

# Bio-based Products from Lignocellulosic Waste Biomass: A State of the Art



This work is licensed under a Creative Commons Attribution 4.0 International License

M. Tišma,\* A. Bucić-Kojić, and M. Planinić

Josip Juraj Strossmayer University of Osijek,  
Faculty of Food Technology Osijek, Franje Kuhača 18,  
HR-31000 Osijek, Croatia

doi: <https://doi.org/10.15255/CABEQ.2021.1931>

Review

Received: May 6, 2021

Accepted: February 15, 2021

This review presents data on the chemical composition of harvest residues and food industry by-products as widely abundant representatives of lignocellulosic waste biomass. Pretreatment methods, with special emphasis on biological methods, are presented as an important step in utilization of lignocellulosic waste biomass for the production of sustainable biofuels and high-value chemicals. Special attention was paid to the methods of lignin isolation and its possible utilization within lignocellulosic biorefinery. The objectives of circular bioeconomy and the main aspects of lignocellulosic biorefinery are highlighted. Finally, current data on industrial, pilot, and research and development plants used in Europe for the production of a variety of bio-based products from different feedstocks are presented.

*Key words:*

biorefinery, circular bioeconomy, lignocellulosic biomass, sustainable development

## Introduction

Lignocellulosic biomass (LB) comes from natural sources or processes that are constantly being replenished. Mostly, it is used for bioenergy, but in recent years considerable attention is given to LB as a source for the production of high-value chemicals. Thus, LB is considered as a renewable, abundant, and economical alternative to fossil resources.<sup>1</sup>

LB is found in large quantities almost everywhere in the world. It is estimated that 181.5 billion tonnes of LB are produced annually on Earth. Only 8.2 billion tonnes are currently used, of which 7 billion tonnes are mainly produced from dedicated agricultural, grass, and forest land.<sup>2</sup> In contrast, non-renewable sources, such as oil, gas, and coal, can be found only in a certain number of countries in the world. Their exploitation causes pollution and climate change accompanied by gradual depletion. Although LB can be used for the production of sustainable biofuels, chemicals, and materials, the majority of the world's energy sources and material products, especially chemicals, still come from fossil fuels, mainly oil and natural gas.<sup>3</sup> Sustainable processes of LB utilization to produce bio-based products that achieve “zero concept” waste must be established.<sup>4</sup> For that purpose, the concept of biorefineries has been proposed.<sup>3</sup> The goal of the biorefinery is to transition to a more sustainable economic system that uses resources more efficiently, reduces

overall waste generation, and allows the recycling of unavoidable waste as a source for the production of new products. However, finding efficient and, at the same time, sustainable technologies is a demanding task.

There are different biorefinery pathways from feedstock to product, depending on the composition and availability of the feedstock, the conversion technologies applied, and the production of the desired products.<sup>1</sup> Several technological, logistical, and economical aspects should be solved before LB finds large application for sustainable biofuels and high-value chemicals production. A significant effort is dedicated to biological pretreatment methods by the use of white-rot fungi.<sup>5</sup> Additionally, novel, eco-friendly, and natural deep eutectic solvents are explored for LB fractionation, lignin isolation, extraction of value-added products from lignin, and biotransformation.<sup>6–8</sup>

This review focuses on the general chemistry of LB and the chemical composition of typical representatives of the widely abundant lignocellulosic waste biomass, such as harvest residues and food industry by-products. The objective of this review is to increase understanding of the chemical complexity of LB waste resources, their availability, and challenges for potential lignocellulosic biorefinery applications. Recent research on the use of lignocellulosic waste biomass is discussed and divided into the following parts: LB pretreatment methods, lignin isolation methods, and the use of lignin in the production of various bio-products.

\*Corresponding author: E-mail: [marina.tisma@ptfos.hr](mailto:marina.tisma@ptfos.hr)

## Lignocellulosic biomass chemistry

Lignocellulosic biomass is mainly comprised of lignin, cellulose, and hemicellulose, which are present in varying amounts and ratios, depending on the origin of the biomass. It also contains small amounts of pectin, protein, extractives, and inorganic compounds.<sup>9</sup> The schematic representation of lignin, cellulose, and hemicellulose as the main components of LB is presented in Fig. 1.

Cellulose is the most abundant component of LB. It is a linear polymer of hundreds to over ten thousand glucose molecules linked by  $\beta$ -1,4 glycosidic bonds. The repeating unit of cellulose is cellobiose. Hydroxyl groups in cellulose are involved in several intra- and intermolecular hydrogen bonds, which result in various ordered crystalline arrangements. Unlike the crystalline region, the amorphous region of cellulose is easily degradable.<sup>10,11</sup> Hemicellulose is a heteropolymer consisting of short, linear, and highly branched chains of different hexoses, pentoses, and sugar acids.<sup>12</sup> Common hemicelluloses are galactans, xylans, mannans, and arabans. Hemicelluloses can be more easily enzymatically degraded compared to cellulose. However, certain oligomeric structures are recalcitrant due to the complex branching and acetylation patterns.<sup>11</sup>

Lignin is a complex, amorphous, and structurally diverse aromatic heteropolymer, with cross-linked racemic macromolecules, and is relatively hydrophobic. It fills the space between hemicellulose and covers the cellulose skeleton making lignocellulosic matrices.<sup>1</sup> Predominant structural compo-

nents of lignin are monolignols (phenylpropanoid aryl-C3 units): *p*-coumaryl alcohol (H, 4-hydroxyl phenyl), coniferyl alcohol (G, guaiacyl), and sinapyl alcohol (S, siringyl) linked by C–O and C–C bonds. These three units differ in the number of methoxy groups in their phenolic rings. Their ratio within the polymer varies among different plants, wood tissues, and cell wall layers.<sup>13</sup> For example, grass contains all three subunits (H, G and S), hardwood contains G and S subunits, while softwood is mostly comprised of G subunits.<sup>1</sup> The G unit contains monomethoxy phenoxide, the S unit contains dimethoxy phenoxide, and the H unit contains the non-substituted phenoxide moiety. Predominant linkages in lignin are  $\beta$ -aryl ether ( $\beta$ -O-4) bonds. The other linkages are phenylcoumaran ( $\beta$ -5), biphenyl (5-5), 1,2-diaryl ether (4-O-5),  $\beta$ - $\beta$  linked structures, structures condensed in 2- or 6- positions, glyceraldehyde-2-aryl ether.<sup>13</sup> These linkages are formed by the addition of the phenol group of one monolignol to the propyl chain of the second monolignol. Monolignols not only possess anti-inflammatory and antinociceptive activities, but also carry a functional allyl alcohol species that have been evaluated as lignin-derived platform chemicals for the synthesis of natural products, pharmaceuticals, and functional materials.<sup>14</sup>

Lignin can be classified as natural, which is described previously, and technical or industrial lignin. Industrial lignin has diverse macromolecular structures due to various chemical modifications, and contains impurities depending on the applied LB treatments.<sup>13</sup>

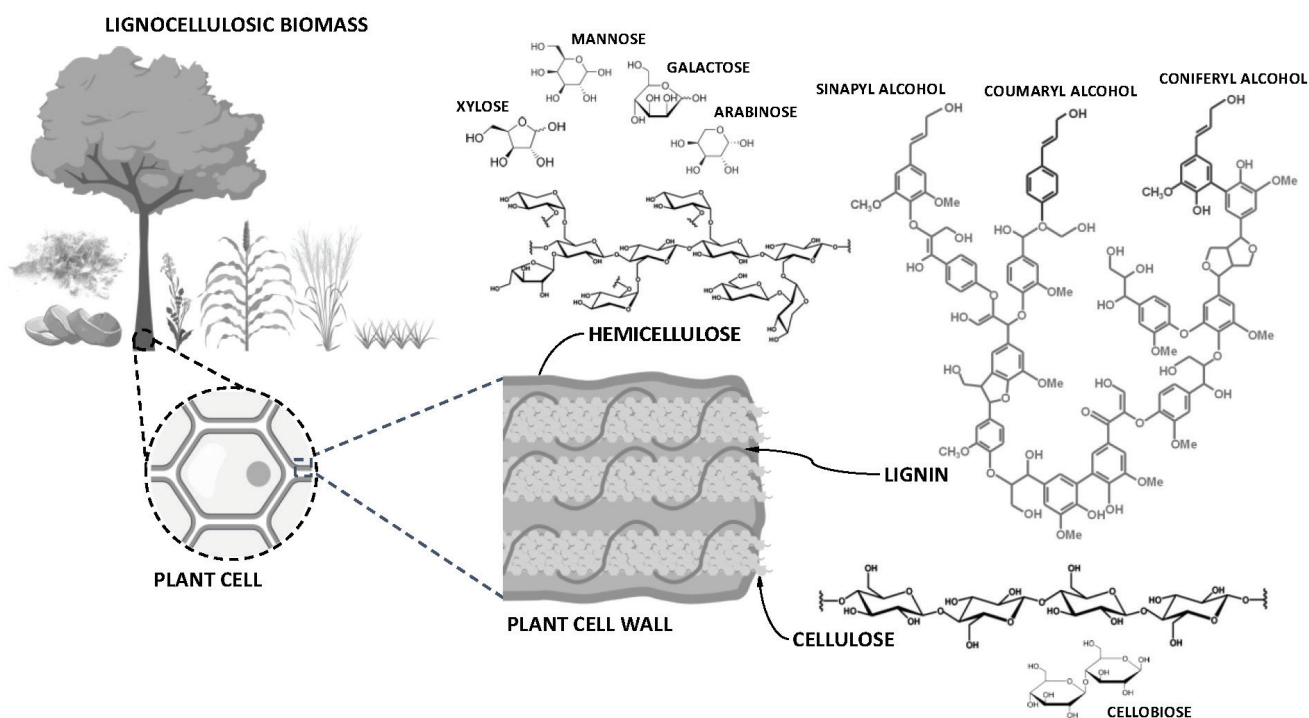


Fig. 1 – Schematic representation of lignin, cellulose, and hemicellulose as the main components of lignocellulosic biomass

## Harvest residues and food industry by-products

Knowledge of the chemical composition of the LB is very important because the selection of pretreatment method depends on the type and composition of biomass.<sup>15</sup> Generally, in modern bioenergy systems, LB supply chain can be divided into forest biomass (treetops, branches, and unmerchantable stems, wood processing residues such as wood chips, sawdust, and shavings), harvest residues, food or feed processing residues, energy crops (including food crops such as sugar cane, oil palm, corn), waste of biological origin (manure) and household, commercial or municipal organic waste.<sup>16</sup> In this paper, the possibility of using harvest residues and food processing residues (food industry by-products) in biorefineries is considered. Therefore, the literature data on the lignin, cellulose, and hemicellulose content in different harvest residues and food industry by-products are reviewed and shown in Table 1 and Table 2, respectively. For those materials, the combined term “agro-food waste” is also often found in literature. As seen from the composition of polymers (Table 1 and Table 2), they differ for the same type of material. Plant variety, agronomic measures of cultivation, weather conditions, harvesting methods, and storage conditions are all factors that influence the chemical composition of harvest residues.<sup>17</sup> In the case of food industry by-products, in addition to the aforementioned, industrial process conditions also contribute to the chemical composition of the resulting waste or by-products. A good example is the chemical composition of brewer’s spent grains, which is strongly influenced by the brewing process, which depends on the type of beer produced and the specific brewing processes, unique to each brewery.

## Lignocellulosic biomass pretreatment methods and lignin isolation

The best way to utilize LB is a cascade process, since it considers the composition characteristics and the nature of cellulose, hemicellulose and lignin.<sup>1</sup> To achieve cascade utilization, the pretreatment step is required. After pretreatment, the conventional separation methods (extraction, regeneration, centrifugation, filtration, distillation, drying) are used. Separation is followed by the process of producing high-value chemicals from the individual components.<sup>1</sup> Cellulose is mainly hydrolyzed to glucose, which can be further converted to different chemicals of biofuels, while hemicellulose is mainly hydrolyzed to xylose and converted to xylitol. Regarding lignin, no efficient approach or protocol has yet been developed to ensure high conversion of lignin into desired products. Much research has been devoted to the separation of lignin and its use for a variety of useful products by chemical, thermochemical or biochemical routes.<sup>67–69</sup>

Generally, LB pretreatment methods can be divided into physical, chemical, physicochemical, biological methods, performed alone or in various combinations.<sup>68–71</sup> However, not all of those methods are eco-friendly or sustainable. Most of them have a negative influence on the environment due to a large amount of chemicals used in the process, and/or are energy-intensive. Physical methods are mechanical (grinding, milling, chopping), sonication, mechanical extrusion, freezing, ozonolysis, pyrolysis, and more recently, pulsed-electric field pretreatments.<sup>71–73</sup> Physical pretreatment methods require high energy utilization, and are therefore expensive for large-scale implementation.<sup>1</sup> Among chemical methods, acid and alkali pretreatment are

Table 1 – Chemical composition of different harvest residues

Harvest residue	Cellulose, % <sub>DM</sub>	Hemicellulose, % <sub>DM</sub>	Lignin, % <sub>DM</sub>
Barley straw <sup>18–23</sup>	37.7 – 40.1	22.2 – 26.7	5.5 – 19.4
Canola straw <sup>24</sup>	44.0	6.2	14.7
Corn stalk <sup>22,25</sup>	35.0 – 39.0	16.8 – 42.0	7.0 – 7.3
Corn stalk, maize stover <sup>22,25</sup>	37.5 – 40.4	16.5 – 42.0	8.3
Oat straw <sup>19,22,23,28,29</sup>	31.7 – 39.4	23.4 – 28.2	4.1 – 23.6
Rice husk <sup>24</sup>	17.3	37.7	19.7
Rice straw <sup>20,22,30–33</sup>	19.6 – 40.2	19.0 – 50.4	1.8 – 14.7
Rye straw <sup>22,29,34</sup>	37.4 – 37.6	30.5	19.0 – 30.8
Soya stalks <sup>35</sup>	34.5	24.8	19.8
Soybean straw <sup>24</sup>	51.7	9.5	10.2
Spelt straw <sup>36</sup>	38.3	24.3	14.8
Sunflower stalks <sup>22,37</sup>	38.5 – 42.1	29.7 – 33.5	13.4 – 17.5
Wheat straw <sup>18–20,22,38–40</sup>	8.9 – 37.0	32.9 – 49.8	20.5 – 25.5

Table 2 – Chemical composition of different food industry by-products

Industrial by-products	Cellulose, % <sub>DM</sub>	Hemicellulose, % <sub>DM</sub>	Lignin, % <sub>DM</sub>
Apple pomace <sup>41,42</sup>	47.5	27.8	14.8 – 22.4
Barley husk <sup>43,44</sup>	39.0	12.0	22.0
Brewer's spent grain <sup>45–49</sup>	12.0 – 40.2	28.4 – 40.0	11.5 – 27.7
Corn cob <sup>22,50</sup>	33.7	31.9	6.1
Flax oil cake <sup>51</sup>	8.2	4.6	6.0
Grape pomace <sup>52,53</sup>	9.2 – 14.5	4.0 – 10.3	11.6 – 41.3
Hemp oil cake <sup>51</sup>	22.5	14.2	16.7
Hull-less pumpkin oil cake <sup>51</sup>	4.4	6.7	0.7
Olive mill waste <sup>54,53</sup>	24.8 – 33.8	13 – 16.3	13.3 – 15.8
Rapeseed cake <sup>56</sup>	15.9	12.5	6.6
Rice bran <sup>57</sup>	34.0	28.2	24.8
Rye bran <sup>43,58</sup>	5.0 – 6.0	ND	3.5 – 4.4
Sugar beet pulp <sup>59–61</sup>	21.5	30.0	3.9
Sugarcane bagasse <sup>62,63</sup>	36.9 – 45.7	25.6 – 29.6	18.9 – 26.1
Wheat bran <sup>64–66</sup>	9.0 – 12.0	38.9	3.0 – 5.0

ND – not determined

the most commonly used. Although they can achieve high solubilization of cellulose and hemicellulose, and removal of lignin, those methods cause a high environmental burden. The other chemical methods are oxidative and organosolv pretreatment,<sup>76,77</sup> ozonolysis, the use of ionic liquids<sup>73</sup> and novel natural deep eutectic solvents.<sup>6,78,79</sup> Physicochemical pretreatment methods include ammonia fiber explosion, ultrasonication, autohydrolysis, liquid hot water, wet oxidation, and CO<sub>2</sub> explosion pretreatment.<sup>73</sup> Biological pretreatment methods are described in the next chapter.

Based on the applied method of lignin isolation, various types of industrial lignin can be produced. Organosolv and soda lignin are produced during the sulfur-free pulping process, while lignosulfonate and Kraft lignin are produced in the sulfur-containing pulping process. All of these processes are based on the application of chemicals and/or high temperatures, and therefore could not be considered sustainable or environmentally friendly.

Kraft lignin represents 85 % of the world's industrial lignin. It is obtained in the chemical process of pulping wood and non-wood pulp with sodium sulfide (Na<sub>2</sub>S) and sodium hydroxide solution (NaOH) at a temperature of 160–180 °C and pH 9–13.5. Kraft lignin is soluble at pH > 10 and has a lower sulfur content (up to 3 %) compared to lignosulfonate lignin (4–8 %), which is obtained by pulping only a certain wood in the presence of bisulfite ions (HSO<sub>3</sub><sup>-</sup>) at 120–150 °C and pH 2–12 for 1–5 h. Lignosulfonate lignin has high ash content and needs to be purified before further use for the pro-

duction of energy or chemicals. It is soluble in acids, alkali, and polar solvents. Lignosulfonates can be used in the prevention of scaling in hot and cooling waters, and as solvent for micronutrients in liquid fertilizers.<sup>80</sup> Soda lignin is formed in the process of soda pulping non-wood materials such as agricultural waste (straw, bagasse, grass, etc.) using 13–16 % sodium hydroxide solution at a temperature of 150–200 °C and pH 9.5–13. This type of lignin does not contain sulfur, which makes it suitable for the production of adhesives according to environmentally friendly principles. Organosolv lignin is like sulfur-free lignin, and is obtained by pulping fibrous wood residues and food industry residues using organic solvents (mixture of water / ethanol or methanol, acetic acid, etc.) at a temperature of 150–200 °C.<sup>13,81,82</sup> The properties of Organosolv lignin differ from other industrial lignin because it contains fewer impurities, has a lower molecular weight, and is water insoluble.

### Biological pretreatment methods

Biological pretreatments can be performed by selected microorganism, microbial consortium or enzyme(s).<sup>73</sup> A comprehensive review on valorization of harvest residues and food-processing industry by-products by solid-state fermentation using various microorganisms was recently published by Šelo *et al.*<sup>17</sup> The majority of research has been dedicated to fungal-based solid-state pretreatment, particularly to the use of white-rot fungi from the class of Basidiomycetes. White-rot fungi improve the

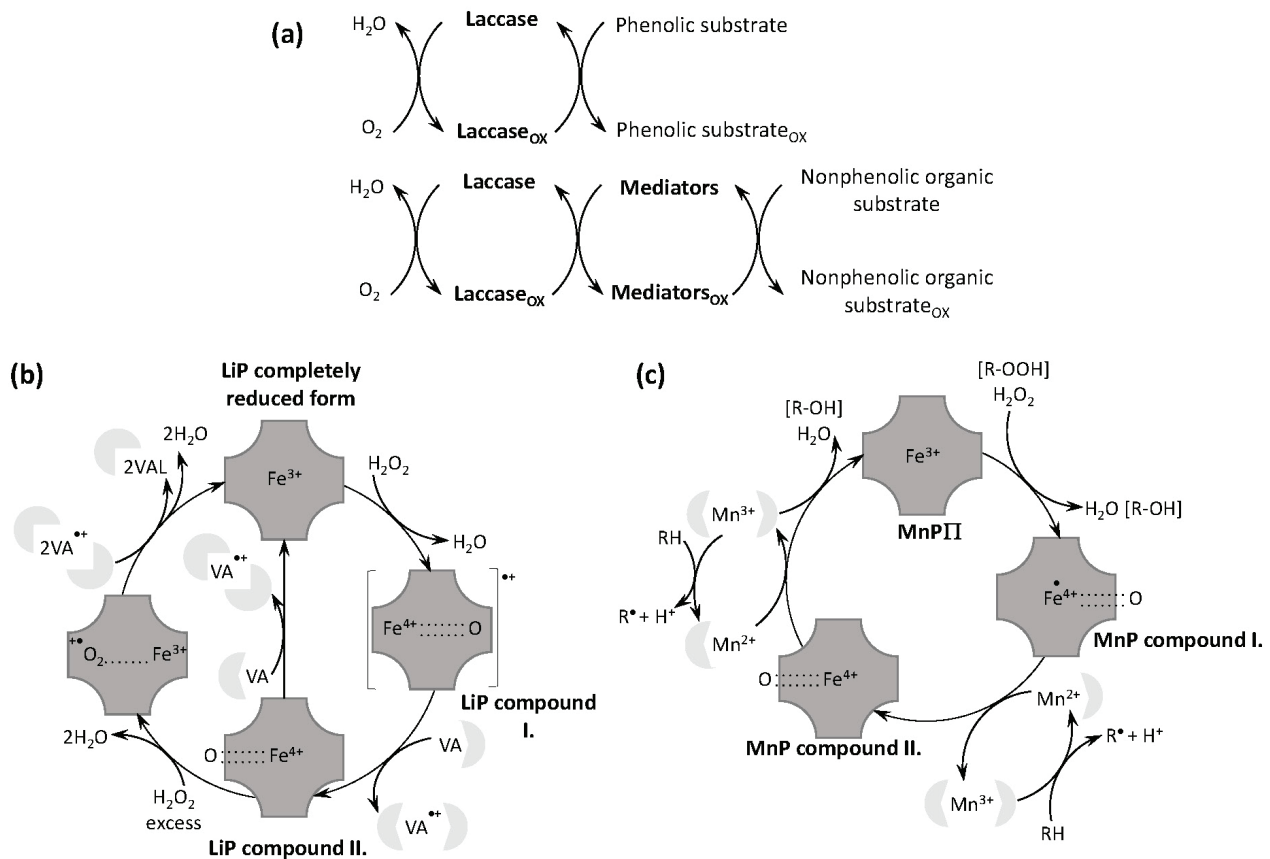


Fig. 2 – Catalytic mechanism of (a) laccase, (b) lignin peroxidase, (c) manganese peroxidase

biodegradability of lignocellulose by increasing the pore size of the material through penetration of the mycelium, and breaking the bonds between polysaccharides and lignin, removing lignin, releasing cellulose, and reducing the degree of polymerization of cellulose.

However, this method has several drawbacks, such as long duration, loss of organic matter during the treatment, technical challenge for the scale-up, and possible contamination.<sup>83,84</sup> Although white-rot fungi break down lignin, they are unable to utilize it as an energy source; therefore, it is assumed that they degrade lignin to access the cellulose.<sup>83</sup> To degrade the lignin, white-rot fungi produce ligninolytic enzymes (LEs). LEs are produced in small amounts and their optimal activities can be achieved through optimization of the media composition by supplementation with salts, low molecular weight phenolic compounds, and nutrition sources. However, the mechanism of LEs function is not completely known. Major LEs are laccase (Lacc), lignin peroxidase (LiP), manganese peroxidase (MnP), and versatile peroxidase (VP).<sup>85</sup> The catalytic mechanism of Lacc in oxidation of phenolic and nonphenolic substrates is presented in Fig. 2a, while the catalytic mechanisms of LiP and MnP are presented in Figs. 2b and 2c, respectively.<sup>86</sup>

Laccase (benzenediol: oxygen oxidoreductase, EC 1.10.3.2) can be considered as the key enzyme involved in lignin oxidation, modification or degradation. Laccases have high redox potential and are active towards a variety of substrates (phenolic and nonphenolic compounds), they can accept molecular oxygen, without the need for costly cofactors.<sup>87,88</sup> Oxidation of phenolic substrates involves removal of one electron from the phenolic hydroxyl groups to form phenyl hydroxyl radicals. With nonphenolic substrates, the use of mediators is essential. The most efficient laccase mediators are 1-hydroxybenzotriazole (HBT), *N*-hydroxyphthalimide (HPI), violuric acid (VLA), *N*-hydroxyacetanilide (NHA), *N*-hydroxyacetanilide (HAA) and 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO).<sup>13</sup> There have been many reviews in the last few years on laccase application for analytical, industrial, and environmental purposes.<sup>89–93</sup>

While laccases are involved in the degradation of lignin, cellulose and hemicellulose are degraded by cellulases and hemicellulases, respectively (Figs. 3a and 3b). The product of depolymerization of cellulose is glucose, whereas the degradation of hemicelluloses releases a mixture of different hexoses and pentoses.<sup>11</sup> There are three types of cellulases (Fig. 3a), namely, endoglucanases (carboxymethyl

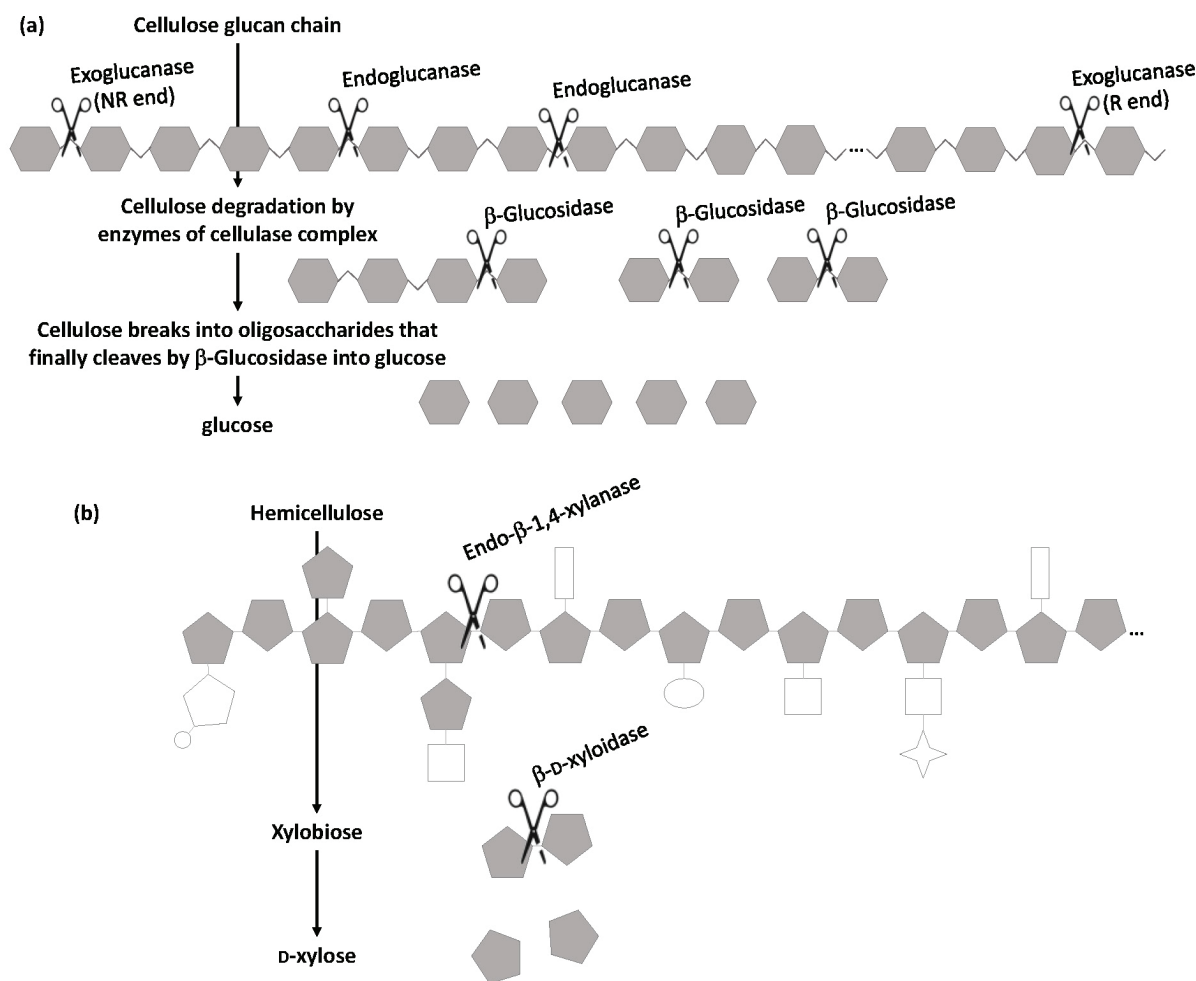


Fig. 3 – Enzymes involved in cellulose and hemicellulose degradation

cellulase), exoglucanases (cellobiohydrolase), and  $\beta$ -glucosidase.<sup>94</sup> To completely hydrolyze cellulose to glucose, all three enzymes are required. Endoglucanases randomly dissociate amorphous parts of cellulose, whereas exoglucanases extract cellobiose from crystalline parts of cellulose.  $\beta$ -glucosidases transform cellobiose to glucose, which can be used, e.g., for bioplastics and biofuels production.<sup>95</sup> However, the high cost and low efficiency of cellulases are the major issues in industrial-scale LB enzymatic degradation.

Large amounts of enzymes are required to produce concentrated glucose solutions due to substrate and product inhibition. Thermally stable cellulases and the immobilization of enzymes on solid supports have been investigated to improve the economics of cellulose degradation.<sup>95</sup> Additionally, protein engineering and directed evolution are powerful technologies to improve enzyme properties such as increased activity, decreased product inhibition, increased thermal stability, improved performance in nonconventional media, and pH stability.<sup>96</sup>

Hemicellulases include a group of enzymes involved in the hydrolysis of galactans, xylans, mannanans, and arabans. The major hemicellulases are endoxylanase (1,4- $\beta$ -D xylan xylanohydrolase), which hydrolyzes  $\beta$ -D-xylano pyranosyl linkages of xylan to form xylo-oligosaccharides, and  $\beta$ -D xylo-sidase (xylobiase), which catalyzes hydrolysis of xylobiose or xylo-oligosaccharides from the nonreducing end, releasing D-xylose in the hydrolysates (Fig. 3b). Xylose is a low-calorie sweetener and versatile feedstock for xylitol production. Many cellulases and hemicellulases that act on insoluble substrates have catalytic domain connected by a flexible peptide linker to a carbohydrate-binding module, which anchors the enzyme to the solid substrate. Carbohydrate-binding modules assist biomass hydrolysis by effectively increasing the concentration of their enzymes near the substrate surface and, depending on amino acid sequence and resulting shape, provide specificity to a certain substrate or substrate region (such as reducing or nonreducing ends).<sup>97,98</sup>

## Lignocellulosic biorefinery in sustainable development and circular bioeconomy

The concept and objectives of the circular economy and the bioeconomy overlap, hence the combined term circular bioeconomy is introduced.<sup>99</sup> The circular bioeconomy is based solely on the use of natural sources, and enables greater environmental sustainability compared to the use of fossil resources. It promotes human imitation of natural processes and activities, and seeks to make all processes circular by reusing the waste produced and using all outputs as inputs to other processes. Bioeconomies are highly dependent on the availability of resources and logistics. Therefore, the development of bioeconomies may depend on strong cooperation between regions that are rich in bioresources and regions that have appropriate technology but insufficient resources.<sup>100</sup> Effective biomass utilization through the strategic use of resources is essential for the production of valuable products, sustainable development, and the maximization of environmental and socioeconomic benefits.<sup>99</sup>

Biorefineries are industrial processes that aim to produce multiple value-added industrial products, fuels, and chemicals from various feedstocks.<sup>101</sup> Biorefineries are generally developed in response to the instability of the petrochemical industry, and out of concern for sustainable energy development and climate change. Biorefinery operations can be made more competitive by using lignocellulosic feedstocks and integrating multiple revenue streams, which is then referred to as lignocellulosic biorefineries.<sup>102</sup> In lignocellulosic biorefineries no single microorganism can catalyze all process steps. By combining specific strains and targeting multiple products, full biomass valorization could be achieved.<sup>103</sup> It is important to emphasize that lignocellulosic biomass as a feedstock in lignocellulosic biorefineries can only be considered after intensive evaluation of production costs, availability, and market value.<sup>9</sup> Although various lignocellulosic energy crops are often used in biorefineries, much effort is dedicated to the use of lignocellulosic waste biomass. In order to develop a sustainable biorefinery, it is important to take an integrated approach to biofuel production and the production of high value-added chemicals, which is explained further herein, where high value-added chemicals refer to those produced from lignin.

### Biofuels from lignocellulosic biomass

Considering the feedstock and technologies used for biofuel production, both liquid (bioethanol, biobutanol, biodiesel) and gas biofuels (biogas, hy-

drogen, syngas) are classified into four generations.<sup>104</sup> The 1st generation biofuels come from biomass which is also a food source, which is the main drawback. The 2nd generation biofuels come from non-food biomass, the 3rd generation fuels use algae, and the 4th generation biofuels are the result of developments in plant biology and biotechnology (metabolic engineering) in carbon capture and storage technology.

Lignocellulosic waste biomass is a non-food biomass and is used as a feedstock for 2nd generation biofuel production. There are still some technical and economic hurdles to overcome before 2nd generation of biofuels becomes more positioned at an industrial scale. The first challenge is related to the availability, storage, and transport of lignocellulosic waste biomass to the biofuel plant, in case it is not available near the plant. The second problem is technological, due to lignocellulose recalcitrant structure resistant to degradation. Most of the efficient pretreatment methods are not environmentally friendly, while those that are, suffer from some disadvantages as described previously. To solve the first challenge, harvesting, transporting, storing, and delivering large volumes of high-quality LB throughout the whole year to a biofuel plant requires careful logistical analysis before plant investment and construction. Transportation of a massive volume of feedstock in an energy-saving manner to the biorefineries is a challenge.<sup>104</sup> To solve the second problem, technological, integration of the process of biofuel production together with the production of other products (e.g. feed or high-value chemicals) should be considered to be located at one place in lignocellulosic biorefineries.<sup>9</sup>

### High-value lignin-based products

Due to the high content of carbon (up to 80 %), hydrogen (up to 6 %), and high C/O ratio, lignin is a potential source of highly-valued aromatic compounds (phenols, vanillin, polymer building blocks), synthetic gas (syngas), and hydrogen. It can be used as an additive/binder in the production of cement and biofuels.<sup>1,81,105–106</sup> Furthermore, lignin can be used in the development of packaging materials (e.g., food packaging), in the production of polymer, and bioplastics. It can also be used for therapeutic purposes due to its antioxidant, antimicrobial, and anticancer effects.<sup>80,105</sup>

The conversion of lignin into value-added products involves three steps: isolation, depolymerization and final upgrading of the obtained platform chemicals.<sup>76</sup> However, the isolation of lignin from lignocellulose is not easy due to its complex structure, poor solubility, and unclear reactivity. Therefore, the industrial use of lignin for the production of value-added products is still limited, and almost

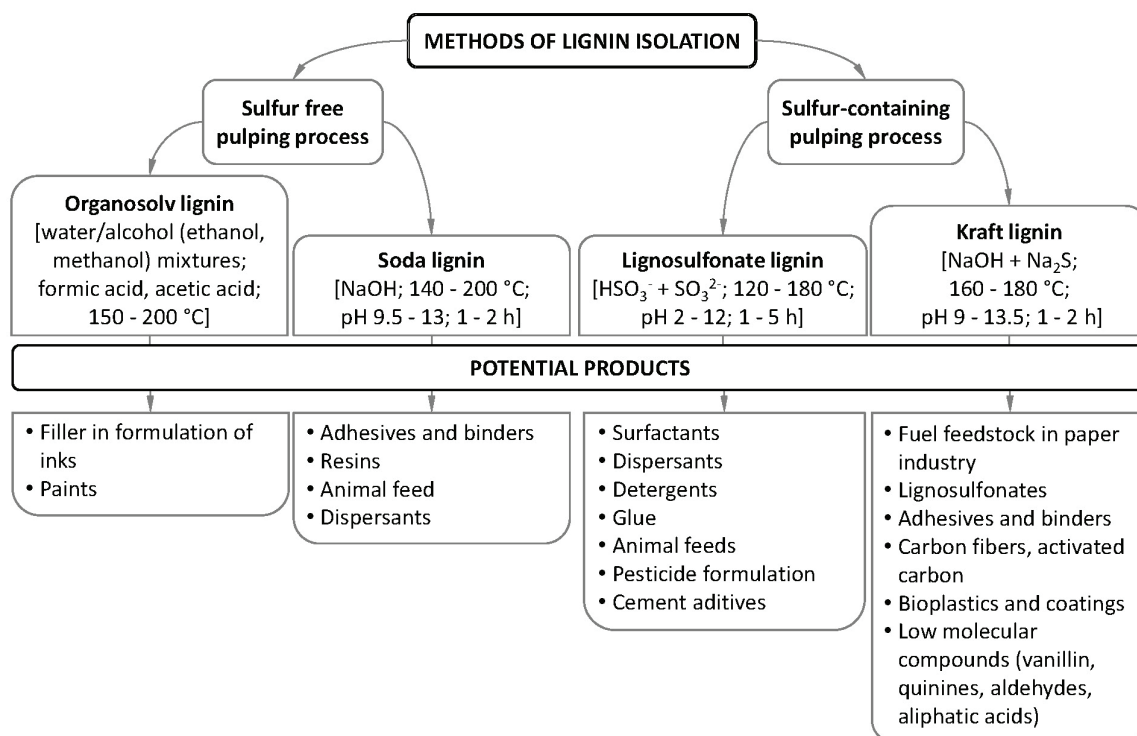


Fig. 4 – Type of lignin based on the methods of isolation and potential bioproducts from lignin

all industrial lignin is combusted to produce heat and electricity, whereas only 1–2 % of lignin is chemically transformed for industrial application.<sup>107</sup>

Industrial lignin is mostly produced by extraction from lignocellulosic biomass or industrial by-products using mechanical, chemical, or enzymatic methods.<sup>80,108</sup> In paper production and wood processing, after the separation of cellulose and hemicellulose, considerable amounts (10–50 %) of black (spent) fluid remain as a by-product, from which industrial lignin can be extracted. There are different types of black liquor (Kraft spent liquor, soda spent liquor, neutral sulfite semi-chemical spent liquor, etc.) depending on the raw material, the pulping process, and the cooking method used in the paper production. The properties of black liquor have influence on the further production process of desired products.

A schematic overview of lignin isolation together with the potential products is presented in Fig. 4.<sup>13,107,109–111</sup>

Although industrial lignin can be used directly in the production of certain chemicals (e.g., polyols), lignin must be modified or fragmented (by depolymerization or modification of functional or hydroxyl groups) for better use in the production of high-value products, as this increases the reactivity of lignin and creates new active sites. The most common methods for modifying lignin are oxidation, pyrolysis, hydrogenation, hydrolysis, gasification, and microbial transformation (Fig. 5).<sup>13,81,82,103,108,112</sup>

One of the most important products of lignin modification are aromatic compounds, such as benzene, toluene, xylene, and phenolic compounds, which are precursors of various highly-valued lignin-based products, such as resins, polyesters, nylon fibers, and polyesters, among others.<sup>113</sup>

The best known aromatic compound obtained with lignin modification by oxidation or microbial transformation, is vanillin. Vanillin is a precursor for the synthesis of various polymers.<sup>113,114</sup> Microbial conversion offers a novel, inexpensive route to the production of high-value products, but the valorization of lignin in this way can be hindered by the tendency of the degraded lignin fractions to undergo repolymerization and condensation reactions.<sup>106</sup>

Kraft lignin and lignosulfonate lignin have the highest commercial application. Kraft lignin products are shown in Fig. 4. They include lignosulfonates, technical carbons, bioplastics and coatings, binders and adhesives, and low-molecular weight compounds such as vanillin, quinines, aldehydes, etc.<sup>107,109</sup> The important high-value products made from Kraft lignin are carbon fibers. They are characterized by high strength, low mass, high thermal and chemical stability, and corrosion resistance. Therefore, they are suitable for the manufacture of sports equipment and composite materials. They find their application in the automobile and aircraft industries. The advantage of lignin in the production of carbon fibers over nonrenewable materials such as polyacrylonitrile (PAN) and pitch, is its non-toxicity, lower melting point, and faster stabili-



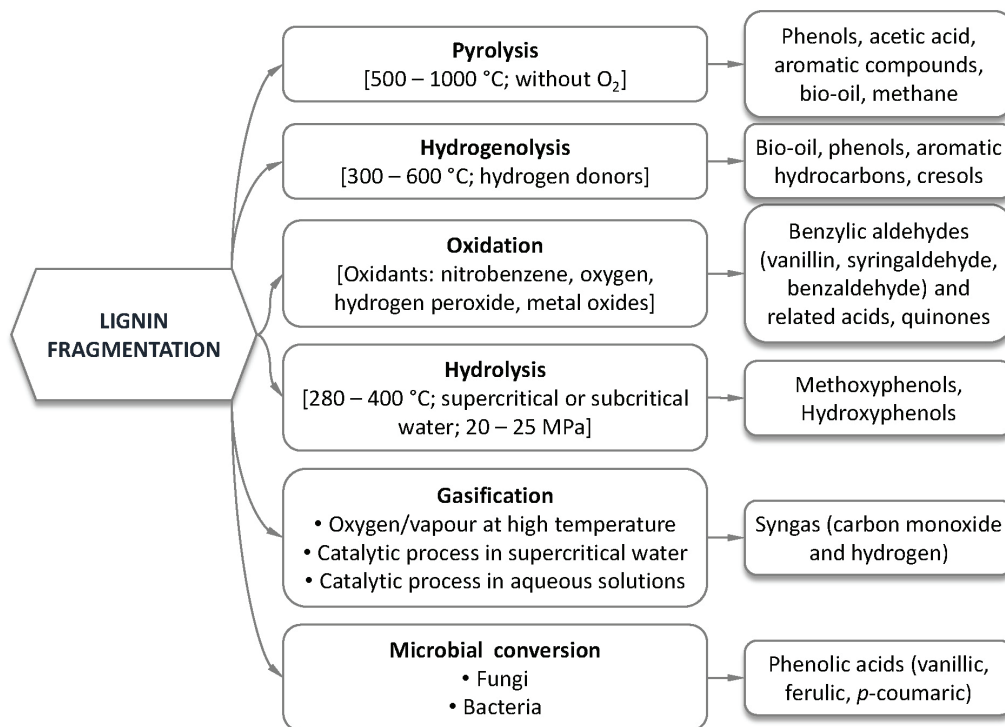


Fig. 5 – Most common processes of lignin fragmentation and potential bio-based products

zation. For the production of lignin-derived carbon fibers, it is necessary to extrude the isolated lignin into fibers, stabilize the fibers by oxidation, and finally pyrolyze them.<sup>113,115,116</sup>

Among the possibilities for lignin utilization is the application of lignin in the production of plastics and new composites. Lignin-based plastics can be obtained by chemically modifying lignin by changing its properties such as viscosity and elasticity or by mixing lignin with certain polymers, e.g., with poly(ethylene oxide) or with acrylonitrile butadiene, which is used as a thermoplastic resin in the automotive industry, in the manufacture of toys, etc. The main limitation in the preparation of plastics using lignin is the immiscibility of lignin with most polymers, as the interactions between them are weak compared to the interactions between lignin molecules due to the large number of polar functional groups of lignin. However, by adding various coupling agents (e.g. polyalkylene oxide, polyvinyl alcohol, ethylene vinyl-acetate copolymer, etc.), it is possible to improve the dispersion and mixing of lignin with a particular polymer.

Kraft lignin can be used as a dispersant (e.g., in the manufacture of pesticides, cement, ceramics) and coagulant (e.g., in the removal of dyes from solvents in the textile industry), but with prior modification to lignosulfonates to increase its solubility in aqueous medium and increase charge density.<sup>117</sup> Some of the modification processes are carboxymethylation,<sup>118</sup> sulfomethylation,<sup>119</sup> phenolation fol-

lowed by sulfonation with sulfuric acid and sodium sulfite.<sup>120</sup> Since lignin contains phenolic units in its structure, it can be used as a substitute in the commercial synthesis of phenol-formaldehyde-based adhesives. Moreover, pine Kraft lignin has been shown to contribute to better water absorption and mechanical properties such as strength, elasticity, etc. in the synthesis of lignin-phenol-formaldehyde compared to phenol-formaldehyde resin.<sup>121</sup>

Lignosulfonate lignin is most commonly used as a dispersant and binder in the manufacture of cement and concrete mixes to reduce the water content and increase the rate of hardening. In general, the term dispersant is often used for surfactants, plasticizers or emulsifiers, depending on the field of application.<sup>113</sup>

The dispersibility of lignosulfonates depends on the balance between molecular weight and species, and the number of functional groups. Various modifications of lignosulfonate lignin alter the properties of lignin. For example, oxidation or nitration of lignosulfonate increases the plasticizing ability in concrete. Reducing the sulfur content in lignosulfonate lignin increases hydrophobicity, and thus improves dispersibility. The same effect is achieved by increasing the molecular weight of lignosulfonates (10,000 – 50,000 g mol<sup>-1</sup>) and oxidation reactions leading to increased availability of lignin functional groups and increased dispersibility. Lignosulfonate lignin is used as an additive in animal feed production, where it can have a binding

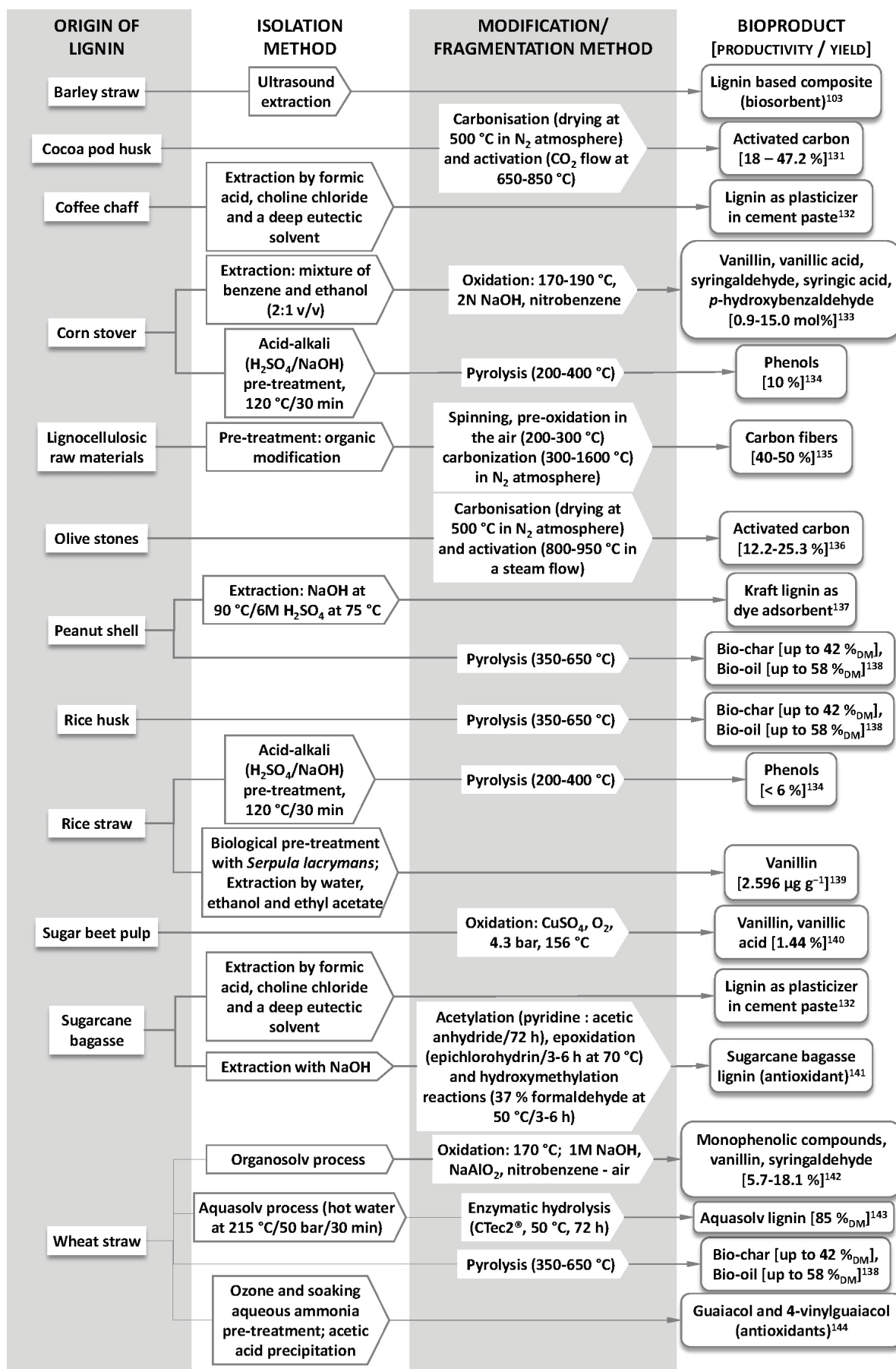


Fig. 6 – Some valuable products obtained from lignin isolated from agro-food waste

function, e.g., in the production of animal feed pellets, or it can be used as an encapsulating agent for fat-soluble vitamins, carotenoids, etc.<sup>122–124</sup>

Organosolv and soda lignin, due to the absence of sulfur, have properties more similar to natural lignin than Kraft and lignosulfonate lignin, and have potential in the development of high-value products according to the environmentally friendly concept. Organosolv lignin is used as an additive for paints, coatings, and as a filler in the formulation of printing inks, while improving the viscosity properties of the products.<sup>125</sup> Although it can be used in the manufacture of most products like Kraft lignin, it is not suitable for binders and adhesives due to its low molecular weight.<sup>97</sup> Soda lignin is used in the production of phenolic resins, animal feed, dispersants, and polymer synthesis.<sup>126</sup>

Phenolic resin, used as a wood adhesive, is commercially prepared on the basis of phenol-formaldehyde. Because of the carcinogenicity of formaldehyde, alternative compounds are being investigated for its replacement, such as aldehyde glyoxal, which is nontoxic and readily biodegradable. Since the structure of lignin is similar to that of phenol-formaldehyde, lignin can partially replace the phenolic part of the resin structure. Comparison of soda lignin and Kraft lignin in the preparation of lignin-phenol-glyoxal resin showed that the use of soda lignin results in a resin that has similar properties to a commercial phenol-formaldehyde resin compared to the resin where Kraft lignin was used. This is due to the better cross-linking of soda lignin with glyoxal due to the higher number of phenolic – OH groups and higher molecular weight compared to Kraft lignin, resulting in higher resin strength and viscosity.<sup>127,128</sup> Sameni *et al.*<sup>129</sup> demonstrated that the addition of soda lignin to high-density polyethylene, used in the packaging and automotive industries, significantly increases the tensile and flexural strength of the polymer due to the low molecular weight, low hydroxyl content, low polar component, and low sulfur content of the soda lignin.

Many compounds from lignin isolated from agro-food waste can be produced, as presented in Fig. 6. However, phenolic compounds are among the most important.<sup>134</sup> They can be used for the production of bioplastics, epoxy- and polyurethane resins, aromatic compound vanillin<sup>133,139,140,142</sup> and its precursor guaiacol.<sup>144</sup> Phenolic acids can be used as food additives to improve the nutritional, organoleptic, and biological properties of food products, as well as in the pharmaceutical sector. Carbon fibers<sup>135</sup> from lignin have great industrial potential (they have yet to be commercially applicable) due to their strength and wide applicability (e.g., in the automotive industry).<sup>80</sup> Activated charcoal<sup>131,136</sup> has good properties as an adsorbent and finds applica-

tion in deodorization and purification of process streams.<sup>80</sup> Lignin-based biocomposites show good properties in heavy metal adsorption.<sup>130</sup>

## Distribution of the bio-based industry in EU

There is an extensive database of EU facilities at pilot and industrial scale, or laboratory level that produce different categories of bio-based products, available on Data portal of agro-economics Modelling – DataM: DataBio-based industry and biorefineries.<sup>145</sup> Bio-based products are categorized as chemicals, liquid biofuels, composites, and fibers, biomethane, pulp and paper, sugar, starch, and timber. Fig. 7 presents bio-based products from lignocellulosic biomass produced at industrial, pilot-scale and laboratory level in European Union. It is clearly visible that most of the feedstocks used for the commercial production of bio-based products originates from the forestry with pulp and paper, and timber as main products. It is interesting to observe that only one industrial scale facility uses forestry feedstock for biomethane production. The majority of commercial liquid biofuels and bio-based chemicals originates from the agricultural feedstock. Ninety-six pilot-scale facilities operate in the field of liquid fuels production using forestry or grasses and short rotation feedstocks.

## Conclusion and future perspective

Lignocellulosic waste biomass is a valuable, renewable feedstock that can be used in lignocellulosic biorefineries for the production of bio-based products to reach sustainable development goals following the principles of circular bioeconomy. The production of multiple products from lignocellulosic biomass requires integration of various processes.

Considering the heterogeneous chemical composition of lignocellulose, the industry faces many challenges, such as the availability of a single type of biomass throughout the year. High processing cost, huge capital investment including transportation and storage cost for lignocellulosic biomass, efficient and sustainable lignocellulosic pretreatment and fractionation techniques focusing on lignin isolation, fractionation and modification are some of the main barriers for profitable biorefineries based on lignocellulosic waste as feedstock.

## ACKNOWLEDGMENTS

*This work was supported by the European Regional Development Fund (ERDF) (Grant KK.01.1.1.04.0107).*

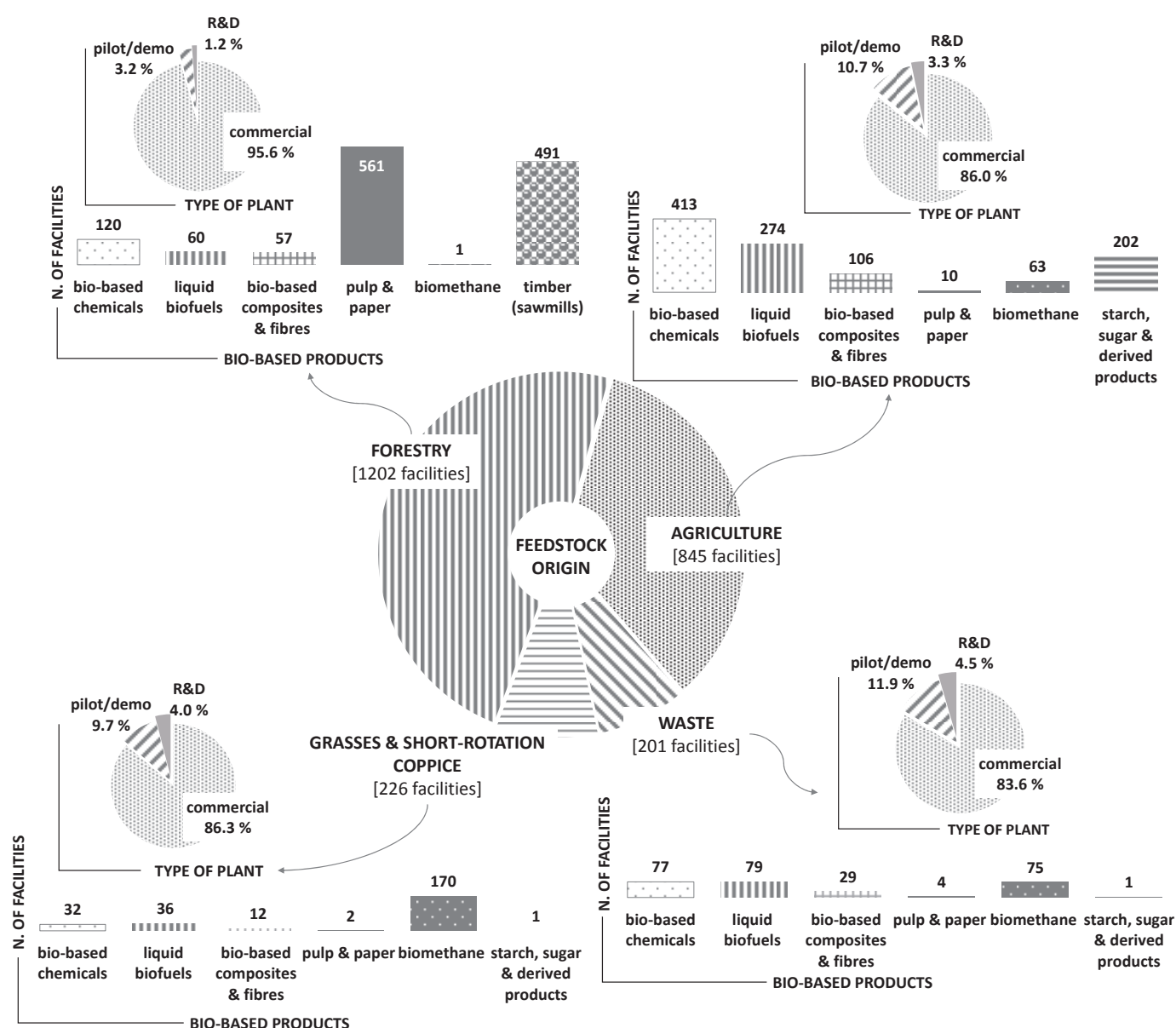


Fig. 7 – Production of bio-based products from lignocellulosic biomass at industrial, pilot-scale and laboratory level in European Union

## References

- Liu, Y., Nie, Y., Lu, X., Zhang, L., Zhang, X., He, H., Pan, F., Zhou, L., Liu, X., Ji, X., Zhang, S., Cascade utilization of lignocellulosic biomass to high-value products, *Green Chem.* **21** (2019) 3499. doi: <https://doi.org/10.1039/C9GC00473D>
- Dahmen, N., Lewandowski, I., Zibek, S., Weidtmann, A., Integrated lignocellulosic value chains in a growing bio-economy: Status quo and perspectives, *GCB Bioenergy* **11** (2019) 107. doi: <https://doi.org/10.1111/gcbb.12586>
- Cherubini, F., Jungmeier, G., Wellisch, M., Willke, T., Skiadas, I., Van Ree, R., de Jong, E., Toward a common classification approach for biorefinery systems, *Biofuels, Bioprod. Biorefin.* **3** (2009) 534. doi: <https://doi.org/10.1002/bbb.172>
- Arevalo-Gallegos, A., Ahmad, Z., Asgher, M., Parra-Saldivar, R., Iqbal, H. M. N., Lignocellulose: A sustainable material to produce value added products with a zero waste approach – A review, *Int. J. Biol. Macromol.* **99** (2017) 308. doi: <https://doi.org/10.1016/j.ijbiomac.2017.02.097>
- Wan, C., Li, Y., Fungal pretreatment of lignocellulosic biomass, *Biotechnol. Adv.* **30** (2012) 147. doi: <https://doi.org/10.1016/j.biotechadv.2012.03.003>
- Satlewal, A., Agrawal, R., Bhagia, S., Sangoro, J., Ragauskas A. J., Natural deep eutectic solvents for lignocellulosic biomass pretreatment: Recent developments, challenges and novel opportunities, *Biotechnol. Adv.* **36** (2018) 2032. doi: <https://doi.org/10.1016/j.biotechadv.2018.08.009>
- Procentese, A., Raganati, F., Olivieri, G., Russo, M. E., Rehman, L., Marzocchella, A., Deep eutectic solvents pre-

- treatment of agro-industrial food waste, *Biotechnol. Biofuels* **37** (2018) 3788.  
doi: <https://doi.org/10.1186/s13068-018-1034-y>
8. Panić, M., Andlar, M., Tišma, M., Rezić, T., Šibalić, D., Cvjetko Bubalo, M., Radojčić Redovniković, I., Natural deep eutectic solvent as a unique solvent for valorisation of orange peel waste by the integrated biorefinery approach, *Waste Manage.* **120** (2021) 340.  
doi: <https://doi.org/10.1016/j.wasman.2020.11.052>
  9. De Bhowmick, G., Sarmah, A. K., Sen, R., Lignocellulosic biorefinery as a model for sustainable development of bio-fuels and value added products, *Bioresour. Technol.* **247** (2018) 1144.  
doi: <https://doi.org/10.1016/j.biortech.2017.09.163>
  10. Park, S., Baker, J. O., Himmel, M. E., Parilla, P. A., Johnson, D. K., Cellulose crystallinity index: Measurement techniques and their impact on interpreting cellulase performance, *Biotechnol. Biofuels* **3** (2010) 10.  
doi: <https://doi.org/10.1186/1754-6834-3-10>
  11. Horn, S. J., Vaaje-Kolstad, G., Westereng, B., Novel enzymes for the degradation of cellulose, *Biotechnol. Biofuels* **5** (2012) 45.  
doi: <https://doi.org/10.1186/1754-6834-5-45>
  12. Zabed, H. M., Akter, S., Yun, J., Zhang, G., Awad, F. N., Qi, X., Sahu, J. N., Recent advances in biological pretreatment of microalgae and lignocellulosic biomass for biofuel production, *Renew. Sust. Energ. Rev.* **105** (2019) 105.  
doi: <https://doi.org/10.1016/j.rser.2019.01.048>
  13. Asina, F. N. U., Brzonova, I., Koliak, E., Kubátova, A., Ji, Y., Microbial treatment of industrial lignin: Successes, problems and challenges, *Renew. Sust. Energ. Rev.* **77** (2017) 1179.  
doi: <https://doi.org/10.1016/j.rser.2017.03.098>
  14. Jiankui, S., Helong, L., Ling-Ping, X., Xuan, G., Yunming, F., Run-Cang, S., Guoyong S., Fragmentation of woody lignocellulose into primary monolignols and their derivatives, *ACS Sustainable Chem. Eng.* **7** (2019) 4666.  
doi: <https://doi.org/10.1021/acssuschemeng.8b04032>
  15. Kumari, D., Singh, R., Pretreatment of lignocellulosic wastes for biofuel production: A critical review, *Renew. Sust. Energ. Rev.* **90** (2018) 877.  
doi: <https://doi.org/10.1016/j.rser.2018.03.111>
  16. Blair, M. J., Gagnon, B., Klain, A., Kulišić, B., Contribution of biomass supply chains for bioenergy to sustainable development goals, *Land* **10** (2021) 181.  
doi: <https://doi.org/10.3390/land10020181>
  17. Šelo, G., Planinić, M., Tišma, M., Tomas, S., Koceva Komlenić, D., Bucić-Kojić, A., A comprehensive review on valorization of agro-food industrial residues by solid-state fermentation, *Foods* **10** (2021) 927.  
doi: <https://doi.org/10.3390/foods10050927>
  18. Abegaz, A., van Keulen, H., Oosting, S. J., Feed resources, livestock production and soil carbon dynamics in Teghane, Northern Highlands of Ethiopia, *Agr. Syst.* **94** (2007) 391.  
doi: <https://doi.org/10.1016/j.agsy.2006.11.001>
  19. Aqsha, A., Tijani, M. M., Moghtaderic, B., Mahinpey, N., Catalytic pyrolysis of straw biomasses (wheat, flax, oat and barley) and the comparison of their product yields, *J. Anal. Appl. Pyrolysis* **125** (2017) 201.  
doi: <https://doi.org/10.1016/j.jaap.2017.03.022>
  20. Balsari, P., Menardo, S., Airoldi, G., Effect of Physical and Thermal Pre-treatments on Biogas Yield of Some Agricultural By-products, in *Progress in Biogas Stuttgart-Hohenheim 2011*, Vol. 1, German Society for Sustainable Biogas and Bioenergy, Stuttgart, 2011, pp 89-94.
  21. Laborel-Préneron, A., Magniont, C., Aubert, J. E., Characterization of barley straw, hemp shiv and corn cob as resources for bio aggregate based building materials, *Waste Biomass Valor.* **9** (2018) 1095.  
doi: <https://doi.org/10.1007/s12649-017-9895-z>
  22. Mussatto, S. I., Ballesteros, L. F., Martins, S., Teixeira, J. A., Use of Agro-Industrial Wastes in Solid-State Fermentation Processes, in Show, K.-Y. and Guo, X. (Eds.) *Industrial Waste*, InTech, Rijeka, 212, pp 121-140.  
doi: <https://doi.org/10.5772/36310>
  23. Pronyk, C., Mazza, G., Fractionation of triticale, wheat, barley, oats, canola, and mustard straws for the production of carbohydrates and lignins, *Bioresour. Technol.* **106** (2012) 117.  
doi: <https://doi.org/10.1016/j.biortech.2011.11.071>
  24. Nasehi, M., Torbatinejad, N. M., Zerehdaran, S., Safaie, A. R., Effect of solid-state fermentation by oyster mushroom (*Pleurotus florida*) on nutritive value of some agro by-products, *J. Appl. Anim. Res.* **45** (2016) 221.  
doi: <https://doi.org/10.1080/09712119.2016.1150850>
  25. Daud, Z., Zainuri, M., Hatta, M., Kassim, A. S. M., Awang, H., Aripin, A. M., Analysis the chemical composition and fiber morphology structure of corn stalk, *Aust. J. Basic Appl. Sci.* **7** (2013) 401.
  26. Atuhaire, M., Kabi, F., Okello, S., Mugerwa, S., Ebong, C., Optimizing bio-physical conditions and pre-treatment options for breaking lignin barrier of maize stover feed using white rot fungi, *Anim. Nutr.* **2** (2016) 361.  
doi: <https://doi.org/10.1016/j.aninu.2016.08.009>
  27. Omer, H. A. A., Ahmed, S. M., El-Kady, R. I., El-Shahat, A. A., El-Ayek, M. Y., El-Nattat, W. S., Morad, A. A. A., Nutritional impact of partial or complete replacement of clover hay by untreated or biologically treated rice straw and corn stalks on: 1. growth performance and economic evaluation of growing New Zealand (NZW) White rabbits, *Bull. Natl. Res. Cent.* **43** (2019) 192.  
doi: <https://doi.org/10.1186/s42269-019-0235-2>
  28. Gomez-Tovar, F., Celis, L. B., Razo-Flores, E., Alatryste-Mondragón, F., Chemical and enzymatic sequential pretreatment of oat straw form ethane production, *Bioresour. Technol.* **116** (2012) 372.  
doi: <https://doi.org/10.1016/j.biortech.2012.03.109>
  29. Masłowski, M., Miedzianowska, J., Strąkowska, A., Strzelec, K., Szyrkowska, M. I., The use of rye, oat and triticale straw as fillers of natural rubber composites, *Polym. Bull.* **75** (2018) 4607.  
doi: <https://doi.org/10.1007/s00289-018-2289-y>
  30. El-Sayed, M. A., El-Samni, T. M., Physical and chemical properties of rice straw ash and its effect on the cement paste produced from different cement types, *J. King. Saud. Univ. Eng. Sci.* **19** (2006) 21.  
doi: [https://doi.org/10.1016/S1018-3639\(18\)30845-6](https://doi.org/10.1016/S1018-3639(18)30845-6)
  31. Jia, J., Chen, H., Wu, B., Cui, F., Fang, H., Wang, H., Ni, Z., Protein production through microbial conversion of rice straw by multi-strain fermentation, *Appl. Biochem. Biotechnol.* **187** (2019) 253.  
doi: <https://doi.org/10.1007/s12010-018-2792-5>
  32. Nazli, M. H., Halim, R. A., Abdullah, A. M., Hussin, G., Samsudin, A. A., Potential of feeding beef cattle with whole corn crop silage and rice straw in Malaysia, *Trop. Anim. Health Prod.* **50** (2018) 1119.  
doi: <https://doi.org/10.1007/s11250-018-1538-2>
  33. Zayed, M. S., Enhancement the feeding value of rice straw as animal fodder through microbial inoculants and physical treatments, *Int. J. Recyc. Org. Waste Agricult.* **7** (2018) 117.  
doi: <https://doi.org/10.1007/s40093-018-0197-7>

34. Wang, L., Skreiberg, Ø., Becidan, M., Li, H., Sintering of rye straw ash and effect of additives, *Energy Procedia* **61** (2014) 2008.  
doi: <https://doi.org/10.1016/j.egypro.2014.12.063>
35. Singh nee' Nigam, P., Gupta, N., Anthwal, A., Pre-treatment of agro-industrial residues, in Singh nee' Nigam, P. and Pandey, A. (Eds.) *Biotechnology for Agro-Industrial Residues Utilisation*, Springer, Dordrecht, 2009, pp 13-33.  
doi: [https://doi.org/10.1007/978-1-4020-9942-7\\_2](https://doi.org/10.1007/978-1-4020-9942-7_2)
36. de Barros, R. R. O., Becarelli, P., de Oliveira, R. A., Tognotti, L., da Silva Bon, E. P., Triticum spelta straw hydrothermal pretreatment for the production of glucose syrups via enzymatic hydrolysis, *Biochem. Eng. J.* **151** (2019) 107340.  
doi: <https://doi.org/10.1016/j.bej.2019.107340>
37. Abd alla Idris, F. E., Osman, S., Idris, S., Alfa, M., Influence of irrigation system on characteristics of pulp and paper manufactured from sunflower stalks, *Int. J. Sci. Technol.* **1** (2012) 248.
38. Mouthier, T. M. B., de Rink, B., van Erven, G., de Gijssel, P., Schols, H. A., Kabel, M. A., Low liquid ammonia treatment of wheat straw increased enzymatic cell wall polysaccharide biodegradability and decreased residual hydroxycinnamic acids, *Bioresour. Technol.* **272** (2019) 288.  
doi: <https://doi.org/10.1016/j.biortech.2018.10.025>
39. Sharma, B., Agrawal, R., Singhania, R. R., Satlewal, A., Mathur, A., Tuli, D., Adsul, M., Untreated wheat straw: Potential source for diverse cellulolytic enzyme secretion by *Penicillium janthinellum* EMS-UV-8 mutant, *Bioresour. Technol.* **196** (2015) 518.  
doi: <https://doi.org/10.1016/j.biortech.2015.08.012>
40. Sharma, R. K., Arora, D. S., Bioprocessing of wheat and paddy straw for their nutritional up-gradation, *Bioprocess Biosyst. Eng.* **37** (2014) 1437.  
doi: <https://doi.org/10.1007/s00449-013-1116-y>
41. Baray Guerrero, M. R., Salinas Gutiérrez, J. M., Meléndez Zaragoza, M. J., López Ortiz, A., Collins-Martínez, V., Optimal slow pyrolysis of apple pomace reaction conditions for the generation of a feedstock gas for hydrogen production, *Int. J. Hydrogen Energy* **41** (2016) 23232.  
doi: <https://doi.org/10.1016/j.ijhydene.2016.10.066>
42. Kosmala, M., Kozodziejczyk, K., Zdunczyk, Z., Juskiewicz, J., Boros, D., Chemical composition of natural and polyphenol-free apple pomace and the effect of this dietary ingredient on intestinal fermentation and serum lipid parameters in rats, *J. Agric. Food Chem.* **59** (2011) 9177.  
doi: <https://doi.org/10.1021/jf201950y>
43. Berger, K., Falck, P., Linninge, C., Nilsson, U., Axling, U., Grey, C., Stålbrand, H., Nordberg Karlsson, E., Nyman, M., Holm, C., Adlercreutz, P., Cereal byproducts have prebiotic potential in mice fed a high-fat diet, *J. Agric. Food Chem.* **62** (2013) 8169.  
doi: <https://doi.org/10.1021/jf502343v>
44. Kohli, D., Garg, S., Jana, A. K., Thermal and morphological properties of chemically treated barley husk fiber, *IJRMET* **3** (2013) 153.
45. Carvalho, F., Esteves, M. P., Parajo, J. C., Pereira, H., Girio, F. M., Production of oligosaccharides by autohydrolysis of brewery's spent grain, *Bioresour. Technol.* **91** (2004) 93.  
doi: [https://doi.org/10.1016/S0960-8524\(03\)00148-2](https://doi.org/10.1016/S0960-8524(03)00148-2)
46. Khidzir, K. M., Abdullah, N., Agamuthu, P., Brewery spent grain: Chemical characteristics and utilization as an enzyme substrate, *Malays. J. Sci.* **29** (2010) 41.  
doi: <https://doi.org/10.22452/mjs.vol29no1.7>
47. Mathias, T. R. S., Alexandre, V. M. F., Cammarota, M. C., de Mello, P. P. M., Sérvulo, E. F. C., Characterization and determination of brewer's solid wastes composition, *J. Inst. Brew.* **121** (2015) 400.  
doi: <https://doi.org/10.1002/jib.229>
48. Mussatto, S. I., Roberto, I. C., Chemical characterization and liberation of pentose sugars from brewer's spent grain, *J. Chem. Technol. Biotechnol.* **81** (2006) 268.  
doi: <https://doi.org/10.1002/jctb.1374>
49. Xiros, C., Topakas, E., Katapodis, P., Christakopoulos, P., Hydrolysis and fermentation of brewer's spent grain by *Neurospora crassa*, *Bioresour. Technol.* **99** (2008) 5427.  
doi: <https://doi.org/10.1016/j.biortech.2007.11.010>
50. Anukam, A. I., Goso, B. P., Okoh, O. O., Mamphweli, S. N., Studies on characterization of corn cob for application in a gasification process for energy production, *J. Chem.* **8** (2017) 1.  
doi: <https://doi.org/10.1155/2017/6478389>
51. Budžaki, S., Strelec, I., Krnić, M., Alilović, K., Tišma, M., Zelić, B., Proximate analysis of cold-press oil cakes after biological treatment with *Trametes versicolor* and *Humicola grisea*, *Eng. Life Sci.* **18** (2018) 924.  
doi: <https://doi.org/10.1002/elsc.201800033>
52. Manara, P., Zabaniotu, A., Vanderghem, C., Richel, A., Lignin extraction from Mediterranean agro-waste: Impact of pretreatment conditions of lignin chemical structure and thermal degradation behaviour, *Catal. Today* **22** (2014) 25.  
doi: <https://doi.org/10.1016/j.cattod.2013.10.065>
53. Zheng, Y., Lee, C., Yu, C., Cheng, Y., Simmons, C. W., Zhang, R., Jenkis, B. M., Vander Gheynst, J. S., Ensilage and bioconversion of grape pomace into fuel ethanol, *J. Agric. Food Chem.* **60** (2012) 11128.  
doi: <https://doi.org/10.1021/jf303509v>
54. Ducom, G., Gautier, M., Pietraccini, M., Tagutchou, J. P., Lebouil, D., Gourdon, R., Comparative analyses of three olive mill solid residues from different countries and processes for energy recovery by gasification, *Renew. Energ.* **145** (2020) 180.  
doi: <https://doi.org/10.1016/j.renene.2019.05.116>
55. Roig, A., Cayuela, M. L., Sánchez-Monedero, M. A., An overview on olive mill wastes and their valorisation methods, *Waste Manage.* **26** (2006) 960.  
doi: <https://doi.org/10.1016/j.wasman.2005.07.024>
56. Eriksson, T., Murphy, M., Ciszuk, P., Burstedt, E., Nitrogen balance, microbial protein production, and milk production in dairy cows fed fodder beets and potatoes, or barley, *J. Dairy Sci.* **87** (2004) 1057.  
doi: [https://doi.org/10.3168/jds.S0022-0302\(04\)73252-X](https://doi.org/10.3168/jds.S0022-0302(04)73252-X)
57. Kodali, B., Pogaku, R., Pretreatment studies of rice bran for the effective production of cellulose, *Elec. J. Env. Agric. Food Chem.* **5** (2006) 1253.
58. Kamal-Eldin, A., Nygaard Lærke, H., Knudsen, K. E. B., Lampi, A. M., Piironen, V., Adlercreutz, H., Katina, K., Poutanen, K., Per Aman, P., Physical, microscopic and chemical characterisation of industrial rye and wheat brans from the Nordic countries, *Food Nutr. Res.* **53** (2009) 1912.  
doi: <https://doi.org/10.3402/fnr.v53i0.1912>
59. Clark, P., Armentano, L., Influence of particle size on the effectiveness of beet pulp fiber, *J. Dairy Sci.* **80** (1997) 898.  
doi: [https://doi.org/10.3168/jds.S0022-0302\(97\)76012-0](https://doi.org/10.3168/jds.S0022-0302(97)76012-0)
60. Iconomou, D., Kandyli, K., Isralidies, C., Nikokyris, P., Protein enhancement of sugar beet pulp by fermentation and estimation of protein degradability in the rumen of sheep, *Small Ruminant Res.* **27** (1998) 55.  
doi: [https://doi.org/10.1016/S0921-4488\(97\)00027-8](https://doi.org/10.1016/S0921-4488(97)00027-8)

61. *Selim, S., Hussein, E.*, Production performance, egg quality, blood biochemical constituents, egg yolk lipid profile and lipid peroxidation of laying hens fed sugar beet pulp, *Food Chem.* **310** (2020) 125864.  
doi: <https://doi.org/10.1016/j.foodchem.2019.125864>
62. *Rocha, G. J. M., Marcos Nascimento, V., Gonçalves, A. R., Silva, V. F. N., Martín, C.*, Influence of mixed sugarcane bagasse samples evaluated by elemental and physical-chemical composition, *Ind. Crops Prod.* **64** (2015) 52.  
doi: <https://doi.org/10.1016/j.indcrop.2014.11.003>
63. *Ahmed, M. H., Babiker, S. A., Elseed, A. E. M. A. F., Mohammed, A. M.*, Effect of urea-treatment on nutritive value of sugarcane bagasse, *ARPN J. Sci. Technol.* **3** (2013) 834.
64. *Cripwell, R. A., Lorenzo, F., Rose, S. H., Basaglia, M., Cagnin, L., Sergio, C., Heber van Zyl, W.*, Utilisation of wheat bran as a substrate for bioethanol production using recombinant cellulases and amyolytic yeast, *Appl. Energy* **160** (2015) 610.  
doi: <https://doi.org/10.1016/j.apenergy.2015.09.062>
65. *Kajala, I., Mäkelä, J., Coda, R., Shukla, S., Shi, Q., Maina, N. H., Juvonen, R., Ekholm, P., Goyal, A., Tenkanen, M., Katina, K.*, Rye bran as fermentation matrix boosts in situ dextran production by *Weissella confusa* compared to wheat bran, *Appl. Microbiol. Biotechnol.* **100** (2016) 3499.  
doi: <https://doi.org/10.1007/s00253-015-7189-6>
66. *Onipe, O. O., Jideani, A. I. O., Beswa, D.*, Composition and functionality of wheat bran and its application in some cereal food products, *Int. J. Food Sci. Technol.* **50** (2015) 2509.  
doi: <https://doi.org/10.1111/ijfs.12935>
67. *Steinbach, D., Kruse, A., Sauer, J.*, Pretreatment technologies of lignocellulosic biomass in water in view of furfural and 5-hydroxymethylfurfural production – A review, *Biomass Conv. Bioref.* **7** (2017) 247.  
doi: <https://doi.org/10.1007/s13399-017-0243-0>
68. *Renders, T., Cooreman, E., Van den Bosch, S., Schutyser, W., Koelwijjn, S.-F., Vangeel, T., Deneyer, A., Van den Bossche, G., Courtin, C. M., Sels, B. F.*, Catalytic lignocellulose biorefining in n-butanol/water: A one-pot approach toward phenolics, polyols, and cellulose, *Green Chem.* **20** (2018) 4607.  
doi: <https://doi.org/10.1039/C8GC01031E>
69. *Nitzsche, R., Budzinski, M., Gröngroft, A.*, Techno-economic assessment of a wood-based biorefinery concept for the production of polymer-grade ethylene, organosolv lignin and fuel, *Bioresour. Technol.* **200** (2016) 928.  
doi: <https://doi.org/10.1016/j.biortech.2015.11.008>
70. *Kovačić, Đ., Kralik, D., Rupčić, S., Jovičić, D., Spajić, R., Tišma, M.*, Soybean straw, corn stover and sunflower stalk as possible substrates for biogas production in Croatia: A review, *Chem. Biochem. Eng. Q.* **31** (2017) 187.  
doi: <https://doi.org/10.15255/CABEQ.2016.985>
71. *Kovačić, Đ., Kralik, D., Jovičić, D., Rupčić, S., Popović, B., Tišma, M.*, Thermal pretreatment of harvest residues and their use in anaerobic co-digestion with dairy cow manure, *Appl. Biochem. Biotechnol.* **184** (2018) 471.  
doi: <https://doi.org/10.1007/s12010-017-2559-4>
72. *Chen, H., Liu, J., Chang, X., Chen, D., Xue, Y., Liu, P., Lin, H., Han, S.*, A review on the pretreatment of lignocellulose for high-value chemicals, *Fuel Process. Technol.* **16** (2017) 196.  
doi: <https://doi.org/10.1016/j.fuproc.2016.12.007>
73. *Kumari, D., Singh, R.*, Coupled green pretreatment of petha wastewater and rice straw, *Environ. Sustainability Indic.* **5** (2020) 100013.  
doi: <https://doi.org/10.1016/j.indic.2019.100013>
74. *Kumar, A. K., Sharma, S.*, Recent updates on different methods of pretreatment of lignocellulosic feedstocks: A review, *Bioresour. Bioprocess.* **4** (2017) 7.  
doi: <https://doi.org/10.1016/j.indic.2019.100013>
75. *Kovačić, Đ., Rupčić, S., Kralik, D., Jovičić, D., Spajić, R., Tišma, M.*, Pulsed electric field: An emerging pretreatment technology in a biogas production, *Waste Manage.* **120** (2021) 467.  
doi: <https://doi.org/10.1016/j.wasman.2020.10.009>
76. *Jasiukaitytė-Grojzdek, E., Huš, M., Grilc, M., Likožar, B.*, Acid-catalysed  $\alpha$ -O-4 aryl-ether bond cleavage in methanol/(aqueous) ethanol: Understanding depolymerisation of a lignin model compound during organosolv pretreatment, *Sci. Rep.* **10** (2020) 11037.  
doi: <https://doi.org/10.1021/acssuschemeng.0c06099>
77. *Jasiukaitytė-Grojzdek, E., Huš, M., Grilc, M., Likožar, B.*, Acid-catalyzed  $\alpha$ -O-4 aryl-ether cleavage mechanisms in (aqueous)  $\gamma$ -valerolactone: Catalytic depolymerization reactions of lignin model compound during organosolv pretreatment, *ACS Sustainable Chem. Eng.* **8** (47) (2020) 17475.  
doi: <https://doi.org/10.1021/acssuschemeng.0c06099>
78. *Bjelić, A., Hočevár, B., Grilc, M., Novak, U., Likožar, B.*, A review of sustainable lignocellulose biorefining applying (natural) deep eutectic solvents (DESs) for separations, catalysis and enzymatic biotransformation processes, *Rev. Chem. Eng.* 2020.  
doi: <https://doi.org/10.1515/revce-2019-0077>
79. *Mamilla, J. L. K., Novak, U., Grilc, M., Likožar, B.*, Natural deep eutectic solvents (DES) for fractionation of waste lignocellulosic biomass and its cascade conversion to value-added bio-based chemicals, *Biomass Bioenergy* **120** (2019) 417.  
doi: <https://doi.org/10.1016/j.biombioe.2018.12.002>
80. *Bajpai, P.*, Value-Added Products from Lignin, *Bajpai, P. (Ed.)*, *Biotechnology for Pulp and Paper Processing*, Springer, Singapore, 2018, pp 561-571.  
doi: [https://doi.org/10.1007/978-981-10-7853-8\\_25](https://doi.org/10.1007/978-981-10-7853-8_25)
81. *Arapova, O. V., Chistyakov, A. V., Tsodikov, M. V., Moiseev, I. I.*, Lignin as a renewable resource of hydrocarbon products and energy carriers (A review), *Pet. Chem.* **60** (2020) 227.  
doi: <https://doi.org/10.1134/S0965544120030044>
82. *Al-Kaabi, Z., Pradhan, R. R., Thevathasan, N., Chiang, Y. W., Gordon, A., Dutta, A.*, Potential value added applications of black liquor generated at paper manufacturing industry using recycled fibers, *J. Cleaner Prod.* **149** (2017) 156.  
doi: <https://doi.org/10.1016/j.jclepro.2017.02.074>
83. *Gao, D., Du, L., Yang, J., Wu, W.-M., Liang, H.*, A critical review of the application of white rot fungus to environmental pollution control, *Crit. Rev. Biotechnol.* **30** (2010) 70.  
doi: <https://doi.org/10.3109/07388550903427272>
84. *Tišma, M., Žnidaršič-Plazl, P., Šelo, G., Tolj, I., Šperanda, M., Bucić-Kojić, A., Planinić, M.*, *Trametes versicolor* in lignocellulose-based bioeconomy: State of the art, challenges and opportunities, *Bioresour. Technol.* **330** (2021) 124997.  
doi: <https://doi.org/10.1016/j.biortech.2021.124997>
85. *Kumar, A., Chandra, R.*, Ligninolytic enzymes and its mechanisms for degradation of lignocellulosic waste in environment, *Heliyon* **6** (2020) e03170.  
doi: <https://doi.org/10.1016/j.heliyon.2020.e03170>
86. *Kulikova, N. A., Klein, O. I., Stepanova, E. V., Koroleva, O. V.*, Use of basidiomycetes in industrial waste processing and utilization technologies: Fundamental and applied aspects (review), *Appl. Biochem. Microbiol.* **47** (2011) 565.  
doi: <https://doi.org/10.1134/S000368381106007X>

87. Tišma, M., Zelić, B., Vasić-Rački, Đ., Žnidaršič-Plazl, P., Plazl, I., Modelling of laccase-catalyzed L-DOPA oxidation in a microreactor, *Chem. Eng. J.* **149** (2009) 383. doi: <https://doi.org/10.1134/S000368381106007X>
88. Tišma, M., Žnidaršič-Plazl, P., Plazl, I., Zelić, B., Vasić-Rački, Đ., Modelling of L-DOPA oxidation catalyzed by laccase, *Chem. Biochem. Eng. J.* **22** (2008) 307. doi: <https://doi.org/10.1016/j.ccej.2009.01.025>
89. Agrawal, K., Chaturvedi, V., Verma, P., Fungal laccase discovered but yet undiscovered, *Bioresour. Bioprocess.* **5** (2018) 4. doi: <https://doi.org/10.1186/s40643-018-0190-z>
90. Zerva, A., Simić, S., Topakas, E., Nikodinovic-Runic, J., Applications of microbial laccases: Patent review of the past decade (2009–2019), *Catalysts* **9** (2019) 1023. doi: <https://doi.org/10.3390/catal9121023>
91. Mayolo-Deloya, K., González-González, M., Rito-Palomares, M., Laccases in food industry: Bioprocessing, potential industrial and biotechnological applications, *Front. Bioeng. Biotechnol.* **5** (2020) 222. doi: <https://doi.org/10.3389/fbioe.2020.00222>
92. Debnath, R., Saha, T., An insight into the production strategies and applications of the ligninolytic enzyme laccase from bacteria and fungi, *Biocatal. Agric. Biotechnol.* **26** (2020) 101645. doi: <https://doi.org/10.1016/j.bcab.2020.101645>
93. Singh, D., Gupta, N., Microbial laccase: A robust enzyme and its industrial applications, *Biologia* **75** (2020) 1183. doi: <https://doi.org/10.2478/s11756-019-00414-9>
94. Srivastava, N., Srivastava, M., Mishra, P. K., Gupta, V. K., Molina, G., Rodriguez-Couto, S., Manikanta, A., Ramieke, P. W., Applications of fungal cellulases in biofuel production: Advances and limitations, *Renewable Sustainable Energy Rev.* **82** (2018) 2379. doi: <https://doi.org/10.1016/j.rser.2017.08.074>
95. Kobayashia, H., Fukuoka, A., Synthesis and utilisation of sugar compounds derived from lignocellulosic biomass, *Green Chem.* **15** (2013) 1740. doi: <https://doi.org/10.1039/C3GC00060E>
96. Del Mar Contreras, M., Lama-Muñoz, A., Manuel Gutiérrez-Pérez, J., Espínola, F., Moya, M., Castro, E., Protein extraction from agri-food residues for integration in biorefinery: Potential techniques and current status, *Bioresour. Technol.* **280** (2019) 459. doi: <https://doi.org/10.1016/j.biortech.2019.02.040>
97. Obeng, E. M., Adam, S. N. N., Budiman, C., Lignocellulases: A review of emerging and developing enzymes, systems, and practices, *Bioresour. Bioprocess.* **4** (2017) 16. doi: <https://doi.org/10.1186/s40643-017-0146-8>
98. Ferreira, A. M., Passos, H., Okafuji, A., Tavares, A. P. M., Ohno, H., Freire, M. G., An integrated process for enzymatic catalysis allowing product recovery and enzyme reuse by applying thermoreversible aqueous biphasic systems, *Green Chem.* **20** (2018) 1218. doi: <https://doi.org/10.1039/C7GC03880A>
99. Kumar Awasthi, M., Sarsaiya, S., Patel, A., Juneja, A., Prasad Singh, R., Yan, B., Kumar Awasthi, S., Jain, A., Liu, T., Duan, Y., Pandey, A., Zhang, Z., Taherzadeh, M. J., Refining biomass residues for sustainable energy and bio-products: An assessment of technology, its importance, and strategic applications in circular bio-economy, *Renew. Sust. Energy Rev.* **127** (2020) 109876. doi: <https://doi.org/10.1016/j.rser.2020.109876>
100. Salvador, R., Puglieri, F. N., Halog, A., de Andrade, F. G., Piekarski, C., De Francisco, A. C., Key aspects for designing business models for a circular bioeconomy, *J. Cleaner Prod.* **278** (2021) 124341. doi: <https://doi.org/10.1016/j.jclepro.2020.124341>
101. Kumar, B., Verma, P., Biomass-based biorefineries: An important archetype towards a circular economy, *Fuel* **288** (2020) 119622. doi: <https://doi.org/10.1016/j.fuel.2020.119622>
102. FitzPatrick, M., Champagne, P., Cunningham, M. F., Whitney, R. A., A biorefinery processing perspective: Treatment of lignocellulosic materials for the production of value-added products, *Bioresour. Technol.* **101** (2010) 8915. doi: <https://doi.org/10.1016/j.biortech.2010.06.125>
103. Troiano, D., Orsat, V., Dumont, M. J., Status of filamentous fungi in integrated biorefineries, *Renew. Sust. Energy Rev.* **117** (2020) 109472. doi: <https://doi.org/10.1016/j.rser.2019.109472>
104. Sharma, B., Larroche, C., Dussap, C.-G., Comprehensive assessment of 2G bioethanol production, *Bioresour. Technol.* **313** (2020) 123630. doi: <https://doi.org/10.1016/j.biortech.2020.123630>
105. Bhat, R., Ahmad, A., Jöudu, I., Applications of Lignin in the Agri-Food Industry, in Sharma S. and Kumar A. (Eds.), *Lignin, Springer Series on Polymer and Composite Materials*. Springer, Cham., 2020, pp 275-298. doi: [https://doi.org/10.1007/978-3-030-40663-9\\_10](https://doi.org/10.1007/978-3-030-40663-9_10)
106. Wang, H., Pu, Y., Ragauskas, A., Yang, B., From lignin to valuable products – strategies, challenges, and properties, *Bioresour. Technol.* **271** (2019) 449. doi: <https://doi.org/10.1016/j.biortech.2018.09.072>
107. Demuner, L. F., Colodette, J. L., Antonio Jacinto Demuner, A. J., Jardim, C. M., Biorefinery review: Wide-reaching products through kraft lignin, *BioResources* **14** (2019) 7543. doi: <https://doi.org/10.15376/biores.14.3.Demuner>
108. Chio, C., Sain, M., Qin, W., Lignin utilization: A review of lignin depolymerization from various aspects, *Renew. Sust. Energy Rev.* **107** (2019) 232. doi: <https://doi.org/10.1016/j.rser.2019.03.008>
109. Vishtal, A., Kraslawski, A., Challenges in industrial applications of technical lignins, *BioRes.* **6** (2011) 3547. doi: <https://doi.org/10.15376/biores.6.3.vishtal>
110. Al-Kaabi, Z., Pradhan, R., Thevathasan, N., Arku, P., Gordon, A., Dutta, A., Beneficiation of renewable industrial wastes from paper and pulp processing, *AIMS Energy* **6** (2018) 880. doi: <https://doi.org/10.3934/energy.2018.5.880>
111. Hilaes, R. T., Ramos, L., Ahmed, M. A., Ingle, A. P., Chandel, A. K., da Silva, S. S., Woon Choi, J., dos Santos J. C., Valorization of Lignin into Value-added Chemicals and Materials, in Ingle, A. P., Chandel, A. K. and da Silva, S. S. (Eds.), *Lignocellulosic Biorefining Technologies*, John Wiley & Sons Ltd., 2020, pp 247-263. doi: <https://doi.org/10.1002/9781119568858.ch11>
112. Wang, H., Ben, H., Ruan, H., Zhang, L., Pu, Y., Feng, M., Ragauskas, A. J., Yang, B., Effects of lignin structure on hydrodeoxygenation reactivity of pine wood lignin to valuable chemicals, *ACS Sustain. Chem. Eng.* **5** (2017) 1824. doi: <https://doi.org/10.1021/acssuschemeng.6b02563>
113. Jardim, J. M., Hart, P. W., Lucia, L., Jameel, H., Insights into the potential of hardwood kraft lignin to be a green platform material for emergence of the biorefinery, *Polymers* **12** (2020) 1795. doi: <https://doi.org/10.3390/polym12081795>



114. *de Souza, R. E., Gomes, F. J. B., Brito, E. O., Lelis, R. C. C., Batalha, L. A. R., Santos, F. A., Longue, D. J.*, A review on lignin sources and uses, *J. Appl. Biotechnol. Bioeng.* **7** (2020) 100.  
doi: <https://doi.org/10.15406/jabb.2020.07.00222>
115. *Bengtsson, A., Bengtsson, J., Sedin, M., Sjöholm, E.*, Carbon fibers from lignin-cellulose precursors: Effect of stabilization conditions, *ACS Sustainable Chem. Eng.* **7** (2019) 8440.  
doi: <https://doi.org/10.1021/acssuschemeng.9b00108>
116. *Dessbesell, L., Paleologou, M., Leitch, M., Pulkki, R., Xu, C.*, Global lignin supply overview and kraft lignin potential as an alternative for petroleum-based polymers, *Renewable Sustainable Energy Rev.* **123** (2020) 109768.  
doi: <https://doi.org/10.1016/j.rser.2020.109768>
117. *Chen, J., Eraghi Kazzaz, A., AlipoorMazandarani, N., Hosseinpour Feizi, Z., Fatehi, P.*, Production of flocculants, adsorbents, and dispersants from lignin, *Molecules* **23** (2018) 868.  
doi: <https://doi.org/10.3390/molecules23040868>
118. *Konduri, M. K., Kong, F., Fatehi P.*, Production of carboxymethylated lignin and its application as a dispersant, *Eur. Polym. J.* **70** (2015) 371.  
doi: <https://doi.org/10.1016/j.eurpolymj.2015.07.028>
119. *Konduri, M. K. R., Fatehi, P.*, Production of water-soluble hardwood kraft lignin via sulfomethylation using formaldehyde and sodium sulfite, *ACS Sustainable Chem. Eng.* **3** (2015) 1172.  
doi: <https://doi.org/10.1021/acssuschemeng.5b00098>
120. *Gao, W., Inwood, J. P. W., Fatehi, P.*, Sulfonation of phenolated kraft lignin to produce water soluble products, *J. Wood Chem. Technol.* **39** (2019) 225.  
doi: <https://doi.org/10.1080/02773813.2019.1565866>
121. *Ghorbani, M., Liebner, F., van Herwijnen, H. W. G., Pfungen, L., Krahofer, M., Budjav, E., Konnerth, J.*, Lignin phenol formaldehyde resoles: The impact of lignin type on adhesive properties, *BioRes.* **11** (2016) 6727.  
doi: <https://doi.org/10.15376/biores.11.3.6727-6741>
122. *Aro, T., Fatehi, P.*, Production and application of lignosulfonates and sulfonated lignin, *Chem. Sus. Chem.* **10** (2017) 1861.  
doi: <https://doi.org/10.1002/cssc.201700082>
123. *Matsushita, Y.*, Conversion of technical lignins to functional materials with retained polymeric properties, *J. Wood Sci.* **61** (2015) 230.  
doi: <https://doi.org/10.1007/s10086-015-1470-2>
124. *Lora, J.*, Chapter 10 – Industrial Commercial Lignins: Sources, Properties and Applications, Belgacem, M.N. and Gandini, A. (Eds.), *Monomers, Polymers and Composites from Renewable Resources*, Elsevier, Amsterdam, 2008, pp 225–241.  
doi: <https://doi.org/10.1016/B978-0-08-045316-3.00010-7>
125. *Belgacem, M. N., Blayo, A., Gandini, A.*, Organosolv lignin as a filler in inks, varnishes and paints, *Ind. Crops Prod.* **18** (2003) 145.  
doi: [https://doi.org/10.1016/S0926-6690\(03\)00042-6](https://doi.org/10.1016/S0926-6690(03)00042-6)
126. *Hodásová, L., Jablonský, M., Škulcová, A., Ház, A.*, Lignin, potential products and their market value, *Wood Res. (Bratislava, Slovakia)* **60** (2015) 973.
127. *Hazwan Hussin, M., Aziz, A. A., Iqbal, A., Ibrahim, M. N. M., Latif, N. H. A.*, Development and characterization novel bio-adhesive for wood using kenaf core (*Hibiscus cannabinus*) lignin and glyoxal, *Int. J. Biol. Macromol.* **122** (2019) 713.  
doi: <https://doi.org/10.1016/j.ijbiomac.2018.11.009>
128. *Sarika, P. R., Nancarrow, P., Khansaheb, A., Ibrahim, T.*, Bio-based alternatives to phenol and formaldehyde for the production of resins, *Polymers* **12** (2020) 2237.  
doi: <https://doi.org/10.3390/polym12102237>
129. *Sameni, J., Jaffer, S. A., Sain, M.*, Thermal and mechanical properties of soda lignin/HDPE blends, *Composites, Part A* **115** (2018) 104.  
doi: <https://doi.org/10.1016/j.compositesa.2018.09.016>
130. *Mohammadabadi, S. I., Javanbakht, V.*, Lignin extraction from barley straw using ultrasound-assisted treatment method for a lignin-based biocomposite preparation with remarkable adsorption capacity for heavy metal, *Int. J. Biol. Macromol.* **164** (2020) 1133.  
doi: <https://doi.org/10.1016/j.ijbiomac.2020.07.074>
131. *Lu, F., Rodriguez-Garcia, J., Van Damme I., Westwood, N. J., Shaw, L., Robinson, J. S., Warren, G., Chatzifragkou, A., McQueen Mason, S., Gomez, L., Faas, L., Balcombe, K., Srinivasan, C., Picchioni, F., Hadley, P., Charalampopoulos, D.*, Valorisation strategies for cocoa pod husk and its fractions, *Curr. Opin. Green Sustain. Chem.* **14** (2018) 80.  
doi: <https://doi.org/10.1016/j.cogsc.2018.07.007>
132. *Akond, A. U. R., Lynam, J. G.*, Deep eutectic solvent extracted lignin from waste biomass: Effects as a plasticizer in cement paste, *Case Stud. Constr. Mater.* **13** (2020) e00460.  
doi: <https://doi.org/10.1016/j.cscm.2020.e00460>
133. *Min, D., Xiang, Z., Liu, J., Jameel, H., Chiang, V., Jin, Y., Chang, H.*, Improved protocol for alkaline nitrobenzene oxidation of woody and non-woody biomass, *Wood Chem. Technol.* **35** (2015) 52.  
doi: <https://doi.org/10.1080/02773813.2014.902965>
134. *Jung, K. A., Woo, S. H., Lim, S.-R., Park, J. M.*, Pyrolytic production of phenolic compounds from the lignin residues of bioethanol processes, *Chem. Eng. J.* **259** (2015) 107.  
doi: <https://doi.org/10.1016/j.cej.2014.07.126>
135. *Xu, Y., Liu, Y., Chen, S., Ni, Y.*, Current overview of carbon fiber: Toward green sustainable raw materials, *BioRes.* **15** (2020) 7234.  
doi: <https://doi.org/10.15376/biores.15.3.Xu>
136. *Louarrat, M., Enaïme, G., Baçaoui, A., Yaacoubi, A., Blin J., Martin, L.*, Optimization of conditions for the preparation of activated carbon from olive stones for application in gold recovery, *J. S. Afr. Inst. Min. Metall.* **119** (2019) 297.  
doi: <http://dx.doi.org/10.17159/2411-9717/2019/v119n3a9>
137. *Ganguly, P., Sengupta, S., Das, P., Bhowal, A.*, Valorization of food waste: Extraction of cellulose, lignin and their application in energy use and water treatment, *Fuel* **280** (2020) 118581.  
doi: <https://doi.org/10.1016/j.fuel.2020.118581>
138. *Fermanelli, C. S., Córdoba, A., Pierella, L. B., Saux, C.*, Pyrolysis and copyrolysis of three lignocellulosic biomass residues from the agro-food industry: A comparative study, *Waste Manag.* **102** (2020) 362.  
doi: <https://doi.org/10.1016/j.wasman.2019.10.057>
139. *Nurika, I., Suhartini, S., Azizah, N., Barker, G. C.*, Extraction of vanillin following bioconversion of rice straw and its optimization by response surface methodology, *Molecules* **25** (2020) 6031.  
doi: <https://doi.org/10.3390/molecules25246031>
140. *Aarabi, A., Mizani, M., Honarvar, M.*, The use of sugar beet pulp lignin for the production of vanillin, *Int. J. Biol. Macromol.* **94**, Part A (2017) 345.  
doi: <https://doi.org/10.1016/j.ijbiomac.2016.10.004>

141. *Kaur, R., Uppal, S. K., Sharma, P.*, Antioxidant and anti-bacterial activities of sugarcane bagasse lignin and chemically modified lignins, *Sugar Tech.* **19** (2017) 675.  
doi: <https://doi.org/10.1007/s12355-017-0513-y>
142. *Luo, K.-H., Zhao, S.-J., Fan, G.-Z., Cheng, Q.-P., Chai, B., Song, G.-S.*, Oxidative conversion of lignin isolated from wheat straw into aromatic compound catalyzed by NaOH/NaAlO<sub>2</sub>, *Food Sci. Nutr.* **8** (2020) 3504.  
doi: <https://doi.org/10.1002/fsn3.1633>
143. *Gil-Chávez, G. J., Prakash Padhi, S. S., Pereira, C. V., Guerreiro, J. N., Matias, A. A., Smirnova, I.*, Cytotoxicity and biological capacity of sulfur-free lignins obtained in novel biorefining process, *Int. J. Biol. Macromol.* **136** (2019) 697.  
doi: <https://doi.org/10.1016/j.ijbiomac.2019.06.021>
144. *Azadfar, M., Haiming Gao, A., Chen, S.*, Structural characterization of lignin: A potential source of antioxidants guaiacol and 4-vinylguaiacol, *Int. J. Biol. Macromol.* **75** (2015) 58.  
doi: <https://doi.org/10.1016/j.ijbiomac.2014.12.049>
145. URL: [https://datam.jrc.ec.europa.eu/datam/mashup/BIOBASED\\_INDUSTRY/index.html](https://datam.jrc.ec.europa.eu/datam/mashup/BIOBASED_INDUSTRY/index.html)