

## The potential of chives, wild garlic, and garlic plant extracts in suppression of bacterial growth

### Abstract

Food spoilage is phenomenon associated with occurrence of the microorganisms in agricultural products. Due to the ability of several microorganisms to colonize plant material, agrochemicals were proposed to suppress bacterial growth. However, overuse of agrochemicals has detrimental impact on environmental quality. Therefore, the objective of this research was to evaluate the potential of aqueous extracts of garlic (*Allium sativum* L.), wild garlic (*Allium ursinum* L.), and chives (*Allium schoenoprasum* L.) to suppress the growth of *Enterococcus faecalis* and *Escherichia coli*. The antimicrobial activity of these plant extracts was determined using the disc diffusion method. Mueller-Hinton agar was inoculated with bacterial inoculum; six filter paper disks, impregnated with 15 µl aqueous extract (1, 2 and 4%), were placed on each agar surface. The Petri dishes were incubated at 37 °C for 24 hours. The diameter of the zone of inhibition was measured and expressed in millimeters (mm). Almost all aqueous extracts showed a negligible effect on the growth of *E. coli* and *E. faecalis*. Significant suppression of bacterial growth compared with other treatments was observed with the aqueous extract of wild garlic at a concentration of 4%. This research confirms the potential of wild garlic extract in suppression of potential pathogens.

**Key words:** antimicrobial activity, chives, garlic, inhibition zone, wild garlic.

### Introduction

Food spoilage is considered one of the most common causes of serious illness in developing countries (Pirbalouti et al., 2009; Sapkota et al., 2012). Gustavsson et al. (2011) estimated that 40% of global food production is lost annually due to various factors (including food spoilage caused by microorganisms). Microorganisms like bacteria, yeasts, and molds, are often involved in food spoilage (Lianou et al., 2016). They have the ability to utilize food products and produce metabolites responsible for food spoilage (Parlapani et al., 2017). Although food spoilage is associated with many Gram-positive bacteria such as *Enterococcus faecalis*, *Staphylococcus aureus*, and *Bacillus cereus* (Braga et al., 2005), most reports on food spoilage mention the presence of Gram-negative bacteria (Pandey and Singh, 2011). Some of the bacteria are associated with foodborne illness and food contamination. Enterococci are Gram-positive bacteria that are commonly found in aquatic and terrestrial environments as well as in plants because of their adaptability to various abiotic conditions (Badgley et al., 2010). Enterococci produce thermostable amines that can lead to poisoning (Giraffa, 2002). *Escherichia coli*, on the other hand, is a Gram-negative bacterium that is often isolated from fresh plant products because it is transmitted through manure-contaminated soil or contaminated irrigation water (Solomon et al., 2002). The possible bacterial plant infections can be controlled using agrochemicals (Shan et al., 2007).

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However, overuse of agrochemicals leads to the accumulation of residues, which then has impact on public health. In order to suppress the growth of pathogens and increase the shelf life of food (Gonelimali et al., 2018), environmentally friendly methods based on the use of natural additives have been developed (Mostafa et al., 2018). One of the methods is based on the use of aqueous or ethanol-based plant extracts from medicinal and aromatic plants, which have antimicrobial (Mathabe et al., 2005) and antioxidant properties (Talib and Mahasneh, 2010).

The genus *Allium* belongs to the *Alliaceae* family (Chehregani et al., 2007) and is characterized by antimicrobial activity against fungi and a variety of bacteria (Whitemore & Naidu, 2000). A variety of antimicrobial compounds have been isolated and determined from the species of this genus (Lanzotti et al., 2013). Although garlic (*Allium sativum* L.) is the most studied species in the genus *Allium* (Lanzotti et al., 2013), other plant species from the same genus: chives and wild garlic, also have antibacterial activity (Lupoae et al., 2013; Sihombing et al., 2018). Considering the reports by Safithri et al. (2011), Stupar et al. (2022), and Ghasemian et al. (2018) that aqueous extracts of *Allium* showed satisfactory effects on bacterial growth.

The objective of this research was to evaluate the potential of aqueous extracts of garlic (*Allium sativum* L.), wild garlic (*Allium ursinum* L.), and chives (*Allium schoenoprasum* L.) to suppress the growth of *Enterococcus faecalis* and *Escherichia coli*.

## Material and methods

Pure cultures of *Escherichia coli* and *Enterococcus faecalis* belonging to the Laboratory of Microbiology (Faculty of Agricultural and Food Sciences Sarajevo, Bosnia, and Herzegovina) were used for this study. Plant material was collected from Vranica mountain (Central Bosnia and Herzegovina) and Novi Travnik municipality (Central Bosnian Canton, Bosnia and Herzegovina). Samples of wild garlic and chives were collected from Mount Vranica, while the plant material of garlic was obtained in Novi Travnik. Shoots and roots of wild garlic and chives, as well as the garlic clove were used for the preparation of the aqueous extracts. The preparation was performed according to the modified method of Norsworthy (2003); after drying and grinding, the plant materials (3, 6, and 12 g of the dried samples) were homogenized in 300 ml of distilled water. Thus, the final concentrations of plant material were 1, 2, and 4 % of dry weight.

The antimicrobial activity of these plant extracts was determined using the disc diffusion method. Mueller-Hinton agar, previously sterilized in an autoclave at 120°C for 20 min, was inoculated with bacterial inoculum adjusted to 0.5 McFarland standard turbidity (i.e., approximately  $1 \times 10^8$  CFU/ml). After 15 minutes of agar plate inoculation, 6 filter paper disks (6 mm diameter) impregnated with 15  $\mu$ l aqueous extract were placed on each agar surface.

The Petri dishes were incubated at 37 °C in an incubator for 24 hours. The diameter of the zone of inhibition was measured and expressed in millimeters (mm). The interpretation of the obtained results was performed according to EUCAST (2017), where the effect of the extracts is classified as resistant (R), susceptible - increased exposure (I), and susceptible - standard dose (S).

After determining the zone of inhibition, statistical significance ( $P < 0.05$ ) between treatments was determined using SPSS software (v. 22, SPSS Inc., Chicago, IL, USA).

## Results and discussion

The results of this study show that the zone of inhibition depends on the bacterial species, the type of plant extracts, and their concentration (Tables 1-4). Tables 1 and 3 show the average diameters of the zone of inhibition of *Escherichia coli* and *Enterococcus faecalis*, respectively.

**Table 1.** The average diameter of the inhibition zone of bacteria *Escherichia coli*  
**Tablica 1.** Prosječni promjer zone inhibicije bakterije *Escherichia coli*

Plant extract	The concentration of aqueous extracts (%)	Inhibition zone diameter (mm)	EUCAST classification
garlic	1	6.0	R
	2	6.0	R
	4	6.0	R
wild garlic	1	6.0	R
	2	6.0	R
	4	7.5	I
chives	1	6.0	R
	2	6.0	R
	4	6.0	R

As shown in Table 1, the diameter of the inhibition zone of *E. coli* was not affected by the aqueous extract of chives and garlic. All concentrations of these extracts resulted in an inhibition zone of 6.0 mm for the same bacteria. Although several studies confirm the potential of aqueous chive extract in suppressing the growth of *E. coli* (Agustin et al., 2022; Sihombing et al., 2018), our results are not consistent with these findings. However, compared with this study, the presented authors used higher concentrations of chives extracts. Similar results were observed by Oyawoye et al. (2022) with garlic extracts. In our study, aqueous *Allium* extracts were used. Shirzadi Karamolah et al. (2017) reported that antimicrobial activity is often determined by the type of plant extracts. Moreover, Oyawoye et al. (2022) showed that ethanol *Allium* plant extracts exhibited a stronger inhibitory effect on the growth of various bacteria compared with aqueous extracts.

**Table 2.** Significance of obtained differences for *E. coli* growth using the Tukey test  
**Tablica 2.** Značajnost dobijenih razlika za *E. coli* primjenom Tukey testa

Concentration (%)	Plant extracts			AIZD (mm) for concentrations
	garlic	wild garlic	chive	
1	6.000	6.000	6.000	6.000a
2	6.000	6.000	6.000	6.000a
4	6.000	7.500	6.000	6.500b
AIZD (mm) for plant extracts	6.000a	6.500b	6.000a	-

AIZD - Average inhibition zone diameter

Application of the highest concentration of the wild garlic extract resulted in a significant increase in the diameter of the *E. coli* inhibition zone compared with the lower

concentrations (Table 2). However, the *E. coli* average inhibition zone at the highest and lowest extract concentrations was 7.5 and 6.0 mm, respectively. The weak antimicrobial potential of wild garlic extracts was previously described by Stupar et al. (2022). Therefore, according to the EUCAST criteria (2017), *E. coli* can be classified as resistant in all treatments except the 4% wild garlic treatment (Table 1).

**Table 3.** The average diameter of the inhibition zone of the bacteria *Enterococcus faecalis*  
**Tablica 3.** Prosječni promjer zone inhibicije bakterije *Enterococcus faecalis*

Plant extract	The concentration of aqueous extracts (%)	Inhibition zone diameter (mm)	EUCAST classification
garlic	1	6.0	R
	2	6.0	R
	4	6.0	R
wild garlic	1	6.0	R
	2	6.0	R
	4	12.0	I
chives	1	6.0	R
	2	6.0	R
	4	6.0	R

**Table 4.** Significance of obtained differences for *E. faecalis* growth using the Tukey test  
**Tablica 4.** Značajnost dobivenih razlika za *E. faecalis* primjenom Tukey testa

Concentration (%)	Plant extracts			AIZD (mm) for concentrations
	garlic	wild garlic	chive	
1	6.000	6.000	6.000	6.000a
2	6.000	6.000	6.000	6.000a
4	6.000	12.000	6.000	8.000b
AIZD (mm) for plant extracts	6.000a	8.000b	6.000a	-

AIZD - Average inhibition zone diameter

From the data presented in Table 3, it can be seen that the aqueous garlic and chives extracts showed no significant effect on suppressing the growth of *E. faecalis*. Although different concentrations of garlic extract showed a reduction in the viability of *E. faecalis*, no significant differences were found between the concentrations used (Octavia et al., 2019). In contrast, Birring et al. (2015) found that garlic extract was able to disrupt *E. faecalis* biofilm formation. Moreover, the best results were obtained with a garlic concentration of 70%, which is a significantly higher concentration than in this study. The significantly highest inhibition zone diameter was observed when 4% garlic extract was used compared with other plant extracts and concentrations (Table 4). Although Stupar et al. (2022) reported the moderate effect of wild garlic extract on the growth of *E. faecalis*, Pavlović et al. (2012) showed that *Allium ursinum* extract exhibited strong antimicrobial performance against the same bacteria. However, previous reports have shown contradictory results

regarding the effects of wild garlic extracts against Gram-positive and Gram-negative bacteria (Ivanova et al., 2009; Lupoae et al., 2013; Putnoky et al., 2013). Stupar et al. (2022) suggest that conflict results can be explained by differences in extraction, solvents and plant parts used for the extract production. As in the experiment with *E. coli*, *E. faecalis* can be considered resistant in all treatments, except for the treatment with the highest wild garlic extract concentration (Table 3).

## Conclusion

According to the results of this study, almost all aqueous extracts showed a negligible effect on the growth of *E. coli* and *E. faecalis*. Significant suppression of bacterial growth compared with other treatments was observed with the aqueous extract of garlic at a concentration of 4%. Further research will focus on the chemical characterization of this extract and the effects of higher concentrations against other bacterial genera.

## References

- Agustin, K. C., Fatmaria, Augustina, I. (2022) Effectiveness of antibacterial extract Bawang Suna (*Allium schoenoprasum* L.) against methicillin-resistant staphylococcus aureus (MRSA) using total plate count. *Traditional Medicine Journal*, 27 (2), 101-110. <https://doi.org/10.22146/mot.71730>
- Badgley, B. D., Nayak, B. S., Harwood, V.J. (2010) The importance of sediment and submerged aquatic vegetation as potential habitats for persistent strains of enterococci in a subtropical watershed. *Water Research*, 44, 5857-5866. <https://doi.org/10.1016/j.watres.2010.07.005>
- Bialonska, D., Ramnani, P., Kasimsetty, S. G., Muntha, K. R., Gibson, G. R., Ferreira, D. (2010) The influence of pomegranate by-product and punicalagins on selected groups of human intestinal microbiota. *International Journal of Food Microbiology*, 140, 175-182. DOI: 10.1016/j.ijfoodmicro.2010.03.038
- Birring, O. J. S., Vilorio, I. L., Nunez, P. (2015) Anti-microbial efficacy of *Allium sativum* extract against *Enterococcus faecalis* biofilm and its penetration into the root dentin: an in vitro study. *Indian Journal of Dental Research*, 26 (5), 477-482. DOI: 10.4103/0970-9290.172041
- Braga L.C., Shupp J.W., Cummings C., Jett M., Takahashi J.A., Carmo L.S. (2005) Pomegranate extract inhibits Staphylococcus aureus growth and subsequent enterotoxin production. *Journal of Ethnopharmacology*, 96, 335-339. DOI: 10.1016/j.jep.2004.08.034
- Chehregani, A., Azimishad, F., Haj Alizade, H. (2007) Study on antibacterial effect of some Allium species from Hamedan-Iran. *International Journal of Agriculture and Biology*, 9(6), 873-876.
- EUCAST - The European Committee on Antimicrobial Susceptibility Testing (2017). Available at: [www.eucast.org](http://www.eucast.org)
- Ghasemian, A., Shokouhi Mostafavi, S. Kh., Vafaei, M., Nojoomi, F. (2018) Antimicrobial effects of aqueous and alcoholic extracts of *Allium schoenoprasum* on some bacterial pathogens. *Infection Epidemiology and Microbiology*, 4(1), 1-4.
- Giraffa G. (2002) Enterococci from foods. *FEMS Microbiology Reviews*, 26, 163-171. <https://doi.org/10.1111/j.1574-6976.2002.tb00608.x>
- Gonellimali, F. D., Lin, J., Miao, W., Xuan, J., Charles, F., Chen, M., Hatab, S. R. (2018) Antimicrobial properties and mechanism of action of some plant extracts against food pathogens and spoilage microorganisms. *Frontiers in Microbiology*, 9, 1639. <https://doi.org/10.3389/fmicb.2018.01639>
- Gustavsson J., Cederberg C., Sonesson U., Otterdijk R., Maybeck A. (2011) Global food losses and food waste: extent, causes, and prevention. Dusseldorf: FAO.
- Ivanova, A., Mikhova, B., Najdenski, H., Tsvetkova, I., Kostova, I. (2009) Chemical composition and antimicrobial activity of wild garlic *Allium ursinum* of bulgarian origin. *Natural Products Communication*, 4, 1059-1062. doi: 10.1177/1934578X0900400808
- Kalem I. K., Bhat Z. F., Kumar S., Desai A. (2017) Terminalia arjuna: a novel natural preservative for improved lipid oxidative stability and storage quality of muscle foods. *Food Science and Human Wellness*, 6, 167-175. <https://doi.org/10.1016/j.fshw.2017.08.001>
- Lanzotti, V., Bonanomi, G., Scala, F. (2013) What makes Allium species effective against pathogenic microbes? *Phytochemistry Reviews*, 12, 751-772. <https://doi.org/10.1007/s11101-013-9295-3>
- Lianou A., Panagou E. Z., Nychas G.-J. E. (2016) Microbiological spoilage of foods and beverages. U: P. Subramaniam, ur. *The stability and shelf-life of food*. Cambridge: Woodhead Publishing, 3-42.
- Lupoae, M., Coprean, D., Dinica, R., Gurau, G., Bahrim, G. (2013) Antimicrobial activity of extracts of wild garlic (*Allium ursinum*) from Romanian spontaneous flora. *Scientific Study and Research*, 14(4), 221-227.
- Mathabe, M. C., Nikolova, R. V., Lall, N., Nyazema, N. Z. (2005) Antibacterial activities of medicinal plants used for the treatment of diarrhea in Limpopo Province, South Africa. *Journal of Ethnopharmacology*, 105, 286-293. <https://doi.org/10.1016/j.jep.2006.01.029>
- Mostafa, A. A., Al-Askar, A. A., Almaary, K. S., Dawoud, T. M., Sholkamy, E. N., Bakri, M. M. (2018) Antimicrobial activity of some plant extracts against bacterial strains causing food poisoning diseases. *Saudi Journal of Biological Sciences*, 25 (2), 361-366. <https://doi.org/10.1016/j.sjbs.2017.02.004>
- Norsworthy, J. K. (2003) Allelopathic potential of wild radish (*Raphanus raphanistrum*). *Weed Technology*, 17 (2), 307-313. [https://doi.org/10.1614/0890-037X\(2003\)017\[0307:APOWRR\]2.0.CO;2](https://doi.org/10.1614/0890-037X(2003)017[0307:APOWRR]2.0.CO;2)
- Octavia, A., Budiardjo, S. B., Indliarti, I. S., Fauziah, E., Suharsini, M., Sutadi, H., Rizal, M. F. (2019) Garlic extract efficacy against the viability of *Enterococcus faecalis* (in vitro). *International Journal of Applied Pharmaceutics*, 11, Special Issue 1,

194-197. DOI: <http://dx.doi.org/10.22159/ijap.2019.v11s1.17351>

Oyawoye, O. M., Olotu, T. M., Nzekwe, S. C., Idowu, J. A., Abdullahi, T. A., Babatunde, S. O., Ridwan, I. A., Batiha, G. E., Idowu, N., Alorabi, M., Faidah, H. (2022) Antioxidant potential and antibacterial activities of *Allium cepa* (onion) and *Allium sativum* (garlic) against multidrug-resistant bacteria. *Bulletin of the National Research Centre*, 46, 214. <https://doi.org/10.1186/s42269-022-00908-8>

Pandey, A., Singh, P. (2011) Antibacterial activity of *Syzygium aromaticum* (Clove) with metal ion effect against foodborne pathogens. *Asian Journal of Plant Science and Research*, 1 (2), 69-80.

Parlapani, F. F., Mallouchos, A., Haroutounian, S. A., Boziaris, I. S. (2017) Volatile organic compounds of microbial and non-microbial origin produced on model fish substrate un-inoculated and inoculated with gilt-head sea bream spoilage bacteria. *LWT Food Science and Technology*, 78, 54-62. <https://doi.org/10.1016/j.lwt.2016.12.020>

Pavlović, D. R., Veljković, M., Stojanović, N. M., Gočmanac-Ignjatović, M., Mihailov-Krstev, T., Branković, S., Sokolović, D., Marčetić, M., Radulović, N., Radenković, M. (2017) Influence of different wild-garlic (*Allium ursinum*) extracts on the gastrointestinal system: spasmolytic, antimicrobial and antioxidant properties. *Journal of Pharmacy and Pharmacology*, 69, 1208-1218. doi: 10.1111/jphp.12746

Pirbalouti, A. G., Chaleshtori, A. R., Tajbakhsh, E., Momtaz, H., Rahimi, E., Shahin, F. (2009) Bioactivity of medicinal plants extracts against *Listeria monocytogenes* isolated from food. *Journal of Food and Agriculture Environment*, 7, 132-135.

Putnoky, S., Caunii, A., Butnariu, M. (2013) Study on the stability and antioxidant effect of the *Allium ursinum* water extract. *Chemistry Central Journal*, 7, 21. doi: 10.1186/1752-153X-7-21

Safithri, M., Bintang, M., Poeloengan, M. (2011) Antibacterial activity of garlic extract against some pathogenic animal bacteria. *Media Peternakan*, 2011, 155-158. DOI: 10.5398/medpet.2011.34.3.155

Sapkota, R., Dasgupta, R., Nancy, Rawat, D. S. (2012) Antibacterial effects of plants extracts on human microbial pathogens & microbial limit tests. *International Journal of Research in Pharmacy and Chemistry*, 2 (4), 926-936.

Shan, B., Cai, Y., Brooks, J. D., Corke, H. (2007) The in vitro antibacterial activity of dietary spice and medicinal herb extracts. *International Journal of Food Microbiology*, 117, 112-119. <https://doi.org/10.1016/j.ijfoodmicro.2007.03.003>

Shirzadi Karamolah, K., Mousavi, F., Mahmoudi, H. (2017) Antimicrobial inhibitory activity of aqueous, hydroalcoholic and alcoholic extracts of leaves and stem of *Daphne mucronata* on the growth of oral bacteria. *GMS Hygiene and Infection Control*, 12, 1-7. DOI: 10.3205/dgkh000301

Sihombing, D.R., Rusmarilin, H., Suryanto, D., Tampubolon, S. D. R., Yanti, S. F. (2018) Antimicrobial effects of chive extracts against bacteria pathogen and *Lactobacillus acidophilus*. *IOP Conference Series: Earth and Environmental Science*, 205, 012049. doi:10.1088/1755-1315/205/1/012049

Solomon, E. B., Yaron, S., Matthews, K. R. (2002) Transmission of *Escherichia coli* O157:H7 from contaminated manure and irrigation water to lettuce plant tissue and its subsequent internalization. *Applied and Environmental Microbiology*, 68 (1), 397-400. DOI: 10.1128/AEM.68.1.397-400.2002

Stupar, A., Šarić, L., Vidović, S., Bajić, A., Kolarov, V., Šarić, B. (2022) Antibacterial potential of *Allium ursinum* extract prepared by the green extraction method. *Microorganisms*, 10 (7), 1358. DOI: 10.3390/microorganisms10071358

Talib W. H., Mahasneh A. M. (2010) Antimicrobial, cytotoxicity, and phytochemical screening of Jordanian plants used in traditional medicine. *Molecules* 15 1811-1824. DOI: 10.3390/molecules15031811

Whitemore, B. B., Naidu, A. S. (2000). Thiosulfonates. U: A. S. Naidu, ur. *Natural food antimicrobial systemes*. Boca Raton, FL: CRC Press, 265-380.

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## Potencijal biljnih ekstrakata vlasca, medvjedeg luka i češnjaka u supresiji bakterijskog rasta

### Sažetak

Kvarenje hrane je pojava povezana sa prisustvom mikroorganizama u poljoprivrednim proizvodima. Zbog sposobnosti nekih mikroorganizama da koloniziraju biljni materijal, predložena je upotreba agrokemikalija za suzbijanje rasta bakterija. Međutim, prekomjerna uporaba agrokemikalija ima štetan utjecaj na kvalitetu okoliša. Stoga je cilj ovog istraživanja bio procijeniti potencijal vodenih ekstrakata češnjaka (*Allium sativum* L.), medvjedeg luka (*Allium ursinum* L.) i vlasca (*Allium schoenoprasum* L.) u suzbijanju rasta *Enterococcus faecalis* i *Escherichia coli*. Antimikrobna aktivnost ovih biljnih ekstrakata određena je disk difuzijskom metodom. Mueller-Hinton agar inokuliran je bakterijskim inokulantom; šest diskova filter papira, impregniranih sa 15 µl vodenog ekstrakta (1, 2 i 4%), postavljeno je na svaku površinu agara. Petrijeve zdjelice inkubirane su na 37 °C u trajanju od 24 h. Promjer zone inhibicije izmjeren je i izražen u milimetrima (mm). Svi vodeni ekstrakti pokazali su inhibitorni učinak na rast *E. coli* i *E. faecalis* Vodeni ekstrakt medvjedeg luka u koncentraciji od 4% značajno je suzbio rast bakterija u usporedbi sa drugim tretmanima. Ovo istraživanje potvrđuje potencijal ekstrakta medvjedeg luka u suzbijanju potencijalnih patogena.

**Ključne riječi:** antimikrobna aktivnost, vlasac, češnjak, zona inhibicije, medvjedi luk