

Clinical relevance of natural killer cells in breast cancer

Klinička važnost prirodnoubilačkih stanica u karcinomu dojke

Tamara Gulić¹⁺, Damir Grebić^{2,3+}, Alma Starčević⁴, Manuela Avirović^{3,5}, Petra Valković Zujic^{3,6}, Danijela Veljković Vujaklija⁶, Gordana Blagojević Zagorac^{1*}

Abstract. Breast carcinoma is the most common malignant disease in the female and one of the leading causes of death among women worldwide. Chronic inflammation and immunosuppressive environment achieved by the interaction of tumour and infiltrating immune cells are the hallmarks of cancers, including breast cancer. Natural killer cells are one of innate immune cells in tumour bed that possess great antitumour potential due to their ability to kill tumour cells without prior sensitization. They also play an important role in priming of type I immune responses and are considered to be main source of Interferon- γ . Unfortunately, antitumour activity of natural killer cells is often suppressed by pro-inflammatory cytokines secreted in tumour microenvironment and suppression of NK cells activity often correlates with therapy response and general outcome of patient with breast carcinoma. Different types of breast cancer are associated with different subsets of natural killer cells that possess different immunoregulatory function. Identification of specific tumour associated natural killer cell subset endowed with different functional capabilities might help practitioners in therapy decisions, therapy response monitoring, as well as in predicting overall prognosis of breast cancer patients. The purpose of this review is to give an overview of natural killer cells and their subsets, their role in pathogenesis of breast cancer and to discuss NK cells as potential useful therapy and prognostic tool for breast carcinomas.

Key words: breast neoplasms; immunotherapy; killer cells, natural

Sažetak. Karcinom dojke je maligna bolest u žena i jedan je od vodećih uzroka smrti žena širom svijeta. Kronična upala i imunosupresivni okoliš koji nastaje kao posljedica interakcije tumorskih stanica i infiltrirajućih imunoloških stanica obilježja su karcinoma, uključujući karcinom dojke. Prirodnoubilačke stanice (NK stanice) jedna su od subpopulacija urođenih imunoloških stanica u tumorskom sijelu, koje posjeduju antitumorski potencijal zbog svoje sposobnosti direktnog ubijanja tumorskih stanica bez prethodne senzibilizacije. Također, igraju važnu ulogu u pripremi imunološkog odgovora tipa 1 i smatraju se glavnim izvorom interferona- γ . Nažalost, antitumorska aktivnost NK stanica često je potisnuta proupalnim citokinima koji se izlučuju u tumorskom mikrookolišu, a supresija aktivnosti NK stanica obično korelira s terapijskim odgovorom i općim lošijim ishodom kod bolesnica s karcinomom dojke. Surogati molekularnih podtipova karcinoma dojke povezani su s različitim subpopulacijama NK stanica, koje posjeduju različite imunoregulacijske funkcije. Identifikacija specifične subpopulacije NK stanica pridruženih s tumorom, koje posjeduju različite imunološke funkcije, može pomoći liječnicima u donošenju odluka o terapiji, praćenju terapijskog odgovora, kao i u predviđanju ukupne prognoze pacijenata s karcinomom dojke. Svrha je ovog preglednog članka istaknuti važnost NK stanica i njihovih subpopulacija u patogenezi karcinoma dojke te diskutirati terapijske i prognostičke mogućnosti korištenja NK stanica u osoba oboljelih od karcinoma dojke.

Ključne riječi: imunoterapija; novotvorina dojke; ubilačke stanice, prirodne

* Authors equally contributed to this work

¹ University of Rijeka, Faculty of Medicine, Department of Physiology and Immunology, Rijeka, Croatia

² Clinical Hospital Center Rijeka, Department of General and Oncological Surgery, Rijeka, Croatia

³ University of Rijeka, Faculty of Medicine, Rijeka, Croatia

⁴ Clinical Hospital Center Rijeka, Clinical Institute for Transfusion Medicine, Tissue Typing Laboratory, Rijeka, Croatia

⁵ University of Rijeka, Faculty of Medicine, Department of Pathology, Rijeka, Croatia

⁶ Clinical Hospital Center Rijeka, Department of Radiology, Rijeka, Croatia

***Corresponding author:**

Gordana Blagojević Zagorac, Associated Professor
University of Rijeka, Faculty of Medicine,
Department of Physiology and Immunology
Brace Branchetta 20, 51000 Rijeka, Croatia
E-mail: gordana.blagojevic@uniri.hr

<http://hrcak.srce.hr/medicina>

INTRODUCTION

Breast cancer is the leading cause of death in women in developed countries. Age, gender, positive family history, heredity, hormones, dense breasts, precancerous lesions (atypical hyperplasia), ionizing radiation, physical inactivity and obesity after menopause, as well as longer exposure to estrogen (hormone replacement treatment, early menarche, late meno-

The tumour microenvironment is complex, and immune escape is considered an important hallmark of cancer, largely contributing to tumour progression and metastasis. NK cells serve as the main effector cells toward cancer in innate immunity and have become an important factor in the treatment of cancer.

pause) are considered to be major risk factors for the development of the disease^{1,2}. High incidence and still high mortality make breast cancer world health problem. According to data from the International Agency for Research on Cancer, from 2018, there was about 2.1 million newly diagnosed female breast cancer cases with estimated death rate of 6,6%^{1,2}. Early detection of the disease is crucial because more than 90% of breast cancer patients can be cured if the disease is detected at early stage^{2,3}. Prognostic factors include number of effected armpit lymph nodes, tumour size, hormone receptor status, and degree of tumour differentiation (grade). Breast carcinomas have ductal or lobular morphology and invasive ductal carcinoma is the most common type of breast cancer that presents with a distinct morphology and clinical behaviour in comparison to invasive lobular carcinoma³. Molecular classification differentiates four subtypes of breast cancers based on expression of estrogen and progesterone receptors, human epidermal growth factor receptor 2 (HER2) and Ki-67. They include luminal A (ER⁺, PR⁺, Ki-67 < 20; HER2⁻), luminal B (ER⁺, PR⁺, Ki-67 > 20, HER2^{+/-}), HER2 positive (ER⁻ and PR⁻), and triple negative (ER⁻, PR⁻ and HER2⁻)⁴. Prognosis and treatment strategies are based on this clas-

sification⁵. Luminal A type is associated with best prognosis, while triple negative is the most malignant type associated with poor survival and relapse is common⁶. Multifactorial therapy, including radiation, surgery, and chemotherapy is the treatment of choice nowadays that has shown the highest percentage survival in patients with breast cancer^{4,5}. At the moment surgical approach is the therapy choice for luminal A subtype while neoadjuvant chemotherapy is used for more malignant breast cancers such as HER2-positive and triple negative tumours and can be combined with immunotherapy^{7,8}. Despite the acceptable results achieved by classical therapy procedures, multidrug resistance continues to be a clinical challenge and development of new diagnostic and therapy approaches is necessary^{7,8}. Recently, the focus of research has been shifted from tumour cells itself to tumour microenvironment (TME) that has been shown to be a key triggering factor for the molecular etiopathogenetic mechanisms of tumour development and growth^{9,10}. The TME shows immunosuppressive characteristics that are partly achieved by the action of tumour cells and infiltrating immune cells¹¹. The composition of tumour infiltrating immune cells can vary according to cancer type^{10,11}. Different types of infiltrating immune cells involved in both innate and adaptive immunity have diverse effects on tumour plasticity and behaviour with either protumoural or antitumoural consequences¹¹⁻¹³. The innate immune cells infiltrating the tumour bed are mostly tumour associated macrophages (TAMs) and natural killer (NK) cells which exert essential roles in the development and regulation of tumour growth and dissemination potential^{11,12}. Besides their ability to kill tumour cells without prior sensitization, NK cells are also involved in the priming of type 1 T helper-cell (Th1) responses and are the major source of IFN γ *in vivo*¹³. Presence of NK cells has been correlated with clinical outcomes in some types of solid cancer and NK cells are not found in large numbers in advanced human neoplasms¹⁴⁻¹⁶. This suggests that low NK cell number could be a reason for the escape of metastatic cells from the immune system^{16,17} and frequency of NK

cells may represent a new predictive biomarker of response to chemotherapy in cancer patients¹⁴⁻¹⁷. Even in tumours in which the number of NK cells is normal, their activity is often suppressed by TME. Hence, NK cells, as key effectors of the cytotoxic antitumour responses and strong inducers of the adaptive immune response, represent important target for immunotherapy of different types of tumours, including breast cancer^{16,17}. Great advantage of their use in immunotherapy is the fact that they can be efficiently expanded *in vitro* and used in autologous or allogeneic NK cell-based immunotherapies¹³⁻¹⁵. In this review, we will give an overview on the NK cells, their activation status in breast cancer and we will discuss present and potential use of immunotherapy that is based on natural killer cells.

NK CELLS CHARACTERISTICS

NK cells are the first line of innate immunity defence with both cytotoxicity and cytokine-producing effector functions against invading pathogens and various tumours^{13,18}. Consistent with their function as innate sentinels, NK cells are widespread throughout lymphoid and non-lymphoid tissues. In most tissues, NK cells represent a minor fraction of total lymphocytes (from 2% in mouse spleen to 10% in mouse lung and from 2% to 18% in human peripheral blood). Human NK cell turnover in blood is around 2 weeks^{18,19}. The NK cell phenotype is defined upon their expression of NCAM1 (CD56) and lack of CD3 expression. The role of adhesion molecule CD56 could mediate interactions between NK cells and target cells, whereas the cross-linking of the low-affinity Fc receptor CD16 responsible for antibody-dependent cell cytotoxicity (ADCC), the most powerful way to initiate NK cell-mediated killing^{18,19}.

In humans, NK cells can be divided into CD56^{dim} and CD56^{bright} NK cell subsets, which differ in their homing properties^{20,21}. Around 90% of peripheral blood and spleen NK cells are CD56^{dim}CD16⁺ that expresses perforin. These CD56^{dim} NK cell are cytotoxic and produce IFN- γ upon interaction with tumour cells *in vitro*^{20,21}. In contrast, most NK cells in lymph nodes and tonsils are CD56^{bright}-

CD16⁻ and lack perforin^{20,22}. Classification of NK cell receptors is based on their effects: activating, inhibitory and mixed functions^{20,22}. The anti-tumour activity of NK cells depends on the interplay of activating and inhibiting receptors repertoire which recognize changes in the expression patterns of tumour cells²³⁻²⁵. The major activating receptors expressed on human NK cells include NKG2D receptor, C-type lectin receptors (CD94/NKG2C) and the natural cytotoxicity receptors (NCRs) NKp30, NKp44, NKp46 and NKp80. Notably, NCRs mediate recognition and killing of tumour cell and cells infected with viruses^{25,26}. Namely, the activating receptor NKG2D is associate with setting of the activation threshold for NK cell activation^{27,28}. In contrast, receptors that inhibit NK cell activation are important for self-tolerance²⁹. This group of receptors includes inhibitory killer immunoglobulin-like receptors (KIR), the C-type lectin inhibitory receptor CD94/NKG2A and the nectin/nectin-like binding receptors TIGIT and CD96 suppressing activation signalling processes in NK cells^{29,30}. In addition, NKG2A was revealed to be an important checkpoint controlling NK cell mediating T-cell activation³⁰. This important mechanism plays a crucial role in the immune escape of certain tumours^{31,32}.

Activated NK cells secrete a wide variety of cytokines such as IFN- γ , TNF- α , IL-10, IL-5, and IL-13 and chemokines such as MIP-1, IL-8, and RANTES³¹⁻³³. IFN- γ is one of the most potent effector cytokines secreted by NK cells and has been shown to modulate FasL, and TRAIL expression and activates antitumour immunity³⁴. Signaling through activating receptor NKG2D on NK cell has been shown to promote the release of IFN- γ ^{33,34}. These studies point out that apart from the NK cell cytotoxic function; cytokines secreted by the NK cells also provide a significant boost to their antitumour immunity³⁵. Similarly, the cytokines secreted by other immune cells or stromal cells in the tumour microenvironment can positively or negatively influence the antitumour function of NK cells^{35,36}. Functional deficiencies and phenotypic alterations of the NK cell fraction have been found in patients with autoimmune and malignant diseases¹³.

NK CELLS IN BREAST CANCER

NK cells influence tumour growth and progression either by direct interaction with tumour cells or indirectly true modulation of function of other immune cells in TME^{37, 38}. Although NK cells possess high anti-tumour potential, their activity is often suppressed by TME, resulting in decreased IFN- γ , perforin and granulysin production, decreased expression of activating receptors on surface of NK cells and their decreased killing po-

A future challenge for the implementation of NK cell-based therapy for breast cancer is to better define NK cell populations and to identify respective markers, as well as functional and regulatory pathways in each subgroup, thereby using different therapeutic strategies for the treatment of tumours infiltrated with different NK cells.

tential which is associated with higher rate of tumour recidivism and with poor prognosis in general³⁷⁻³⁹. TME can modulate NK cells activity in several ways, mainly by high production of pro-inflammatory cytokines Transforming growth factor beta (TGF- β), Interleukin 10 (IL-10) and Prostaglandin E2 (PGE2) that are responsible for inhibition properties of NK cells and are abundantly found in TME^{40, 41}. These cytokines are produced by tumour cells, TAMs, tumour associated fibroblasts and dendritic cells (DCs)^{40, 41}. Immunosuppressive influence of TME on NK cells is shown in Figure 1.

Inhibition of NK cells is supported by low concentrations of oxygen, glucose and glutamine in tumour tissue⁴². Several studies confirm that the immune profile of NK cells in TME of solid tumours is characterized by an increased expression of inhibitory receptors (CD94/NKG2A)^{31, 37} and decreased gene expression of cytotoxic mediators³²⁻³⁵. Furthermore, the main groups of activating receptors such as NKG2D receptor and a family of natural killer receptors (NKp30, NKp44, NKp46 and DNAM-1) are decreased in many tumours (such as gastric cancer, colorectal carcinoma, and metastatic melanoma)³⁵⁻³⁷.

In patients with breast carcinoma CD56^{bright} tumour-infiltrating NK cells are more abundant in

tumour microenvironment^{19, 20}. Some research groups showed positive correlation of CD56^{bright} subset frequency in tumour tissue and invasiveness of breast cancers^{19, 21-23}. Accordingly, in the peripheral blood of patients with breast cancer the amount of circulating CD56^{bright} NK cells increased respectively to the grade of malignancy, suggesting tolerogenic properties on the systemic level^{19, 22}. Mamesier et al.³⁸ have demonstrated the importance of tolerogenic NK cells in triple negative breast cancer which result in a weakened cellular immunity. Crucial receptor for the inhibitory signal after recognition of tumour cells and therefore tolerogenic properties of NK cells is NKG2A receptor whose inhibitory activity is described in breast cancer^{21, 31}. Namely, higher NKG2A expression in NK cells infiltrating tumour tissue is associated with more invasive types of breast cancers^{31, 32}. Moreover, activated, mature CD16⁺ NK cells although overexpress activating receptors like NCRs (NKp46, NKp30) and NKG2D have tolerogenic properties when NKG2A expression is high^{23, 32}.

Frazao et al.¹⁹ observed CD16⁺NKG2A⁺ NK cell population that is associated with higher TMS stage and lymph node invasion in patients with ductal carcinoma, subtype luminal A or B. We found increased expression of NKG2A in infiltrating NK cells of triple negative breast cancer in comparison to Luminal A and B (unpublished data). We also observed that the expression of activating receptors NKG2D, NKp44 and NKG2C is decreased in different molecular subtypes of breast cancer (luminal A, luminal B and triple negative) (unpublished data). Another mechanism that tumours may escape NK cell immunosurveillance, beside downregulation of activating and upregulation of inhibitory receptors, is the deviation of the NKG2D receptor/NKG2DL ratio⁴³. Moreover, expression of non-signalling Fas receptor or TRAIL decoy receptors protects tumour cell from FasL (Fas ligand) or TRAIL (tumour necrosis factor-related apoptosis inducing ligand)-mediated apoptosis due to evading NK cell cytotoxicity⁴⁴. Considering that function of NK cells in TME is one of the very important factors that will determine chemotherapy response and overall prognosis of patients with breast carcinoma, restore of their activity could be of choices for anti-tumour therapy.

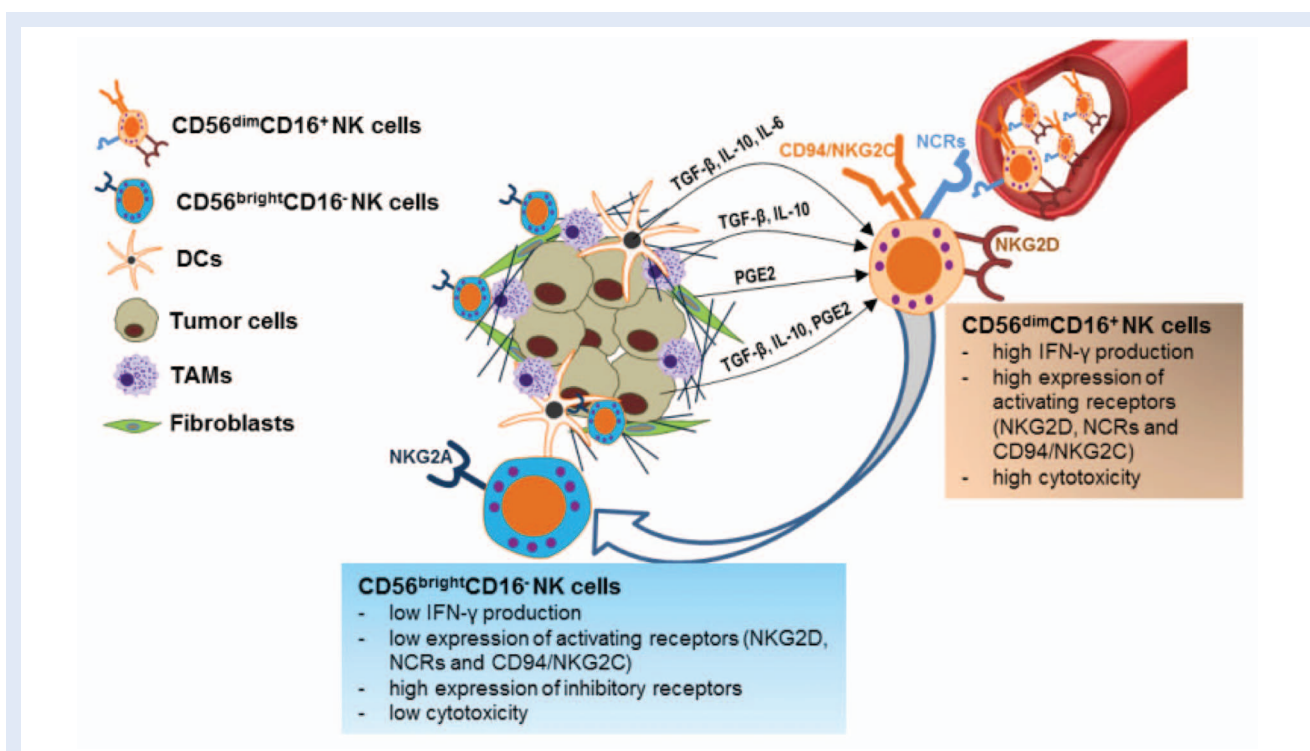


Figure 1. Mechanisms of NK cells suppression in tumour microenvironment.

Most of the NK cells in the blood stream have high cytotoxicity potential, produce high amounts of IFN- γ and expression of activating receptors (CD56^{dim}CD16⁺ NK cells) on their surface is high. In the TME, under the influence of immunosuppressive cytokines, NK cells transform into the cells that possess low cytotoxic potential, produce small amounts of IFN- γ and expression of inhibitory receptors (CD56^{bright}CD16⁻ NK cells) on their surface is high. The most important immunosuppressive cytokines that impair NK cells activity in TME are PGE2 (secreted mostly by fibroblasts), TGF- β and IL-10 (both secreted mostly by DCs, tumour cells and TAMs).

THERAPY POTENTIAL OF NK CELLS

In the last years, harnessing NK cells for the therapeutic purpose is an attractive option and has received rejuvenating interest in recent times⁴⁵. NK cells immunotherapy offers several advantages. First, the use of NK cells will bypass the need of antigen-specific T cells then they can directly kill tumour cells and can also rapidly secrete proinflammatory cytokines that can potentiate the adaptive immune response^{17,45}. In addition, NK cells are easy to isolate, manipulate and to transfer in the patient's body either as autologous (from the same patient) or allogeneic (from other healthy donors) transfer⁴⁵. Despite expectations, transfusion of *ex vivo* activated and expanded autologous NK cells in different solid tumours such as breast cancer, metastatic melanoma, and gastrointestinal cancer patients did not lead to favourable anticancer response⁴⁶. The failure of autologous NK cell adoptive thera-

py to produce positive clinical outcome insisted on the use of allogeneic NK cells. The advantage of use of allogeneic over autologous NK cells is due to inhibition of NK cell-mediated recognition of self-MHC molecules. Unfortunately, early phase of clinical trial using the infusion of activated allogeneic NK cells in ovarian and breast cancer patients did not show a significant expansion of transfused NK cells and therefore did not result in regression of disease⁴⁷. In addition, contradictory results were observed with use of *ex vivo* expanded NK cells expressing chimeric antigen receptors (CAR)⁴⁷. Those results raised the need for new therapy approaches. One of ideas was to use tumour-specific antibodies that will promote ADCC reaction as an effective way to intensify NK cell activity. This approach showed good results in the therapy of metastatic breast cancers^{45,48-50}. Considering that impaired function of NK cells in TME is result of action of secreted cytokines, mainly TGF- β , IL-10 and PEG2, that induce de-

crease in expression of activating and increase of expression of inhibitory receptors on cell surface of NK cells targeting that points is the new step in NK cells-based cancer therapy. And really, use of antagonistic mAbs against inhibitory NK cells receptors in order to promote NK cells cytotoxicity already showed promising results^{45,48}. Similarly, clinical trials in which TGF- β blockade therapy is used in patients with solid tumours demonstrated acceptable safety and preliminary evidence of antitumour activity⁵¹. Two independent studies showed good results of TGF- β blockade when used together with anti-PD-1/PD-L1 therapy on murine EMT6 breast mammary carcinoma and colorectal cancer⁵². At the moment, many companies are conducting clinical trials considering effects of PGE2 receptor (EP2 and EP4) antagonists in treatment of patients with cancers and results are eagerly awaited due to fact that many studies showed that EP2 is abnormally expressed in many cancers, including breast cancer^{53,54} in which it can regulate metastasis⁵⁵.

CONCLUSION

Despite a significant progress that was made over the last two decades in understanding the interrelation and complexity of causal events between a tumour microenvironment and NK cells there are still many questions yet to be answered. NK cell-based immunotherapy of solid tumours posed several challenges but combining NK cell-based immunotherapy with approaches that can target immunosuppressed tumour microenvironment may provide benefits in the treatment of solid tumours, especially those resistant to current T cell-based immunotherapy. NK cell-based immunotherapy is still being intensively tested in clinical trials that include patients with solid cancers. Although, the use of immunotherapy mediated by NK cells is not a standard part of breast cancer treatment yet, and there are many open questions to be answered, there is hope that this will change in the future considering that all data discussed in this review suggest that NK cells have great therapeutic potential and should be considered as a potential useful tool in therapy protocols for the treatment of patients with solid tumours, including breast cancer.

Funding: The investigation was supported by grants from the University of Rijeka (uniri-primed-19-171498; uniri-biomed-18-229-1392; uniri-18.07.2.1.02).

Conflicts of Interest: Authors declare no conflicts of interest.

REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68:394-424.
2. Ghoncheh M, Pournamdar Z, Salehiniya H. Incidence and Mortality and Epidemiology of Breast Cancer in the World. *Asian Pac J Cancer Prev* 2016;17:43-6.
3. Fechner RE, Mills SE. *Breast Pathology*. 1st Edition. Chicago: ASCP Press, 1990;89-106.
4. Sørlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H et al. Gene expression patterns of breast carcinomas distinguish tumour subclasses with clinical implications. *Proc Natl Acad Sci U S A* 2001;98:10869-74.
5. Parker JS, Mullins M, Cheang MC, Leung S, Voduc D, Vickery T et al. Supervised risk predictor of breast cancer based on intrinsic subtypes. *J Clin Oncol* 2009;27:1160-7.
6. Strehl JD, Wachter DL, Fasching PA, Beckmann MW, Hartmann A. Invasive Breast Cancer: Recognition of Molecular Subtypes. *Breast Care* 2011;6:258-264.
7. Ellis MJ. Mutational analysis of breast cancer: guiding personalized treatments. *Breast* 2013;22:19-21.
8. Mohamed A, Krajewski K, Cakar B, MA CX. Targeted therapy for breast cancer. *Am J Pathol* 2013;183:1096-1112.
9. Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumour microenvironment. *Cancer Cell* 2012;21:309-22.
10. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011;144:646-74.
11. Bense RD, Sotiriou C, Piccart-Gebhart MJ, Haanen JBAG, van Vugt MATM, de Vries EGE et al. Relevance of Tumour-Infiltrating Immune Cell Composition and Functionality for Disease Outcome in Breast Cancer. *J Natl Cancer Inst* 2016;109:192.
12. Mantovani A, Marchesi F, Malesci A, Laghi L, Allavena P. Tumour-associated macrophages as treatment targets in oncology. *Nat Rev Clin Oncol* 2017;14:399-416.
13. Stoll G, Zltvogel L, Kroemer G. Differences in the composition of the immune infiltrate in breast cancer, colorectal carcinoma, melanoma and non-small cell lung cancer: a microarray-based meta-analysis. *Oncoimmunology* 2016;5:1067746.
14. Ishigami S, Natsugoe S, Tokuda K, Nakajo A, Che X, Iwashige H et al. Prognostic value of intratumoural natural killer cells in gastric carcinoma. *Cancer* 2000;88:577-83.
15. Villegas FR, Coca S, Villarrubia VG, Jiménez R, Chillón MJ, Jareño J et al. Prognostic significance of tumour infiltrating natural killer cells subset CD57 in patients with squamous cell lung cancer. *Lung Cancer* 2002;35:23-8.

16. Carrega P, Morandi B, Costa R, Frumento G, Forte G, Altavilla G et al. Natural killer cells infiltrating human non-small-cell lung cancer are enriched in CD56 bright CD16(-) cells and display an impaired capability to kill tumour cells. *Cancer* 2008;112:863-75.
17. Sconocchia G, Eppenberger S, Spagnoli GC, Tornillo L, Droeser R, Caratelli S et al. NK cells and T cells cooperate during the clinical course of colorectal cancer. *Oncoimmunology* 2014;3:952197.
18. Salgado R, Denkert C, Demaria S, Sirtaine N, Klauschen F, Pruneri G et al. International TILs Working Group 2014. The evaluation of tumour-infiltrating lymphocytes (TILs) in breast cancer: recommendations by an International TILs Working Group 2014. *Ann Oncol* 2015;26:259-71.
19. Frazao A, Messaoudene M, Nunez N, Dulphy N, Roussin F, Sedlik C et al. CD16⁺NKG2A^{high} Natural Killer Cells Infiltrate Breast Cancer-Draining Lymph Nodes. *Cancer Immunol Res* 2019;7:208-218.
20. Fehniger TA, Cooper MA, Nuovo GJ, Cella M, Facchetti F, Colonna M et al. CD56bright natural killer cells are present in human lymph nodes and are activated by T cell-derived IL-2: a potential new link between adaptive and innate immunity. *Blood* 2003;101:3052-7.
21. Choucair K, Duff JR, Cassidy CS, Albrethsen MT, Kelso JD, Lenhard A et al. Natural killer cells: a review of biology, therapeutic potential and challenges in treatment of solid tumours. *Future Oncol* 2019;15:3053-69.
22. Michel T, Poli A, Cuapio A, Briquemont B, Iserentant G, Ollert M et al. Human CD56bright NK Cells: An Update. *J Immunol* 2016;196:2923-31.
23. Kamiya T, Seow SV, Wong D, Robinson M, Campana D. Blocking expression of inhibitory receptor NKG2A overcomes tumour resistance to NK cells. *J Clin Invest* 2019;129:2094-2106.
24. Wu Y, Tian Z, Wei H. Developmental and Functional Control of Natural Killer Cells by Cytokines. *Front Immunol* 2017;8:930.
25. Jelenčić V, Šestan M, Kavazović I, Lenartić M, Marinović S, Holmes TD et al. **NK cell receptor NKG2D sets activation threshold for the NCR1 receptor early in NK cell development.** *Nat Immunol* 2018;19:1083-92.
26. Stabile H, Fionda C, Gismondi A, Santoni A. Role of Distinct Natural Killer Cell Subsets in Anticancer Response. *Front Immunol* 2017;8:293.
27. Kruse PH, Matta J, Ugolini S, Vivier E. Natural cytotoxicity receptors and their ligands. *Immunol Cell Biol* 2014;92:221-9.
28. Mirjačić Martinović KM, Babović NLJ, Džodić RR, Jurišić VB, Tanić NT, Konjević GM. Decreased expression of NKG2D, NKP46, DNAM-1 receptors, and intracellular perforin and STAT-1 effector molecules in NK cells and their dim and bright subsets in metastatic melanoma patients. *Melanoma Research* 2014;24:295-304.
29. Sternberg-Simon M, Brodin P, Pickman Y, Onfelt B, Kärre K, Malmberg KJ et al. **Natural killer cell inhibitory receptor expression in humans and mice: a closer look.** *Front Immunol* 2013;4:65.
30. Venstrom JM, Pittari G, Gooley TA, Chewning JH, Spellman S, Haagenson M et al. HLA-C-dependent prevention of leukemia relapse by donor activating KIR2DS1. *N Engl J Med* 2012;367:805-16.
31. McWilliams EM, Mele JM, Cheney C, Timmerman EA, Fiazuddin F, Strattan EJ et al. Therapeutic CD94/NKG2A blockade improves natural killer cell dysfunction in chronic lymphocytic leukemia. *Oncoimmunology* 2016;5:1226720.
32. Schleypen JS, Von Geldern M, Weiss EH, Kotzias N, Rohrmann K, Schendel DJ et al. **Renal cell carcinoma-infiltrating natural killer cells express differential repertoires of activating and inhibitory receptors and are inhibited by specific HLA class I allotypes.** *Int J Cancer* 2003;106:905-12.
33. Freeman BE, Raué HP, Hill AB, Slifka MK. Cytokine-Mediated Activation of NK Cells during viral Infection. *J Virol* 2015;89:7922-31.
34. Paul S, Lal G. The Molecular Mechanism of Natural Killer Cells Function and Its Importance in Cancer Immunotherapy. *Front Immunol* 2017;8:1124.
35. Rocca YS, Roberti MP, Arriaga JM, Amat M, Bruno L, Pampena MB et al. Altered phenotype in peripheral blood and tumour-associated NK cells from colorectal cancer patients. *Innate Immun* 2013;19:76-85.
36. Levy EM, Roberti MP, Mordoh J. Natural killer cells in human cancer: from biological functions to clinical applications. *J Biomed Biotechnol* 2011;2011:676198.
37. Han B, Mao FY, Zhao YL, Lv YP, Teng YS, Duan M et al. Altered Nkp30, Nkp46, NKG2D, and DNAM-1 Expression on Circulating NK Cells Is Associated with Tumour Progression in Human Gastric Cancer. *J Immunol Res* 2018;3:6248590.
38. Mamessier E, Sylvain A, Thibult ML, Houvenaeghel G, Jacquemier J, Castellano R et al. Human breast cancer cells enhance self tolerance by promoting evasion from NK cell antitumour immunity. *J Clin Invest*. 2011;121:3609-22.
39. Smyth MJ, Cretney E, Kelly JM, Westwood JA, Street SE, Yagita H et al. **Activation of NK cell cytotoxicity.** *Mol Immunol* 2005;42:501-10.
40. Konjević GM, Vuletić AM, Mirjačić Martinović KM, Larsen AK, Jurišić VB. The role of cytokines in the regulation of NK cells in the tumour environment. *Cytokine* 2019;117:30-40.
41. Stojanovic A, Correia MP, Cerwenka A. Shaping of NK cell responses by the tumour microenvironment. *Cancer Microenviron* 2013;6:135-46.
42. Garner WL, Minton JP, James AG, Hoffmann CC. Human breast cancer and impaired NK cell function. *J Surg Oncol* 1983;24:64-6.
43. Duan S, Guo W, Xu Z, He Y, Liang C, Mo Y et al. Natural killer group 2D receptor and its ligands in cancer immune escape. *Mol Cancer* 2019;18:29.
44. Garofano F, Gonzalez-Carmona MA, Skowasch D, Schmidt-Wolf R, Abramian A, Hauser S et al. **Clinical Trials with Combination of Cytokine-Induced Killer Cells and Dendritic Cells for Cancer Therapy.** *Int J Mol Sci* 2019;20:4307.
45. Hu W, Wang G, Huang D, Sui M, Xu Y. Cancer Immunotherapy Based on Natural Killer Cells: Current Progress and New Opportunities. *Front Immunol* 2019;10:1205.
46. Miller JS, Soignier Y, Panoskaltsis-Mortari A, McNearney SA, Yun GH, Fautsch SK et al. **Successful adoptive transfer and in vivo expansion of human haploidentical NK cells in patients with cancer.** *Blood* 2005;105:3051-7.

47. Geller MA, Cooley S, Judson PL, Ghebre R, Carson LF, Argenta PA et al. A phase II study of allogeneic natural killer cell therapy to treat patients with recurrent ovarian and breast cancer. *Cytotherapy* 2011;13:98-107.
48. Tallero R, Conti L, Lanzardo S, Sottile R, Garofalo C, Wagner AK et al. **NK cells control breast cancer and related cancer stem cell hematological spread.** *Oncoimmunology* 2017;6:1284718.
49. Waks AG, Winer EP. Breast Cancer Treatment: A Review. *JAMA* 2019;321:288-300.
50. Hu W, Wang G, HUANG D, Sui M, Xu Y. Cancer Immunotherapy Based on Natural Killer Cells: Current Progress and New Opportunities. *Front Immunol* 2019;10:1205.
51. Morris JC, Tan AR, Olencki TE, Shapiro GI, Dezube BJ, Reiss M et al. Phase I study of GC1008 (fresolimumab): a human anti-transforming growth factor-beta (TGFβ) monoclonal antibody in patients with advanced malignant melanoma or renal cell carcinoma. *PLoS One* 2014;9:90353.
52. Mariathasan S, Turley SJ, Nickles D, Castiglioni A, Yuen K, Wang Y et al. TGFβ attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. *Nature* 2018;554:544-548.
53. Take Y, Koizumi S, Nagahisa A. Prostaglandin E Receptor 4 Antagonist in Cancer Immunotherapy: Mechanisms of Action. *Front Immunol* 2020;11:324.
54. Ching MM, Reader J, Fulton AM. Eicosanoids in Cancer: Prostaglandin E₂ Receptor 4 in Cancer Therapeutics and Immunotherapy. *Front Pharmacol* 2020;11:819.
55. Cheuk IW, Shin VY, Siu MT, Tsang JY, Ho JC, Chen J et al. Association of EP2 receptor and SLC19A3 in regulating breast cancer metastasis. *Am J Cancer Res* 2015;5:3389-99.