



Original scientific paper

## Novel electrochemical sensing platform for detection of hydrazine based on modified screen-printed graphite electrode

Farideh Mousazadeh<sup>1</sup>, Sayed Zia Mohammadi<sup>2,✉</sup>, Maryam Mohammadhasani-Pour<sup>2</sup>

<sup>1</sup>School of Medicine, Bam University of Medical Sciences, Bam, Iran

<sup>2</sup>Department of Chemistry, Payame Nour University, Tehran, Iran

Corresponding author: ✉ [szmohammadi@yahoo.com](mailto:szmohammadi@yahoo.com)

Received: April 26, 2022; Accepted: July 29, 2022; Published: October 25, 2022

### Abstract

The current work aimed to fabricate a screen-printed graphite electrode (SPGE) modified by MnO<sub>2</sub> nanorods (MnO<sub>2</sub> NRs) for sensing hydrazine. Thus, a facile protocol was adopted to construct the MnO<sub>2</sub> nanorods that were subsequently applied to modify the SPGE surface directly. As-synthesized MnO<sub>2</sub> NRs/SPGE sensor exhibited a strong sensing behavior towards the hydrazine, with a large peak current and small oxidation potential. This electrochemical sensor in the optimized conditions to detect the hydrazine possessed a low detection limit (0.02 μM), a broad linear dynamic range (0.05-275.0 μM) and an admirable sensitivity (0.0625 μA μM<sup>-1</sup>). The sensor applicability was practically estimated in real water samples, which revealed successful recovery values.

### Keywords

MnO<sub>2</sub> nanorods; electrochemical sensor; hydrazine diffusion coefficient

### Introduction

Hydrazine, a vital chemical reagent, has attracted widespread research interest due to its industrial application and poisonousness. It is a robust reducing agent with various applications in the production of pesticides and medicine [1-3]. It is also a chemical deoxidizer with broad applications as an oxygen scavenger in water boilers [4]. Hydrazine is a key raw material in the construction of rockets and explosives [5,6]. Although hydrazine plays a great role in human production and life, it is easy to be absorbed by living organisms. The persistent contact with this agent may be associated with complications, such as some disturbances in the reproductive system, central nervous system, liver, kidneys and lungs [7-9]. According to the U.S. Environmental Protection Agency (US EPA), hydrazine is positioned in a class of possibly carcinogenic to humans, with a recommended threshold limit value (TLV) of less than 10 ppb [10].

Accordingly, it is substantial to achieve a practical and efficient sensing protocol for the determination of this agent in various media. In this regard, different analytical and instrumental

protocols have been designed to detect hydrazine so far, such as gas chromatography, capillary electrophoresis, chemiluminescence, high-performance liquid chromatography, liquid chromatography, colorimetry, and spectrophotometry [11-17]. Despite the unique benefits of each of these techniques, they typically require precision instrumentation and length of time. Among these, electrochemical approaches have been concerned by many researchers owing to their peculiar traits, which are simplicity, rapidity, affordability and short response time [18-34]. Voltammetric measurements typically use economical manners to detect analytes by recording variations in response current over alteration in electrode potential [35-42].

The recent development of electrochemical approaches has been greatly simplified by screen-printed electrodes (SPEs) due to their high sensitivity, functionality and versatility [43-45]. The main performance features of a sensor are all gathered in SPEs, including cost-effectiveness, minimal sample preparation, ease of operation, high speed, small size, limited background, and comfortable surface modification. There is always a need for further research to develop electrode materials to improve the selectivity and sensitivity of electrochemical sensors [46-51]. Many advances in nanotechnology have been made in diverse fields in recent years [52-75], which have led to the introduction of highly efficient sensing platforms. Types of nano-scale materials have so far been identified with distinct physicochemical properties that can be employed in the electrochemical sensors to detect various analytes exhibiting admirable results [76-82].

A popular oxide material is manganese dioxide ( $\text{MnO}_2$ ), whose behavior can be enhanced by changing its morphology and surface area.  $\text{MnO}_2$  is a polymorph owing to an octahedral [ $\text{MnO}_6$ ] spatial arrangement. Nano-sized  $\text{MnO}_2$  exhibits commendable benefits due to a larger surface-to-volume ratio and further reactive surface for electrochemical reactions. The diverse application of this substance in electrochemistry and sensor fabrication can be attributed to the simple reduction of  $\text{MnO}_2$  to  $\text{Mn}_2\text{O}_3$  and  $\text{MnO}$  and, at the proper potential, the re-oxidation to  $\text{MnO}_2$  as a catalytic circle for electrochemical detection [83-87].

The current work aimed to fabricate a new screen-printed graphite electrode (SPGE) supported by  $\text{MnO}_2$  nanorods ( $\text{MnO}_2$  NRs/SPGE) for sensitively sensing hydrazine. The sensor applicability was tested in real water samples, the results of which revealed successful recovery values.

## Experimental

### *Chemicals and instrumentations*

All materials with analytical grades applied throughout this work were supplied from Aldrich and Merck. Electrochemical experiments were recorded using a PGSTAT-302N Autolab potentiostat/galvanostat (Eco Chemie, The Netherlands). The control of all experiments was carried out by a General Purpose Electrochemical System (GPES) software. The SPGEs were purchased from DropSens (Spain) and consisted of an Ag pseudo-reference electrode, graphite auxiliary electrode, and graphite working electrode. All pH values were measured by a digital Metrohm 710 pH meter.

### *Synthesis of $\text{MnO}_2$ nanorods*

The  $\text{MnO}_2$  NRs were obtained by dissolving  $\text{KMnO}_4$  (0.316 g) in deionized water (30 mL) while vigorously stirring, followed by the addition of 3 M HCl (1.4 mL) under vigorous stirring for another half hour. Then, the solution was placed in a 50-mL Teflon-lined autoclave at 160 °C for six hours. Next, the products were cooled down to room temperature and subsequently centrifuged and thoroughly rinsed with ethanol and deionized water to clean any impurities, followed by drying at 60 °C for 12 h.

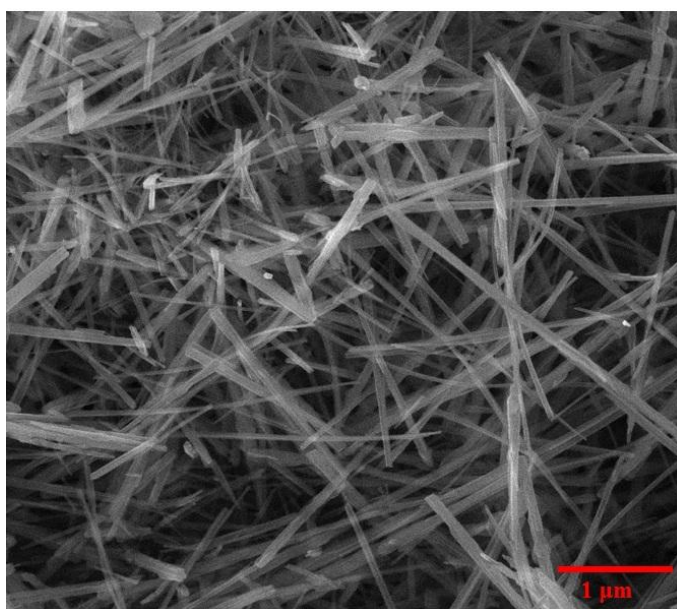
### Preparation of MnO<sub>2</sub> NRs/SPGE

First, 1 mg of prepared MnO<sub>2</sub> nanorods was added into an aqueous solution (1 ml), followed by sonication for 30 min to give a homogeneous solution. Then, 4 μL of MnO<sub>2</sub> NRs was dispersed on the surface of SPGE dropwise. Following the solvent's evaporation, the sensor's surface was washed several times with deionized water to clean free modifier molecules and subsequently air-dried. The obtained electrode was noted as MnO<sub>2</sub> NRs/SPGE.

## Results and discussion

### Characterization of MnO<sub>2</sub> nanorods

Figure 1 illustrates the FE-SEM images captured for the as-fabricated MnO<sub>2</sub> NRs, and observing them confirmed rod-shaped MnO<sub>2</sub> nanorods with a thickness ranging from 15 to 25 nm and a length of about 3 μm. The MnO<sub>2</sub> NRs showed an almost uniform size distribution.

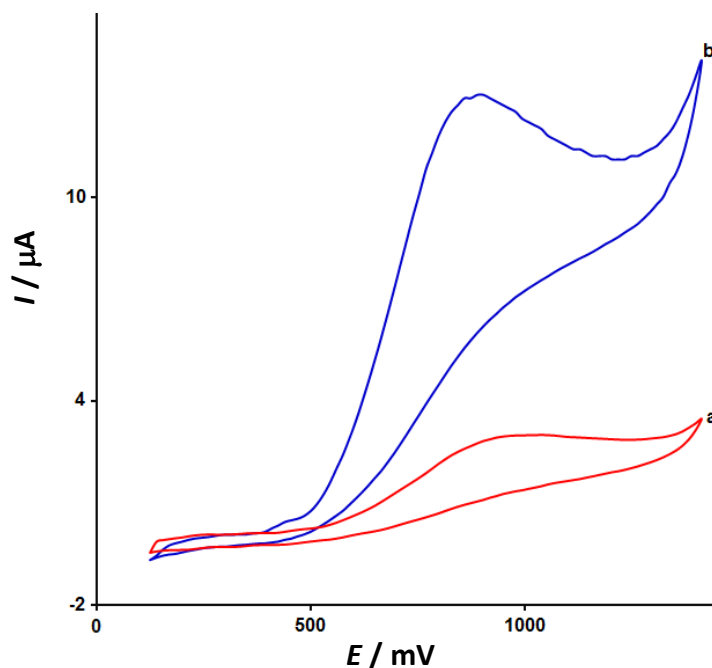


**Figure 1.** The FESEM image of synthesized MnO<sub>2</sub> nanorods

### Electrochemical response of hydrazine at different electrodes

The differential pulse voltammetry (DPV) method was recruited to study the effect pH value of electrolyte solution in different pH values (2.0-9.0) in the presence of 40.0 μM of hydrazine in phosphate buffer solution (0.1 M PBS) on the MnO<sub>2</sub> NRs/SPGE surface. The peak current of hydrazine oxidation was maximum at the pH value of 7.0, thereby selecting this value as the optimum pH in the hydrazine detection.

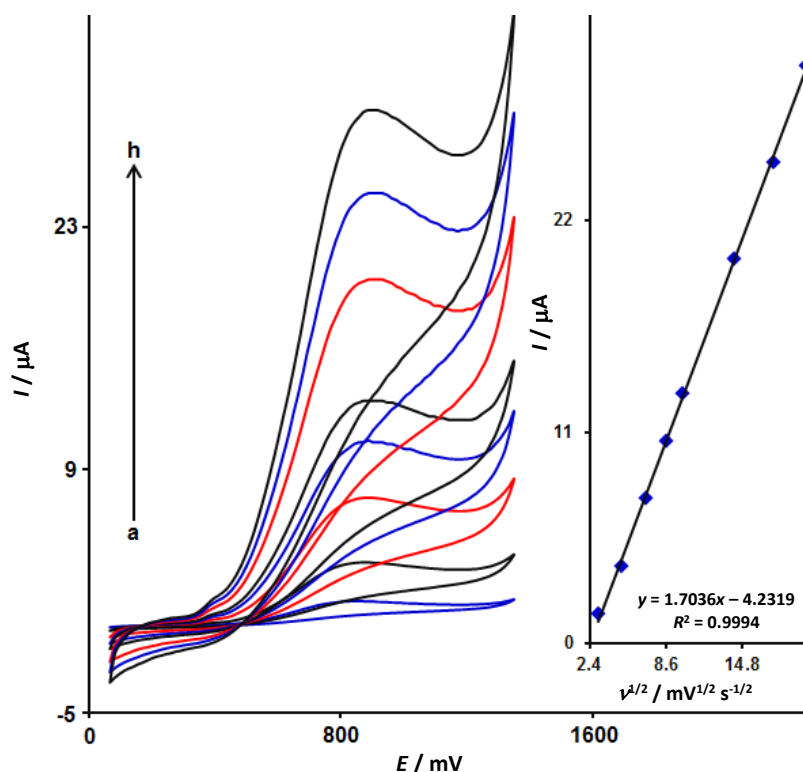
Figure 2 shows the application of the cyclic voltammetry (CV) method to evaluate the electrochemical behavior of 200.0 μM hydrazine at different electrodes (unmodified SPGE, and MnO<sub>2</sub> NRs/SPGE) in PBS (0.1 M, pH 7.0) at the scan rate of 50 mV/s. Based on the results, there was an oxidation peak on the surfaces of the electrodes, but no reduction peak, highlighting an irreversible electrochemical response of hydrazine on the electrodes. A relatively wide and weak peak current ( $I_{pa}$ ) of hydrazine oxidation was found on the unmodified SPGE (at 1000 mV with 3.0 μA), which reveals that the electrochemical oxidation does not happen spontaneously due to high activation overpotential. The hydrazine  $I_{pa}$  on MnO<sub>2</sub> NRs/SPGE, when compared with unmodified SPGE, displayed further elevation to 13.0 μA, meaning an increase up to 4.3 times that on the unmodified SPGE. In addition, hydrazine oxidation occurred at a lower potential than unmodified SPGE.



**Figure 2.** CV curves of unmodified SPGE (curve a), and MnO<sub>2</sub> NRs/SPGE (curve b) in 0.1 M PBS containing 200.0 μM hydrazine; scan rate: 50 mV s<sup>-1</sup>

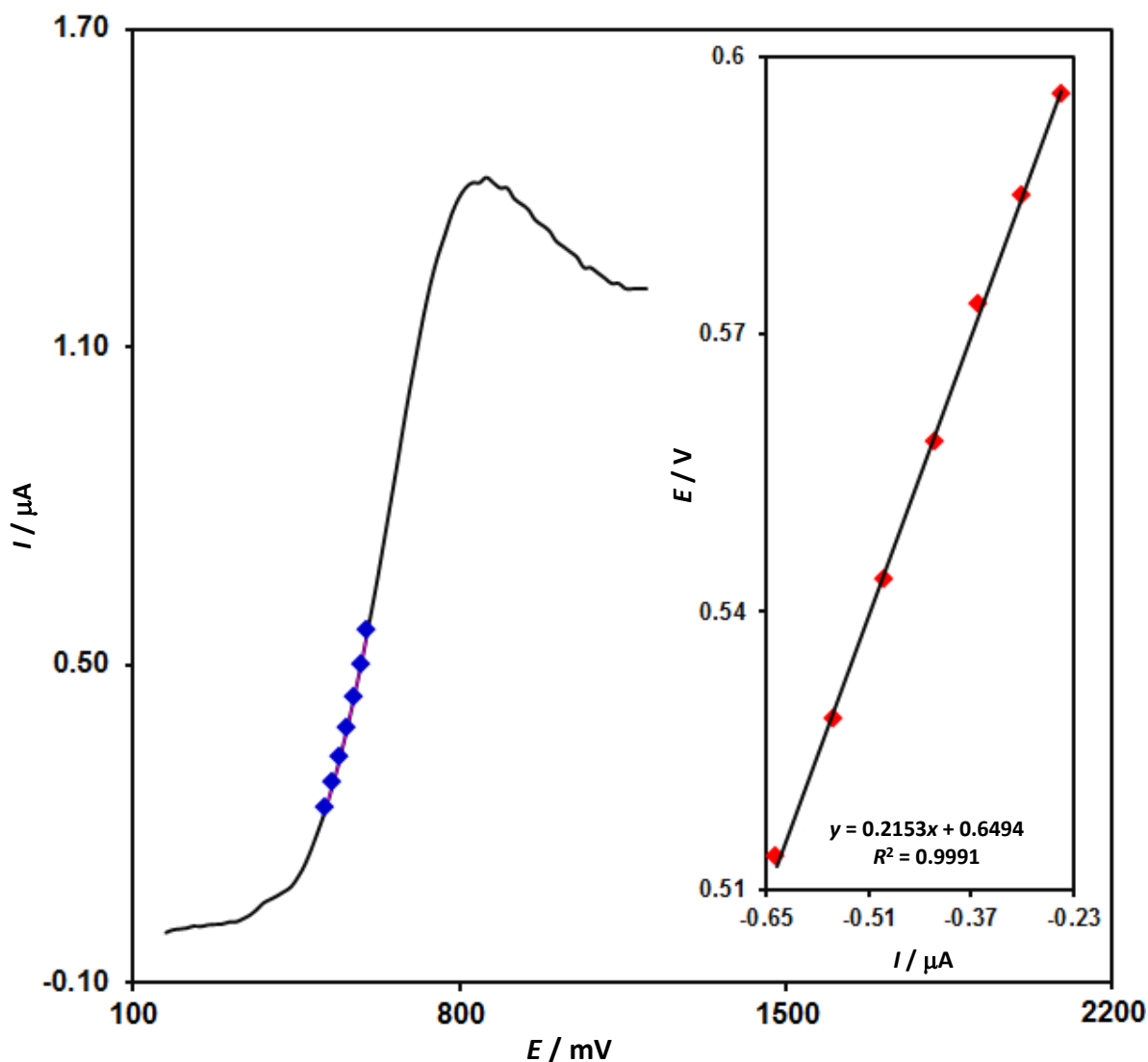
#### Effect of the scan rate ( $\nu$ ) on the results

The influence of various scan rates between 10 and 400 mV/s on the anodic peak currents for hydrazine (100.0 μM) was studied using the MnO<sub>2</sub> NRs/SPGE (Figure 3). The regression equation was  $I_{pa}(\text{hydrazine}) = 1.7036 \nu^{1/2} - 4.2319$  ( $R^2=0.9994$ ) (Figure 3, inset). This result indicates that the oxidation process is controlled by diffusion. Further, there was a shift in the oxidation peak potential of hydrazine toward a more positive potential by increasing the scan rates.



**Figure 3.** CV curves of 100.0 μM hydrazine in 0.1 M PBS (pH 7.0) at a scan rate of 10 to 400 mV s<sup>-1</sup> for MnO<sub>2</sub> NRs/SPGE (a-h refer to 10, 25, 50, 75, 100, 200.0, 300.0 and 400.0 mV s<sup>-1</sup>)  
Inset: plot of the square root of the scan rate vs. the oxidation peak current of hydrazine

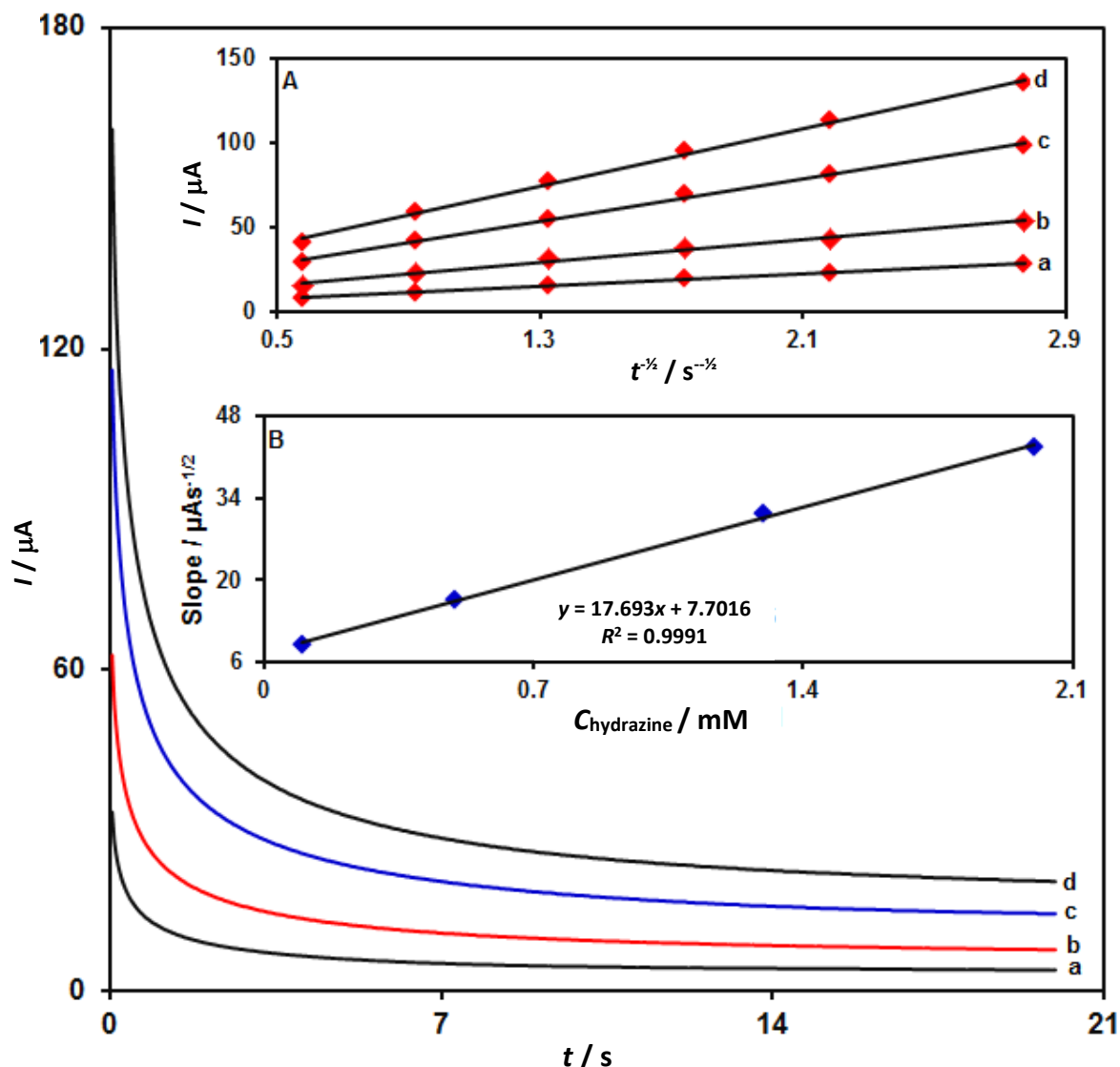
To study the rate-determining step as shown in Figure 4, the data of the rising part of the current-voltage curve obtained at 10 mV/s scan rate were applied to draw a Tafel plot for 100.0  $\mu\text{M}$  of hydrazine. The linearity of the  $E$  versus  $\log I$  plot, implies the intervention of the kinetics of the electrode process. The slope of this plot was utilized to estimate the number of electrons transferred in the rate-determining step. Figure 4 shows the Tafel slope of 0.2153 V for the linear section of the plot, which means the rate-limiting step of one-electron transfer with a transfer coefficient of  $\alpha = 0.72$ .



**Figure 4.** CV response for 100.0  $\mu\text{M}$  hydrazine with 10  $\text{mVs}^{-1}$  scan rate and the inset is the Tafel plot derived from the rising part of the corresponding voltammogram

#### Chronoamperometric analysis

Chronoamperometric determinations for different concentrations of hydrazine on the surface of  $\text{MnO}_2$  NRs/SPGE were measured by adjusting the potential of the working electrode at 0.94 V in PBS (0.1 M, pH 7.0), see Figure 5. For an electroactive material (hydrazine in this case) with a diffusion coefficient of  $D$ , the current observed for the electrochemical reaction at the mass transport limited condition is described by the Cottrell equation [88]. As shown in Figure 5A,  $I$  versus  $t^{-1/2}$  plots were used with the optimal fit for various hydrazine concentrations. We drew the slopes from straight lines against different concentrations of hydrazine, see Figure 5B. According to the Cottrell equation and obtained slope, the mean  $D$  value was  $2.7 \times 10^{-5} \text{ cm}^2/\text{s}$ .



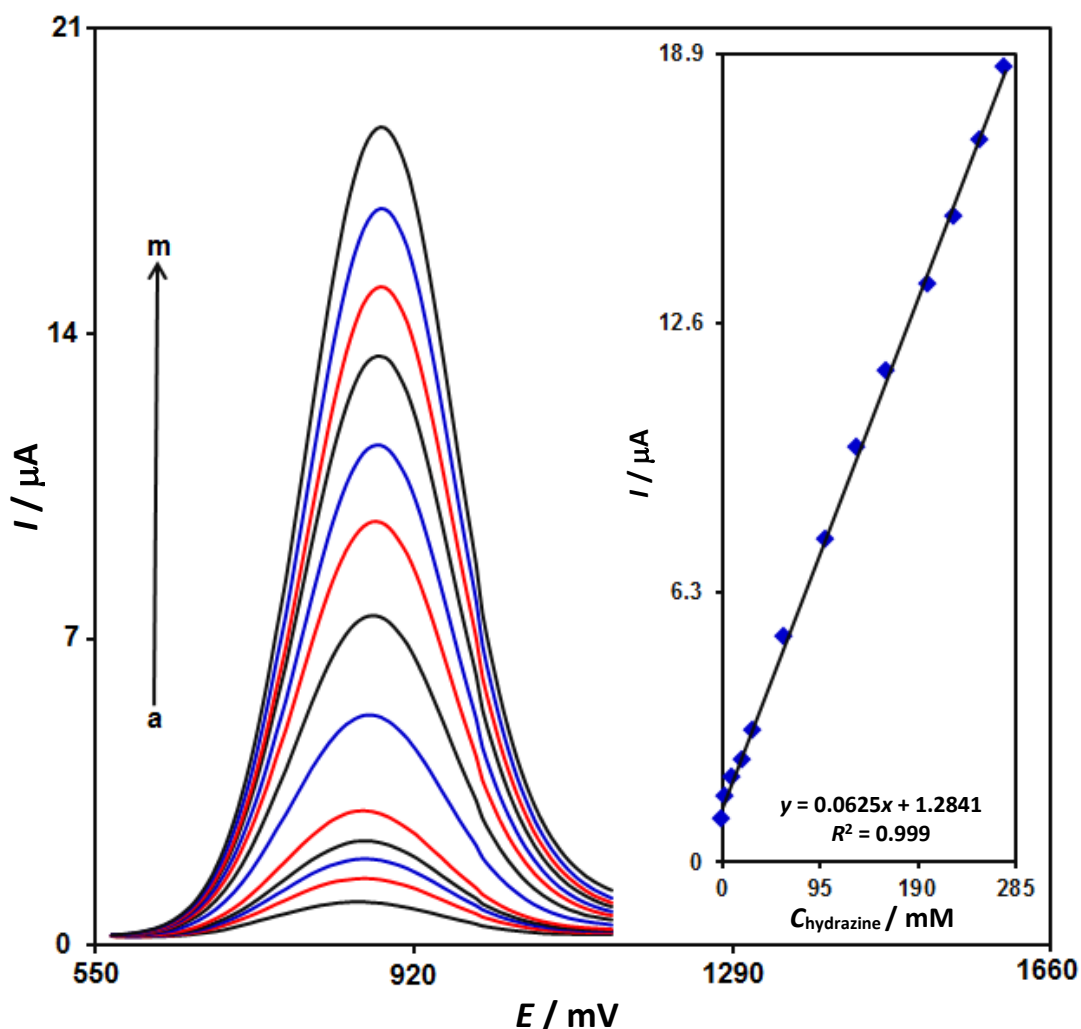
**Figure 5.** The chronoamperograms obtained at  $\text{MnO}_2$  NRs/SPGE in 0.1 M PBS at pH of 7.0 for different concentrations of hydrazine. a-d are related to 0.1, 0.5, 1.3, and 2.0 mM of hydrazine  
 Inset A: the  $I$  plot versus  $t^{-1/2}$  observed by chronoamperograms a to d.  
 Inset B: slope plot of the straight line vs. concentration of hydrazine

*Calibration curve, linear range and detection limit*

$\text{MnO}_2$  NRs/SPGE sensor was used to electrochemically detect different hydrazine concentrations (Figure 6). A gradual elevation was observed for the peak currents of hydrazine oxidation by raising its concentrations, which means an advanced performance of our sensor in the electrocatalytic oxidation of hydrazine. Oxidation peak currents of hydrazine versus  $C_{\text{hydrazine}}$  (Figure 6, inset) showed a wide linear range from 0.05 to 275.0  $\mu\text{M}$ . The detection limit ( $\text{LOD} = 3\sigma/S$ ; where  $\sigma$  is the standard deviation of blank response, and  $S$  is the slope of the calibration curve with a linear range of concentrations of the analyte) was calculated to be 0.02  $\mu\text{M}$ .

*Interference study*

The effect of some interference species on the determination of hydrazine was studied. The results show that the interfering effects of glucose, sucrose, urea, uric acid,  $\text{Na}^+$ ,  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{pb}^{2+}$ , and  $\text{Ag}^+$  on the anodic peak current of hydrazine is less than 5%. Hence, the  $\text{MnO}_2$  NRs/SPGE has a superior selectivity for hydrazine.



**Figure 6.** DPV responses of hydrazine on  $\text{MnO}_2$  NRs/SPGE at different hydrazine concentrations (a-m refer to: 0.05, 3.0, 10.0, 20.0, 30.0, 60.0, 100.0, 130.0, 160.0, 200.0, 225.0, 250.0 and 275.0  $\mu\text{M}$ ) in 0.1 M PBS (pH 7.0). Inset: The relationship between the oxidation peak currents and [hydrazine]

#### Analytical application

The detection of hydrazine in the water samples (drinking water and tap water) was performed using  $\text{MnO}_2$  NRs/SPGE sensor. The concentration values of hydrazine were calculated via the method of standard addition. Attained findings are summarized in Table 1, the recovery is between 96.7 and 102.5 %, and the relative standard deviations are all less than or equal to 3.0%. The experimental results confirmed that the  $\text{MnO}_2$  NRs/SPGE sensor has great potential for analytical application.

**Table 1.** Determining hydrazine in water samples through  $\text{MnO}_2$  NRs/SPGE (n=3)

Sample	C / $\mu\text{M}$		Recovery, %	RSD, %
	Spiked	Found		
Drinking water	0	-	-	-
	4.0	4.1	102.5	3.2
	6.0	5.8	96.7	2.4
Tap water	0	-	-	-
	5.0	4.9	98.0	1.9
	7.0	7.1	101.4	3.0

## Conclusion

The present work utilized an ultra-facile protocol to construct MnO<sub>2</sub> nanorods-modified SPGE (MnO<sub>2</sub> NRs/SPGE) for the electrochemical determination of hydrazine. According to CV findings, the as-fabricated sensor exhibited an electrocatalytic performance compared with the unmodified SPGE for the oxidation of hydrazine. The linear current response to the hydrazine level was between 0.05 and 275.0 μM, and the limit of detection was 0.02 μM with a sensitivity of 0.0625 μA μM<sup>-1</sup>. The diffusion coefficient for hydrazine using MnO<sub>2</sub> NRs/SPGE, 2.7×10<sup>-5</sup> cm<sup>2</sup> s<sup>-1</sup>, was obtained. The developed sensor applicability was practically tested to detect the concentrations of hydrazine in real water samples, which revealed successful recovery values.

## References

- [1] K. F. Khaled, *Applied Surface Science* **252** (2006) 4120-4128. <https://doi.org/10.1016/j.apsusc.2005.06.016>
- [2] U. Ragnarsson, *Chemical Society Reviews* **30** (2001) 205-213. <https://doi.org/10.1039/B010091A>
- [3] V. Rosca, M. T. M. Koper, *Electrochimica Acta* **53** (2008) 5199-205. <https://doi.org/10.1016/j.electacta.2008.02.054>
- [4] I. Cruz Vieira, K. Omuro Lupetti, O. Fatibello-Filho, *Analytical Letters* **35** (2002) 2221-2231. <https://doi.org/10.1081/AL-120016097>
- [5] R. Lan, J. T. S. Irvine, S. Tao, *International Journal of Hydrogen Energy* **37** (2012) 1482-1494. <https://doi.org/10.1016/j.ijhydene.2011.10.004>
- [6] A. Serov, C. Kwak, *Applied Catalysis B: Environmental* **98**(1-2) (2010) 1-9. <https://doi.org/10.1016/j.apcatb.2010.05.005>
- [7] C. A. Reilly, S. D. Aust, *Chemical Research in Toxicology* **10** (1996) 328-334. <https://doi.org/10.1021/tx960189j>
- [8] S. Garrod, M. E. Bollard, A. W. Nicholls, S. C. Connor, J. Connelly, J. K. Nicholson, E. Holmes, *Chemical Research in Toxicology* **18** (2005) 115-122. <https://doi.org/10.1021/tx0498915>
- [9] B. Toth, *Journal of Environmental Science and Health - Part C Environmental Carcinogenesis and Ecotoxicology Reviews* **2** (2008) 51-102. <https://doi.org/10.1080/10590508409373321>
- [10] G. Choudhary, H. Hansen, *Chemosphere* **37** (1998) 801-843. [https://doi.org/10.1016/S0045-6535\(98\)00088-5](https://doi.org/10.1016/S0045-6535(98)00088-5)
- [11] J. R. Holtzclaw, S. L. Rose, J. R. Wyatt, *Analytical Chemistry* **56** (1984) 2952-2956. <https://doi.org/10.1021/ac00278a074>
- [12] M. Khan, S. Kumar, K. Jayasree, K. K. Reddy, P. K. Dubey, *Chromatographia* **76** (2013) 801-809. <https://doi.org/10.1007/s10337-013-2467-x>
- [13] J. Liu, J. Jiang, Y. Dou, F. Zhang, X. Liu, J. Qu, Q. Zhu, *Organic and Biomolecular Chemistry* **17** (2019) 6975-6979. <https://doi.org/10.1039/C9OB01407A>
- [14] V. M. Dhalape, S. T. Khadangale, R. V. Pinjari, *Materials Today: Proceedings* **23** (2020) 400-409. <https://doi.org/10.1016/j.matpr.2020.02.060>
- [15] R. R. Gopireddy, A. Maruthapillai, J. A. Selvi, S. Mahapatra, *Materials Today: Proceedings* **34** (2021) 430-436. <https://doi.org/10.1016/j.matpr.2020.02.659>
- [16] N. Pourreza, R. Abdollahzadeh, *Microchemical Journal* **150** (2019) 104067. <https://doi.org/10.1016/j.microc.2019.104067>
- [17] D. S. Kosyakov, A. S. Amosov, N. V. Ul'yanovskii, A. V. Ladesov, Y. G. Khabarov, O. A. Shpigun, *Journal of Analytical Chemistry* **72** (2017) 171-177. <https://doi.org/10.1134/S106193481702006X>

- [18] Y. Pei, M. Hu, Y. Xia, W. Huang, Z. Li, S. Chen, *Sensors Actuators B* **304** (2020) 127416. <https://doi.org/10.1016/j.snb.2019.127416>
- [19] N. S. K. Gowthaman, S. Shankar, S. A. John, *ACS Sustainable Chemistry & Engineering* **6** (2018) 17302-17313. <https://doi.org/10.1021/acssuschemeng.8b04777>
- [20] S. Tajik, H. Beitollahi, H. Won Jang, M. Shokouhimehr, *Talanta* **232** (2021) 122379. <https://doi.org/10.1016/j.talanta.2021.122379>
- [21] I. Sheikhsaoie, H. Beitollahi, *Food and Chemical Toxicology* **162** (2022) 112864-112864. <https://doi.org/10.1016/j.fct.2022.112864>
- [22] H. Mahmoudi-Moghaddam, S. Tajik, H. Beitollahi, *Food Chemistry* **286** (2019) 191-196. <https://doi.org/10.1016/j.foodchem.2019.01.143>
- [23] H. Karimi-Maleh, H. Beitollahi, P. S. Kumar, S. Tajik, P. M. Jahani, F. Karimi, C. Karaman, Y. Vasseghian, M. Baghayeri, J. Rouhi, P. L. Show, S. Rajendran, L. Fu, N. Zare, *Food and Chemical Toxicology* **164** (2022) 112961. <https://doi.org/10.1016/j.fct.2022.112961>
- [24] F. G. Nejad, I. Sheikhsaoie, H. Beitollahi, *Food and Chemical Toxicology* **162** (2022) 112864. <https://doi.org/10.1016/j.fct.2022.112864>
- [25] S. Tajik, Z. Dourandish, F. G. Nejad, H. Beitollahi, A. A. Afsha, P. M. Jahani, A. Di Bartolomeo, *Journal of The Electrochemical Society* **169** (4) (2022) 046504. <https://doi.org/10.1149/1945-7111/ac62c3>
- [26] S. S. Moshirian-Farahi; H.A. Zamani; M. Abedi, *Eurasian Chemical Communications* **2**(9) (2020) 702-711. [http://www.echemcom.com/article\\_105259.html](http://www.echemcom.com/article_105259.html)
- [27] M. Payehghadr; Y. Taherkhani; A. Maleki; F. Nourifard, *Eurasian Chemical Communications* **2**(9) (2020) 982-990. [http://www.echemcom.com/article\\_114589.html](http://www.echemcom.com/article_114589.html)
- [28] A. Hosseini Fakhrabad; R. Sanavi Khoshnood; M.R. Abedi; M. Ebrahimi, *Eurasian Chemical Communications* **3**(9) (2021) 627-634. [http://www.echemcom.com/article\\_134775.html](http://www.echemcom.com/article_134775.html)
- [29] J. Mohanraj, D. Durgalakshmi, R. A. Rakkesh, S. Balakumar, S. Rajendran, H. Karimi-Maleh, *Journal of Colloid and Interface Science* **566** (2020) 463-472. <https://doi.org/10.1016/j.jcis.2020.01.089>
- [30] H. Karimi-Maleh, A. F. Shojaei, K. Tabatabaeian, F. Karimi, S. Shakeri, R. Moradi, *Biosensors and Bioelectronics*, **86** (2016) 879-884. <https://doi.org/10.1016/j.bios.2016.07.086>
- [31] T. Eren, N. Atar, M. L. Yola, H. Karimi-Maleh, *Food Chemistry* **185** (2015) 430-436. <https://doi.org/10.1016/j.foodchem.2015.03.153>
- [32] H. Karimi-Maleh, R. Darabi, M. Shabani-Nooshabadi, M. Baghayeri, F. Karimi, J. Rouhi, M. Alizadeh, O. Karaman, Y. Vasseghian, C. Karaman, *Food and Chemical Toxicology* **162** (2022) 112907. <https://doi.org/10.1016/j.fct.2022.112907>
- [33] J.D. Lović, *Journal of Electrochemical Science and Engineering* **12** (2022) 275-282. <https://doi.org/10.5599/jese.1166>
- [34] N. M. Abdul Khader Jailani, M. Chinnasamy, N. S. K. Gowthaman, *Journal of Electrochemical Science and Engineering* **12** (2022) 275-282. <https://doi.org/10.5599/jese.1207>
- [35] A. Hosseini Fakhrabad, R. Sanavi Khoshnood, M. R. Abedi, M. Ebrahimi, *Eurasian Chemical Communications* **3** (2021) 627-634. <http://dx.doi.org/10.22034/ecc.2021.288271.1182>
- [36] H. Karimi-Maleh, F. Tahernejad-Javazmi, A. A. Ensafi, R. Moradi, S. Mallakpour, H. Beitollahi, *Biosensors and Bioelectronics* **60** (2014) 1-7. <https://doi.org/10.1016/j.bios.2014.03.055>
- [37] Z. Dourandish, S. Tajik, H. Beitollahi, P. M. Jahani, F. G. Nejad, I. Sheikhsaoie, A. Di Bartolomeo, *Sensor* **22** (2022) 2238. <https://doi.org/10.3390/s22062238>
- [38] H. Karimi-Maleh, Q. A. Arotiba, *Journal of Colloid and Interface Science* **560** (2020) 208-212. <https://doi.org/10.1016/j.jcis.2019.10.007>

- [39] M. R. Aflatoonian, S. Tajik, B. Aflatoonian, M. S. Ekrami-Kakhki, K. Divsalar, I. Sheikh Shoaie, Z. Dourandish, M. Sheikhshoaie, *Eurasian Chemical Communications* **2**(4) (2020) 505-515. [http://www.echemcom.com/article\\_99027.html](http://www.echemcom.com/article_99027.html)
- [40] H. Karimi-Maleh, A. Khataee, F. Karimi, M. Baghayeri, L. Fu, J. Rouhi, C. Karaman, O. Karaman, R. Boukherroub, *Chemosphere* **291** (2022) 132928. <https://doi.org/10.1016/j.chemosphere.2021.132928>
- [41] H. Pyman; H. Roshanfekr; S. Ansari, *Eurasian Chemical Communications* **2**(2) (2020) 213-225. [http://www.echemcom.com/article\\_92411.html](http://www.echemcom.com/article_92411.html)
- [42] M. R. Aflatoonian; B. Aflatoonian; R. Alizadeh; R. Abbasi Rayeni, *Eurasian Chemical Communications* **2** (2020) 35-43. [http://www.echemcom.com/article\\_96655.html](http://www.echemcom.com/article_96655.html)
- [43] M. A. Tapia, C. Perez-Rafols, R. Gusmao, N. Serrano, Z. Sofer, J. M. Díaz-Cruz, *Electrochimica Acta* **362** (2020) 137144. <https://doi.org/10.1016/j.electacta.2020.137144>
- [44] W. Shi, J. Li, J. Wu, Q. Wei, C. Chen, N. Bao, H. Gu, *Analytical and Bioanalytical Chemistry* **412** (2020) 7275-7283. <https://doi.org/10.1007/s00216-020-02860-w>
- [45] S.Z. Mohammadi, H. Beitollahi, T. Rohani, H. Allahabadi, *Journal of Electrochemical Science and Engineering* **9** (2019) 113-123. <https://doi.org/10.5599/jese.637>
- [46] S. Tajik, H. Beitollahi, F. Garkani-Nejad, M. Safaei, P. Mohammadzadeh Jahani, *International Journal of Environmental Analytical Chemistry* **102**(7) (2020) 1477-1490. <https://doi.org/10.1080/03067319.2020.1738418>
- [47] O. C. Bodur, S. Dinç, M. Özmen, F. Arslan, *Biotechnology and Applied Biochemistry* **68** (2021) 20-29. <https://doi.org/10.1002/bab.1886>
- [48] T. Jamali, H. Karimi-Maleh, M. A. Khalilzadeh, *LWT - Food Science and Technology* **57** (2014) 679-685. <https://doi.org/10.1016/j.lwt.2014.01.023>
- [49] P. Prasad, N. Y. Sreedhar, *Chemical Methodologies* **2** (2018) 277-290. <https://doi.org/10.1016/j.lwt.2014.01.023>
- [50] F. Tahernejad-Javazmi, M. Shabani-Nooshabadi, H. Karimi-Maleh, *Talanta* **176** (2018) 208-213. <https://doi.org/10.1016/j.talanta.2017.08.027>
- [51] S. Tajik, Y. Orooji, F. Karimi, Z. Ghazanfari, H. Beitollahi, M. Shokouhimehr, R. S. Varma, H. W. Jang, *Journal of Food Measurement and Characterization* **15**(5) (2021) 4617-4622. <https://doi.org/10.1007/s11694-021-01027-0>
- [52] A. A. Ensafi, H. Karimi-Maleh, S. Mallakpour, *Colloids Surface B* **104** (2013) 186-193. <https://doi.org/10.1016/j.colsurfb.2012.12.011>
- [53] V. Karthika, P. Kaleeswaran, K. Gopinath, A. Arumugam, M. Govindarajan, N. S. Alharbi, G. Benelli, *Materials Science and Engineering C* **90** (2018) 589-601. <https://doi.org/10.1016/j.msec.2018.04.094>
- [54] R. Jabbari, N. Ghasemi, *Chemical Methodologies* **5** (2021) 21-29. <https://doi.org/10.22034/chemm.2021.118446>
- [55] A. A. Ensafi, H. Bahrami, B. Rezaei, H. Karimi-Maleh, *Materials Science and Engineering C* **33** (2013) 831-835. <https://doi.org/10.1016/j.msec.2012.11.008>
- [56] S. Tajik, A. A. Afshar, S. Shamsaddini, M. B. Askari, Z. Dourandish, F. Garkani Nejad, H. Beitollahi, A. Di Bartolomeo, *Industrial & Engineering Chemistry Research* **62** (2023) 4473-4480. <https://doi.org/10.1021/acs.iecr.2c00370>
- [57] S. Gupta, M. Lakshman, *Journal of Medicinal and Chemical Sciences* **2** (2019) 51-54. <https://doi.org/10.26655/JM-CHEMSCI.2019.3.3>
- [58] H. Beitollahi, M. Shahsavari, I. Sheikhshoaie, S. Tajik, P. M. Jahani, S. Z. Mohammadi, A. A. Afshar, *Food and Chemical Toxicology* **161** (2022) 112824. <https://doi.org/10.1016/j.fct.2022.112824>
- [59] S. Tajik, H. Beitollahi, S. Shahsavari, F. G. Nejad, *Chemosphere* **291** (2022) 132736. <https://doi.org/10.1016/j.chemosphere.2021.132736>

- [60] H. Karimi-Maleh, F. Karimi, Y. Orooji, G. Mansouri, A. Razmjou, A. Aygun, F. Sen, *Scientific reports* **10**(1) (2020) 11699. <https://doi.org/10.1038/s41598-020-68663-2>
- [61] S. Tajik, Z. Dourandish, F. G. Nejad, A. Aghaei Afshar, H. Beitollahi, *Micromachines* **13**(3) (2022) 369. <https://doi.org/10.3390/mi13030369>
- [62] H. Karimi-Maleh, M. Sheikhshoaie, I. Sheikhshoaie, M. Ranjbar, J. Alizadeh, N. W. Maxakato, A. Abbaspourrad, *New Journal of Chemistry* **43**(5) (2019) 2362-2367. <https://doi.org/10.1039/C8NJ05581E>
- [63] S. Tajik, A. Lohrasbi-Nejad, P. Mohammadzadeh Jahani, M. B. Askari, P. Salarizadeh, H. Beitollahi, *Journal of Food Measurement and Characterization* **16**(1) (2022) 722-730. <https://doi.org/10.1007/s11694-021-01201-4>
- [64] S. A. Alavi-Tabari, M. A. Khalilzadeh, H. Karimi-Maleh, *Journal of Electroanalytical Chemistry* **811** (2018) 84-88. <https://doi.org/10.1016/j.jelechem.2018.01.034>
- [65] S. Tajik, M. B. Askari, S. A. Ahmadi, F. G. Nejad, Z. Dourandish, R. Razavi, H. Beitollahi, A. Di Bartolomeo, *Nanomaterials* **12**(3) (2022) 491. <https://doi.org/10.3390/nano12030491>
- [66] H. Karimi-Maleh, C. Karaman, O. Karaman, F. Karimi, Y. Vasseghian, L. Fu, M. Baghayeri, J. Rouhi, P. Senthil Kumar, P. L. Show, S. Rajendran, *Journal of Nanostructure in Chemistry* **12** (2022) 429-439. <https://doi.org/10.1007/s40097-022-00492-3>
- [67] Y. Orooji, P. N. Asrami, H. Beitollahi, S. Tajik, M. Alizadeh, S. Salmanpour, M. Baghayeri, J. Rouhi, A. L. Sanati, F. Karimi, *Journal of Food Measurement and Characterization* **15**(5) (2021) 4098-4104. <https://doi.org/10.1007/s11694-021-00982-y>
- [68] S. Tajik, Y. Orooji, Z. Ghazanfari, F. Karimi, H. Beitollahi, R. S. Varma, H. W. Jang, M. Shokouhimehr, *Journal of Food Measurement and Characterization* **15**(4) (2021) 3837-3852. <https://doi.org/10.1007/s11694-021-00955-1>
- [69] M. Miraki, H. Karimi-Maleh, M. A. Taher, S. Cheraghi, F. Karimi, S. Agarwal, V. K. Gupta, *Journal of Molecular Liquids* **278** (2019) 672-676. <https://doi.org/10.1016/j.molliq.2019.01.081>
- [70] S. Mafi, K. Mahanpoor, *Eurasian Chemical Communications* **2**(1) (2020) 59-77. [http://www.echemcom.com/article\\_96613.html](http://www.echemcom.com/article_96613.html)
- [71] S. S. Mohammadi; N. Ghasemi; M. Ramezani, *Eurasian Chemical Communications* **2**(1) (2020) 87-102. [http://www.echemcom.com/article\\_96615.html](http://www.echemcom.com/article_96615.html)
- [72] F. Raoufi; H. Aghaei; M. Ghaedi, *Eurasian Chemical Communications* **2**(2) (2020) 226-233. [http://www.echemcom.com/article\\_92414.html](http://www.echemcom.com/article_92414.html)
- [73] S. Sarli; N. Ghasemi, *Eurasian Chemical Communications* **2**(3) (2020) 302-318. [http://www.echemcom.com/article\\_96625.html](http://www.echemcom.com/article_96625.html)
- [74] M. Ozdal, S. Gurkok, Recent advances in nanoparticles as antibacterial agent, *ADMET and DMPK* **10**(2) (2022) 115-129. <https://doi.org/10.5599/admet.1172>
- [75] S. Staroverov, S. Kozlov, A. Fomin, K. Gabalov, V. Khanadeev, D. Soldatov, I. Domnitsky, L. Dykman, S.V. Akchurin, O. Guliy, *ADMET and DMPK* **9**(4) (2021) 255-266. <https://doi.org/10.5599/admet.1023>
- [76] M. R. Ganjali, F. Garkani-Nejad, S. Tajik, H. Beitollahi, E. Pourbasheer, B. Larijani, *International Journal of Electrochemical Science* **12** (2017) 9972-9982. <https://doi.org/10.20964/2017.11.49>
- [77] M. Taei, H. Salavati, M. Fouladgar, E. Abbaszadeha, *Quarterly Journal of Iranian Chemical Communication* **8** (2020) 67-79. <https://doi.org/10.30473/ICC.2019.45932.1543>
- [78] S. Tajik, H. Beitollahi, F. Garkani-Nejad, I. Sheikhshoaie, A. Sugih Nugraha, H. Won Jang, Y. Yamauchi, M. Shokouhimehr, *Journal of Materials Chemistry A* **9** (2021) 8195-8220. <https://doi.org/10.1039/D0TA08344E>

- [79] H. Karimi-Maleh, M. Alizadeh, Y. Orooji, F. Karimi, M. Baghayeri, J. Rouhi, S. Tajik, H. Beitollahi, S. Agarwal, V. K. Gupta, *Journal of Industrial and Engineering Chemistry* **60** (2021) 816-823. <https://doi.org/10.1021/acs.iecr.0c04698>
- [80] D. Vishnu, B. Dhandapani, S. R. Ramakrishnan, P. K. Pandian, T. Raguraman, *Journal of Nanostructure in Chemistry* **11** (2021) 215-228. <https://doi.org/10.1007/s40097-020-00360-y>
- [81] H. Karimi-Maleh, K. Cellat, K. Arıkan, A. Savk, F. Karimi, F. Şen, *Materials Chemistry and Physics* **250** (2020) 123042. <https://doi.org/10.1016/j.matchemphys.2020.123042>
- [82] S. Sepahi, M. Kalae, S. Mazinani, M. Abdouss, S. M. Hosseini, *Journal of Nanostructure in Chemistry* **11** (2021) 245-258. <https://doi.org/10.1007/s40097-020-00362-w>
- [83] S. Ghafour, Taher, K. Abdulkareem Omar, B. Mohammed Faqi-Ahmed, *Asian Journal of Green Chemistry* **4** (2019) 231-238. <https://dx.doi.org/10.22034/AJGC/2020.2.10>
- [84] N. Jaiswal, I. Tiwari, C. W. Foster, C. E. Banks, *Electrochimica Acta* **227** (2017) 255-266. <https://doi.org/10.1016/j.electacta.2017.01.007>
- [85] M. A. Prathap, B. Satpati, R. Srivastava, *Sensors and Actuators B* **186** (2013) 67-77. <https://doi.org/10.1016/j.snb.2013.05.076>
- [86] F. T. Goh, Z. Liu, X. Ge, Y. Zong, G. Du, T. A. Hor, *Electrochimica Acta* **114** (2013) 598-604. <https://doi.org/10.1016/j.electacta.2013.10.116>
- [87] Y. Huang, C. Cheng, X. Tian, B. Zheng, Y. Li, H. Yuan, M. M. Choi, *Electrochimica Acta* **89** (2013) 832-839. <https://doi.org/10.1016/j.electacta.2012.11.071>
- [88] A. J. Bard, L. R. Faulkner, *Electrochemical Methods: Fundamentals and Applications*, John Wiley & Sons, New York, 2<sup>nd</sup> edition, 2001. ISBN: 978-0-471-04372-0