Review

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1

# Science-based evidence on pathways and effects of human exposure to micro- and nanoplastics

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Human exposure to plastic particles has raised great concern among all relevant stakeholders involved in the protection of human health due to the contamination of the food chain, surface waters, and even drinking water as well as due to their persistence and bioaccumulation. Now more than ever, it is critical that we understand the biological fate of plastics and their interaction with different biological systems. Because of the ubiquity of plastic materials in the environment and their toxic potential, it is imperative to gain reliable, regulatory-relevant, science-based data on the effects of plastic micro- and nanoparticles (PMNPs) on human health in order to implement reliable risk assessment and management strategies in the circular economy of plastics. This review presents current knowledge of human-relevant PMNP exposure doses, pathways, and toxic effects. It addresses difficulties in properly assessing plastic exposure and current knowledge gaps and proposes steps that can be taken to underpin health risk perception, assessment, and mitigation through rigorous science-based evidence. Based on the existing scientific data on PMNP adverse health effects, this review brings recommendations on the development of PMNP-specific adverse outcome pathways (AOPs) following the AOP Users' Handbook of the Organisation for Economic Cooperation and Development (OECD).

KEY WORDS: adverse outcome; plastic particles; regulatory-relevant data; risk assessment, risk management

Plastic pollution has long been one of the top environmental issues due to its abundance and persistence in the aquatic environment, which threatens to double over the next 20 years in a business-as-usual scenario (1), whether it slowly degrades to or is released into the environment as industrial plastic micro- and nanoparticles (PMNPs) (e.g., paints, adhesives, electronics, and cosmetics) (2–6).

Chronic exposure to plastic particles raises great concern for all life, as we interact with them without completely understanding how they affect our planet and much less our bodies through the food chain and surface or even drinking water (3–10). The smaller they are the more easily they penetrate blood, endothelial, intestinal, blood-brain, and blood-placental barriers to reach and accumulate in tissues and organs with different adverse outcomes. Their uptake by different body compartments also depends on their shape and surface chemistry (11).

Because of the ubiquity of plastic materials and their toxic potential, it is imperative to gather reliable, regulatory-relevant, and science-based evidence on how PMNPs affect human health to properly assess and manage risks in the context of circular economy. The main regulatory pathways to collect such data should follow

Corresponding author: Nikolina Kalčec, Institute for Medical Research and Occupational Health, Ksaverska cesta 2, 10000 Zagreb, Croatia E-mail: nkalee@imi.hr. ORCID: 0000-0002-3914-3063 the Integrated Approaches to Testing and Assessment (IATA) supported by the adverse outcome pathway (AOP) concept (12, 13). In 2019, the World Health Organization (WHO) reviewed the state of evidence on PMNPs in drinking water (14), while the European Commission adopted its strategy for plastics in a circular economy in 2018 (15). However, the main challenge is the lack of sensitive analytical methods to detect and quantify PMNPs in different biological and environmental matrices. It is impossible to assess PMNP exposure, toxicokinetics, and toxicodynamics without measuring the number and concentration of particles that reach and accumulate in biological fluids, cells, tissues, and organs. So far, dose-response data in humans are very scarce.

This review aims to synthesise current knowledge on PMNP exposure doses, pathways, and health effects in humans. We also highlight current knowledge gaps and the challenges in assessing plastic exposure and bring recommendations for the development of PMNP-specific AOPs based on the existing scientific evidence on PMNP adverse health effects.

# METHODOLOGY

The first step towards collecting current science-based evidence of PMNP effects on human health was a comprehensive literature search across the Web of Science Core Collection (WoSCC) database, using the search phrase "(micro OR nano) AND (plast\*) AND (tox\*) AND ("in vivo" OR human)". The search was further refined by the additional phrases "("mur\*" OR "mouse" OR "rat")", to obtain results published on in vivo rodent animal models. The database was accessed on 28 March 2023, and the search identified a total of 343 papers. They were screened, and non-relevant papers excluded to finally get 43 relevant papers reporting qualitative data on PMNP health effects and quantitative data on PMNP uptake, of which 26 were review articles. Due to the scarcity of humanrelated experimental data, we decided to include and analyse data from in vivo and in vitro studies to fully appreciate the scope of health issues that could be associated with plastic materials and PMNPs. Therefore, our analysis includes all data relating to the magnitude of exposure (i.e., number of particles taken up by cells, tissues, or organisms) and PMNP effects on cell or tissue metabolism.

In addition to the literature search in the WoSCC database, we reviewed information in the AOP-Wiki (16) to identify AOPs that may be associated with the health effects reported for PMNPs. This work was based on comparing adverse effects reported in the 43 WoSCC papers (Figure 1) with molecular initiating events (MIEs) or key events (KEs) described in the AOP-Wiki for different AOPs. The value of the AOP framework lies in the fact that AOPs are not specific for particular stressors but can define downstream PMNP effects if proper MIEs or KEs are identified. AOPs can facilitate risk assessment and management by placing mechanistic knowledge in hazard characterisation in a specific regulatory context (17, 18). Moreover, identifying AOPs relevant for PMNPs can further inform the choice of the most suitable assays for assessing specific adverse outcomes (19). As already demonstrated by several IATA OECD case studies describing AOPs for non-genotoxic carcinogens, skin sensitisation, chemical-induced liver steatosis, and neural development (17), implementation of the AOP framework may improve non-animal *in vitro* testing for future hazard characterisation of PMNPs.

## **RESULTS AND DISCUSSION**

# Routes of exposure and estimated human uptake of PMNPs

Data on routes of human exposure (Figure 2) show that inhalation and ingestion are much more relevant for PMNP uptake than the transdermal route (20, 21). There are many sources of inhalation exposure, depending on the type of contamination of indoor and outdoor air. Inhaled PMNPs can translocate into the lung tissue or enter the digestive system via mucociliary clearance (7, 22, 23). Oral uptake is mostly owed to contamination of bottled drinks or different food items including table salt (24, 25). PMNPs can also contaminate food during food processing and packaging.

For all exposure routes, the mechanism of uptake depends on PMNP size, shape, solubility, and surface characteristics as well as on biological factors, such as the site of particle deposition (7, 26). Depending on the size, PMNPs can spread through passive diffusion, paracellular transfer, or active cellular uptake (7), which also depends on the cell type, e.g., Peyer's patches in the intestine or alveolar macrophages in the lung (7, 27). Even though dermal

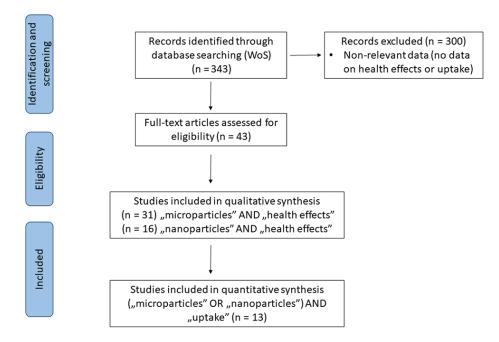


Figure 1 Flow chart of literature search in the Web of Science Core Collection (WoSCC) database Bakan B, et al. Science-based evidence on pathways and effects of human exposure to micro- and nanoplastics Arh Hig Rada Toksikol 2024;75:1-14

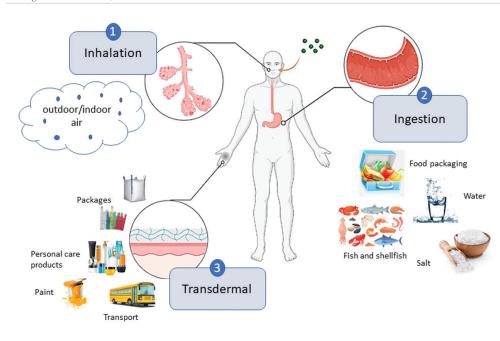


Figure 2 Human exposure routes to plastic micro- and nanoparticles in humans (created with BioRender. com)

contact accounts for a relatively small fraction of PMNP uptake, some studies suggest that PMNPs can be absorbed through hair follicles and sweat ducts, which could lead to systemic exposure (28–30).

Estimations of PMNP intake by humans significantly vary between studies (Table 1) and depend on the type of PMNP and exposure route(s) considered by a particular study. For example, individual dietary PMNP intake in the USA is estimated to 39,000-52,000 particles per year, while the total intake increases to 74,000-121,000 particles per year if we include the inhalation route (31). It is important to note that all estimations presented in Table 1 are based on limited data and may not be representative of a specific population or geographic region. Furthermore, reliable, accurate, and precise methods for identification, characterisation, and quantification of PMNPs in complex matrices like environmental and biological samples are still scarce (8, 14, 15, 20, 32). This problem is especially frustrating for the development and implementation of particle dosimetry in PMNP risk assessment and management whose aim is to quantify the number of plastic particles in a particular analytical sample.

Due to the resistance of plastic materials to degradation, bioaccumulation should be considered a serious health issue, as inhaled and ingested plastic particles may remain in different tissues for long and cause chronic health effects (28–30, 33). Deng et al. (34) reported that plastic microparticles in the liver, kidney, and gut tissue accumulate in a range of 0.07–0.41 mg of plastic material per gram of tissue. Another animal study (23) reported very low accumulation in the intestinal cell layer and no plastic materials in the liver, spleen, or kidney. Unfortunately, most studies report difficulties in properly assessing the toxicokinetic/toxicodynamic profiles of PMNPs (35).

#### Health effects of PMNPs

According to the reviewed scientific literature (Figure 1), PMNPs may cause various detrimental effects on cell viability, inflammatory response, lipid metabolism, oxidative status, intestinal microbiota, ion transport, signalling pathways, DNA integrity, hepatic function, and more. Table 2 lists the reported PMNP effects on human and animal cells and tissues. The sheer number of different adverse health effects following exposure to PMNPs raises concern about the ever-increasing plastic pollution.

# Linking PMNP-induced health effects to existing AOPs for chemicals

The AOP framework (13) enables collection and logical organisation of experimental data from different sources for efficient identification of essential biological events affected by exposure of living organisms to chemical stressors. Here we used the AOP framework to identify molecular initiating events (MIEs) and key events (KEs) associated with biological effects of PMNPs given in Table 2. The main aim was to generate hypothesis-based AOPs relevant to PMNPs by linking PMNP-induced toxicity endpoints reported so far with existing AOPs in the AOP-Wiki (16). However, it is important to note that many of these AOPs are under development.

Our analysis shows that PMNPs reported to induce biological effects have already been described either as MIEs or as KEs in 170 different AOPs (Table 3), which is an alarming finding that urges for more focused risk assessment of plastic across all steps in its value chain.

Biological models	Exposure data	Reference
Ecotoxicity, human cells, humans	Ingestion route: 11,000 particles from shellfish, 4000 particles from drinking water, and 7–1000 particles from edible sea salt per person per year	(36)
Ecotoxicity, human cells, humans	Ingestion route. Sources: seafood, tea bags, honey, sugar, beverage drinks, commercial salts, milk, beer, tap and bottled drinking water	(37)
Ecotoxicity, humans	Ingestion sources: drinking water, food containing plastic particles or weathering from plastic containers, salts and honey, and beer	(5)
Ecotoxicity, human cells and exposure	Ingestion through the food chain	(27, 31, 38–45)
Human exposure	Ingestion route: $\leq 30$ particles/day from tap water and beverages, 37–100 particles/year from sea salt; in total $\leq 250$ pg/kg body weight per day for an adult from tap water, beverages, and sea salt	(11)
Rodent model	5-day or al exposure to 60 nm polystyrene particles: 10 $\%$ of the dose found in the gas trointestinal tract	(46)
In vitro models	Exposure experiments with red blood cells, peripheral blood mononuclear cells, and mast cells	(47)
In vitro models	Exposure experiments with intestinal epithelial cell lines, LS174T, HT-29, and CaCo-2	(22, 48)
In silico models	/	(49)
Invertebrates and vertebrates	Exposure through the food chain	(50)
In vitro models	Exposure experiments with epithelial HeLa and cerebral T98G cells	(51)
In vivo model organisms	Ingestion route: 12,000–204,000 particles per person per year via plastic-contaminated seafood (fish and shellfish), beer, table salt, sugar, and honey	
In vitro model of the whole digestive system	Annual ingestion: 123,000 particles for adults (714 mg/day), 81,000 particles for children (449 mg/day).	(53)
<i>In vitro</i> models, animal models, plants	Ingestion via the food chain	(54, 55)
Human exposure	Annual ingestion: 11,000 particles from shellfish for European top consumer. Ingestion sources: 50.97 particles/L of beer, 24.53 particles/L of soft drinks, 5.79 particles/L of energy drinks, 5.26 particles/L of cold tea, 3 to 11 particles/L of milk; the highest mean concentrations of particles from drink consumption in the US (9.24–11.8 particles/L) and the lowest in Germany (0.91–1.29 particles)	
<i>In vivo</i> animal models, mammals	The primary route of exposure: ingestion of food and water contaminated with PMNP; annual consumption of 39,000 and 52,000 particles per person in the US	(57)
In vivo	Ingestion of PMNP does not provide a significant contribution to the transfer of absorbed chemicals from the water to the biota via the gut	(10)
In vitro, in vivo	The oral bioavailability of 50 nm-sized polystyrene particles differs between 0.2 and 2 % in rodents ( <i>in vivo</i> ) and humans ( <i>in vitro</i> ); a relationship between the particle's composition, size, and uptake has not yet been established	
In vivo ecotoxicity (fish)	Estimated exposure from the consumption of <i>Sufflamen fraenatus</i> , <i>Heniochus acuminatus</i> , <i>Pseudotriacanthus</i> , <i>Leiognathus brevirostris</i> , and <i>Atropus atropus</i> for adults: 456, 310, 213, 156 and 121 particles per week, respectively and children: 68, 45, 32, 25 and 19 of particles per week, respectively	
In vivo, human	Annual ingestion via shellfish: 11,000 particles for the European population. It is also reported that a regular consumer of sea salt ingests approximately 37 synthetic fibres daily	
I <i>n vitro, in vivo</i> , human	Estimated exposure for adults: 258 to 312 particles daily	
In vitro intestinal barrier	Uptake of ≤0.144 % of polystyrene microparticles across the Caco-2 monolayer	(2, 61)
In vitro	Absorption of more than 70 $\%$ of nanoparticles with a significant reduction of rate to 30–50 $\%$ for microparticles	(62)

Table 1 Overview of human-relevant exposure pathways for plastic micro- and nanoparticles with numerical data for levels of intake where available

Models	Type of plastics	PMNP biological effects	Reference
Ecotoxicity, human cells, human exposure	PE-, PS, PVC-, PET- and PLGA- MPs; PS-NPs	Reactive oxygen species (ROS) production (63, 64), activation of antioxidant enzymes (50), increase in glutathione S-transferase (GST) activity (65) and mitogen-activated proteins kinase signalling pathways (66), a decline in lipid digestion and inhibition of digestive enzymatic activities (67), impact on the cell morphology and cell proliferation of immune cells (68, 69)	(36)
Ecotoxicity, human cells, human exposure	PE- and PS- MPs	DNA damage via oxidative stress, distortion of cellular proteins involved in cell division, an aberration in signalling responses, down-regulation of transcriptional genes related to apoptotic expressions (70), increased DNA fragmentation in liver tissue, altered activity of antioxidant enzymes and increased lipid peroxidation (71, 72)	(37)
Ecotoxicity, human cells	PS-MPs; PS-NPs	Disturbance in lipid metabolism, oxidative stress and neurotoxicity (34), aggregation of serum proteins (73)	(5)
Ecotoxicity, human blood cells	PS-NPs	Conformational changes in blood proteins, cytotoxic and genotoxic effects in lymphocytes and erythrocytes	(74)
Ecotoxicity	MPs of few microns or less	Adsorption of proteins and local inflammation in the gastrointestinal system (75)	(27)
Ecotoxicity, human exposure	Nylon fibres of a respirable size; PE-MPs	Persistent inflammation (7)	(41)
Ecotoxicity, animal models, human exposure	PS- and PE- MPs; PS-NPs	Decrease in hepatic triglyceride and total cholesterol levels, decrease in gene expression related to lipogenesis and triglyceride synthesis in liver, reduction of intestinal mucus secretion (76), metabolic disorders due to alteration of intestinal microbiota (77), induction of IL-6 and IL-8 expression in gastric adenocarcinoma cells (68), induction of oxidative stress inT98G cells (63), increased AChE activity in the liver (34)	(42)
In vivo animal models	PS-MPs	Absence of histologically detectable lesions and no inflammatory responses in mice (23)	(43)
In vitro CaCo-2 & THP-1 / T98G & HeLa human cells	PS- and PE- MPs	No cytotoxicity (23, 63)	(39)
In vitro model	PS-NPs	Inflammation, oxidative stress, lysosomal dysfunction and apoptosis in cultured cells	(11)
<i>In vitro</i> (mouse hepatocytes)	PS-NPs	Oxidative stress and DNA damage (78)	(39)
Invertebrates and vertebrates	PS-, PE-, HDPE-MPs	Decreased mucus secretion and mucus secretion-related gene expression (79), down-regulation of genes related to ion transport (76), modified serum levels of IL1 $\alpha$ and granulocyte colony-stimulating factor G-CSF, decreased regulatory T cell count and increased the proportion of Th17 cells in splenocytes (80), blood neutrophil counts and IgA levels elevated in dams, and spleen lymphocytes were altered in both dams and offspring (81)	(46)
<i>In vitro</i> [red blood cells, peripheral blood mononuclear cells (PBMCs), mast cells]	PS-MPs	No cytotoxic effects on PBMCs and mast cells, haemolysis of erythrocytes, increased IL-6 production	(47)
<i>In vitro</i> (intestinal epithelial cell lines, LS174T, HT-29, and Caco-2)	PS-NPs	Reduced cell viability	(22)
Rodents	PS-MPs	Accumulation in the liver, kidney, and gut; energy and lipid metabolism disorders, liver inflammation (34), decreased intestinal mucus, changes in intestinal biota (77, 79)	(82)
<i>In vitro</i> (epithelial HeLa and cerebral T98G cells) and rodents	PS-MPs	Cells: binding of blood plasma coagulation factors VII and IX leading to a decrease in thrombin generation Mice: accumulation in the liver, kidney and gut with evidence of oxidative stress, energy balance disturbance, and neurotoxicity (83, 84)	(51)

# Table 2 Science-based evidence of adverse health effects of plastic micro and nanoparticles (PMNP)

Models	Type of plastics	PMNP biological effects	Reference
In vivo model organisms	PS- and PE- MPs	Blood clots (85), blood cell cytotoxicity (86), oxidative stress (34, 87)	(50)
In vitro (CaCo-2 cells)	PET-NPs	No effect on cell viability and membrane integrity, no significant change in apoptosis and necrosis	(48)
In vivo, animal models	PS-MPs	Altered hepatic lipid metabolism, decrease in body weight, liver weight, decreased serum triglycerides and total cholesterol, mucus secretion, changes in gut microbiota, impairment of bile acid metabolism (77, 79)	(2)
In vivo, animal models	PS-MPs	Mechanical injury, false satiation, low growth rate, increased immune response, energy depletion, blocked enzyme production, decreased fecundity, oxidative stress, morbidity and mortality (88, 89)	(55)
<i>In vivo</i> , animal models	PS-MPs; PS-NPs	Inflammatory response, reduced intestinal mucus secretion, damage to the intestinal barrier function leading to an increase in the permeability of the gut mucosa, trigger an imbalance of gut microbiota, and alter metabolism, such as lipogenesis, triglyceride synthesis (77, 79), induction of pro-inflammatory responses (pro-inflammatory cytokines IL6, IL8 and IL1β), and inhibition of cell viability (68)	(54)
In vivo	LDPE-MPs	No toxic effects	(90)
In vitro (CaCo-2 cells)	PLA particles	No cytotoxicity and no altered cell viability	(9)
<i>In vivo</i> , human	Plastic fibres; PS-MPs	Bioaccumulation of synthetic microfibres in the gastro-intestinal tract and lungs of humans (91), inflammation, genotoxicity, oxidative stress, and apoptosis in the human body (92)	(60)
In vitro, in vivo	PS-MPs	<i>In vitro</i> : no effects on the phosphorylation of STAT-1 and STAT-6, no effect on the expression of CXCL10 and CCL22, CD209 and CD206 genes. Mice – no statistically significant effects on body and organ weights, no effect on tissue morphology	
In vitro (CaCo-2 cells)	PS-MPs; PS-NPs	Increase in intracellular ROS levels, mitochondrial depolarisation, and increased cytotoxicity	(93)
Ecotoxicity	PS-NPs	Up-regulation of cytokines involved in gastric pathologies (68), disruption of iron transport (94), induction of apoptosis (22), endoplasmic reticulum stress (95) and oxidative stress (96)	
In vitro	Molecularly imprinted polymers; PS-NPs	No significant toxicity	(98, 99)

Table 2 continued

HDPE – high-density polyethylene; LDPE – low-density polyethylene; MPs – microplastics; NPs – nanoplastics; PE – polyethylene; PET – polyethylene terephthalate; PLA – polylactic acid; PLGA – poly(lactic-co-glycolic acid); PS – polystyrene; PVC – polyvinyl chloride

## CONCLUSIONS

The reviewed literature clearly evidences adverse health effects of PMNPs, even though they are not fully understood in humans. Many studies demonstrate that PMNPs can accumulate and cause inflammation in various tissues and organs or disrupt cellular processes.

It is important to consider all routes to determine aggregate exposure and include all routes in future studies of PMNP health effects. Risk assessment should also include interactions between PMNPs and other environmental pollutants that may act as a Trojan horse introducing hazardous substances into the body. Research and action are needed to address all these issues and minimise the risks associated with the ever-increasing use of plastic materials. Further studies are needed to fully understand the mechanisms by which PMNPs affect human health and the extent of their toxicity at various exposure levels. Long-term studies are also needed to determine their chronic effects in order to develop effective risk assessment and management strategies and inform policy decisions aimed at minimising exposure to these particles and protecting human health.

#### Conflicts of interest

None to declare.

Table 3 AOPs in AOP-Wiki related to the observed PMNP effects found in the literature search (see Figure 1)[Reported biological effects of PMNP are denoted as molecular initiating events (MIE) or key events (KE) for each AOP. AOPs marked in the last column with an asterisk are "under development"]

Reported biological effect of PMNP	Adverse outcome (AO)	Total number of AOPs	Event type in identified AOPs
	Decreased population growth rate/Decreased population size	8	<b>MIE</b> in AOP 444*; <b>KE</b> in AOP 386*, AOP 387*, AOP 396*, AOP 299*, AOP 311*, AOP 325*, AOP 326*
	Lung fibrosis/Lung cancer/ Decreased lung function/ Dysfunction of the respiratory system	7	MIE in AOP 481*; KE in AOP 382*, AOP 319*, AOP 303*, AOP 416*, AOP 451*, AOP 418*
	Liver fibrosis/Cholestasis/ Steatohepatitis/Liver injury	5	<b>KE</b> in AOP 383*, AOP 27*, AOP 213 (open for adoption), AOP 273*, AOP 494*
	Breast cancer	2	<b>MIE</b> in AOP 294*; <b>KE</b> in AOP 293*
Increased reactive oxygen species (ROS); oxidative stress induction	Increased mortality	7	MIE in AOP 327*, AOP 328*, AOP 329*, AOP 330*, AOP 186*; <b>KE</b> in AOP 377*, AOP 413 (open for citation & comment)
	Reproductive failure/Decreased fertility/ Decreased reproductive success/Impaired fertility	4	<b>KE</b> in AOP 207 <b>*</b> , AOP 476*, AOP 473*, AOP 492*
	Treatment-resistant gastric cancer	1	MIE in AOP 298 (under review)
	Decreased cognitive function	1	MIE in AOP 488*
	Inflammatory events in light- exposed tissues	1	MIE in AOP 282 (under review)
	Insulin resistance	2	<b>KE</b> in AOP 457*, AOP 497*
	Chronic kidney disease	1	<b>KE</b> in AOP 384*
	Apoptotic cell death	1	MIE in AOP 423*
	Increased oxidative damage	1	<b>KE</b> in AOP 26 (open for adoption)
	Increased mesotheliomas	1	<b>KE</b> in AOP 409*
	Acute myeloid leukaemia	1	<b>KE</b> in AOP 432*
	Decreased population growth rate/Decreased population size/ Decreased population trajectory	6	<b>KE</b> in AOP 388 <b>*</b> , AOP 444*, AOP 216*, AOP 238*, AOP 396*, AOP 435*
	Lung cancer	5	<b>KE</b> in AOP 303*, AOP 451*, AOP 416*, AOP 417*, AOP 272 (Approved)
	Breast cancer	4	MIE in AOP 443*, AOP 293*, AOP 294* KE in AOP 200 (open for adoption)
	Decreased fertility/Reduced sperm count	2	<b>KE</b> in AOP 476 <b>*</b> , AOP 322*
DNA damage/DNA strand breaks	Increased mesotheliomas	1	<b>KE</b> in AOP 409*
	Acute myeloid leukaemia	1	<b>KE</b> in AOP 432*
	Microcephaly	1	<b>KE</b> in AOP 441 <b>*</b>
	Apoptotic cell death	1	<b>KE</b> in AOP 423 <b>*</b>
	Increased chromosomal aberrations	1	<b>KE</b> in AOP 296 (endorsed)
	Cataracts	1	<b>KE</b> in AOP 478 (open for citation & comment)
	Learning and memory impairment	1	KE in AOP 483 (open for citations & comments)
	Vascular remodelling	1	KE in AOP 470 (open for citations & comments)
T 11''1 '1'	Increased mortality	2	KE in AOP 413 (open for citation & comment), AOP 329*
Increased lipid peroxidation	Impaired fertility	1	<b>KE</b> in AOP 492*

Reported biological effect of PMNP	Adverse outcome (AO)	Total number of AOPs	Event type in identified AOPs
	Decreased population growth	3	<b>KE</b> in AOP 340*, AOP 341*, AOP 444*
	Decreased fertility/Reproductive failure/Decreased reproduction	4	<b>KE</b> in AOP 70*, AOP 71*, AOP 207*, AOP 476*
	Orofacial cleating	2	<b>KE</b> in AOP 460*, AOP 491*
	Breast cancer	2	KE in AOP 200 (open for adoption), AOP 439*
	Decreased lung function/ Chronic obstructive pulmonary disease	2	<b>KE</b> in AOP 419*, AOP 452*
Apoptosis	Prostate cancer	1	<b>KE</b> in AOP 495*
	Apoptosis/Necrosis	1	AO in AOP 205 (open for comment)
	Testicular atrophy	1	KE in AOP 212 (endorsed)
	Liver injury	1	<b>KE</b> in AOP 285*
	Thyroid hormone interference	1	<b>KE</b> in AOP 393*
	Neurodegeneration	1	<b>KE</b> in AOP 260*
	Kidney failure	1	<b>KE</b> in AOP 447 <b>*</b>
T 1.1' '	Kidney toxicity	1	<b>KE</b> in AOP 257 <b>*</b>
Lysosomal disruption	Liver fibrosis	1	<b>KE</b> in AOP 144 (under review)
Decreased locomotor activity	Decreased population growth rate	1	<b>KE</b> in AOP 218 (not under active development)
Decrease in body weight	Decreased body weight	1	AO in AOP 6 (endorsed)
Glutathione depletion	Impaired fertility	1	<b>KE</b> in AOP 492 <b>*</b>
	Learning and memory impairment	4	<b>KE</b> in AOP 12 (endorsed), AOP 17 (endorsed), AOI 48 (endorsed), AOP 490*
	Parkinsonian motor deficits	2	<b>KE</b> in AOP 3 (endorsed), AOP 464*
	Lung fibrosis/ Bronchiolitis obliterans/ Chronic obstructive pulmonary disease	3	<b>KE</b> in AOP 206*, AOP 280*, AOP 452*
	Cholestasis/Immune-mediated hepatitis/ Increased liver steatosis	2	<b>KE</b> in AOP 413*, AOP 362* <b>MIE</b> in AOP 62*
Inflammation/	Increase, papillomas/ carcinomas	2	<b>KE</b> in AOP 114*, AOP 115*
Neuroinflammation	Psoriatic skin disease	1	<b>KE</b> in AOP 313*
	Increased mortality	1	<b>KE</b> in AOP 377*
	Hyperinflammation	1	<b>AO</b> in AOP 392*
	Memory Loss	1	<b>KE</b> in AOP 429 <b>*</b>
	Kidney failure	1	<b>KE</b> in AOP 447 <b>*</b>
	Metabolically unhealthy obesity	1	<b>KE</b> in AOP 493*
	Increased mesotheliomas	1	<b>KE</b> in AOP 171 <b>*</b>
	Breast cancer	1	<b>KE</b> in AOP 439*
	Chronic obstructive pulmonary disease	1	<b>KE</b> in AOP 452*
	Respiratory dysfunction	1	<b>KE</b> in AOP 481*
Pulmonary inflammation	Increased mesotheliomas	1	<b>KE</b> in AOP 409*
	Increased mortality	1	<b>KE</b> in AOP 377 <b>*</b>
	Hypersensitivity response	1	<b>KE</b> in AOP 39*
	Atherosclerosis	1	<b>KE</b> in AOP 237 <b>*</b>

#### Table 3 continued

Bakan B, et al. Science-based evidence on pathways and effects of human exposure to micro- and nanoplastics Arh Hig Rada Toksikol 2024;75:1-14

Table 3 continued

Total Reported biological effect Adverse outcome (AO) number Event type in identified AOPs of PMNP of AOPs Increased neutrophil activation Increased thrombo-inflammation 1 **KE** in AOP 412\* KE in AOP 77 (open for comment), AOP 78 (open for comment), AOP 79 (open for comment), AOP 80 Death/Failure, Colony 6 (open for comment), AOP 87 (open for comment), AOP 178\* Decreased population growth rate 3 MIE in AOP 326\*, AOP 325\*, AOP 324\* Heart failure/Increased 3 KE in AOP 479\*, AOP 480\*, AOP 377\* mortality Mitochondrial dysfunction 2 **KE** in AOP 447\*, AOP 437\* Kidney failure/Kidney toxicity 2 Liver fibrosis/Liver injury KE in AOP 144 (under review), AOP 273\* 1 Learning and memory impairment KE in AOP 48 (endorsed) Parkinsonian motor deficits 1 KE in AOP 3 (endorsed) 1 Breast cancer KE in AOP 200 (open for adoption) Apoptotic cell death 1 **KE** in AOP 423\* T-cell-dependent antibody Impaired IL-1R signalling 1 MIE in AOP 277 (under review) response impairment Energy reserves depletion/ Decreased population growth rate 1 KE in AOP 97\* Decreased fatty acid beta-Decreased body weight 1 KE in AOP 6 (endorsed) oxidation T-cell-dependent antibody 1 KE in AOP 154 (endorsed) Blocked enzyme production response impairment (inhibition of aromatase/ Metastasis, Breast Cancer 1 KE in AOP 443\* calcineurin activity) Decreased fertility 1 MIE in AOP 153\* AO in AOP 73\*, AOP 126\*, AOP 345\*, AOP 476\*, Decreased fecundity/Decreased Decreased fecundity/ AOP 64\*, AOP 66\*, AOP 67\*, AOP 68\*, AOP 70\*, 13 population growth rate/ AOP 71\*, AOP 74\*, AOP 153\* fertility Decreased fertility KE in AOP 444\* Oxidation of iron in 1 MIE in AOP 31\* Cyanosis haemoglobin Non-alcoholic fatty liver 1 **KE** in AOP 454\* disease/Tumorigenesis Endoplasmic reticulum stress Parkinsonian motor deficits 1 KE in AOP 464\* 2 **KE** in AOP 51\*, AOP 18 (under review) Impaired fertility Decreased cholesterol/altered Decreased sperm quantity and/ 1 MIE/KE in AOP 69\* cholesterol metabolism or quality in the adult testis Decreased cognitive function 1 KE in AOP 487\* 1 KE in AOP 401\* Steatohepatitis Altered lipid metabolism Pancreatic acinar tumours 1 KE in AOP 166\* Metabolic syndrome/Insulin 1 AO in AOP 497\* resistance Metabolic syndrome/stress 1 KE in AOP 81 (open for comment) Death/Failure, Colony Increased blood CCK level Pancreatic acinar cell tumours 1 KE in AOP 316\* (satiety) KE in AOP 48 (endorsed), AOP 13 (endorsed), AOP Learning and memory impairment 5 12 (endorsed), AOP 17 (endorsed), AOP 490\* KE in AOP 38 (endorsed), AOP 144 (under review), Liver fibrosis/Liver injury/ 5 AOP 278\*, AOP 273\*, AOP 494\* Cell injury 3 KE in AOP 265\*, AOP 264\*, AOP 266\* Decreased growth Heart failure/Increased mortality 2 KE in AOP 479\*, AOP 377\*

1

Neurodegeneration

KE in AOP 281\*

Reported biological effect of PMNP	Adverse outcome (AO)	Total number of AOPs	Event type in identified AOPs
Inhibition of digestive enzymatic activities (trypsin inhibition)	Pancreatic acinar cell tumours	1	<b>MIE</b> in AOP 316*
Alteration in intestinal microbiota	Gut dysbiosis	1	<b>KE</b> in AOP 428*
Decrease in gene expression related to liver	Increased hepatocellular adenomas and carcinomas	1	<b>KE</b> in AOP 107 (under review)
	Sensitisation of skin	1	MIE in AOP 40 (endorsed)
Covalent protein binding	Increased allergic respiratory hypersensitivity response	1	MIE in AOP 39*
Meiotic spindle disorganisation	Increased aneuploid offspring	1	<b>KE</b> in AOP 106 (EAGMST under review)
Increased mortality	Increased mortality	23	AO in AOP 16*, AOP 96*, AOP 104*, AOP 113*, AOP 160*, AOP 161*, AOP 138*, AOP 177*, AOP 186*, AOP 312*, AOP 155 (endorsed), AOP 156 (endorsed), AOP 157 (endorsed), AOP 158 (endorsed), AOP 159 (endorsed), AOP 363 (under review), AOP 377*, AOP 364 *, AOP 365 *, AOP 399 (open for citation & comment), AOP 413 (open for citation & comment), AOP 410 (open for citation & comment), AOP 450*
Decreased population growth rate	Decreased population growth rate	54	AO in AOP 23 (endorsed), AOP 25 (endorsed), AOP 29*, AOP 30 (under review), AOP 100*, AOP 122(under development), AOP 123*, AOP 155 (endorsed), AOP 156 (endorsed), AOP 157 (endorsed), AOP 158 (endorsed), AOP 159 (endorsed), AOP 101*, AOP 312*, AOP 63*, AOP 103*, AOP 310*, AOP 16*, AOP 312*, AOP 334*, AOP 336*, AOP 337*, AOP 338*, AOP 339*, AOP 340*, AOP 341*, AOP 297*, AOP 346*, AOP 326*, AOP 325*, AOP 324*, AOP 363 (under review), AOP 348*, AOP 376 (open for citation & comment), AOP 386*, AOP 387*, AOP 388*, AOP 389*, AOP 364*, AOP 365*, AOP 399 (open for citation & comment), AOP 410 (open for citation & comment), AOP 216 *, AOP 238*, AOP 299*, AOP 311*, AOP 444*, AOP 138*, AOP 177*, AOP 97*, AOP 203*, AOP 218 (not under active development), AOP 219 (not under active development), AOP 323 (open for citation & comment)

#### Table 3 continued

AOP – adverse outcome pathway; KE – key event; MIE – molecular initiating event

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### REFERENCES

- Kumar R, Verma A, Shome A, Sinha R, Sinha S, Jha PK, Kumar R, Kumar P, Shubham, Das S, Sharma P, Prasad PVV. Impacts of plastic pollution on ecosystem services, sustainable development goals, and need to focus on circular economy and policy interventions. Sustain 2021;13:9963. doi: 10.3390/su13179963
- Paul MB, Stock V, Cara-Carmona J, Lisicki E, Shopova S, Fessard V, Braeuning A, Sieg H, Böhmert L. Micro- and nanoplastics – current state of knowledge with the focus on oral uptake and toxicity. Nanoscale Adv 2020;2:4350–67. doi: 10.1039/D0NA00539H
- Sharma S, Chatterjee S. Microplastic pollution, a threat to marine ecosystem and human health: a short review. Environ Sci Pollut Res Int 2017;24:21530–47. doi: 10.1007/s11356-017-9910-8
- Lehner R, Weder C, Petri-Fink A, Rothen-Rutishauser B. Emergence of nanoplastic in the environment and possible impact on human health. Environ Sci Technol 2019;53:1748–65. doi: 10.1021/acs. est.8b05512
- Allouzi MMA, Tang DYY, Chew KW, Rinklebe J, Bolan N, Allouzi SMA, Show PL. Micro (nano) plastic pollution: The ecological influence on soil-plant system and human health. Sci Total Environ 2021;788:147815. doi: 10.1016/j.scitotenv.2021.147815

- Barboza LGA, Dick Vethaak A, Lavorante BRBO, Lundebye AK, Guilhermino L. Marine microplastic debris: An emerging issue for food security, food safety and human health. Mar Pollut Bull 2018;133:336–48. doi: 10.1016/j.marpolbul.2018.05.047
- Wright SL, Kelly FJ. Plastic and human health: A micro issue? Environ Sci Technol 2017;51:6634–47. doi: 10.1021/acs.est.7b00423
- Briffa SM. Looking at the bigger picture-considering the hurdles in the struggle against nanoplastic pollution. Nanomaterials (Basel) 2021;11(10):2536. doi: 10.3390/nano11102536
- Shopova S, Sieg H, Braeuning A. Risk assessment and toxicological research on micro- and nanoplastics after oral exposure via food products. EFSA J 2020;18(S1): e181102. doi: 10.2903/j.efsa.2020. e181102
- Rainieri S, Barranco A. Microplastics, a food safety issue? Trends Food Sci Technol 2019;84:55–7. doi: 10.1016/j.tifs.2018.12.009
- Heddagaard FE, Møller P. Hazard assessment of small-size plastic particles: is the conceptual framework of particle toxicology useful? Food Chem Toxicol 2020;136:111106. doi: 10.1016/j.fct.2019.111106
- OECD. Overview of Concepts and Available Guidance related to Integrated Approaches to Testing and Assessment (IATA), Series on Testing and Assessment No. 329. Paris: OECD; 2020 [displayed 7 June 2023]. Available at https://www.oecd.org/chemicalsafety/riskassessment/concepts-and-available-guidance-related-to-integratedapproaches-to-testing-and-assessment.pdf
- OECD. Users' Handbook supplement to the Guidance Document for developing and assessing Adverse Outcome Pathways. Paris: OECD; 2016 [displayed 7 June 2023]. Available at https://www.oecdilibrary.org/docserver/5jlv1m9d1g32-en.pdf?expires= 1709568200&id=id&accname=guest&checksum=9BC272DE8E3A 4E49221B07080FA6BC9A
- WHO. Microplastics in drinking water. Geneva: WHO; 2019 [displayed 8 June 2023]. Available at https://iris.who.int/bitstream/hand le/10665/326499/9789241516198-eng.pdf?sequence=5&is Allowed=y
- 15. EUR-Lex. A European Strategy for Plastics in a Circular Economy. Communication from The Commission to The European Parliament, The Council, The European Economic and Social Committee and The Committee of The Regions. Brusseles: EC; 2018 [displayed 8 June 2023]. Available at https://eur-lex.europa.eu/legal-content/EN/ TXT/?uri=COM%3A2018%3A28%3AFIN
- AOP-Wiki. Adverse Outcome Pathway Wiki [displayed 8 June 2023]. Available at https://aopwiki.org/
- 17. Bajard L, Adamovsky O, Audouze K, Baken K, Barouki R, Beltman JB, Beronius A, Bonefeld-Jørgensen EC, Cano-Sancho G, de Baat ML, Di Tillio F, Fernández MF, FitzGerald RE, Gundacker C, Hernández AF, Hilscherova K, Karakitsios S, Kuchovska E, Long M, Luijten M, Majid S, Marx-Stoelting P, Mustieles V, Negi CK, Sarigiannis D, Scholz S, Sovadinova I, Stierum R, Tanabe S, Tollefsen KE, van den Brand AD, Vogs C, Wielsøe M, Wittwehr C, Blaha L. Application of AOPs to assist regulatory assessment of chemical risks Case studies, needs and recommendations. Environ Res 2023;217:114650. doi: 10.1016/j. envres.2022.114650
- Russell WMS, Burch RL. The Principles of Humane Experimental Technique. London: Methuen & Co. Ltd.; 1959.
- van der Zalm AJ, Barroso J, Browne P, Casey W, Gordon J, Henry TR, Kleinstreuer NC, Lowit AB, Perron M, Clippinger AJ. A framework for establishing scientific confidence in new approach methodologies. Arch Toxicol 2022;96:2865–79. doi: 10.1007/s00204-022-03365-4

- Zarus GM, Muianga C, Hunter CM, Pappas RS. A review of data for quantifying human exposures to micro and nanoplastics and potential health risks. Sci Total Environ 2021;756:144010. doi: 10.1016/j. scitotenv.2020.144010
- Yee MS-L, Hii L-W, Looi CK, Lim W-M, Wong S-F, Kok Y-Y, Tan B-K, Wong C-Y, Leong C-O. Impact of microplastics and nanoplastics on human health. Nanomaterials (Basel) 2021;11(2):496. doi: 10.3390/ nano11020496
- 22. Inkielewicz-Stepniak I, Tajber L, Behan G, Zhang H, Radomski MW, Medina C, Santos-Martinez MJ. The role of mucin in the toxicological impact of polystyrene nanoparticles. Materials (Basel) 2018;11(5):724. doi: 10.3390/ma11050724
- Stock V, Böhmert L, Lisicki E, Block R, Cara-Carmona J, Pack LK, Selb R, Lichtenstein D, Voss L, Henderson CJ, Zabinsky E, Sieg H, Braeuning A, Lampen A. Uptake and effects of orally ingested polystyrene microplastic particles *in vitro* and *in vivo*. Arch Toxicol 2019;93:1817–33. doi: 10.1007/s00204-019-02478-7
- Liebezeit G, Liebezeit E. Synthetic particles as contaminants in German beers. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 2014;31:1574–8. doi: 10.1080/19440049.2014.945099
- Yang D, Shi H, Li L, Li J, Jabeen K, Kolandhasamy P. Microplastic pollution in table salts from China. Environ Sci Technol 2015;49:13622– 7. doi: 10.1021/acs.est.5b03163
- Stapleton PA. Toxicological considerations of nano-sized plastics. AIMS Environ Sci 2019;6:367–78. doi: 10.3934/environsci.2019.5.367
- Cox KD, Covernton GA, Davies HL, Dower JF, Juanes F, Dudas SE. Human consumption of microplastics. Environ Sci Technol 2019;53:7068–74. doi: 10.1021/acs.est.9b01517
- Revel M, Châtel A, Mouneyrac C. Micro(nano)plastics: A threat to human health? Curr Opin Environ Sci Heal 2018;1:17–23. doi: 10.1016/j.coesh.2017.10.003
- Sangkham S, Faikhaw O, Munkong N, Sakunkoo P, Arunlertaree C, Chavali M, Mousazadeh M, Tiwari A. A review on microplastics and nanoplastics in the environment: Their occurrence, exposure routes, toxic studies, and potential effects on human health. Mar Pollut Bull 2022;181:113832. doi: 10.1016/j.marpolbul.2022.113832
- Schneider M, Stracke F, Hansen S, Schaefer UF. Nanoparticles and their interactions with the dermal barrier. Dermatoendocrinol 2009;1:197–206. doi: 10.4161/derm.1.4.9501
- Chang X, Xue Y, Li J, Zou L, Tang M. Potential health impact of environmental micro- and nanoplastics pollution. J Appl Toxicol 2020;40:4–15. doi: 10.1002/jat.3915
- 32. World Health Organization. Call for experts on human health risks from exposure to microplastic, 2020 [displayed 11 March 2024]. Available at https://www.who.int/news-room/articles-detail/call-forexperts-on-human-health-risks-from-exposure-to-micropalstic
- 33. Bradney L, Wijesekara H, Palansooriya KN, Obadamudalige N, Bolan NS, Ok YS, Rinklebe J, Kim KH, Kirkham MB. Particulate plastics as a vector for toxic trace-element uptake by aquatic and terrestrial organisms and human health risk. Environ Int 2019;131:104937. doi: 10.1016/j.envint.2019.104937
- Deng Y, Zhang Y, Lemos B, Ren H. Tissue accumulation of microplastics in mice and biomarker responses suggest widespread health risks of exposure. Sci Rep 2017;7(1):46687. doi: 10.1038/ srep46687
- van Raamsdonk LWD, van der Zande M, Koelmans AA, Hoogenboom RLAP, Peters RJB, Groot MJ, Peijnenburg AACM, Weesepoel YJA. Current insights into monitoring, bioaccumulation, and potential

health effects of microplastics present in the food chain. Foods 2020;9(1):72. doi: 10.3390/foods9010072

- Ali I, Cheng Q, Ding T, Yiguang Q, Yuechao Z, Sun H, Peng C, Naz I, Li J, Liu J. Micro- and nanoplastics in the environment: Occurrence, detection, characterization and toxicity – A critical review. J Clean Prod 2021;313:127863. doi: 10.1016/j.jclepro.2021.127863
- Alimba CG, Faggio C, Sivanesan S, Ogunkanmi AL, Krishnamurthi K. Micro(nano)-plastics in the environment and risk of carcinogenesis: Insight into possible mechanisms. J Hazard Mater 2021;416:126143. doi: 10.1016/j.jhazmat.2021.126143
- Bouwmeester H, Hollman PCH, Peters RJB. Potential health impact of environmentally released micro- and nanoplastics in the human food production chain: Experiences from nanotoxicology. Environ Sci Technol 2015;49:8932–47. doi: 10.1021/acs.est.5b01090
- Barbosa F, Adeyemi JA, Bocato MZ, Comas A, Campiglia A. A critical viewpoint on current issues, limitations, and future research needs on micro- and nanoplastic studies: From the detection to the toxicological assessment. Environ Res 2020;182:109089. doi: 10.1016/j. envres.2019.109089
- Hesler M, Aengenheister L, Ellinger B, Drexel R, Straskraba S, Jost C, Wagner S, Meier F, von Briesen H, Büchel C, Wick P, Buerki-Thurnherr T, Kohl Y. Multi-endpoint toxicological assessment of polystyrene nano- and microparticles in different biological models *in vitro*. Toxicol In Vitro 2019;61:104610. doi: 10.1016/j.tiv.2019.104610
- Fred-Ahmadu OH, Bhagwat G, Oluyoye I, Benson NU, Ayejuyo OO, Palanisami T. Interaction of chemical contaminants with microplastics: Principles and perspectives. Sci Total Environ 2020;706:135978. doi: 10.1016/j.scitotenv.2019.135978
- Campanale C, Massarelli C, Savino I, Locaputo V, Uricchio VF. A detailed review study on potential effects of microplastics and additives of concern on human health. Int J Environ Res Public Health 2020;17(4):1212. doi: 10.3390/ijerph17041212
- Das RK, Sanyal D, Kumar P, Pulicharla R, Brar SK. Science-societypolicy interface for microplastic and nanoplastic: Environmental and biomedical aspects. Environ Pollut 2021;290:117985. doi: 10.1016/j. envpol.2021.117985
- De Nicola M, Mirabile Gattia D, Traversa E, Ghibelli L. Maturation and demise of human primary monocytes by carbon nanotubes. J Nanoparticle Res 2013;15(6):1711. doi: 10.1007/s11051-013-1711-5
- 45. Dobaradaran S, Akhbarizadeh R, Javad Mohammadi M, Izadi A, Keshtkar M, Tangestani M, Moazzen M, Shariatifar N, Mahmoodi M. Determination of phthalates in bottled milk by a modified nano adsorbent: Presence, effects of fat and storage time, and implications for human health. Microchem J 2020;159:105516. doi: 10.1016/j. microc.2020.105516
- Hirt N, Body-Malapel M. Immunotoxicity and intestinal effects of nano- and microplastics: a review of the literature. Part Fibre Toxicol 2020;17(1):57. doi: 10.1186/s12989-020-00387-7
- Hwang J, Choi D, Han S, Jung SY, Choi J, Hong J. Potential toxicity of polystyrene microplastic particles. Sci Rep 2020;10(1):7391. doi: 10.1038/s41598-020-64464-9
- Magrì D, Sánchez-Moreno P, Caputo G, Gatto F, Veronesi M, Bardi G, Catelani T, Guarnieri D, Athanassiou A, Pompa PP, Fragouli D. Laser ablation as a versatile tool to mimic polyethylene terephthalate nanoplastic pollutants: Characterization and toxicology assessment. ACS Nano 2018;12:7690–700. doi: 10.1021/acsnano.8b01331
- Jeong J, Choi J. Development of AOP relevant to microplastics based on toxicity mechanisms of chemical additives using ToxCast<sup>TM</sup> and

deep learning models combined approach. Environ Int 2020;137:105557. doi: 10.1016/j.envint.2020.105557

- Jeong C-B, Won E-J, Kang H-M, Lee M-C, Hwang D-S, Hwang U-K, Zhou B, Souissi S, Lee S-J, Lee J-S. Microplastic size-dependent toxicity, oxidative stress induction, and p-JNK and p-p38 activation in the monogonont rotifer (*Brachionus koreanus*). Environ Sci Technol 2016;50:8849–57. doi: 10.1021/acs.est.6b01441
- Kik K, Bukowska B, Sicińska P. Polystyrene nanoparticles: Sources, occurrence in the environment, distribution in tissues, accumulation and toxicity to various organisms. Environ Pollut 2020;262:114297. doi: 10.1016/j.envpol.2020.114297
- Kirstein IV, Gomiero A, Vollertsen J. Microplastic pollution in drinking water. Curr Opin Toxicol 2021;28:70–5. doi: 10.1016/j. cotox.2021.09.003
- Liao Y-L, Yang J-Y. Microplastic serves as a potential vector for Cr in an *in-vitro* human digestive model. Sci Total Environ 2020;703:134805. doi: 10.1016/j.scitotenv.2019.134805
- Matthews S, Mai L, Jeong CB, Lee JS, Zeng EY, Xu EG. Key mechanisms of micro- and nanoplastic (MNP) toxicity across taxonomic groups. Comp Biochem Physiol Part - C Toxicol Pharmacol 2021;247:109056. doi: 10.1016/j.cbpc.2021.109056
- Petersen F, Hubbart JA. The occurrence and transport of microplastics: The state of the science. Sci Total Environ 2021;758:143936. doi: 10.1016/j.scitotenv.2020.143936
- Borm PJ, Robbins D, Haubold S, Kuhlbusch T, Fissan H, Donaldson K, Schins R, Stone V, Kreyling W, Lademann J, Krutmann J, Warheit D, Oberdorster E. The potential risks of nanomaterials: a review carried out for ECETOC. Part Fibre Toxicol 2006;3:11. doi: 10.1186/1743-8977-3-11
- 57. Rahman A, Sarkar A, Yadav OP, Achari G, Slobodnik J. Potential human health risks due to environmental exposure to nano- and microplastics and knowledge gaps: A scoping review. Sci Total Environ 2021;757:143872. doi: 10.1016/j.scitotenv.2020.143872
- Sana SS, Dogiparthi LK, Gangadhar L, Chakravorty A, Abhishek N. Effects of microplastics and nanoplastics on marine environment and human health. Environ Sci Pollut Res Int 2020;27:44743–56. doi: 10.1007/s11356-020-10573-x
- Selvam S, Manisha A, Roy PD, Venkatramanan S, Chung SY, Muthukumar P, Jesuraja K, Elgorban AM, Ahmed B, Elzain HE. Microplastics and trace metals in fish species of the Gulf of Mannar (Indian Ocean) and evaluation of human health. Environ Pollut 2021;291:118089. doi: 10.1016/j.envpol.2021.118089
- Singh RP, Mishra S, Das AP. Synthetic microfibers: Pollution toxicity and remediation. Chemosphere 2020;257:127199. doi: 10.1016/j. chemosphere.2020.127199
- Vardakas P, Skaperda Z, Tekos F, Trompeta A-F, Tsatsakis A, Charitidis CA, Kouretas D. An integrated approach for assessing the *in vitro* and *in vivo* redox-related effects of nanomaterials. Environ Res 2021;197:111083. doi: 10.1016/j.envres.2021.111083
- Li P, Wang X, Su M, Zou X, Duan L, Zhang H. Characteristics of plastic pollution in the environment: A review. Bull Environ Contam Toxicol 2021;107:577–84. doi: 10.1007/s00128-020-02820-1
- Schirinzi GF, Pérez-Pomeda I, Sanchís J, Rossini C, Farré M, Barceló D. Cytotoxic effects of commonly used nanomaterials and microplastics on cerebral and epithelial human cells. Environ Res 2017;159:579–87. doi: 10.1016/j.envres.2017.08.043
- Alomar C, Sureda A, Capó X, Guijarro B, Tejada S, Deudero S. Microplastic ingestion by *Mullus surmuletus* Linnaeus, 1758 fish and its

potential for causing oxidative stress. Environ Res 2017;159:135–42. doi: 10.1016/j.envres.2017.07.043

- 65. Jeong CB, Kang HM, Lee MC, Kim DH, Han J, Hwang DS, Souissi S, Lee SJ, Shin KH, Park HG, Lee JS. Adverse effects of microplastics and oxidative stress-induced MAPK/Nrf2 pathway-mediated defense mechanisms in the marine copepod *Paracyclopina nana*. Sci Rep 2017;7:41323. doi: 10.1038/srep41323
- Yu P, Liu Z, Wu D, Chen M, Lv W, Zhao Y. Accumulation of polystyrene microplastics in juvenile *Eriocheir sinensis* and oxidative stress effects in the liver. Aquat Toxicol 2018;200:28–36. doi: 10.1016/j. aquatox.2018.04.015
- Tan H, Yue T, Xu Y, Zhao J, Xing B. Microplastics reduce lipid digestion in simulated human gastrointestinal system. Environ Sci Technol 2020;54:12285–94. doi: 10.1021/acs.est.0c02608
- Forte M, Iachetta G, Tussellino M, Carotenuto R, Prisco M, De Falco M, Laforgia V, Valiante S. Polystyrene nanoparticles internalization in human gastric adenocarcinoma cells. Toxicol In Vitro 2016;31:126–36. doi: 10.1016/j.tiv.2015.11.006
- Xia T, Kovochich M, Liong M, Zink JI, Nel AE. Cationic polystyrene nanosphere toxicity depends on cell-specific endocytic and mitochondrial injury pathways. ACS Nano 2008;2:85–96. doi: 10.1021/ nn700256c
- Avio CG, Gorbi S, Milan M, Benedetti M, Fattorini D, D'Errico G, Pauletto M, Bargelloni L, Regoli F. Pollutants bioavailability and toxicological risk from microplastics to marine mussels. Environ Pollut 2015;198:211–22. doi: 10.1016/j.envpol.2014.12.021
- Liu Z, Yu P, Cai M, Wu D, Zhang M, Chen M, Zhao Y. Effects of microplastics on the innate immunity and intestinal microflora of juvenile *Eriocheir sinensis*. Sci Total Environ 2019;685:836–46. doi: 10.1016/j.scitotenv.2019.06.265
- Qiao R, Sheng C, Lu Y, Zhang Y, Ren H, Lemos B. Microplastics induce intestinal inflammation, oxidative stress, and disorders of metabolome and microbiome in zebrafish. Sci Total Environ 2019;662:246–53. doi: 10.1016/j.scitotenv.2019.01.245
- Mohr K, Sommer M, Baier G, Schöttler S, Okwieka P, Tenzer S, Landfester K, Mailänder V, Schmidt M, Meyer RG. Aggregation behavior of polystyrene-nanoparticles in human blood serum and its impact on the *in vivo* distribution in mice. J Nanomed Nanotechnol 2014;5(2):1000193. doi: 10.4172/2157-7439.1000193
- 74. Gopinath PM, Saranya V, Vijayakumar S, Mythili Meera M, Ruprekha S, Kunal R, Pranay A, Thomas J, Mukherjee A, Chandrasekaran N. Assessment on interactive prospectives of nanoplastics with plasma proteins and the toxicological impacts of virgin, coronated and environmentally released-nanoplastics. Sci Rep 2019;9(1):8860. doi: 10.1038/s41598-019-45139-6
- Powell JJ, Thoree V, Pele LC. Dietary microparticles and their impact on tolerance and immune responsiveness of the gastrointestinal tract. Br J Nutr 2007;98(Suppl 1):59–63. doi: 10.1017/S0007114507832922
- Jin Y, Xia J, Pan Z, Yang J, Wang W, Fu Z. Polystyrene microplastics induce microbiota dysbiosis and inflammation in the gut of adult zebrafish. Environ Pollut 2018;235:322–9. doi: 10.1016/j. envpol.2017.12.088
- Jin Y, Lu L, Tu W, Luo T, Fu Z. Impacts of polystyrene microplastic on the gut barrier, microbiota and metabolism of mice. Sci Total Environ 2019;649:308–17. doi: 10.1016/j.scitotenv.2018.08.353
- Zheng T, Yuan D, Liu C. Molecular toxicity of nanoplastics involving in oxidative stress and desoxyribonucleic acid damage. J Mol Recognit 2019;32(11):e2804. doi: 10.1002/jmr.2804

- Lu L, Wan Z, Luo T, Fu Z, Jin Y. Polystyrene microplastics induce gut microbiota dysbiosis and hepatic lipid metabolism disorder in mice. Sci Total Environ 2018;631–632:449–58. doi: 10.1016/j. scitotenv.2018.03.051
- Li L-L, Amara R, Souissi S, Dehaut A, Duflos G, Monchy S. Impacts of microplastics exposure on mussel (*Mytilus edulis*) gut microbiota. Sci Total Environ 2020;745:141018. doi: 10.1016/j. scitotenv.2020.141018
- Park E-J, Han J-S, Park E-J, Seong E, Lee G-H, Kim D-W, Son H-Y, Han H-Y, Lee B-S. Repeated-oral dose toxicity of polyethylene microplastics and the possible implications on reproduction and development of the next generation. Toxicol Lett 2020;324:75–85. doi: 10.1016/j.toxlet.2020.01.008
- Jiang B, Kauffman AE, Li L, McFee W, Cai B, Weinstein J, Lead JR, Chatterjee S, Scott GI, Xiao S. Health impacts of environmental contamination of micro- and nanoplastics: A review. Environ Health Prev Med 2020;25(1):29. doi: 10.1186/s12199-020-00870-9
- Ašmonaitė G, Sundh H, Asker N, Carney Almroth B. Rainbow trout maintain intestinal transport and barrier functions following exposure to polystyrene microplastics. Environ Sci Technol 2018;52:14392–401. doi: 10.1021/acs.est.8b04848
- Wang Y, Zhang D, Zhang M, Mu J, Ding G, Mao Z, Cao Y, Jin F, Cong Y, Wang L, Zhang W, Wang J. Effects of ingested polystyrene microplastics on brine shrimp, *Artemia parthenogenetica*. Environ Pollut 2019;244:715–22. doi: 10.1016/j.envpol.2018.10.024
- Jones AE, Watts JA, Debelak JP, Thornton LR, Younger JG, Kline JA. Inhibition of prostaglandin synthesis during polystyrene microsphere-induced pulmonary embolism in the rat. Am J Physiol Cell Mol Physiol 2003;284(6):L1072–81. doi: 10.1152/ ajplung.00283.2002
- Çobanoğlu H, Belivermiş M, Sıkdokur E, Kılıç Ö, Çayır A. Genotoxic and cytotoxic effects of polyethylene microplastics on human peripheral blood lymphocytes. Chemosphere 2021;272:129805. doi: 10.1016/j.chemosphere.2021.129805
- Lu Y, Zhang Y, Deng Y, Jiang W, Zhao Y, Geng J, Ding L, Ren H. Uptake and accumulation of polystyrene microplastics in zebrafish (*Danio rerio*) and toxic effects in liver. Environ Sci Technol 2016;50:4054–60. doi: 10.1021/acs.est.6b00183
- Wright SL, Thompson RC, Galloway TS. The physical impacts of microplastics on marine organisms: A review. Environ Pollut 2013;178:483–92. doi: 10.1016/j.envpol.2013.02.031
- Sussarellu R, Suquet M, Thomas Y, Lambert C, Fabioux C, Pernet ME, Le Goïc N, Quillien V, Mingant C, Epelboin Y, Corporeau C, Guyomarch J, Robbens J, Paul-Pont I, Soudant P, Huvet A. Oyster reproduction is affected by exposure to polystyrene microplastics. Proc Natl Acad Sci U S A 2016;113:2430–5. doi: 10.1073/ pnas.1519019113
- Rainieri S, Conlledo N, Larsen BK, Granby K, Barranco A. Combined effects of microplastics and chemical contaminants on the organ toxicity of zebrafish (*Danio rerio*). Environ Res 2018;162:135–43. doi: 10.1016/j.envres.2017.12.019
- Henry B, Laitala K, Klepp IG. Microfibres from apparel and home textiles: Prospects for including microplastics in environmental sustainability assessment. Sci Total Environ 2019;652:483–94. doi: 10.1016/j.scitotenv.2018.10.166
- 92. Prata JC, da Costa JP, Lopes I, Duarte AC, Rocha-Santos T. Environmental exposure to microplastics: An overview on possible

human health effects. Sci Total Environ 2020;702:134455. doi: 10.1016/j.scitotenv.2019.134455

- 93. Wang Q, Bai J, Ning B, Fan L, Sun T, Fang Y, Wu J, Li S, Duan C, Zhang Y, Liang J, Gao Z. Effects of bisphenol A and nanoscale and microscale polystyrene plastic exposure on particle uptake and toxicity in human Caco-2 cells. Chemosphere 2020;254:126788. doi: 10.1016/j. chemosphere.2020.126788
- Mahler GJ, Esch MB, Tako E, Southard TL, Archer SD, Glahn RP, Shuler ML. Oral exposure to polystyrene nanoparticles affects iron absorption. Nat Nanotechnol 2012;7:264–71. doi: 10.1038/ nnano.2012.3
- Chiu H-W, Xia T, Lee Y-H, Chen C-W, Tsai J-C, Wang Y-J. Cationic polystyrene nanospheres induce autophagic cell death through the induction of endoplasmic reticulum stress. Nanoscale 2015;7:736–46. doi: 10.1039/C4NR05509H
- 96. Ruenraroengsak P, Tetley TD. Differential bioreactivity of neutral, cationic and anionic polystyrene nanoparticles with cells from the

human alveolar compartment: robust response of alveolar type 1 epithelial cells. Part Fibre Toxicol 2015;12:19. doi: 10.1186/s12989-015-0091-7

- 97. Wang L, Wu W, Bolan NS, Tsang DCW, Li Y, Qin M, Hou D. Environmental fate, toxicity and risk management strategies of nanoplastics in the environment: Current status and future perspectives. J Hazard Mater 2021;401:123415. doi: 10.1016/j. jhazmat.2020.123415
- Roshanzadeh A, Park S, Ganjbakhsh SE, Park J, Lee D-H, Lee S, Kim E-S. Surface charge-dependent cytotoxicity of plastic nanoparticles in alveolar cells under cyclic stretches. Nano Lett 2020;20:7168–76. doi: 10.1021/acs.nanolett.0c02463
- Canfarotta F, Waters A, Sadler R, McGill P, Guerreiro A, Papkovsky D, Haupt K, Piletsky S. Biocompatibility and internalization of molecularly imprinted nanoparticles. Nano Res 2016;9:3463–77. doi: 10.1007/s12274-016-1222-7

#### Znanstveno utemeljeni dokazi o načinima i učincima ljudske izloženosti mikro- i nanoplastici

Izloženost ljudi česticama plastike izaziva veliku zabrinutost svih značajnih sudionika koji su uključeni u zaštitu ljudskoga zdravlja zbog onečišćenja prehrambenog lanca, površinskih voda, čak i pitke vode, kao i zbog njihove postojanosti i bioakumulacije. Stoga je ključno bolje razumijevanje biološke sudbine plastike i njezine interakcije s različitim biološkim sustavima. Zbog učestalosti materijala plastike u okolišu i njihove potencijalne toksičnosti, važno je prikupiti pouzdane, regulatorno značajne i znanstveno utemeljene podatke o učincima mikro- i nanoplastike na ljudsko zdravlje kako bi se pravilno primijenile pouzdane strategije procjene rizika i upravljanja rizicima u kružnoj ekonomiji lanca vrijednosti plastike. Ovaj pregledni rad predstavlja trenutačno znanje o dozama izloženosti mikro- i nanoplastici relevantnima za ljude, oblicima izloženosti, kao i o njezinim toksičnim učincima. Istaknute su poteškoće u pravilnoj procjeni problema izloženosti plastici i trenutačni nedostatci u znanju te su predloženi potencijalni koraci koji se mogu poduzeti kako bi se podržala percepcija, procjena i ublažavanje rizika za zdravlje strogim znanstveno utemeljenim dokazima. Na temelju postojećih znanstvenih podataka o štetnim učincima mikro- i nanoplastike na zdravlje, daju se preporuke za razvoj specifičnih načina nepoželjnih ishoda (eng. *adverse outcome pathway*, krat. AOP) za mikro- i nanoplastiku prema priručniku korisnika AOP-a Organizacije za ekonomsku suradnju i razvoj (izv. Organisation for Economic Co-operation and Development, krat. OECD).

KLJUČNE RIJEČI: materijali plastike; nepovoljan ishod; procjena rizika; regulacijski značajni podatci; upravljanje rizikom