

PSYCHOSES BY ATTACKS FROM SUBVERTED MAST CELLS: A ROLE FOR ARTERIAL INTRAMURAL FLOW BADLY STEERED BY THE NASAL GANGLIA?

Gottfried R. S. Treviranus

Psychiatrische Praxis am Unitobler Campus, Berne, Switzerland

SUMMARY

Mechanisms of cortical psychoses are approached by complementing big data-driven genetics and imaging with a putatively subverted neurovascular “reverse plumbing” by arteries. The “cortical spread” of grey matter loss in schizophrenia and the mid-pericallosal “congestion” in fMRI of periodic catatonia - treatable electromagnetically along arteries - are interpreted in terms of the fastest interstitial outflow through the Cerebral IntraMural Reverse Arterial Flow-engine (CIMURAF, Treviranus 2018-19) draining “waste” via arterio-adventitial lymphatics to the neck. Such repetitively sliding segments of CIMURAF are wrung downstream by muscles likely steered by the neurovascular pterygopalatine ganglion. At the pericallosal artery, along its ideal long straight segment, this likely happens diverging from the mid-callosum towards the front and the back. In the case of a convergent inversion a mid-callosal clash will result, which is observable in psychoses as a mid-callosal high-flow-spot simultaneously with hyper-perfusions of branches and “backwatering” of pial vessels with reactive waste - till date interpreted psycho-mathematically. CIMURAF might also accelerate the perivascular intrusion of MCs by flushing autocrine signals (of which electro-magnetism moves the dipoles) through a putative periadventitial counter-current. Psychoses plausible occur through tryptase-mediated attacks operated by mast cells against oligodendrocytes’ cytoskeleton (Medic 2009) and probably via complement-4 (Schizophrenia WG, 2014) against neurons. Usually MCs are essential long-lived “orchestrators” of homeostases and immune or barrier defences interacting with nerves, immunocytes, organs, and routes. MCs after somatic programming as to “destination & destiny” (Treviranus 2017a, 6.2., 2018) rapidly intrude also into the brain’s parenchyma, first within the lymphatics and then putatively by crossing-over to extraluminal arterial routes. MCs transverse the BBBs, while macrophages only trespass in “disease” (Faraco et al. 2017). Both can be “subverted” by a list of microbes (and putatively blown up by COVID-19 within walls). Enuresis and MCs’ reactions to clozapine add to the interactive support from (epi-)genetics and imaging.

Key words: psychoses - Intramural Peri-Arterial Drainage - microbial subversion of mast cells - Covid-19 - clozapine and enuresis

* * * * *

INTRODUCTION

“This is the great error of our day (...): the physicians separate the soul from the body.”
Plato

Research on psychoses struggles to reconcile big data with macro-physiological mechanisms (Ruzzo & Geschwind 2016) and thus witnesses a “near complete absence of clearly associated biological changes.” (Dhindsa & Goldstein 2016). Here a convergent “tangible”, yet little tested, theory, involving the most rapid (neurovascular) interstitial fluid outflow within in the cerebral arterial walls (CIMURAF) and its external reverse acceleration of adventitial mast cells (Treviranus 2019a,p) is proposed as a main cause of cortical immuno-vascular dysfunction in the context of “psychoses”. Mast cells (MCs) are prototype neuro-immune partners (Forsythe 2019), and emerge as ubiquitous players (Singh et al. 2016, Daniel 2019), also inside the brain (Neumann 1890, Olsson 1964, Dines & Powell 1997, Skaper et al. 2012, 2018, Gilfillan et al. 2011, Karagkouni et al. 2013, Silver & Curley 2013). MCs guard the lymphatic and blood vessels (Kunder et al. 2012), and the BB-barrier (Theoharides 1996), which they can pass, what macrophages in health cannot (Faraco et al. 2017).

Yet MCs also act as assailants of oligodendrocytes (ODCs; Medic 2009) and probably of neurons. In SCZ ODCs indeed are diminished in layer 3 (BA9) and VI of (BA9) of the PFC by 20-25% and in WM (Vostrikov et al. 2018).

Here some “real-biological” hypotheses are applied to the symmetrical mounting of cortical damage in SCZ (Sun et al. 2009) and the perfusion-related findings in periodic catatonia (perCat; Foucher et al. 2019).

THE INTRAMURAL FLOW OF ARTERIES DRAINING BRAINS’S INTERSTITIUM TO ADVENTITIAL LYMPHATICS

Arteries resurface in neuropsychiatry (Meynert 1867/-68, Reynolds & Trimble 2009, iCross-Disorder Group 2019) and have joined psycho-neuro-immunology (Hanson & Gottesman 2005, Maes et al. 2000) while feeding the cerebral 17-fold performing capillaries (Wilhelm et al. 2016) falling ill in SCZ (Uranova et al. 2010, Delgado-Marín et al. 2019). Brain cells are – without buffering – dependent on vessels for heat and waste removal and the supply of energy, oxygen, water, and nutrition. The interstitial one among the extra-cellular spaces separates brain cells by 1/5 of total brain volume and hereby distances adhesive and charged cells

by around 40 nm so that waste products may escape by diffusion (Kaur et al. 2020).

Arteries' role here is reactivated through a model (Cerebral IntraMURal Reverse Flow; details: Treviranus 2018p, 2019a,p) postulating their pervasive "aortic blueprint" (whereby neuron-driven muscles only switch co-axial tubes; see Figure 1) to realize brain's quickest "Intramural PeriArterial Drainage (IPAD)" (Albargothy, 2018) of interstitial fluid - between sliding peri-muscular basement membranes. Besides histo-topological correspondences (Figure 1, 2) CIMURAF added the energetic drive from vascular smooth muscles (VSMCs) – now a part of IPAD; Aldea et al. 2019). Atrophy by being correlated in psychoses with the rarely far, but usual "one-hop" *dys*-functional connectivity support atrophy (somehow) to be driven by the latter (Shafiei 2019) – or by another cause: While the "epicenter" here points to the cingulate, where both salience and DM-networks border, it hereby also points to the long, straight pericallosal artery (pCallA), where CIMURAF should deploy accelerations. Conversely at the subgenual u-turn of the pCallA depressant MCs could "fly off the adventitial track», e.g. in depressant mastocytosis (Boddaert et al. 2017). Here the neuronal massacre of depression occurs (Drevets et al. 2008), nullifying ECT (Liu et al. 2015, Qiu et al. 2018).

The VSMCs only (repetitively) twist membranous tubes co-axially in a segment which thereby is moved *up* stream (!) under neural command, hereby simultaneously creating and undoing pairs of slight hyperboloid "candy-cracker" stenoses (interiorly constraining the arterial wall enough to move fluid upstream within a sliding closed segmented "ring" loaded with "waste"-clearing fluid to move between the membranes wrapping the VSMCs, Figure 2, a segment which nevertheless has radially open shutters towards the perivascular space creating a parallel flow overcoming less resistance). Since such accelerations of the segment (as

in the guts' peristalsis) are thought to be directed by neurovascular commands (e. g. from PPG), intramural (intralaminar) pressure should build up and be relieved by the moving stenoses to an average. Segments moved by contrary commands instead should pressurize a section between them more, and eventually transmit peaks to branches, which thus would be injected by "waste", which could well transport various principles leading e. g. to degranulations from MC-guardians at the BBB. Conversely divergent moves of two segmented "rings" would cause a negatively pressured segment eventually suctioning from the wall of branches.

Vascular, arterial factors at first sight seem to be non-contributing to psychoses (Keshavan & Kaneko 2013) since they occur at similar rates in vascular and Alzheimer's dementia, the latter worsening under blocked drainage (Wang et al. 2019). Yet CADASIL a small vessel and subcortical stroke disorder with enlarged perivascular Virchow-Robin spaces (VRS) with intact BBB (Rajani et al. 2019) tells otherwise, as it seems to be interpretable in terms of CIMURAF: The proposed sliding surfaces between the membranous wrapping of VSMCs of its "aortic blueprint" could act as a molecular grinding mill between these VSMCs (Liu et al. 2017), and such could cause the molecular fragmentation of Notch-3 noted in CADASIL (Young et al. 2019). Notch-molecules replace MCs' canonical c-kit at every step of adhesion (Murata et al. 2019) and vascular remodeling (van Engeland et al. 2019). CIMURAF was the first model to propose a muscular suction driving force for the observed diffusive ECS solute transport - besides not qualified (Asgari et al. 2016) pulsation - and actually inverse trans-astrocyte flow (Treviranus 2019). CIMURAF also questions macro-electromagnetic therapies (Argyelan et al. 2019, McWirther et al. 2015), fluctuations influencing fMRI (Wittaker et al. 2019) or outlasting ECT (Cabral-Calderin et al. 2016, Bächinger et al. 2017), as well as EEG-microstates (da Cruz et al. 2020).

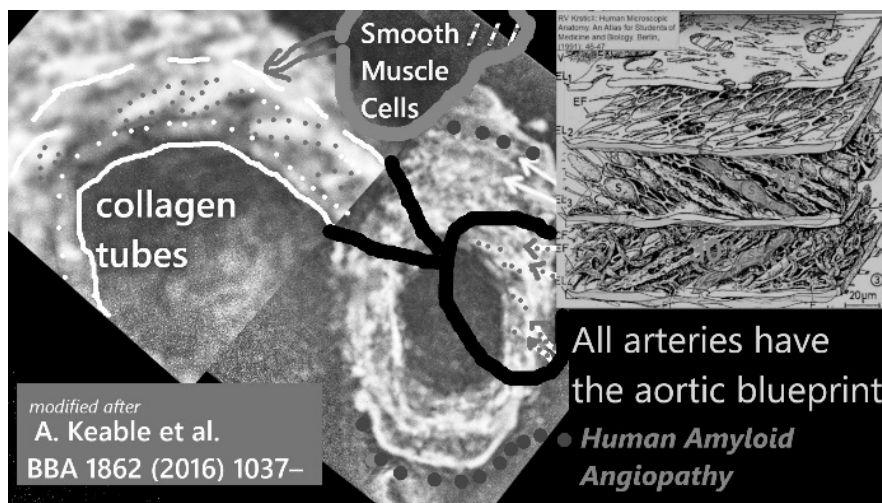


Figure 1. The „pervasive aortic blueprint“ of the CIMURAF-model allows to identify probable obliquely running vascular smooth muscle cells from transverse histological cuts of cerebral arteries attained by Human Cerebral Amyloid Angiopathy aggregating A β flowing between the basement membrane wrappings of the VSMCs (Keable et al. 2016). RUQ: Krstić 1984

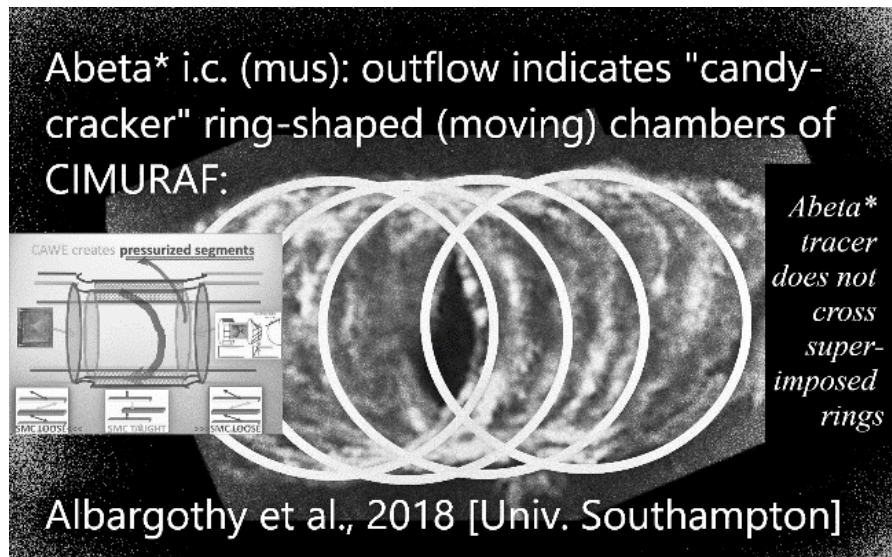


Figure 2. The image from the leading intramural IPAD-model (Albargothy et al. 2018; Fig 1e; CCBY) “shows A β tracer (...) within the wall of the artery in a spiral or ladder-type distribution” - which by superimposing white rings seems more compatible with *separate rings of tracer* as CIMURAF would predict them to be created by sliding twisted segments (see insert)

THE PERICALLOSAL ARTERY & INTRAMURAL ARTERIAL OUTFLOW

The pCallA-complex next to (peri-)callosal regions supplies the anterior 2/3 of the medial and superomedial hemispherical surfaces. The corpus callosum (CCALL) in front is supplied by the pCallA, alias segments A2 to A5 of the anterior cerebral artery (ACA). While in 1/3 the pCallA ends on the precuneus, in nearly 2/3 the frontal supply is continuous with branches from the posterior cerebral artery (PCA) via the outer perisplenial circle (and an inner plexus cooling the fibres) fed in 88% by the occipital medial artery (Blaauw & Meiners 2020).

The callosal drainage occurs via the internal cerebral veins (Kahilogullari et al. 2008, Wolfram-Gabel & Maillot 1992). The centerfold pCallA's networks branch out laterally (Kakou et al. 2000) providing a topological alternative to psycho-mathematics.

Subregions like the subgenual one of the anterior cingulate cortex (sgACC) (Touroutoglou & Dickerson, 2019) constitute driving hubs of functional macroconnectivity (FC). The posterior CC (pCC) provides internally-directed cognition within the DMN, while its dorsal part tunes the brain's metastability and focus (Leech & Sharp 2014).

The pCallA supplies the inter-hemispheric linker CCALL (Roland et al. 2017) and the gyrus cinguli related to effort and need (Heilbronner & Hayden 2016) averaging rewards (Aberg et al. 2020). In SCZ the WM of the CCALL shows early and lasting (cognition-hampering; Ohoshi et al. 2019) micro- and only late, posterior macro-losses (Madigand et al. 2019) - as if by an early (maybe arterial) hit followed by progression (Thompson et al. 2009).

The spectrum towards schizophrenia

A cortical “spread of psychosis” emerges from longitudinal imaging of schizophrenia (SCZ; Velakoulis et al. 2006, Bellanis et al. 2010, Pantelis et al. 2003, 2005, Dietsche et al. 2017) through at times revealing sagittally shifted windows of neuro-progression (Lewandowski et al. 2020).

Besides cortical processes, such of the striatum are key to prodromata and SCZ (Hubl et al. 2017, McCutcheon et al. 2019), while still adding the L-sided hyperperfusion of sensorimotor cortex to that of the putamen (Foucher et al. 2018).

Not parkinsonism or dyskinesia, but other neuro-motor anomalies like (maybe only arterially explainable) “soft signs” (NSS) are prevalent in SCZ and in homogenous proportion among patient's relatives (sparing complex fine movements; Schäppi et al. 2018) – whilst predicting grey matter-losses (GM) in FEP. These unexplained symptoms fluctuate independently (Bachmann & Schröder 2018). NSS do not correlate with cortico-subcortical disbalance (Schröder et al. 1998), but with schizotypal traits (Galindo et al. 2016), whereby these manifest clearer cingulate than subcortical changes (Derome et al. 2020a,b).

Periodic catatonia

In up to 12% of SCZ the WKL-classification (Foucher et al. 2020) distinguishes the in 27% familial periodic catatonia (perCAT) as a notably qualitatively disordered motricity with initially insufficient compromise of other systems. PerCAT shows a (in 5/6 also affective) bipolar dimensional, recurrent-progressive course (Cerele d'excellence 2020). Among segregated cortico-subcortico-thalamo-cortical circuits (CSTCs) perCat seems to

spread from those to the lateral (LMA) and supplementary (SMA; Walther et al. 2017) motor areas (Haber & Calzavara 2008), to the one to ACC (destabilizing effort), and earlier to that to OFC (destabilizing mood), yet potential deficits of the caudal cingulate motor area of cingulate cortex (CgCx) match its core kinetic deviations including mutism.

After averaging fMRT-signals during 6 tasks the remnants mirrored the, possibly vascular, “trait” (Foucher et al. 2018) as a high-flow-spot on the mid-medial CgCx with hyperperfusion on the L side in the same coronal segment: of the precentral gyrus, posterior Broca, and supplementary motor area (SMA) (Foucher et al. 2020). The Bernese perCat-similar patients replicated this with a slightly more anterior high-flow-spot plus again an orthogonal catatonia-correlated hyperperfusion of both SMA and L ventral premotor focus, “compensating” (Walther et al. 2017) for a disconnected 1st CSTC through ACC or rather 5th through SMA (Numbering backwards on Figure 3, Alexander et al. 1986). These high-flow-spots could result from “clashing” inversions of normally divergent CIMURAFs: both to the mid-callosum (instead towards the front and rear ends).

Patients with perCAT are relieved by a robotized rTMS (Zorn et al. 2012) tracing an individual path (probably) along the MCA-branches (communicating over the top with those of the pCallA; Ugur 2005). On fMRT/PET-overlap images during Alzheimer’s hallucinations less perfusion and tissue of the R anterior insula (as from a distal MCA-superior division disorder) again correlate with the L mid-cingulate gyrus (Blanc et al. 2014).

The intriguing link between enuresis and severe mental disorder

Cerebral micturition control (Griffiths 2015) refers to storage and voiding (Harvie et al. 2019), and the thalamus, active in both, relays bladder sensations from the brainstem’s PAG towards the prefrontal decider to void (Arya & Weissbart 2017): in childhood enuresis this path is disconnected on the L. Equally the L medial orbital superior frontal gyrus, in front of the knee of the CCALL – a hub amidst all other players like the CgCx above the “knee” - was less active (Zhu et al. 2019).

The rate of (strongly hereditary) childhood enuresis in SCZ (as noted since Kraepelin) is twice that in siblings and thrice the basic prevalence (21% vs. 11% vs. 7%), while implicating an (early spreaded) prolongation of prodromata by 2 years, and worse cognition. While cognition suffered only in the fluency tests, late micturition control correlated *per se* with less GM. In SCZ this was most significant at a spot on the R superior frontal gyrus. In controls likely GM scars persist and a strong urge comes with a broad activation of the PFC manifesting a zone of GM loss again from the medial PFC down to the sgACC (Hyde et al. 2008). Late onset of enuresis in SCZ (36%) has been twice that in BPAD and predicted negative symptoms (Hollis 2003).

All these areas (besides some lesser temporal and occipital ones) could be reached by a toxic intrusion along the pCallA and to the left MCA (since it supplies the routes of bladder interoception signals through ACC and insula). This unique developmental and also more actual and variegated “intoxication” could stem from adventitially intruding MCs, if they would ascend within the carotid sheath from the crossing of the lymphatic thoracic duct with the aortic arch – assuming that ascending lymphatics at that level would open up to expel the fluid column in order to hasten transport of epitopes to lymph nodes from some bladder or nail infection. Acute episodes of SCZ in non-adults do present with 20% UTIs versus 13% in MDD (e. g. Carson et al. 2017), and bladder infections can be latent (Gilbert et al. 2019).

AN IMMUNO-LYMPHO-ARTERIAL FRAMEWORK OF CONCATENATED HYPOTHESES

The overall framework used evolves as follows: **A.** The perfusion patterns deranged in psychoses, as referred to the sagittal supply of the brain branching off laterally over the cortex from the pericallosal artery (pCallA), **B.** are put into relation with the intramural reverse CIMURAF maximally generated within the straight pCallA. **C.** The pseudo-peristaltic neurovascular steering by the pterygo-palatine ganglion (PPG) instructs, **D.** the CIMURAF to wring out - by “candy-cracker twists” - interstitial fluid towards arterio-adventitial lymph vessels (Drosz 2008, Xu 2007, Bocharov 1968), while the meningo-lymphatics reach the same carotid sheath (jugular) deep cervical lymph nodes along veins (Aspelund et al. 2015, Louveau et al. 2018, Pal et al. 2020, Absinta 2017). **E.** A putative adventitial perivascular flow parallel to CIMURAF (by open radial shutters of such sliding wrung segments) would trigger known counter-flow movements of immunocytes like MCs (by flushing all but rear autocrine signals; Treviranus 2019b,p). **F.** Long-lived MCs - after chemico-physical programming (Csaba 2014) at bodily barriers as to “destination & destiny” (6.2, Treviranus 2017b) – reach the brain via lympho-adventitio-arterial routes. **G.** The MCs’ “orchestration” of also cerebral agendas for metabolic, morphological, “Freudian” sexual, reproductive, stress-, and defense responses is often hidden - as in acupuncture (Jung et al. 2017). **H.** MCs modulate the complex double-layer-dural immune responses (Coles et al. 2017) and move along arteries, lymphatics, and calvarial channels (Cai et al. 2019). **I.** MCs invade the thalamus balancing sleep and the «cognitive complexity divide» (Treviranus 2017, 2018, Morrow & Fitzpatrick 2017). **J.** MCs irritate the cortical surfaces escalating “epileptoid” symptoms towards psychosis (Blumer et al. 2004, Bob et al. 2006) and, **K.** after descending via the arachnoid trabeculae (Engelhardt & Marchetti 2020). **L.** by their proteases (Douahier et al. 2014) attack

oligodendrocytes (ODCs) and probably also neurons coated with complement-4 (C4), as reflected by genetics (Schizophrenia WG, 2014). **M.** MCs as first defendants against unknown germs at barriers and nail folds (Ito et al. 2015) are targeted by their “subversions”.

Mast cells: the reason for the might of clozapine?

While Clozapine (CLZ) is uniquely effective at least in early SCZ and perCAT (Lin et al. 2016), e. g. by dampening thalamo-cortical glutamatergic transmission (Fukuyama et al. 2019), therapeutic consumers are threatened by unwanted side-effects mostly from its meta-

bolites (which can be modulated e.g. by fluvoxamine; Polcwiartek & Nielsen 2016, Lu et al. 2018): weight gain, enuresis and obstipation are common (Every-Palmer et al. 2017, Luche & Francois 2020), and myocarditis or hematological can be rapidly lethal (Leung et al. 2019, De Berardis et al. 2018). Unfortunately many attempts to work around these costs through variant molecules have been unfruitful, while its receptor-profile and short stay on the D2-receptor are shared by various competitors (Nucifora et al. 2017). CLZ via the clathrin-mediated endocytosis prevents MC-activation via IL-37 and MrgX2 (Murakami et al. 2018) - and cell entry by COVID-19 - and inhibits the focal adhesion protein paxillin mediating MCs' cytoskeleton responses

Table 1. Subversive (self-serving) manipulations of mast cells by pathogens

Pathogen	Mechanism	Source (et al.)
Dengue virus	IgG-enhanced uptake	Rathore & St John 2020
COVID-19 ?	IL-1-excess, HA, proteases	Kritas 2020
COVID-19 ?	? Cytokine storm	Kunder 2011
COVID-19 ?	Pulmonary endothelitis	Theoharides 2020
COVID-19 ?	MC sustain B-cell function	Palm 2016
COVID-19 – NO RISK?	MCAS, mastocytosis	Valent 2020
MIS-C/Takotsubo	MC-Etosis (Treviranus 2019)	Riollano-Crus 2020
HIV-1	CXCR4, CCR5, function, death	Taub 2004
HIV-1	Colonize MCs and switch to Tc	Jiang 2015
HHV-8 Kaposi sarcoma	Angiogenic, MC-IgE-reaction	Ayers 2018, Byakwaga 2020
Cytomegalovirus (CMV)	TRAIL-triggers apoptosis	Smith 2013
Listeria monocytogenes	Broad changes	Jobbins 2013
Bartonella spp., Brucella spp.	Genome manipulation, LPS anti TRL4	Ben-Tekaya 2013
Bartonella quintana	LPS: Protects VSMCs from MC via Toll-like receptors	den Dekker 2012 Matera 2008
Salmonella typhimurium	SptP suppresses MCs	Choi 2013
Staphylococcus aureus	Own uptake into MC	Abel 2011
Staphylococcus aureus	Invasion via VEGF, release SP, IL-33	Johnzon 2016
Group B Staphylococcus	Provoked degranulation	Gendrin 2015
Chlamydia pneumoniae	In Mφs metabolic switch	Itoh 2014
Chlamydia pneumoniae	MCs attack beta-cells	Rodriguez 2015
Chlamydia pneumoniae	Metabolic switches	Rodriguez 2015
Mycobacterium tuberc.	Tbc stops extracellular traps	Campillo-Navarro 2018
Mycobacterium tuberc.	Tbc enters MC	Muñoz 2009
Mycobacterium tuberc.	T7S protein, survival in Mφs	Jin 2019
Mycobacterium marinum	Colonizes MCs	Siad 2016
Toxoplasma gondii	Block IgE-response via PLCγ	Smith 2013
Toxoplasma gondii	Adenosine production	Mahamed 2012
Candida albicans	Temporal MC response	Lopes 2015
Candida albicans	Protective uptake into MC	Trevisan 2014
Toxocara canis	Increase of MC number	Carlos 2011
Parasitic worms	ES-62 targets PKC	Bell 2015

MCs are the rapid initial opponents of most microbes – e. g. via extracellular traps (Möllerherm et al. 2016); whereafter – by e.g. potentiating IL-33 (Zhou et al. 2020) – they orchestrate adaptive immune responses (Toniato et al. 2017) not only against worms (Shimokawa et al. 2017) like the top SCZ-associated *Toxocara canis* (Arias et al. 2012, Carlos et al. 2011), as antigen presenters and activators of distant lymph-nodes via cytokine-pellets (Kunder 2009). Thus MCs qualify as prime targets of “subversion” by microbes, which includes ligands of MC receptors, intrusive molecules, colonization and metabolic upset of MCs, and changes to their niches

to IgE-signaling (Han et al. 2015). CLZ is indeed unique among antipsychotics for inhibiting MC-degranulation (Seol et al. 2004), but also dopaminergic modulation of innate immunity (Mackie et al. 2018). The response of SCZ to CLZ follows one of the few polymorphisms: that of protein G-protein subunit-beta 3 (GNB3; Samanaite et al. 2020) which again is involved in various MC-related pathologies and in thermogenesis (Özdemir et al. 2017, Zhang et al. 2019).

In lymphocytes of persons with SCZ epigenetic DNA-methylations (CpG) seem to be both anomalous in certain segments (Teroganova et al. 2016) and specifically changed by CLZ: a) particularly at genes involved in „cell substrate -” and “cell matrix-adhesion” allowing for integrin-centered migration along routes, while such related b) to CREB-binding protein (CREBBP) uniquely (among 29.134 CpG!) seemed to halve symptoms linearly after CLZ (Kinoshita et al. 2017). Only a weak, yet rare support resulted for CREBB from a parallel investigation and for GNB3 (and some transmitter-related genes) from a meta-analysis (Gillespie 2019).

Among countless processes (Pardo et al. 2017) CREBBP constrains B-cell response and MHC-class-II-activation (Jiang et al. 2017). Importantly, since it acetylates the oncogen K-ras (Dixon et al. 2016), it likely inhibits the key MC-activating c-kit-receptor pathways (Khalaf et al. 2007). While kit supports the adhesion and survival of MCs, Notch only supports adhesion (Murata et al. 2014, 2019), the latter being low in SCZ and BPAD (Hoseth et al. 2018).

Both weak effects on CREBB and GNB3 – awaiting replications – also match requirements of the “Destination & Destiny”-hypothesis (Treviranus 2013, 2019), which implies that MCs are programmed to advance along arteries (like kanban-transfer charts along rail-switches) through serial decisions at bifurcations according to a set of adherence molecules “imprinted” at the start to match differential matrix partner molecules leading to cerebral destinations.

Since MCs are likely to modulate CSTCs not only in the 1st medial loop to ACC (Treviranus 2017, 2018), an analogous action of MCs on the 2nd medial loop to the dorsolateral PFC (dlPFC; a functional area subserving 4D-modeling, i.e. Thought) as a possible strong contributor to SCZ could explain the effects of CLZ via MC-stabilization, since it selectively reactivates dlPFC (Samanaite et al. 2020).

Mast cells: How great agents become subverted by microbes?

MCs like macrophages (Féger et al. 2002, Conti et al. 2019) become subversively targeted by pathogens (Table 1) and then - also via the lymphatics (Tsunoda 2017) - do derange the brain (Girolamo et al. 2017, Severance & Yolken 2020, Radua et al. 2018). In Covid-19 persons with already subverted MCs could be most at risk, and a MC-stabilizer is a candidate drug (Theoharides 2020).

CONCLUSIONS AND OUTLOOK

The above imaginable mechanistic steps draw some interactive support and thus are hoped to deploy some traction towards more real-biological psychosis research.

Acknowledgements: None.

Conflict of interest: None to declare.

References

1. Abel J, Goldmann O, Ziegler C, Smeltzer MS, Cheung AL, Bruhn D, et al.: *Staphylococcus aureus evades the extracellular antimicrobial activity of mast cells by promoting its own uptake. J Innate Immun* 2011; 3:495-507
2. Aberg KC, Kramer EE, Schwartz S: *Interplay between midbrain and dorsal anterior cingulate regions arbitrates lingering reward effects on memory encoding. Nat Commun* 2020; 11:1829
3. Absinta M, Ha SK, Nair G, Sati P, Luciano N, Palisoc M et al.: *Human and nonhuman primate meninges harbor lymphatic vessels that can be visualized noninvasively by MRI. Elife* 2017; 6. pii: e29738
4. Afrin LB, Pöhlau D, Raithel M, Haenisch B, Dumoulin FL, Homann J et al.: *Mast cell activation disease: An underappreciated cause of neurologic and psychiatric symptoms and diseases. Brain Behav Immun.* 2015; 50: 314-21
5. Albargothy NJ, Johnston DA, MacGregor-Sharp M, Weller RO, Verma A, Hawkes CA, Carare RO: *Convective influx/lymphatic system: tracers injected into the CSF enter and leave the brain along separate periarterial basement membrane pathways. Acta Neuropathol* 2018; 136:139–52
6. Aldea R, Weller RO, Wilcock DM, Carare RO, Richardson G: *Cerebrovascular smooth muscle cells as the drivers of Intramural Periarterial Drainage of the brain. Front Aging Neurosci* 2019; 11:1
7. Alexander GE, DeLong MR, Strick PL: *Parallel organization of functionally segregated circuits linking basal ganglia and cortex. Annu Rev Neurosci* 1986; 9:357-81
8. Argyelan M, Oltedal L, Deng ZD, Wade B, Bikson M, Joannanne A, et al.: *Electric field causes volumetric changes in the human brain. Elife* 2019; 8:e49115
9. Arias I, Sorlozano A, Villegas E, de Dios Luna J, McKenney K, Cervilla J, et al.: *Infectious agents associated with schizophrenia: a meta-analysis. Schizophr Res* 2012; 136:128-36
10. Asgari M, de Zélicourt D, Kurtcuoglu V: *Glymphatic solute transport does not require bulk flow. Sci Rep* 2016; 6:38635
11. Aspelund A, Antila S, Proulx ST, Karlsen TV, Karaman S, Detmar M, et al.: *A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. J Exp Med* 2015; 212:991-9
12. Arya NG & Weissbart SJ: *Central control of micturition in women: Brain-bladder pathways in continence and urgency urinary incontinence. Clin Anat* 2017; 30:373-84

13. Ayers LW, Barbachano-Guerrero A, McAllister SC, Ritchie JA, Asiago-Reddy E, Bartlett LC, et al.: Mast cell activation and KSHV infection in Kaposi sarcoma. *Clin Cancer Res* 2018; 24:5085-97
14. Bachmann S & Schröder J: Neurological soft signs in schizophrenia: An update on the state- versus trait-perspective. *Front Psychiatry* 2018; 8:272
15. Schizophrenia WG of the Psychiatric Genomics Consortium: Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 2014; 511:421-7
16. Bächinger M, Zerbi V, Moisa M, Polania R, Liu Q, Mantini D, et al.: Concurrent tACS-fMRI reveals causal influence of power synchronized neural activity on resting state fMRI connectivity. *J Neurosci* 2017; 37:4766-77
17. Bell KS, Al-Riyami L, Lumb FE, Britton GJ, Poole AW, Williams CM, et al.: The role of individual protein kinase C isoforms in mouse mast cell function and their targeting by the immunomodulatory parasitic worm product, ES-62. *Immunol Lett* 2015; 168:31-40
18. Bellani M, Dusi N, Brambilla P: Longitudinal imaging studies in schizophrenia: the relationship between brain morphology and outcome measures. *Epidemiol Psychiatr Soc* 2010; 19:207-10
19. Ben-Tekaya H, Gorvel JP, Dehio C: Bartonella and Brucella-weapons and strategies for stealth attack. *Cold Spring Harb Perspect Med* 2013; 3:pil.a010231
20. Blaauw J & Meiners LC: The splenium of the corpus callosum: embryology, anatomy, function and imaging with pathophysiological hypothesis. *Neuroradiol* 2020; 62:563-85
21. Blanc F, Noblet V, Philippi N, Cretin B, Foucher J, Armspach JP, Rousseau F; Alzheimer's Disease Neuroimaging Initiative: Right anterior insula: Core region of hallucinations in cognitive neurodegenerative diseases. *PLoS ONE* 2014; 9:e114774
22. Blumer D, Montouris G, Davies K: The interictal dysphoric disorder: recognition, pathogenesis, and treatment of the major psychiatric disorder of epilepsy. *Epilepsy Behav* 2004; 5:826-40
23. Bob P, Glaslova K, Susta M, Jasova D, Raboch J: Traumatic dissociation, epileptic-like phenomena, and schizophrenia. *Neuro Endocrinol Lett* 2006; 27:321-6
24. Bocharov, V: [Intramural blood and lymphatic vessels of the aorta, inferior caval, portal and hepatic veins in the human]. *Arkh Anat Gistol Embriol* 1968; 54:72-82. [www.medlit.ru/en/]
25. Boddaert N, Salvador A, Chandesris MO, Lemaître H, Grévent D, Gauthier C, et al.: Neuroimaging evidence of brain abnormalities in mastocytosis. *Transl Psychiatry* 2017; 7:e1197
26. Byakwaga H, Barbachano-Guerrero A, Wang D, McAllister S, Naphri K, Laker-Oketta M et al.: Immunoglobulin E (IgE) levels are associated with Kaposi's sarcoma in HIV-infected African adults [online 2020 Jun 19]. *J Infect Dis* 2020; jiaa340
27. Cabral-Calderin Y, Williams KA, Opitz A, Dechent P, Wilke M: Transcranial alternating current stimulation modulates spontaneous low frequency fluctuations as measured with fMRI. *Neuroimage* 2016; 141:88-107
28. Cai R, Pan C, Ghasemigharagoz A, Todorov MI, Förster B, Zhao S et al.: Panoptic imaging of transparent mice reveals whole-body neuronal projections and skull-meninges connections. *Nat Neurosci* 2019; 22:317-27
29. Carlos D, Machado ER, De Paula L, Sá-Nunes A, Sorgi CA, Jamur MC, et al.: Evidence for eosinophil recruitment, leukotriene B4 production and mast cell hyperplasia following *Toxocara canis* infection in rats. *Braz J Med Biol Res* 2011; 44:319-26
30. Carson CM, Phillip N, Miller BJ: Urinary tract infections in children and adolescents with acute psychosis. *Schizophr Res* 2017; 183:36-40
31. Choi HW, Brooking-Dixon R, Neupane S, Lee CJ, Miao EA, Staats HF, Abraham SN: Salmonella typhimurium impedes innate immunity with a mast-cell-suppressing protein tyrosine phosphatase. *Spt P Immunity* 2013; 39:1108-20
32. Coles JA, Stewart-Hutchinson PJ, Myburgh E, Brewer JM: The mouse cortical meninges are the site of immune responses to many different pathogens, and are accessible to intravital imaging. *Methods* 2017; 127:53-61
33. Conti P, Caraffa A, Ronconi G, Frydas I, Theoharides TC: Recent progress on pathophysiology, inflammation and defense mechanism of mast cells against invading microbes: inhibitory effect of IL-37. *Cent Eur J Immunol* 2019; 44:447-54
34. Cross-Disorder Group of the Psychiatric Genomics Consortium: Genomic relationships, novel loci, and pleiotropic mechanisms across eight psychiatric disorders. *Cell* 2019; 179:1469-82.e11
35. Csaba G: Immunoendocrinology: faulty hormonal imprinting in the immune system. *Acta Microbiol Immunol Hung* 2014; 61:89-106
36. da Cruz JR, Favrod O, Roinishvili M, Chkonia E, Brand A, Mohr C, et al.: EEG microstates are a candidate endophenotype for schizophrenia. *Nat Commun* 2020; 11:3089
37. De Berardis D, Rapini G, Olivieri L, Tomasetti C, Valchera A, Fornaro M, et al.: Safety of antipsychotics for the treatment of schizophrenia: a focus on the adverse effects of clozapine. *Ther Adv Drug Saf* 2018; 9:237-56
38. Delgado-Marín L, Basmadjian OM, Occhieppo VB, Marchese NA, Bregonzio C, Baiardi G: Vascular alterations in mental disorders: Focus in angiotensin II role. *Psychiatr Neurosci Update*. Cham ZG: Springer, 2019M p. 10-12
39. den Dekker WK, Tempel D, Bot I, Biessen EA, Joosten LA, Netea MG et al.: Mast cells induce vascular smooth muscle cell apoptosis via a toll-like receptor 4 activation pathway. *Arterioscler Thromb Vasc Biol* 2013; 32:1960-69
40. Derome M, Tonini E, Zöllner D, Schaer M, Eliez S, Debbané M: Developmental trajectories of cortical thickness in relation to schizotypy during adolescence. *Schizophr Bull* 2020a; sbaa020 [online, 2020 Mar 5]
41. Derome M, Zöllner D, Modinos G, Schaer M, Eliez S, Debbané M: Developmental trajectories of subcortical structures in relation to dimensional schizotypy expression along adolescence. *Schizophr Res* 2020b; 218: 76-84
42. Dhindsa, R. & Goldstein, D: Schizophrenia: From genetics to physiology at last. *Nature* 2016; 530:162-3
43. Dietsche B, Kircher T, Falkenberg I: Structural brain changes in schizophrenia at different stages of the illness: A selective review of longitudinal magnetic resonance imaging studies. *Aust N Z J Psychiatr* 2017; 51:500-8
44. Dines KC & Powell HC: Mast cell interactions with the nervous system: relationship to mechanisms of disease. *J Neuropathol Exp Neurol* 1997; 56: 627-40

45. Dixon ZA, Nicholson L, Zeppetbauer M, Matheson E, Sinclair P, Harrison CJ, Irving JA: CREBBP knockdown enhances RAS/RAF/MEK/ERK signaling in Ras pathway mutated acute lymphoblastic leukemia but does not modulate chemotherapeutic response. *Haematologica* 2017; 102:736-45
46. Douaiher J, Succar J, Lancerotto L, Gurish MF, Orgill DP, Hamilton MJ et al.: Development of mast cells and importance of their tryptase and chymase serine proteases in inflammation and wound healing. *Adv Immunol* 2014; 122:211-52
47. Drevets WC, Savitz J, Trimble M: The subgenual anterior cingulate cortex in mood disorders. *CNS Spectr* 2008; 13:663-81
48. Drozd K, Janczak D, Dziegiel P, Podhorska M, Patrzalek D, Ziolkowski P, et al.: Adventitial lymphatics of internal carotid artery in healthy and atherosclerotic vessels. *Folia Histochem Cytobiol* 2008; 46:433-6
49. Engelhardt B & Marchetti L: Immune cell trafficking across the blood-brain barrier in the absence and presence of neuroinflammation. *Vascular Biology* 2020; 2:H1-H18
50. Every-Palmer S, Inns SJ, Grant E, Ellis PM: Effects of Clozapine on the gut: cross-sectional study of delayed gastric emptying and small and large intestinal dysmotility. *CNS Drugs* 2019; 33:81-91
51. Faraco G, Park L, Anrather J, Iadecola C: Brain perivascular macrophages: characterization and functional roles in health and disease. *J Mol Med (Berl)* 2017; 95:1143-52
52. Féger F, Varadaradjalou S, Gao Z, Abraham SN, Arock M: The role of mast cells in host defense and their subversion by bacterial pathogens. *Trends Immunol* 2002; 23:151-8
53. Fitzpatrick CJ & Morrow JD: Thalamic mast cell activity is associated with sign-tracking behavior in rats. *Brain Behav Immun* 2017; 65:222-9
54. Forsythe P: Mast cells in neuroimmune interactions. *Trends Neurosci* 2019; 42:43-55
55. Foucher JR, de Billy C, Jeanjean LC, Obrecht A, Mainberger O, Clauss JME, et al.: A brain imaging-based diagnostic biomarker for periodic catatonia: Preliminary evidence using a Bayesian approach. *Neuropsychobiol* 2020; 79:352-65
56. Foucher JR, Gawlik M, Roth JN, de Crespín de Billy C, Jeanjean LC, Obrecht A, et al.: Wernicke-Kleist-Leonhard phenotypes of endogenous psychoses: a review of their validity. *Dialogues Clin Neurosci* 2020; 22:37-49
57. Foucher JR, Zhang YF, Roser M, Lamy J, De Sousa PL, Weibel S, et al.: A double dissociation between two psychotic phenotypes: Periodic catatonia and cataplasia. *Prog Neuro-Psychopharmacol Biol* 2018; 86:363-69
58. Fukuyama K, Kato R, Murata M, Shiroyama T, Okada M: Clozapine normalizes a glutamatergic transmission abnormality induced by an impaired NMDA receptor in the thalamocortical pathway via the activation of a group III metabotropic glutamate receptor. *Biomolecules* 2019; 9:234
59. Galindo L, Pastoriza F, Bergé D, Mané A, Picado M, Bulbena A, Robledo P, et al.: Association between neurological soft signs, temperament and character in patients with schizophrenia and non-psychotic relatives. *PeerJ* 2016; 4:e1651
60. Gendrin C, Vornhagen J, Ngo L, Whidbey C, Boldenow E, Santana-Ufret V, et al.: Mast cell degranulation by a hemolytic lipid toxin decreases GBS colonization and infection. *Sci Adv* 2015; 1:e1400225
61. Gilbert NM & Lewis AL: Covert pathogenesis: Transient exposures to microbes as triggers of disease. *PLoS Pathog* 2019; 15:e1007586
62. Gilfillan AM, Austin SJ, Metcalfe DD: Mast cell biology: introduction and overview. *Adv Exp Med Biol* 2011; 716:2-12
63. Gillespie AL: Trajectories of DNA methylation associated with clozapine exposure and clinical outcomes. Kings College London, thesis, 2019 [kclpure.kcl.ac.uk]
64. Girolamo F, Coppola C, Ribatti D: Immunoregulatory effect of mast cells influenced by microbes in neurodegenerative diseases. *Brain Behav Immun* 2017; 65:68-89
65. Griffiths D: Neural control of micturition in humans: A working model. *Nature Reviews Urology* 2015; 12
66. Haber SN & Calzavara R: The cortico-basal ganglia integrative network: the role of the thalamus. *Brain Res Bull* 2009; 78:69-74
67. Han SY, Choi YJ, Kang MK, Park JH, Kang YH: Resveratrol suppresses cytokine production linked to FcεRI-MAPK activation in IgE-antigen complex-exposed basophilic mast cells and mice. *Am J Chin Med* 2015; 43:1605-23
68. Hanson DR & Gottesman II: Theories of schizophrenia: a genetic-inflammatory-vascular synthesis. *BMC Med Genet* 2005; 6:7
69. Harvie C, Weissbart SJ, Kadam-Halani P, Rao H, Arya LA: Brain activation during the voiding phase of micturition in healthy adults: A meta-analysis of neuroimaging studies. *Clin Anat* 2019; 32:13-9
70. Heilbronner SR & Hayden BY: Dorsal anterior cingulate cortex: A bottom-up view. *Ann Rev Neurosci* 2016; 39:149-170
71. Hollis C: Developmental precursors of child- and adolescent-onset schizophrenia and affective psychoses: diagnostic specificity and continuity with symptom dimensions. *Br J Psychiatry* 2003; 182:37-44
72. Hoeselt EZ, Krull F, Dieset I, Mørch RH, Hope S, Gardsjord ES, et al.: Attenuated Notch signaling in schizophrenia and bipolar disorder. *Sci Rep* 2018; 8:5349
73. Hubl D, Schultze-Lutter F, Hauf M, Dierks T, Federspiel A, Kaess M, et al.: Striatal cerebral blood flow, executive functioning, and fronto-striatal functional connectivity in clinical high risk for psychosis. *Schizophr Res* 2018; 201:231-6
74. Hyde TM, Deep-Soboslay A, Iglesias B, Callicott JH, Gold JM, Meyer-Lindenberg A, et al.: Enuresis as a premorbid developmental marker of schizophrenia. *Brain* 2008; 131(Pt 9):2489-98
75. Ito T, Ito N, Saathoff M, Stampachiachiere B, Bettermann A, Bulfone-Paus S, et al.: Immunology of the human nail apparatus: the nail matrix is a site of relative immune privilege. *J Invest Dermatol* 2005; 125:1139-48
76. Itoh R, Murakami I, Chou B, Ishii K, Soejima T, Suzuki T, et al.: Chlamydia pneumoniae harness host NLRP3 inflammasome-mediated caspase-activation for optimal intracellular growth in murine macrophages. *Biochem Biophys Res Commun* 2014; 452:689-94
77. Jiang AP, Jiang JF, Wei JF, Guo MG, Qin Y, Guo Q et al.: Human mucosal mast cells capture HIV-and mediate viral trans-infection of CD4+ T cells. *J Virol* 2015; 90:2928-37

78. Jin C, Wu X, Dong C, Li F, Fan L, Xiong S, Dong Y: *EspR promotes mycobacteria survival in macrophages by inhibiting MyD88 mediated inflammation and apoptosis. Tuberculosis (Edinb)* 2019; 116:22-31
79. Jobbings CE, Sandig H, Whittingham-Dowd JK, Roberts IS, Bulfone-Paus S: *Listeria monocytogenes alters mast cell phenotype, mediator and osteopontin secretion in a listeriolyisin-dependent manner. PLoS One* 2013; 8:e57102
80. Johnson CF, Rönnberg E, Guss B, Pejler G: *Live Staphylococcus aureus induces expression and release of vascular endothelial growth factor in terminally differentiated mouse mast cells. Front Immunol* 2016; 7:247
81. Jung SJ, Song H, Kim YY, Kim J, Kim S, Song YK, et al.: *Distribution of mast cells and locations, depths, and sizes of the putative acupoints CV 8 and KI 16. Evid Based Complement Alternat Med* 2017; 2017:2953278
82. Kahilogullari G, Comert A, Arslan M, Esmer AF, Tuccar E, Elhan A, et al.: *Callosal branches of the anterior cerebral artery: an anatomical report. Clin Anat* 2008; 21:383-8
83. Kakou M, Destrieux C, Velut S: *Microanatomy of the pericallosal arterial complex. J Neurosurg* 2000; 93:667-75
84. Karagkouni A, Alevizos M, Theoharides TC: *Effect of stress on brain inflammation and multiple sclerosis. Autoimm Rev* 2013; 12:947-53
85. Kaur J, Davoodi-Bojd E, Fahmy LM, et al.: *Magnetic resonance imaging and modeling of the glymphatic system. Diagnostics (Basel)* 2020; 10:344
86. Keable A, Fenna K, Yuen HM, Johnston DA, Smyth NR, Smith C, et al.: *Deposition of amyloid β in the walls of human leptomeningeal arteries in relation to perivascular drainage pathways in cerebral amyloid angiopathy. BBA* 2016; 1862:1037-46
87. Keshavan MS & Kaneko Y: *Secondary psychoses: an update. World Psychiatry* 2013; 12:4-15
88. Khalaf WF, Yang FC, Chen S, White H, Bessler W, Ingram DA, et al.: *K-ras is critical for modulating multiple c-kit-mediated cellular functions in wild-type and Nf1 +/- mast cells. J Immunol* 2007; 178:2527-34
89. Kritas SK, Ronconi G, Caraffa A, Gallenga CE, Ross R, Conti P: *Mast cells contribute to coronavirus-induced inflammation: new anti-inflammatory strategy. J Biol Regul Homeost Agents* 2020; 34
90. Krstić RV: *Illustrated Encyclopedia of Human Histology. Heidelberg: J. Springer, 1984*
91. Kunder CA, St John AL, Abraham SN: *Mast cell modulation of the vascular and lymphatic endothelium. Blood* 2011; 118:5383-93
92. Kunder CA, St John AL, Li G, Leong KW, Berwin B, Staats HF, Abraham SN: *Mast cell-derived particles deliver peripheral signals to remote lymph nodes. J Exp Med* 2009; 206:2455-67
93. Leech R & Sharp DJ: *The role of the posterior cingulate cortex in cognition and disease. Brain* 2014; 137:12-32
94. Leung JG, Barreto JN, Thompson CA: *Lymphoma following clozapine exposure: more information needed. Schizophr Res* 2018; 199:420-1
95. Lewandowski KE, Bouix S, Ongur D, Shenton ME: *Neuroprogression across the early course of psychosis: J Psychiatr Brain Sci* 2020; 5:e200002
96. Liu Y, Du L, Li Y, Liu H, Zhao W, Liu D, et al.: *Antidepressant effects of electroconvulsive therapy correlate with subgenual anterior cingulate activity and connectivity in depression. Medicine (Balt.)* 2015; 94:e2033
97. Liu Z, Yago T, Zhang N, et al.: *L-selectin mechanochemistry restricts neutrophil priming in vivo. Nat Commun* 2017; 8:15196
98. Lopes JP, Stylianou M, Nilsson G, Urban CF: *Opportunistic pathogen Candida albicans elicits a temporal response in primary human mast cells. Sci Rep* 2015; 12287
99. Louveau A, Herz J, Alme MN, Salvador AF, Dong MQ, Viar KE, et al.: *CNS lymphatic drainage and neuroinflammation are regulated by meningeal lymphatic vasculature. Nat Neurosci* 2018; 21:1380-91
100. Lu ML, Chen TT, Kuo PH, Hsu CC, Chen CH: *Effects of adjunctive fluvoxamine on metabolic parameters and psychopathology in clozapine-treated patients with schizophrenia: A 12-week, randomized, double-blind, placebo-controlled study. Schizophr Res* 2018; 193:126-33
101. Luche N & Francois D: *Clozapine-induced enuresis: an underrecognized and undertreated problem. Prim Care Compan CNS Disord* 2020; 22
102. Mackie P, Lebowitz J, Saadatpour L, Nickoloff E, Gaskill P, Khoshbouei H: *The dopamine transporter: an unrecognized nexus for dysfunctional peripheral immunity and signaling in Parkinson's disease. Brain Behav Immun* 2018; 70:21-35
103. Madigand J, Tréhout M, Delcroix N, Dollfus S, Leroux E: *Corpus callosum microstructural and macrostructural abnormalities in schizophrenia according to the stage of disease. Psychiatry Res Neuroimaging* 2019; 291:63-70
104. Maes M, Bocchio C, Bignotti S, Battista T, Pioli R, Boin F, et al.: *Effects of atypical antipsychotics on the inflammatory response system in schizophrenic patients resistant to treatment with typical neuroleptics. Eur Neuropsychopharmacol* 2000; 10:119-24
105. Mahamed DA, Mills JH, Egan CE, Denkers EY, Bynoe MS: *CD73-generated adenosine facilitates Toxoplasma gondii differentiation to long-lived tissue cysts in the central nervous system. PNAS USA* 2012; 109:16312-27
106. MAJ: *La catatonie périodique. 2020 [http://www.cercle-d-excellence-psy.org]*
107. Matera G, Liberto M-C, Joosten LAB, Vinci M, Quirino A, Pulicari MC, et al.: *The Janus face of Bartonella quintana recognition by Toll-like receptors (TLRs): a review. European cytokine network* 2008; 19:113-8
108. McCutcheon RA, Abi-Dargham A, Howes OD: *Schizophrenia, dopamine and the striatum: From biology to symptoms. Trends Neurosci* 2019; 42:205-20
109. McWhirter L, Carson A, Stone J: *The body electric: a long view of electrical therapy for functional neurological disorders. Brain* 2015; 138:1113-20
110. Medic N, Lorenzon P, Vita F, Trevisan E, Marchioli A, Soranzo MR, et al.: *Mast cell adhesion induces cytoskeletal modifications and programmed cell death in oligodendrocytes. J Neuroimmunol* 2010; 218:57-66
111. Meyer JM & Stahl SM: *The Clozapine Handbook. Cambridge Univ Pr, 2019*
112. Meynert T: *Der Bau der Großhirnrinde und seine örtlichen Verschiedenheiten, nebst einem pathologisch-anatomischen Korollarium. Vierteljahrsschrift für Psychiatrie* 1867-1868, 1:77-93, 126-170, 198-217; 2:88-113
113. Möllerherm H, von Köckritz-Blickwede M, Branitzki-Heinemann K: *Antimicrobial activity of mast cells: role and relevance of extracellular DNA traps. Front Immunol* 2016; 7:265

114. Muñoz S, Rivas-Santiago B, Enciso JA: *Mycobacterium tuberculosis* entry into mast cells through cholesterol-rich membrane microdomains. *Scand J Immunol* 2009; 70:256-63
115. Murata A, Hikosaka M, Yoshino M, Zhou L, Hayashi SI: Kit-independent mast cell adhesion mediated by Notch. *Int Immunol* 2019; 31:69-79
116. Murata A, Yoshino M, Hikosaka M, Okuyama K, Zhou L, Sakano S, et al.: An evolutionary-conserved function of mammalian notch family members as cell adhesion molecules. *PLoS One* 2014; 9:e108535. [correction 2014; 9:e11581]. *PLoS One* 2014; 9:e108535
117. Neumann J: Über das Vorkommen der sogenannten „Mastzellen“ bei pathologischen Veränderungen des Gehirns. *Virchows Arch Pathol Anat* 1890; 122:378-80
118. Nucifora FC Jr, Mihaljevic M, Lee BJ, Sawa A: Clozapine as a Model for Antipsychotic Development. *Neurotherapeutics* 2017; 14:750-61
119. Özdemir AC, Wynn GM, Vester A, Weitzmann MN, Neigh GN, Srinivasan S, Rudd MK: GNB3 overexpression causes obesity and metabolic syndrome. *PLoS One* 2017; 12:e0188763
120. Ohoshi Y, Takahashi S, Yamada S, Ishida T, Tsuda K, Tsuji T, et al.: Microstructural abnormalities in callosal fibers and their relationship with cognitive function in schizophrenia: A tract-specific analysis study. *Brain Behav* 2019; 9:e01357
121. Olsson Y: Mast cells in the nervous system. *Int Rev Cytol* 1968; 24:27-70
122. Pal S, Nath S, Meiningner CJ, Gashev AA: Emerging roles of mast cells in the regulation of lymphatic immuno-physiology. *Front Immunol* 2020; 11:1234
123. Palm AK, Garcia-Faroldi G, Lundberg M, Pejler G, Kleinau S: Activated mast cells promote differentiation of B cells into effector cells. *Sci Rep* 2016; 6:20531
124. Pantelis C, Velakoulis D, McGorry PD, Wood SJ, Suckling J, Phillips LJ et al.: Neuroanatomical abnormalities before and after onset of psychosis: A cross-sectional and longitudinal MRI comparison. *Lancet* 2003; 361:281-88
125. Pantelis C, Yücel M, Wood SJ, Velakoulis D, Sun D, Berger G, et al.: Structural brain imaging evidence for multiple pathological processes at different stages of brain development in schizophrenia. *Schizophr Bull* 2005; 31:672-96
126. Pardo L, Valor LM, Eraso-Pichot A, Barco A, Golbano A, Hardingham GE, et al.: CREB Regulates Distinct Adaptive Transcriptional Programs in Astrocytes and Neurons. *Sci Rep* 2017; 7:6390
127. Polcwiartek C & Nielsen J: The clinical potentials of adjunctive fluvoxamine to clozapine treatment: a systematic review. *Psychopharmacology (Berl)* 2016; 233:741-50
128. Qiu H, Li X, Luo Q, Li Y, Zhou X, Cao H, et al.: Alterations in patients with major depressive disorder before and after electroconvulsive therapy measured by fractional amplitude of low-frequency fluctuations (fALFF). *J Affect Disord* 2019; 244:92-9
129. Radua J, Ramella-Cravaro V, Ioannidis JPA, Reichenberg A, Phiphophatsanee N, Amir T, et al.: What causes psychosis? An umbrella review of risk and protective factors. *World Psychiatry* 2018; 17:49-66
130. Rajani RM, Ratelade J, Domenga-Denier V, Hase Y, Kalimo H, Kalaria RN & Joutel A: Blood brain barrier leakage is not a consistent feature of white matter lesions in CADASIL. *Acta Neuropathol Commun* 2019; 7:187
131. Rathore AP & St John AL: Protective and pathogenic roles for mast cells during viral infections. *Curr Opin Immunol* 2020; 66:74-81
132. Reynolds EH & Trimble MR: Epilepsy, psychiatry, and neurology. *Epilepsia* 2009; 50(S3):50-5
133. Riollano-Cruz M, Akkoyun E, Briceno-Brito E, Kowalsky S, Posada R, Sordillo EM, et al.: Multisystem inflammatory syndrome in Children (MIS-C) related to COVID-19: A New York City Experience [online 2020 Jun 25]. *J Med Virol* 2020; 10.1002/jmv.26224
134. Rodriguez AR, Plascencia-Villa G, Witt CM, Yu JJ, José-Yacamán M, Chambers JP, et al.: Chlamydia pneumoniae promotes dysfunction of pancreatic beta cells. *Cell Immunol* 2015; 295:83-91
135. Rodríguez-López GM, Soria-Castro R, García-Pérez BE, Campillo-Navarro M, Leyva-Paredes K, Donis-Maturano L, et al.: *Mycobacterium tuberculosis* catalase inhibits the formation of mast cell extracellular traps. *Front Immunol* 2018; 9:1161
136. Roland JL, Snyder AZ, Hacker CD, Mitra A, Shimony JS, Limbrick DD, et al.: On the role of the corpus callosum in interhemispheric functional connectivity in humans. *PNAS USA* 2017; 114:13278-83
137. Ruzo EK & Geschwind DH: Schizophrenia genetics complements its mechanistic understanding. *Nat Neurosci* 2016; 19:523-5
138. Samanaite R, Gillespie A, Sendt KV, McQueen G, MacCabe JH, Egerton A: Biological predictors of clozapine response: a systematic review. *Front Psychiatry* 2018; 9:327
139. Schöppl L, Stegmayer K, Viher PV, Walther S: Distinct associations of motor domains in relatives of schizophrenia patients: Different pathways to motor abnormalities in schizophrenia? *Front Psychiatry* 2018; 9:129
140. Schröder J, Silvestri S, Bubeck B, Karr S, Demisch S, Scherrer S, Geider FJ, et al.: D2 dopamine receptor up-regulation, treatment response, neurological soft signs, and extrapyramidal side effects in schizophrenia: a follow-up study with 123I-iodobenzamide single photon emission computed tomography in the drug-naive state and after neuroleptic treatment. *Biol Psychiatry* 1998; 43:660-5
141. Seol IW, Kuo NY, Kim KM: Effects of dopaminergic drugs on the mast cell degranulation and nitric oxide generation in RAW 264.7 cells. *Arch Pharm Res* 2004; 27:94-8
142. Severance EG & Yolken RH: From infection to the microbiome: An evolving role of microbes in schizophrenia. *Curr Top Behav Neurosci* 2020; 44:67-84
143. Shafiei G, Markello RD, Makowski C, Talpalaru A, Kirschner M, Devenyi GA, et al.: Spatial patterning of tissue volume loss in schizophrenia reflects brain network architecture. *Biol Psychiatry* 2019; pii:S0006-3223(19)31785-8
144. Shimokawa C, Kanaya T, Hachisuka M, Ishiwata K, Hisaeda H, Kurashima Y: Mast cells are crucial for induction of group 2 Innate Lymphoid Cells and clearance of helminth infections. *Immunity* 2017; 46:863-74.e4
145. Siad S, Byrne S, Mukamolova G, Stover C: Intracellular localisation of *Mycobacterium marinum* in mast cells. *World J Immunol* 2016; 6:83-95

146. Silver R & Curley JP: Mast cells on the mind: new insights and opportunities. *Trends Neurosci* 2013; 36:513-21
147. Singh J, Shah R, Singh D: Targeting mast cells: Uncovering prolific therapeutic role in myriad diseases. *Int Immunopharmac* 2016; 40:362-84
148. Skaper SD, Facci L, Zusso M, Giusti P: An inflammation-centric view of neurological disease: Beyond the neuron. *Front Cell Neurosci* 2018; 12:72. [correction 2020; 13:578]
149. Skaper SD, Giusti P, Facci L: Microglia and mast cells: two tracks on the road to neuroinflammation. *FASEB J* 2012; 26:3103-17
150. Smith NL, Abi Abdallah DS, Butcher BA, Denkers EY, Baird B, Holowka D: *Toxoplasma gondii* inhibits mast cell degranulation by suppressing phospholipase C γ -mediated Ca(2+) mobilization. *Front Microbiol* 2013; 4:179
151. Smith W, Tomasec P, Aicheler R, Loewendorf A, Nemčovičová I, Wang EC, et al.: Human cytomegalovirus glycoprotein UL14 targets the TRAIL death receptors to thwart host innate antiviral defenses. *Cell Host Microbe* 2013; 13:324-35
152. Sun D, Stuart GW, Jenkinson M, Wood SJ, McGorry PD, Velakoulis D, et al.: Brain surface contraction mapped in first-episode schizophrenia: a longitudinal magnetic resonance imaging study. *Mol Psychiatry* 2009; 14:976-86
153. Taub DD, Mikovits JA, Nilsson G, Schaffer EM, Key ML, Petrow-Sadowski C, Ruscetti FW: Alterations in mast cell function and survival following in vitro infection with human immunodeficiency viruses-through CXCR4. *Cell Immunol* 2004; 230:65-80
154. Teroganova N, Girshkin L, Suter CM, Green MJ: DNA methylation in peripheral tissue of schizophrenia and bipolar disorder: a systematic review. *BMC Genet* 2016; 17:27
155. Theoharides T: Mast cells: The immune gate to the brain. *Life Sc* 1996; 46:607-17
156. Theoharides T: COVID-19, pulmonary mast cells, cytokine storms, and beneficial actions of luteolin. *BioFactors* 2020; 1-3
157. Thompson PM, Bartzokis G, Hayashi KM, Klunder AD, Lu PH, Edwards N, et al.: HGDH Study Group: Time-lapse mapping of cortical changes in schizophrenia with different treatments. *Cereb Cortex* 2009; 19:1107-23
158. Toniato E, Frydas I, Robuffo I, Ronconi G, Caraffa A, Kritas SK, Conti P: Activation and inhibition of adaptive immune response mediated by mast cells. *JBRHA* 2017; 31:543-8. [https://www.biolifesas.org]
159. Touroutoglou A & Dickerson BC: Cingulate-centered large-scale networks: Normal functions, aging, and neurodegenerative disease. *Handb Clin Neurol* 2019; 166:113-27
160. Treviranus GRS: Thoracic aortic rupture: a new sternal rotation-thoracic buckling theory extends „manubrial pinch“ to explain autopsies after side impact. Use in early care and FE-safety engineering. 13th ECTES, Basle, 2012
161. Treviranus GRS: «Real»-Biological Proposals for new paths to bipolarity, mixity, fibromyalgia, and migraine. Presentation, *Int. Review Bipolar Disorders* 2013
162. Treviranus GRS): Rescue of the appropriative „Thought-Action-Mood“ space: Anatomy and mast cells generate „mixable“ dimensions in language and statistics. *Psychiatr Danub* 2018a; 30(Suppl 7):620-9. [http://www.psychiatria-danubina.com/UserDocsImages/pdf/dnb_vol30_sup7/dnb_vol30_sup7_620.pdf]
163. Treviranus GRS: The Co-axial Arterial Wall Engine (CAWE) for depulsion and Cerebral IntraMural Reverse Arterial Flow via neurogenic sliding compartments and macro-nano-links. Poster. 14th Psychoimmunology Expert Meeting 22.-25.3.18 Neuroinflammation in Psychiatry – from basic research to psychopathology. Günzburg, Schloss Reisensburg, 2018p. [www.biposuisse.ch/Gbg18]
164. Treviranus GRS: Mast cell autocrineity near cerebral arterial wall “reverse glymphatic flow” as prime target of electromagnetic effects. *Psychiatr Danub* 2019a; 31(Suppl 3):357-70. [http://www.psychiatria-danubina.com/UserDocsImages/pdf/dnb_vol31_noSuppl%203/dnb_vol31_noSuppl%203_357.pdf]
165. Treviranus GRS: Mast cell autocrineity near Cerebral IntraMural Arterial Flow as target of electromagnetic effects. [www.biposuisse.ch/ect-x-mc-x-art-19-2]. Poster: 7th Biennial Cambridge Int Conf on Mental Health 5th/6th Sept, 2019p. [www.cmhr-cu.org]
166. Trevisan E, Vita F, Medic N, Soranzo MR, Zabucchi G, Borelli V: Mast cells kill *Candida albicans* in the extracellular environment but spare ingested fungi from death. *Inflammation* 2014; 37:2174-2189
167. Tsunoda I: Lymphatic system and gut microbiota affect immunopathology of neuroinflammatory diseases, including multiple sclerosis, neuromyelitis optica and Alzheimer's disease. *Clin Exp Neuroimmunol* 2017; 8:177-9
168. Ugur HC, Kahilogullari G, Coscarella E, Unlu A, Tekdemir I, Morcos JJ, Elhan A, et al.: Arterial vascularization of primary motor cortex (precentral gyrus). *Surg Neurol* 2005; 64(S2):S48-52
169. Uranova NA, Zimina IS, Vikhreva OV, Krukov NO, Rachmanova VI, Orlovskaya DD: Ultrastructural damage of capillaries in the neocortex in schizophrenia. *World J Biol Psychiatr* 2010; 11:567-78
170. Valent P, Akin C, Bonadonna P, Brockow K, Niedozytko M, Niedozytko B, et al.: Risk and management of patients with mastocytosis and MCAS in the SARS-CoV-2 (COVID-19) pandemic: Expert opinions. *J Allergy Clin Immunol* 2020; S0091-6749(20)30839-3
171. van Engeland NCA, Suarez Rodriguez F, Rivero-Müller A, Ristori T, Duran CL, Stassen OMJA, et al.: Vimentin regulates Notch signaling strength and arterial remodeling in response to hemodynamic stress. *Sci Rep* 2019; 9:12415
172. Velakoulis D, Wood SJ, Wong MT, McGorry PD, Yung A, Phillips L, et al.: Hippocampal and amygdala volumes according to psychosis stage and diagnosis: a magnetic resonance imaging study of chronic schizophrenia, first-episode psychosis, and ultra-high-risk individuals. *Arch Gen Psychiatry* 2006; 63:139-49
173. Vostrikov VM & Uranova NA: Defitsit oligodendrotsitov v lobnoï kore pri shizofrenii. *Zh Nevrol Psikhiatr Im S S Korsakova* 2018; 118:100-3
174. Walther S, Schäppi L, Federspiel A, Bohlhalter S, Wiest R, Strik W, Stegmayer K: Resting-state hyperperfusion of the Supplementary Motor Area in catatonia. *Schizophr Bull* 2017; 43:972-81
175. Walther S, Schäppi L, Federspiel A, Bohlhalter S, Wiest R, Strik W, Stegmayer K: Resting-state hyperperfusion of

- the Supplementary Motor Area in catatonia. *Schizophr Bull* 2017; 43:972-81
176. Wang L, Zhang Y, Zhao Y, Marshall C, Wu T, Xiao M: Deep cervical lymph node ligation aggravates AD-like pathology of APP/PS1 mice. *Brain Pathol* 2019; 29:176-92
177. Whittaker JR, Driver ID, Venzi M, Bright MG, Murphy K: Cerebral autoregulation evidenced by synchronized low frequency oscillations in blood pressure and resting-state fMRI. *Front Neurosci* 2019; 13:433. [correction: 2020; 14:544]
178. Wilhelm I, Nyúl-Tóth Á, Suciú M, Hermenean A, Krizbai IA: Heterogeneity of the blood-brain barrier. *Tissue Barr* 2016; 4:e1143544
179. Wolfram-Gabel R & Maillot C: The venous vascularization of the corpus callosum in man. *Surg Radiol Anat* 1992; 14:17-21
180. Xu X, Lin H, Lv H, Zhang M, Zhang Y: Adventitial lymphatic vessels: an important role in atherosclerosis. *Med Hypotheses* 2007; 69:1238-41
181. Young KZ, Lee SJ, Zhang X, Cartee NMP, Torres M, Keep SG et al.: NOTCH3 is non-enzymatically fragmented in inherited cerebral small-vessel disease. *J Biol Chem* 2020; 295:1960-72
182. Zhang X, Wang X, Yin H, Zhang L, Feng A, Zhang QX, et al.: Functional inactivation of mast cells enhances subcutaneous adipose tissue browning in mice. *Cell Rep* 2019; 28:792-803.e4
183. Zhou Z, Yan F, Liu O: Interleukin (IL)-33: an orchestrator of immunity from host defence to tissue homeostasis. *Clin Transl Immunology* 2020; 9:e1146
184. Zhu W, Che Y, Wang Y, Jia Z, Wan T, Wen J, et al.: Study on neuropathological mechanisms of primary monosymptomatic nocturnal enuresis in children using cerebral resting-state functional magnetic resonance imaging. *Sci Rep* 2019; 9:19141
185. Zorn L, Renaud P, Bayle B, Goffin L, Lebossé C, de Mathelin M, Foucher J: Design and evaluation of a robotic system for transcranial magnetic stimulation. *IEEE Trans Biomed Eng* 2012; 59:805-15

Correspondence:

Gottfried R.S. Treviranus, MD
Psychiatrische Praxis am Unitobler Campus
CH 3012 Berne, Switzerland
E-mail: bipo Suisse@bluewin.ch