the formation of sister chromatid exchanges in peripheral

- blood lymphocytes of children. *Br J Anaesth* 2003;90:233–5. doi: 10.1093/bja/aeg030
65. Karabiyik L, Şardaş S, Polat U, Kocabaş NA, Karakaya AE. Comparison of genotoxicity of sevoflurane and isoflurane in human lymphocytes studied *in vivo* using the comet assay. *Mutat Res* 2001;492:99–107. doi: 10.1016/S1383-5718(01)00159-0
 66. Arslan M, Isik B, Kavutcu M, Kurtipek O. Oxidative stress and antioxidant activity of female rat liver tissue after sevoflurane anaesthesia: young versus old. *Bratisl Med J* 2012;113:702–6. doi: 10.4149/bll_2012_159
 67. Türkan H, Aydın A, Sayal A, Eken A, Akay C, Karahalil B. Oxidative and antioxidative effects of desflurane and sevoflurane on rat tissue *in vivo*. *Arh Hig Rada Toksikol* 2011;62:113–9. doi: 10.2478/10004-1254-62-2011-2096
 68. Molin SZFD, Krueh CRP, de Fraga RS, Alboim C, de Oliveira JR, Alvares-da-Silva MR. Differential protective effects of anaesthesia with sevoflurane or isoflurane. *Eur J Anaesthesiol* 2014;31:695–700. doi: 10.1097/EJA.000000000000127
 69. Sivaci R, Kahraman A, Serteser M, Sahin DA, Dilek ON. Cytotoxic effects of volatile anesthetics with free radicals undergoing laparoscopic surgery. *Clin Biochem* 2006;39:293–8. doi: 10.1016/j.clinbiochem.2006.01.001
 70. Erbas M, Demiraran Y, Ak Yildirim H, Sezen G, Iskender A, Karagoz I, Kandis H. Comparison of effects on the oxidant/antioxidant system of sevoflurane, desflurane and propofol infusion during general anesthesia. *Brazilian J Anesthesiol* 2015;65:68–72. doi: 10.1016/j.bjane.2014.05.004
 71. Zhou X, Lu D, Li W da, Chen X hui, Yang X yu, Chen X, Zhou Z bin, Ye JH, Feng X. Sevoflurane affects oxidative stress and alters apoptosis status in children and cultured neural stem cells. *Neurotox Res* 2018;33:790–800. doi: 10.1007/s12640-017-9827-5
 72. Allaouchiche B, Debon R, Goudable J, Chassard D, Duffo F. Oxidative stress status during exposure to propofol, sevoflurane and desflurane. *Anesth Analg* 2001;93:981–5. doi: 10.1097/0000539-200110000-00036
 73. Shen X, Dong Y, Xu Z, Wang H, Miao C, Soriano SG, Sun D, Baxter MG, Zhang Y, Xie Z. Selective anesthesia-induced neuroinflammation in developing mouse brain and cognitive impairment. *Anesthesiology* 2013;118:502–15. doi: 10.1097/ALN.0b013e3182834d77
 74. Yalcin S, Aydoğan H, Yuce HH, Kucuk A, Karahan MA, Vural M, Camuzcuoğlu A, Aksoy N. Effects of sevoflurane and desflurane on oxidative stress during general anesthesia for elective cesarean section. *Wien Klin Wochenschr* 2013;125:467–73. doi: 10.1007/s00508-013-0397-0
 75. Eroglu F, Yavuz L, Ceylan BG, Yilmaz F, Eroglu E, Delibas N, Naziroğlu M. New volatile anesthetic, desflurane, reduces vitamin e level in blood of operative patients via oxidative stress. *Cell Biochem Funct* 2010;28:211–6. doi: 10.1002/cbf.1641
 76. Nogueira FR, Braz LG, Souza KM, Aun AG, Arruda NM, Carvalho LR, Chen CYO, Braz JRC, Braz MG. Comparison of DNA damage and oxidative stress in patients anesthetized with desflurane associated or not with nitrous oxide: A prospective randomized clinical trial. *Anesth Analg* 2018;126:1198–205. doi: 10.1213/ANE.0000000000002729
 77. Cukurova Z, Cetingok H, Ozturk S, Gedikbasi A, Hergunsel O, Ozturk D, Don B, Cefle K, Palanduz S, Ertem DH, Tarantino G. DNA damage effects of inhalation anesthetics in human bronchoalveolar cells. *Medicine (Baltimore)* 2019;98:e16518. doi: 10.1097/MD.00000000000016518
 78. Schallner N, Ulbrich F, Engelstaedter H, Biermann J, Auwaerter V, Loop T, Goebel U. Isoflurane but not sevoflurane or desflurane aggravates injury to neurons *in vitro* and *in vivo* via p75NTR-NF-κB activation. *Anesth Analg* 2014;119:1429–41. doi: 10.1213/ANE.0000000000000488
 79. Türkan H, Bukan N, Sayal A, Aydın A, Bukan MH. Effects of halothane, enflurane, and isoflurane on plasma and erythrocyte antioxidant enzymes and trace elements. *Biol Trace Elem Res* 2004;102:105–12. doi: 10.1385/BTER:102:1-3:105
 80. Husum B, Wulf HC, Neebuhr E, Kyst A, Valentin N. Sister chromatid exchanges in lymphocytes of humans anaesthetized with isoflurane. *Br J Anaesth* 1984;56:559–64. doi: 10.1093/bja/56.6.559
 81. Sanders RD, Weimann J, Maze M. Biologic effects of nitrous oxide: A mechanistic and toxicologic review. *Anesthesiology* 2008;109:707–22. doi: 10.1097/ALN.0b013e3181870a17
 82. van Amsterdam J, Nabben T, van den Brink W. Recreational nitrous oxide use: Prevalence and risks. *Regul Toxicol Pharmacol* 2015;73:790–6. doi: 10.1016/j.yrtph.2015.10.017
 83. Hert SGD. The current place of nitrous oxide in clinical practice. *Eur J Anaesthesiol* 2015;32:517–20. doi: 10.1097/EJA.000000000000264
 84. American Society of Anesthesiologists. *Anesthetic Gases: Information for Management in Anesthetizing Areas and Postanesthesia Care Unit*. Park Ridge (IL): ASA; 1999.
 85. Wrońska-Nofer T, Nofer JR, Jajte J, Dziubałtowska E, Szymczak W, Krajewski W, Waogoneksowicz W, Rydzyski K. Oxidative DNA damage and oxidative stress in subjects occupationally exposed to nitrous oxide (N₂O). *Mutat Res* 2012;731:58–63. doi: 10.1016/j.mrfmmm.2011.10.010
 86. Badner NH, Drader K, Freeman D, Spence JD. The use of intraoperative nitrous oxide leads to postoperative increases in plasma homocysteine. *Anesth Analg* 1998;87:711–3. doi: 10.1213/0000539-199809000-00041
 87. Myles PS, Chan MTV, Kaye DM, McIlroy DR, Lau CW, Symons JA, Chen S. Effect of nitrous oxide anesthesia on plasma homocysteine and endothelial function. *Anesthesiology* 2008;109:657–63. doi: 10.1097/ALN.0b013e31818629db
 88. Badner NH, Freeman D, Spence JD. Preoperative oral B vitamins prevent nitrous oxide-induced postoperative plasma homocysteine increases. *Anesth Analg* 2001;93:1507–10. doi: 10.1097/0000539-200112000-00034
 89. Chai D, Cheng Y, Jiang H. Fundamentals of fetal toxicity relevant to sevoflurane exposures during pregnancy. *Int J Dev Neurosci* 2019;72:31–5. doi: 10.1016/j.ijdevneu.2018.11.001
 90. Tian Y, Wu X, Guo S, Ma L, Wei H, Zhao X. Minocycline attenuates sevoflurane-induced cell injury via activation of Nrf2. *Int J Mol Med* 2017;39:869–78. doi: 10.3892/ijmm.2017.2908
 91. Johnson SC, Pan A, Li L, Sedensky M, Morgan P. Neurotoxicity of anesthetics: Mechanisms and meaning from mouse intervention studies. *Neurotoxicol Teratol* 2019;71:22–31. doi: 10.1016/j.ntt.2018.11.004
 92. Olsen EA, Brambrink AM. Anesthesia for the young child undergoing ambulatory procedures: Current concerns regarding harm to the developing brain. *Curr Opin*

- Anaesthesiol 2013;26:677–84. doi: 10.1097/ACO.000000000000016
93. McCann ME, Berde C, Soriano S, Marmor J, Bellinger D, de Graaff JC, de Graaff JC, Dorris L, Bell G, Morton N, Dorris L, Morton N, Disma N, Giribaldi G, Withington D, Withington D, Grobler A, Stargatt R, Hunt RW, Sheppard SJ, Marmor J, Giribaldi G, Bellinger DC, Hartmann PL, Hardy P, Frawley G, Izzo F, Sternberg BS, Lynn A, Wilton N, Mueller M, Polaner DM, Absalom AR, Szmuk P, Morton N, Berde C, Soriano S, Davidson AJ, for the GAS Consortium. Neurodevelopmental outcome at 5 years of age after general anaesthesia or awake-regional anaesthesia in infancy (GAS): an international, multicentre, randomised, controlled equivalence trial. *Lancet* 2019;393:664–77. doi: 10.1016/S0140-6736(18)32485-1

Oksidacijski stres uzrokovan općom intravenskom i inhalacijskom anestezijom

Ovo je pregledni rad o oksidacijskom stresu uzrokovanom općim anestetima zbog toga što uzrokuje komplikacije nakon operacije i usporava oporavak. Oksidacijski stres nastaje kada je stvaranje reaktivnih kisikovih spojeva (ROS) veće od stanične zaštite antioksidansima. Poznavanje učinka anestetika na oksidacijski stres posebice je važno pri planiranju kirurških zahvata kod kojih je poznato da uzrokuju nastanak veće količine ROS-a. Naime, antagonistički ili aditivni učinak anestetika na oksidacijski stres može kod takvih kirurških zahvata biti ključan za ishod operacije. Brojna su istraživanja o toj temi, no protokoli pokusa na staničnim kulturama i pokusnim životinjama vrlo su različiti. U istraživanjima na ljudima anestetici su upotrebljavani tijekom vrlo raznorodnih kirurških zahvata, često uz primjenu više anestetika, mjereni su različiti parametri oksidacijskoga stresa, a uzorci (najčešće krvi) uzimani su obično prije zahvata i u različitim intervalima nakon njega. Zbog svega je toga ponekad teško razlučiti kakav je učinak nekog anestetika na oksidacijski stres. Općenito intravenozni anestetici imaju antioksidacijsko djelovanje, imaju ga i plinoviti anestetici – uglavnom nakon kratkotrajne anestezije, a dugotrajna anestezija uzrokuje privremeni porast oksidacijskoga stresa.

KLJUČNE RIJEČI: glutation; malondialdehid; reaktivni kisikovi spojevi; SOD; TBARS