Morpho-palynological assessment of the genus *Terminalia* L. (Combretaceae) in Egypt

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Abstract – Eleven *Terminalia* species growing in streets and gardens in Cairo (Egypt) were investigated for their pollen morphology. The study's objectives were to explore the variation among *Terminalia* species' pollen pheno-characters and to categorize the various species within this genus according to these pollen characteristics. The results showed that there are significant variations in the pollen morphological characteristics in the genus under study, and as a result, these traits can be regarded as eurypalynous. Each of the two main categories of *Terminalia* species mono-morphic and di-morphic includes various pollen morphologies, ranging from spheroidal to prolate with hexa-aperture that may be similar or of two types. For each of the eleven species, an identification key and a clustering phenogram for the fifteen major pollen features have been created. The aperture type, aperture similarity, and exine ornamentation are some of the crucial traits for the differentiation of the species examined while the pollen shape class has a poor diagnostic character. According to the unique characteristics of the pollen morphology of the *Terminalia* species, three groups and two subgroups were identified.

Keywords: microscopy, palynology, pollen key, species determination, taxonomy

Introduction

The family Combretaceae is an advanced flowering plant with a worldwide tropical distribution (Hutchinson 1969). The family has a different number of genera, ranging from 16 (Rendle 1956) to 20 (Mabberley 2017). Cronquist (1981) recognized 400 species, while Willis (1966) recognized 600 species within this family. These variations in the number of genera and species are due to a lot of synonyms and the either clumping or splitting of genera (Scott 1979). Classification of Combretaceae has been faced with many opinions since Exell (1954) as the family was divided into two subfamilies, Strephonematoideae with the monotypic genus Strephonema and Combretoideae with the rest of the genera. More recent treatments by Exell and Stace (1966) and Stace (2007), divided the family into two tribes, Laguncularieae and Combreteae. The latter tribe was subdivided into three subtribes: Combretinae, Pteleopsidinae and Terminaliinae. Terminalia is the second largest genus after the genus Combretum under the subtribe Combritenae, tribe Combreteae, family Combretaceae, suborder Myrtineae, order Myrtales, (Stace 2002, Heywood et al. 2007). Species

under this genus are shrubs, trees, and creepers distributed in tropical and subtropical regions, especially in Africa and savannas (Arundhati et al. 2020). Many of its species have high economic value as they are used widely in commercial applications and Chinese traditional medicine (Zhang et al. 2019). The genus name came from the word terminal in Latin, as their leaves are crowded at the tops of the shoots. Recognition of the species under the genus Terminalia was according to fruit and leaf morphological characters (Chakrabarty et al. 2019, Rayane et al. 2022). The species under this genus differ greatly in their internal and external features in addition to their chromosome numbers (Stace 1965, Ohri 1996). Wickens (1973) and Fyhrquist (2007) found that bark, leaves, and fruit morphology can be used as diagnostic features in the identification of Terminalia species. Since species under the genus Terminalia are traditionally used in the treatment of some illnesses, precise identification and recognition of the closely related species need clarification.

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Flowers in the genus Terminalia are grouped in terminal simple racemes with duplicates of five anthers alternating with five petals. Despite the numerous stamens, only a few pollen morphological studies have dealt with taxonomical issues of the genus, e.g. Erdtman (1966), Patel et al. (1984), El-Ghazali et al. (1998), Krachai and Pornpongrungrueng (2015). Other palynological articles were mostly focused on the family level. The taxonomic disagreement concerning the species in the genus Terminalia, compounded by the challenge of accurately identifying certain species based only on macro-morphological characteristics, is the reason for this investigation. The aims of this work are to investigate the variation in the pollen pheno-characters of Terminalia species, and to classify the different species under this genus with respect to their pollen characters. Therefore, the subject of this investigation was the pollen grain classification of eleven Terminalia cultivated species, planted in Cairo parks, streets, and botanical gardens as ornamental trees.

Material and methods

Mature unopened flowers of eleven species were collected from Egyptian botanical gardens and streets during the flowering season (Tab. 1). The flowers were opened carefully using two needles under a stereo-microscope to release the anthers. The obtained anthers were smeared onto cleaned glass slides with a few drops of glycerol, covered, and sealed with paraffin wax. In order to prevent the harmomegathic effects – which occur when chemicals are exposed to aperturate pollen grains during the acetolysis process and the grains fold in response to changes in humidity (Taia 2022), non-acetolysed pollen grains were examined and then measured and photographed using a Motic (B-150D) light microscope (LM) fitted with a USB digital-video camera and computer software with 10× and 40× objective lenses.

All the measurements were based on 20 to 30 pollen grains. A calibrated oculometer was used to obtain the min-

imum and maximum measures and calculate the mean and standard deviation of each parameter and the apocolpium index (polar area index), which was calculated as the ratio of the mean distance between the apices of two colpi in both the main and sub apertures to its equatorial diameter. Values are expressed in micrometers.

For scanning electron microscope (SEM) investigation, non-acetolysed pollen grains were sputtered onto double sticks fixed in cleaned, aluminum labeled stubs, then coated with 20 nm gold in a Polaron JFC-1100 coating unit, finally examined and photographed using JEOL-JSM.I T200 Series Scanning Electron Microscope made available in the electron microscope unit, Faculty of Science, Alexandria University, Egypt. The terminology used for the descriptions is that of Punt et al. (2007).

Fifteen pollen morphological characters have been subjected to cluster analyses using PAST 3 programs V.3. designed by Hammer (1999-2013).

To generate a key, the data obtained from the description of pollen characters were subjected to automated key generation using version 4.12 of the DELTA suite of programs (Dallwitz et al. 2000).

Results

General description of Terminalia pollen grains

For the accurate identification of members of the *Terminalia* species, the morphological characteristics of pollen – such as size, shape, aperture conditions, and exine ornamentation – are crucial. The morphological properties of pollen, both quantitative and qualitative, derived by LM and SEM analysis are compiled in Tab. 2 and Fig. 1 and Fig. 2. The examined species' pollen grains are eurypalynous, meaning they vary greatly in character, shed in monads, apolar or isopolar and radially symmetric. These species are classified as either monomorphic or dimorphic. *T. arjuna, T. bentzoe, T. catappa, T. mantaly*, and *T. muelleri* are examples of monomorphic plants; on the other hand, the

Tab. 1. The localities of collected specimens, geographic coordinates, and the date of collection of the investigated *Terminalia* species, kept at Cairo University Herbarium (CAI).

No	Species	Locality	Date	Collector
1	<i>T. arjuna</i> (Roxb.) Wight & Arn.	Cairo: Zamalek, El-Giza street 30° 02' N 31° 13' E 12	July 2023	Rim Hamdy
2	T. bellirica (Gaertn.) Roxb.	Giza: Zoological garden 30° 01' 27.4" N 31° 12' 52.7" E 7	Nov. 2016	Heba Mostafa
3	<i>T. bentzoe</i> (L.) L. f.	Giza: Zoological garden 30° 01' 34.0" N 31° 12' 46.0" E 7	July 2016	Heba Mostafa
4	T. brownii Fresen.	Giza: Mazhar botanical garden 30° 03' N 31° 08' E 10	Dec. 2016	Heba Mostafa
5	T. catappa L.	Giza: Orman botanical garden 30° 01' N 31° 12' E 7	May 2019	Rim Hamdy
6	T. chebula Retz.	Giza: Zoological garden 30° 01' 34.0" N 31° 12' 46.0" E 7	Sept. 1963	Mohamed El-Mahdi
7	T. laxiflora Engl. & Diels	Cairo: El Zohriya garden 30º 02' N 31º 13' E 16	July 2023	Rim Hamdy
8	T. mantaly H. Perrier	Giza: Mazhar botanical garden 30° 03' N 31° 08' E 10	Feb. 2019	Rim Hamdy
9	T. muelleri Benth.	Giza: Zoological garden 30° 01' 35.2" N 31° 12' 52.4" E 7	Nov. 2016	Heba Mostafa
10	<i>T. myriocarpa</i> Van Heurck & Mull.	Giza Zoological garden 30° 01' 34.8" N 31° 12' 46.0" E 7	Dec. 2019	Heba Mostafa
11	T. sericea Burch. ex DC.	Giza: Zoological garden 30° 01' 34.0" N 31° 12' 46.0" E 7	June 1969	Mohamed El-Mahdi

torial axis diameter (μm), Eth – exine thickness (μm), EO – exine ornamentation, Gr – group, Hlob – hexalobate, M – mean, Mono – monomorphic, Min–Max – minimum and maximum values, No – number, P – polar axis length (μm), P/E – polar axis/equatorial axis, Pr – prolate, RG – rugate granulate, RP – rugate perforate, Sca – scabrate, SD – standard deviation, Sh – shape, SP – sub-prolate, Sph – spheroidal, St – striate rugate. Tr – triangular. Tab. 2. Pollen morphological data of 11 Terminalia species from Cairo, Egypt. AI – apocolpium index, Amb – the outline of pollen grain seen in polar view, Cir – circular, Di – dimorphic, E – equa-

No	Sheries	State		d	[II]	D/F	5	Amb	Eth	ED		Main ar	sertitres			Sub ane	sarut	
		200		4	1	Ì)	No	Type	Length	\mathbf{MI}	No	Type	Length	AI
	T. arjuna	Mono	Min – Max M ± SD	24.8 - 33.6 30.2 ± 3.4	23.8 - 30.2 28.7 ± 1.5	1.12	SP	Hlob, Cir	2.6	StR	3	Colpate	23.8 - 32.6 29.8 ± 2.8	0.25	Э	Colpate	21.2 - 30.2 28.4 ± 1.8	0.34
7	T. bellirica	Di	Min – Max M ± SD	24.8 - 32.2 29.2 ± 3.0	21.8 - 28.2 26.5 ± 1.7	1.15	SP	Πr	2.0	StR	$\tilde{\mathbf{w}}$	Colpate	22.8 - 30.2 29.2 ± 3.0	0.22	3	Colporate	21.5 - 29.8 26.2 ± 1.6	0.30
			Min – Max M ± SD	25.2 - 30.4 28.2 ± 1.8	24.8 - 30.6 29.0 ± 1.6	1.0	Sph	Πr	2.2	StR	3	Colpate	22.0 - 25.4 24.2 ± 1.6	0.21	З	Colporate	20.2 - 24.2 22.8 ± 1.4	0.28
$\tilde{\mathbf{\omega}}$	T. bentzoe	Mono	Min – Max M ± SD	23.8 - 28.2 25.6 ± 2.6	22.4 - 26.0 25.2 ± 1.8	1.1	Sph	Hlob	2.2	Sca	$\tilde{\omega}$	Colpate	21.2 - 26.4 22.6 ± 3.8	0.19	$\tilde{\omega}$	Colporate	20.4 - 2 2.6 22.2 ± 0.4	0.22
4	T. brownii	Di	Min – Max M ± SD	24.2 - 31.6 28.4 ± 3.2	23.2 - 29.8 27.6 ± 2.2	1.12	Sp	Πr	2.2	St	\mathfrak{c}	Colporate	23.2 - 29.6 23.8 ± 5.8	0.16	\mathfrak{S}	Colporate	22.8 - 28.2 23.2 ± 5.0	0.19
			Min – Max M ± SD	25.0 - 32.2 29.8 ± 2.4	21.8 - 29.8 27.6 ± 2.2	1.26	\Pr	Πr	2.2	St	$\tilde{\mathbf{\omega}}$	Colporate	24.2 - 31.2 29.4 ± 1.8	0.14	$\tilde{\mathbf{\omega}}$	Colporate	22.8 - 30.2 26.6 ± 3.6	0.20
5	T. catappa	Mono	Min – Max M ± SD	25.8 - 34.6 33.6 ± 1.0	23.2 - 29.8 27.4 ± 2.4	1.25	\Pr	Πr	2.5	RP	\mathfrak{c}	Colporate	23.4 - 32.8 30.2 ± 2.6	0.21	\mathfrak{S}	Colporate	21.2 - 30.6 28.6 ± 2.0	0.25
9	T. chebula	Di	Min – Max M ± SD	22.4 - 27.6 25.2 ± 2.4	20.0 - 24.2 23.2 ± 1.0	1.12	SP	Cir	2.6	Sca	$\tilde{\omega}$	Colpate	20.4 - 24.8 22.8 ± 2.0	0.24	3	Colporate	16.8 - 18.2 17.0 ± 1.2	0.36
			Min – Max M ± SD	22.2 - 27.2 26.2 ± 1.0	22.6 - 27.8 26.0 ± 1.8	1.0	Sph	Cir	2.4	Sca	$\tilde{\omega}$	Colpate	19.2 - 24.8 22.0 ± 2.8	0.28	3	Colporate	15.4 - 18.2 17.2 ± 1.0	0.41
	T. laxiflora	Di	Min – Max M ± SD	21.8 - 26.8 24.2 ± 2.6	20.0 - 24.8 23.8 ± 1.0	1.1	Sp	Πr	2.2	RG	$\tilde{\mathbf{w}}$	Colpate	19.2 - 24.8 23.6 ± 1.2	0.25	ŝ	Colporoidate	19.0 - 24.2 23.6 ± 0.6	0.26
			Min – Max M ± SD	24.8 - 33.2 29.2 ± 4.0	19.8 - 26.4 24.2 ± 2.2	1.22	\Pr	Hlob	2.2	RG	$\tilde{\mathbf{\omega}}$	Colpate	21.8 - 30.2 28.2 ± 2.0	0.26	ŝ	Colporoidate	20.8 - 29.6 26.8 ± 2.8	0.28
×	T. mantaly	Mono	Min – Max M ± SD	24.4 - 32.2 30.2 ± 1.0	23.8 - 30.4 29.2 ± 1.2	1.12	Sph	Hlob	2.2	StR	$\tilde{\mathbf{\omega}}$	Colpate	20.2 - 28.2 27.8 ± 0.4	0.28	$\tilde{\mathbf{\omega}}$	Colporate	19.2 - 27.2 25.8 ± 1.4	0.32
6	T. muelleri	Mono	$\begin{array}{l} Min - Max \\ M \pm SD \end{array}$	24.6 - 31.8 29.2 ± 2.6	23.4 - 30.8 28.2 ± 2.6	1.12	Sp	Hlob	2.6	StR	$\tilde{\mathbf{\omega}}$	Colpate	22.2 - 29.6 27.2 ± 2.4	0.28	с С	Colporoidate	20.2 - 26.4 24.8 ± 1.6	0.32
10	T. myriocarpa	Di	Min – Max M ± SD	24.8 - 28.2 25.8 ± 2.4	23.2 - 26.4 24.8 ± 1.6	1.1	SP	Cir	2.4	RP	\mathfrak{c}	Colpate	22.2 - 26.4 23.6 ± 2.8	0.34	$\tilde{\omega}$	Colporate	20.0 - 24.2 22.6 ± 1.6	0.38
			Min – Max M ± SD	27.8 - 34.5 32.4 ± 2.1	24.2 - 30.8 28.6 ± 2.0	1.25	\mathbf{Pr}	Cir	2.4	RP	$\tilde{\mathbf{\omega}}$	Colpate	24.2 - 30.8 28.2 ± 2.6	0.32	$\tilde{\mathbf{\omega}}$	Colporate	20.2 - 27.6 25.8 ± 1.8	0.36
11	T. sericea	Di	Min – Max M ± SD	18.6 - 24.4 22.0 ± 2.2	17.2 - 23.4 21.2 ± 2.2	1.12	SP	Hlob	2.2	RP	$\tilde{\omega}$	Colpate	18.8 - 20.4 19.2 ± 1.2	0.32	<i>.</i> ,	Colporoidate	15.2 - 19.8 18.6 ± 1.2	0.42
			Min – Max M ± SD	20.2 - 24.4 23.2 ± 1.2	20.0 - 24.6 23.2 ± 1.4	1.0	Sph	Cir	2.2	RP	$\tilde{\omega}$	Colpate	19.0 - 21.2 20.4 ± 1.2	0.34	3	Colporoidate	18.2 - 21.0 19.8 ± 1.6	0.41

dimorphic species were represented by *T. bellirica, T. brownii, T. chebula, T. laxiflora, T. myriocarpa* and *T. sericea*; indicating that they have two distinct pollen shapes, sizes, apertures, and even exine ornamentation within the pollen grains that are gathered from the same flower.

Shape and size

In their polar view, the pollen grains were triangular, hexa-lobed or circular (Fig. 1 and Fig. 2). In the equatorial view, they were spheroidal (*T. bellirica*, *T. bentzoe*, *T. chebula*, *T. mantaly* and *T. sericea*), subprolate (*T. arjuna*, *T. bellirica*, *T. brownii*, *T. chebula*, *T. laxiflora*, *T. muelleri*, *T. myriocarpa* and *T. sericea*) and prolate (*T. brownii, T. catappa, T. laxiflora* and *T. myriocarpa*). *T. catappa* had the largest pollen diameter on the polar axis (P) at 34.6 μ m, while *T. sericea* had the smallest (18.6 μ m). Similarly, *T. muelleri* and *T. myriocarpa* had the biggest pollen diameter on the equatorial axis (E) (30.8 μ m), whereas *T. sericea* had the smallest diameter (17.2 μ m). Based on the ratio between the lengths of the polar axis (an imaginary straight line connecting the two poles) to the equatorial diameter (P/E ratio), pollen sizes and shape variations were examined; *T. brownii* had the highest P/E value (1.26), while *T. bellirica, T. chebula*, and *T. sericea* had the lowest values (0.1).



Fig. 1. Light microscopy micrographs of the studied *Terminalia* species. (1-2) *T. arjuna*, (3-6) *T. bellirica*, (7-8) *T. bentzoe*, (9-10) *T. brownii*, (11-12) *T. catappa*, (13-16) *T. chebula*, (17-19) *T. laxiflora*, (20) *T. mantaly*, (21-22) *T. muelleri*, (23-25) *T. myriocarpa*, (26-28) *T. sericea*. (A – amb, C – colpus, E – exine, P – pore, scale bar = $10 \mu m$).

Aperture conditions

Apertures are the first characteristic to take into account when identifying pollen. In *T. arjuna*, *T. brownii*, and *T. catappa*, the apertures are iso-colpate; in the other investigated species, the apertures are hetero-colpate; they have two or more types of colpi, one of which varies in length and/or in whether endoapertures are present. The six apertures seen on the pollen grains of the *Terminalia* species



Fig. 2. Scanning electron microscope microphotographs showing the different pollen characters within the studied *Terminalia* species. (29-31) *T. arjuna*, (32-34) *T. bellirica*, (35) *T. bentzoe*, (36-37) *T. brownii*, (38-39) *T. catappa*, (40-41) *T. chebula*, (42-46) *T. laxiflora*, (47-49) *T. mantaly*, (50-52) *T. myriocarpa*. (a – amb, c – colpus, e – exine, p – pore). Scale bar = 1 μm (31, 39, 41, 49), 5 μm (29, 30, 32-38, 40, 43-48, 50-52), 10 μm (42).

under study are composed of three primary, long apertures that alternate with three smaller, possibly distinct, but somewhat shorter apertures.

Pollens of the species *T. arjuna* are hexa-colpate in which the main apertures and the sub-apertures are of the same type. The pollen grains of the species *T. bellirica*, *T. bentzoe*, *T. chebula*, *T. mantaly* and *T. myriocarpa* are hexa-colpate in which the main apertures are colpate and the sub-apertures are colporate. Also, hexa-colpate pollen grains are observed in *T. laxiflora*, *T. muelleri* and *T. sericea* in which the main apertures are colpate and the sub-apertures are colpate in which the main apertures are colpate and the sub-apertures are colpate in which the main apertures are colpate and the sub-apertures are colporoidate. In *T. brownii* and *T. catappa* the pollen grains are hexa-colporate in which the main apertures and the sub-apertures are of the same type.

The colpi are not connected at the poles and the apocolpium indices varied between the studied species from 0.14 in *T. brownii* to 0.42 in *T. sericea*. The aperture membranes are either psilate or granulate. In the case of composite apertures, the pores are either small and in the same colpi direction (lolongate) or oval shaped covered by granulated exinous layer (colporoidate). According to the state of the pollen grains, momo-or di- morphic and aperture types the studied species are grouped mainly in two main groups and four subgroups (Tab. 2).

Exine ornamentation

Exine sculpturing is the most distinguishing character of the species; in this study, the species have some variation in exine ornamentation; the exine is generally tectate, scabrate in *T. bentzoe*, and *T. chebula*; rugate in *T. laxiflora*; rugate perforate in *T. catappa*, *T. myriocarpa* and *T. sericea* with thin ectexine and thick endexine and intine layers as observed by light microscope investigation. Striate ornamentation is observed only in *T. brownii* while striate-rugate ornamentation is recorded in *T. arjuna*, *T. bellirica*, *T. mantaly* and *T. muelleri* (Tab. 2). Exine thickness ranges from the minimum (2.0 µm) in *T. bellirica* to the maximum (2.6 µm) in *T. arjuna*, *T. chebula* and *T. muelleri* (Fig. 1).

Clustering analysis

The phenogram resulting from the cluster analysis of fifteen palynological characters (On-line Suppl. Tab. 1) after coding according to their states (On-line Suppl. Tab. 2) divided the studied taxa into three groups at similarity distance 0.936. Group 1 has one species only, *T. sericea* (11). Group 2 holds eight species, subdivided into two subgroups at similarity distance 0.952, 2A with three species; *T. brownii* (4), *T. laxiflora* (7), *T. myriocarpa* (10); and subgroup 2B with five species; *T. catappa* (5), *T. mantaly* (8), *T. muelleri* (9), *T. arjuna* (1), *T. bellerica* (2). Group 3 includes two species, *T. bentzoe* (3) and *T. chebula* (6) (Fig. 3).

Computer-generated key

1. Pollen shape subprolate or prolate
Pollen shape subprolate or spheroidal
Pollen shape prolate <i>T. catappa</i>
Pollen shape spheroidal4
Pollen shape subprolate5
2. Amb shape triangular or hexalobate; Exine ornamenta-
tion rugate-granulate T. laxiflora
Amb shape triangular; Exine ornamentation striate .
Amb shape circular; Exine ornamentation rugate-per-
forate T. myriocarpa
3. Amb shape circular or hexalobate; Exine ornamenta-
tion rugate-perforate T. sericea
Amb shape triangular; Exine ornamentation striate-
rugate
Amb shape circular: Exine ornamentation scabrate
T. chebula
4. Exine ornamentation scabrate
Exine ornamentation striate rugate T. mantaly
5. Amb shape circular or hexalobate; Sub aperture type
Amb shape havelebate. Sub aparture type colores
And shape nexalobate; sub aperture type colporol-





Discussion

Morpho-palynological studies have proved to be of great importance for plant identification and taxonomic classification. In this study, different palynological features such as variations in pollen characters in the same flower (morphism), polar and equatorial outline and their ratio (P/E), type of aperture, presence or absence of pores, beside exine thickness and ornamentation were investigated in eleven *Terminalia* (Combretaceae) species belonging to 7 sections from Egypt using LM and SEM. Based on palynological features, a taxonomic key was created for simple identification and species delimitations.

Mignot et al. (1994) pointed to the importance of aperture type in angiosperm recognition at any taxonomic level, even down to the intra individual level. Based on the results, Terminalia species may be divided into two main groups: monomorphic and dimorphic. Each group has a variety of pollen shapes, ranging from spheroidal to prolate, with many shapes seen in the pollen sampled from the same flower. Pollen hydration status and environmental conditions may contribute to heterogeneity in pollen shape and size within the same taxon (Grant-Jacob et al. 2022, Taia et al. 2023). El Ghazali (2022) noted differences in the sizes and forms of pollen among the same taxa within the family Combretaceae. Despite this, he defined their pollens as stenopalynous, meaning that their characteristics are either stable or slightly changed. According to this investigation, the Terminalia under study exhibit significant variety in their pollen morphological characteristics, and as a result, they can be classified as eurypalynous.

The highest PA length is seen in *T. myriocarpa* (27.8-34.5 μ m), whereas *T. sericea* has the smallest PA length (18.6-23.2 μ m). El Ghazali et al. (1998) have documented these differences in pollen size and shape. Given that multiple shape types have been observed in a single flower, shape class is determined to have less diagnostic value within the taxa in the material under examination. Taia et al. (2023) have noted this investigation; as a result, it does not play a significant role in the taxonomic discrimination of the examined taxa, except in the case of *T. sericea*, which is grouped separately in the clustering analysis (group A), this separation being due to it having the smallest PA and ED lengths and the smallest main aperture and sub-aperture lengths. This result supports the classification of this species as it belongs to section Psidioides Exell.

Four monomorphic species (*T. catappa, T. mantaly, T. muelleri*, and *T. arjuna*) were gathered by the mono-morphic character within the same taxa and placed in subgroup 2B in close proximity to the dimorphic taxa (*T. bellerica*). This grouping strengthens the taxonomic status of *T. catappa, T. mantaly, T. muelleri* as they belong to the same section Eucatappa Engl. & Diels. Despite *T. arjuna* being a separate section Pentaptera (Roxb.) Engl. & Diels, it is grouped with the former three species according to the mono-morphic pollen character.

According to El Ghazali and Krzywinski (1989), the apertures can be classified as iso- or hetero-colpate, meaning that they have two or more types of colpi, with one type differing in length and/or having endoapertures or not. This previously acknowledged observation was discovered in *Rosa sericea* (Ullah et al. 2022). It was also discovered in *Fagonia* species (Taia et al. 2021). El-Ghazali et al. (1998) noted the heterocolpate pollen grains within the taxa under study. This study noted variations in the length and width of the colpi as well as the existence and dimensions of endo-apertures; this result is in line with previous studies mentioned.

Since T. brownii, T. laxiflora, and T. myriocarpa have triangular or circular amb, the triangular amb gathered them in subgroup 1B. Amb shape was determined to be a valuable character in the building of the identification key. This result is partially in agreement with the taxonomy of T. brownii, T. laxiflora as they belong to the same section, Platycarpae Engl. & Diels, but does not agree with the classification of T. myriocarpa as a separate section Myriocarpae. A triangular amb is also present in T. bellirica, although it is distinct from the preceding three species in terms of other pollen characteristics and is located in subgroup 2B, albeit remaining closely linked to them. Certain species could be distinguished using the hexagonal, triangular, or circular amb, as the identification key illustrates. The two species; T. bellirica and T. bentzoe are belonging to section Billiricae Engl. & Diels, however, the study separated them in different groups; this separation is due to the difference between them in pollen state and amb shape.

Exine thickness ranged from 2.0 μ m to 2.6 μ m; however, there were significant differences in exine ornamentation. The rugate-perforate pattern was seen in *T. myriocarpa* and *T. sericea*, while the striate or striate-rugate pattern was seen in the majority of the investigated species. Only *T. laxiflora* has rugate-granulate ornamentation, but *T. bentzoe* and *T. chebula* have scabrate exine ornamentation; for this reason, the former two species were grouped together in group C although they belong to different sections; *T. bentzoe* sec. Belliricae Engl. & Diels, *T. chebula* sec. Myrobalanus (Gaertn.) Eichl. As a result, exine thickness varies very little among the taxa, whereas exine ornamentation, displayed by the key, is more helpful in identifying the taxa. This observation is consistent with every palynological study conducted since that of Erdtman (1966).

Conclusion

Based on the study's findings, three groups and two subgroups can be identified based on the unique characteristics of the pollen morphology of the *Terminalia* species under investigation. The pollen morphism, aperture homogeneity, amb form, and exine ornamentation are the most crucial characteristics in classifying the investigated taxa.

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References

- Arundhati, D., Akshayakumar, B., Ranjan, R. G., 2020: Molecular characterization and genetic diversity assessment of ten plant species under family Combretaceae using ISSR markers. Research Journal of Biotechnology 15(12), 11–18.
- Chakrabarty, T., Krishna, G., Rasingam, L., 2019: Taxonomic notes on Indian *Terminalia* (Combretaceae). Plant Science Today 6(3), 281–286. https://doi.org/10.14719/pst.2019.6.3.539
- Cronquist, A., 1981: An integrated system of classification of flowering plants. Columbia University Press, New York. https://doi.org/10.2307/2806386
- Dallwitz, M. J., Paine, T. A., Zurcher, E. J., 2000: Onwards. Principles of interactive keys. Website http://delta-intkey. com/ www/interactivekeys. htm [accessed 16 December 2014].
- El Ghazali, G. E., Krzywinski, K.,1989: An attempt to clarify the term heterocolpate. Grana 28(3), 179–186. http://dx.doi. org/10.1080/00173138909427429.
- El Ghazali, G. E., Tsuji, S., El Ghazaly, G. A., Nilsson, S., 1998: Combretaceae R. Brown. World Pollen and Spore Flora 21, 1–40.
- El Ghazali, G. E., 2022: The genus *Buchenavia* Eichl. and its taxonomic affinity to the genus *Terminalia* L. (Combretaceae): Insight from pollen morphology-a review. Review of Palaeobotany and Palynology 301, 104644. https://doi.org/10.1016/j. revpalbo.2022.104644
- Erdtman, G., 1966: Pollen morphology and plant taxonomy. Angiosperms. (An Introduction to Palynology I). Hafner Publishing Company, New York and London.
- Exell, A. W., 1954: Combretaceae. Flora Malesiana 4, 533-589.
- Exell, A. W., Stace, C. A., 1966: Revision of the Combretaceae. Boletim da Sociedade Broteriana sér. 2, 40: 5–26.
- Grant-Jacob, J. A., Praeger, M., Eason, R. W., Mills, B., 2022: Generating images of hydrated pollen grains using deep learning. IOP SciNotes 3(2), 024001. https://doi. org/10.1088/2633-1357/ac6780.
- Heywood, V. H., Brummitt, R. K., Culham, A., Seberg, O., 2007: Flowering Plant Families of the World. Royal Botanic Gardens, Kew. Ontario: Firefly books. https://doi. org/10.1111/j.1467-8748.2007.00585.x.
- Hutchinson, J., 1969: Evolution and phylogeny of flowering plants: Dicotyledons: facts and theory, with over 550 illustrations and maps by the author. (No Title). Academic Press, London.
- Fyhrquist, P., 2007: Traditional medicinal uses and biological activities of some plant extracts of African *Combretum* Loefl., *Terminalia* L. and *Pteleopsis* Engl. Species. Ph.D Thesis, 183p. http://ethesis.helsinki.fi.
- Krachai, P., Pornpongrungrueng, P., 2015: Pollen morphology of Combretaceae from Thailand and its taxonomic significance. Thai Forest Bulletin (Botany) 43, 4–14.
- Mabberley, D. J., 2017: Mabberley's plant-book: a portable dictionary of plants, their classification and uses. Cambridge University Press, Cambridge, UK. https://doi. org/10.1017/9781316335581
- Mignot, A., Hoss, C., Dajoz, I., Leuret, C., Henry, J. P., Dreuillaux, J. M., Heberle-Bors, E., Till-Bottraud, I., 1994: Pollen aperture polymorphism in the angiosperms: importance, possible causes and consequences. Acta Botanica Gallica 141(2), 109–122. https://doi.org/10.1080/12538078.1994.1051 5144
- Ohri, D., 1996: Genome size and polyploidy variation in the tropical hardwood genus *Terminalia* (Combretaceae). Plant

Systematics and Evolution 200, 225-232. https://doi.org/10.1007/BF00984937

- Patel, V. C., Skvarla, J. J., Raven, P. H., 1984: Pollen characters in relation to the delimitation of Myrtales. Annals of the Missouri Botanical Garden 71(3), 858–969. https://doi. org/10.2307/2399170
- Hammer, Ø., 1999-2013: PAST paleontological statistics version 3.0: reference manual. University of Oslo.
- Punt, W., Blackmore, S., Nilsson, S., Thomas, A., 2007: Glossary of pollen and spore terminology. Review of Paleobotany and Palynology 143(1–2), 1–81. https://doi.org/10.1016/j.revpalbo.2006.06.008
- Rayane, T., Ribeiro, M., Maria, I., Loiola, B., Margareth, F. DE. S., 2022: Look at the trees: synopsis of *Terminalia* ss in the Brazilian Amazon with conservation and distribution implications. Anais da Academia Brasileira de Ciências 94(4), 1–19. https://doi.org/10.1590/0001-3765202220210265
- Rendle, A. B., 1956: The Classification of Flowering Plants-Dicotyledons 2. Cambridge University Press, Cambridge, UK. https://doi.org/10.5962/bhl.title.95518
- Scott, A. J., 1979: A revision of Anogeissus (Combretaceae). Kew Bulletin 1, 555–566. https://doi.org/10.2307/4109799
- Stace, C. A., 1965: The significance of the leaf epidermis in the taxonomy of the Combretaceae: I. a general review of tribal, generic and specific characters. Journal of the Linnean Society of London, Botany 59(378), 229–252. https://doi. org/10.1111/j.1095-8339.1965.tb00060.x
- Stace, C. A., 2002: Proposal to conserve Terminalia nom. cons. (Combretaceae) against an additional name Bucida. Taxon 51(1), 193–194. https://doi.org/10.2307/1554985
- Stace, C. A., 2007: Combretaceae. In: K. Kubitzki (ed.), The families and genera of vascular plants, 9, 68–82. Springer Verlag, Berlin, Heidelberg. https://doi.org/10.1007/978-3-540-32219-1_11
- Taia, W. K., 2022: Pollen grain diversity and application in taxonomy and evolution. Taeckholmia 42(1), 27–42. https://doi. org/10.21608/taec.2022.258213
- Taia, W. K., Ibrahim, M., Hassan, S., Asker, A., 2021: Palynological study of the genus *Fagonia* L. (Zygophyllaceae, Zygophylloideae) in Libya. Libyan Journal of Science and Technology 13(1), 29–37. https://doi.org/10.37376/ljst.v13i1.2272
- Taia, W. K., Hamdy, R. S., Abd El-Maged, A. M., 2023: Morphological diversity of pollen grains within the flowering stages of *Tecoma stans* (L.) Juss. ex Kunth. GSC Advanced Research and Reviews 16(1), 280–287. https://doi.org/10.30574/gscarr.2023.16.1.0320
- Ullah, F., Gao, Y. D., Zaman, W., Gao, X. F., 2022: Pollen morphology of *Rosa sericea* complex and their taxonomic contribution. Diversity 14(9), 705–722. https://doi.org/10.3390/ d14090705
- Wickens, G. E., 1973: Combretaceae. In: Polhill, R.M. (ed.), Flora of Tropical East Africa, 1–99. London, Crown Agents for Oversea Governments and Administrations, United Kingdom.
- Willis, J. C., 1966: A dictionary of the flowering plants and ferns. A dictionary of the flowering plants and ferns 7. Cambridge University Press, Cambridge, UK. https://doi.org/10.5962/ bhl.title.1428
- Zhang, X. R., Kaunda, J. S., Zhu, H. T., Wang, D., Yang, C. R., Zhang, Y. J., 2019: The genus *Terminalia* (Combretaceae): An ethnopharmacological, phytochemical and pharmacological review. Natural Products and Bioprospecting 9, 357–392. https://doi.org/10.1007/s13659-019-00222-3