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Approaches for shelf life extension of milk and milk products: at a glance

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ABSTRACT

Milk, as an almost complete food, has been a necessity of infants since their birth. Its balanced nutritional composition acts as a great site for microorganisms. Thus, the threat of its spoilage always persists as soon as it leaves the udder. The preservation of milk has been a concern since the development of humankind to keep it safe for consumption after a while. Earlier development of humans raised the idea of the conversion of milk into different forms i.e. curd, cheese, etc. In the later centuries, the beginning of dairy industry started using various means and technologies to eliminate the contamination from milk and milk products to overcome the issue of shelf life and to assure safety and quality. The heat treatment came in the most popular form as pasteurization and sterilization. However, due to the adverse effect of heat on the nutrition of milk the dairy industry strives for innovation in the process and technology. The current consumer demand for minimally processed, gualitative, and nutritional food makes the application of non-thermal technologies such as highpressure processing (HPP), pulsed electric field (PEF), cold plasma, irradiation, ultrasonication, ohmic heating, microfiltration, bactofugation, etc. to enhance shelf life of products. In this review article, the role of three major aspects of the dairy industry viz. milk processing, use of additives, and various packaging techniques pertaining to the enhancement of the shelf life of milk and milk products are discussed.

KEYWORDS

dairy processing; cold plasma; Ohmic heating; ultrasound; high-pressure processing

KEY CONTRIBUTION

Our article comprehensively discusses the advances in thermal and non-thermal techniques used for milk processing; (ii) discusses the advantages and limitations of all methods, and (iii) performs a comparative analysis of various methods.

Introduction

Food is the basic necessity of life and amongst it, milk plays a vital role from the birth until death of a human being. It is the only food that is considered as an almost complete food on the earth and hence, the sole source of nutrition for new borns (Adeniji and Eyinla, 2019). According to a report issued by OECD-FAO world milk production is about to reach 997 million tons by 2029 (Milk – Worldwide, Statista Market Forecast, 2024). India is the largest milk-producing country and India's milk production rose by 4% to230.58 M Tin 2022-23, of these, liquid milk consumption in India accounts for 50% (Ganguly et al., 2017). Being an animal-origin product of high nutritional value, it comes under the category of perishable food that often undergoes microbial or chemical spoilage (Singh, 2018). In addition, the emerging pathogen causes several diseases across the world that increase the economic burden of industry and government.

The dairy industry has devised various means and technologies to meet consumer requirements and provide safe, nutritious, and good-quality milk products (Neokleous et al., 2022). Thus, the preservation of the nutritional quality of milk paves the way for converting milk into various milk-based products such as cheese, yoghurt, butter, paneer, ice cream, etc., through processing i.e. coagulation, and heat treatment. Shelf-life of milk and milk products has been the most important criterionfor the sustainable growth of the dairy industryin the current era. Therefore, the thermal processing techniques have evolved into higher capacity milk processing lines and equipment with faster production ratesand energy-efficient processes. The various batch operations are being replaced with continuous operations, e.g. OSTA for online milk standardization and pasteurization, TetraPak and Elecstar line with pet bottling equipment for UHT processing, CONTIMAB for butter and ghee manufacturing, TVR and MVR equipped powder plants with auto-pilot modes, and they have reduced thechances of contamination. However, due to the dark side of certain technologies regarding nutritional and physico-chemical aspectsof the product, non-thermal technologies came into the picture. The shelflife of dairy products is majorly affected by three aspects; processing, additives, and packaging. Emerging dairy processing technologies, biological additives, and various packaging systems (Figure 1) are therefore mainly emphasized in this review article.



Figure 1. Classification of shelf life enhancement component

Thermal technologies for processing milk and milk products: pasteurization, sterilization, and

microwave

Preservation of milk can be achieved by various conventional or thermal treatments like pasteurization, sterilization, freezing, chilling, drying, and the addition of chemical preservatives (Amit et al., 2017). Chilling helps in delaying the growth of bacteria by slowing down the metabolic activity whereas drying inhibits bacteria growth by lowering the water activity (a_w). Both pasteurization and sterilization are well-adopted, proven, and practised industrial processes to make milk safe for consumption and to increase shelf life. Pasteurization makes milk free from pathogens and a few spoilage-causing microorganisms whereas sterilization destroys all vegetative cells. The time-temperature employed for pasteurization is 63 °C /30 min or 72 °C/15 sec and for sterilization 115-120 °C/15-20 min or >135 °C/1-3 sec (Ultra High Temperature), respectively (Bezie, 2019).

Another novel version of heat treatment came in the form of microwave (MW). MWs are the electromagnetic waves between the frequency bands of 300 MHz to 300 GHz. The typical characteristic of heat generation within the material is the magnificence of a treatment. Exposure to microwave causes rapid heat generation leading to volumetric heating and a quick increase in temperature. This offers a greater advantage reducing adverse effects on nutrients over the conventional methods.

The usual frequency bands; 2450±50 MHz(domestic) and 915±25 MHz(commercial), have penetration depths of 3 to 8 cm and 8 to 22 cm (Martins et al., 2019). The effectiveness of MW is significantly driven by the dielectric properties of the material. Two mechanisms are proposed for MW heating; 1) lonic conduction; which results from resistance of food molecules to the flow of the ions and hence collisions between molecules, and 2) Dipolar rotation; associated with alternating motion of polar molecules trying to align with the alternating electric field. The effectiveness of the treatment depends on the frequency of waves applied to food, dielectric property, size, shape, and density of food.

Exposure of *burfi* (milk-based traditional Indian sweet) at a power level of 40% (i.e. 400 w) for 7 sec showed acceptability up to 19 days at 30 ± 1 °C and 28 days at 5 ± 1 °C (Kumar et al., 2018), whereas, paneer (a heat and acid coagulated milk product similar to cottage cheese) treatment at a power level of 60% (i.e. 600w) for 32s was found acceptable for 8 d and 15 d at 30 °C and 5 °C (Kumar et al., 2017), respectively. Rasooly et al., 2014 added Shiga toxin Type-2(Stx2) at a conc. of 0.5 mg/ml to milk and exposed it to two different microwave treatments; (1) 25 min, 10% power duty cycle (cycle period of 30 s) for a total of 165 kJ energy and 65 °C at the end of this cycle, (2) 20 min, 20% duty cycle for 198 kJ and 78 °C at the end of this cycle.

They found a Stx2 reducing effect in milkin both methods, i.e. either with conventional heating of milk at 95 °C for 5 min or by increased microwave energy of 198 kJ. MW treatment of milk at 750 W for 75 s reduced the viable count of total mesophiles and enterococci by one log cycle whereas pseudomonas, total coliforms, and *Enterobacteriaceae* viable counts were decreased by two log cycles after the treatment of milk. The faecal coliforms, yeast, and lactic acid bacteria were found less susceptible to the treatment. The treatment at 900 W for 75 s gave an effect equivalent to boiling (Tremonte et al., 2014). *C. sakazakiiwas* inoculated (10⁸ cfu/ml) in reconstituted infant formula followed by its MW treatment in the range of 400-900 W for 0-120 s. After treatment, the samples were stored at 4 °C for 24 h to confirm the recovery of cells. MW at 800 and 900W decreased the *C. sakazakii* below detectable levels, reaching maximum temperatures of 78.8±2.3 °C and 88.1±1.5 °C, respectively, and no cell recovery was observed during post-treatment storage (Pina-Pérez et al., 2014). Other similar studies are shown in Table 1.

Sr.	Type of Food	Treatment	Result	Reference
No.				
1	Burfi	Burfi was treated at 400 W/7 s and samples	Burfi was acceptable up to 19 days	Kumar et al., 2018
		were stored at 30±1 °C and 5±1 °C.	at 30±1 $^\circ\text{C}$ and 28 days at 5±1 $^\circ\text{C}.$	
2	Burfi	Curry leaf (0.05 ppm) and clove bud (0.15	The time-power level combination	Badola et al., 2017
		ppm) added to burfi were kept for MW	of 10%/60 s was found best. The	
		heating at 0-100% power levels for 10-90 s	treated samples were acceptable	
		followed by storage 30±1 $^\circ\text{C}$ and 5±1 $^\circ\text{C}.$	on the 8^{th} and 12^{th} day at 30±1 $^{\circ}\text{C}$	
			and 5±1 °C compared to the	
			control, respectively.	
3	Paneer	Paneer samples were treated at a power	The shelf life found for paneer was	Kumar et al., 2017
		level of 60 (i.e. 600w) for 32s. Samples	8 d and 15 d at 30 $^\circ\mathrm{C}$ and 4 $^\circ\mathrm{C},$	
		were stored at 4 °C and 30 °C.	respectively.	
5	Milk	Shiga toxin (Stx2) at a conc. of 0.5 mg/ml	Conventional heating of milk at 95	Rasooly et al., 2014
		added to milk was exposed to two different	°C for 5 min or at increased	
		microwave treatments; 1) for 25 min, 10%	microwave energy of 198 kJ	
		power duty cycle (cycle period of 30 s) for	reduced the Stx2 activity.	
		a total of 165 kJ energy and 65 $^\circ \! C$ at the end		
		of this cycle, (2) for 20 min, 20% duty cycle		
		for 198 kJ and 78 $^\circ\mathrm{C}$ at the end of this cycle.		
6	Reconstituted	C. sakazakii inoculated (10 ⁸ cfu/ml) RIF	MW at 800 and 900W decreased	Pina-Pérezet al.,
	infant	followed MW treatment in the range of	the C. sakazakii below detectable	2014
	formula	400-900 W for 0-120 s. After treatment,	levels, reaching maximum	
		samples were stored at 4 $^\circ$ C/24 h to	temperatures of 78.8±2.3 °C and	
		confirm the recovery of the cell.	88.1±1.5 °C, respectively. Post-	
			treatment storage confirmed no	
			recovery of cells.	
7	Milk	Effect of MWH (900 W) was compared to	They could not find major	Géczi et al., 2013
		conventional heating methods under	differences between MW heating	
		identical conditions of final temperature	and traditional heating.	
		and treatment time. The initial total viable		
		cell count was 50,000-3,50,000 cfu/cm ³ .		

Table 1. Application of microwave for enhancing the shelf life of milk and milk products

Non-thermal technologies for processing milk and milk products

These processes diminish the nutritional and sensorial profile of milk and milk products. On the other hand, the increased awareness and consumer demands for fresh, minimally processed food, with good nutritional and organoleptic quality, and extended shelf life has forced the food industry to search for the new independent technologies or those combined with traditional ones (Coutinho et al., 2019). This has resulted in an innovative approach of the dairy industry towards non-thermal technologies such as

High-Pressure Processing (HPP), Pulsed Electric Field (PEF), Cold Plasma, Irradiation, Ultrasonication, Ohmic heating, Microfiltration, Bactofugation, etc. to enhance the shelf life of products (Martins et al., 2019; Nikmaram and Keener, 2022). All these processes facilitate their operation at ambient temperature or a temperature less than when conventional heat treatments are used, thereby, retaining the natural content of the milk and milk products. Furthermore, microfiltration helps in removing bacterial cells as well as somatic cells through filters having a pore size of 0.1-10 μ m (Tomasula and Bonnaillie, 2015).

It can be applied in combination with pasteurization for extended shelf-life of products. Bactofugation removes bacterial spores by applying centrifugal force followed by pasteurization. The emerging non-thermal technologies are discussed here with and relevant studies conducted to increase the shelf life of milk and various milk products are summarized in Table 2.

	Food			
		High-pressure	processing (HPP)	
1	Milk	Milk was inoculated with 5 different	HPP (600 MPa for 3 min) significantly reduced	Stratakos
		strains of E. coli, Salmonella, and L.	pathogenic count, total viable counts,	et al., 2019
		monocytogenes treated at 400, 500,	Enterobacteriaceae, LAB, and Pseudomonas spp. in	
		and 600 MPa with a hold time at	milk, thus prolonging the microbiological shelf life	
		pressure of 1, 3 and 5 min at room	of milk by 1 week compared to pasteurized milk.	
		temp. After processing, raw,		
		pasteurized and HPP milk was		
		stored in one-litre bottles at 4 \pm 0.5		
		°C for ta28-day shelf life study.		
2	UF-cheese	UF-cheeses were manufactured	Both UF-cheeses (made with RC or BR) showed a	Ribeiro et
		with recombinant chymosin (RC) or	significant increase in syneresis over time.	al., 2018
		bovine rennet (BR), processed at	Regarding texture, HPP yielded firmer cheeses.	
		600 MPa/ 5 min/ 25 °C, and stored	After 1 day of storage, HP-processed cheeses made	
		at 7 °C for up to 56 days.	with RC and BR had a reduction of 42.8 and 52.5%	
			in proteolysis, respectively, in comparison with the	
			controls. Moreover, HPP promoted a reduction in	
			the psychrotrophic count.	
3	Milk	Two pathogenic strains – L.	After 10 days of storage, 2 cfu/ mL of E.coli ATCC	Liepa et
		monocytogenes ATCC 7644 (LM)	25922 and L. monocytogenes ATCC 7644 were	al., 2018
		and <i>E. coli</i> ATCC 25922 (EC)	detected in analysed samples with lower ability to	
		inoculated UHT milk weretreated at	repair was detected in milk samples treated with	
		400, 500, 550, and 600 MPa/ 15	600 MPa/ 15 min/ with inlet temperature 20 °C)	
		min/ with inlet temperatures 20 °C,		
		and then stored at 4 \pm 2 °C for 10 d.		

Table 2. Non-thermal technologies used for preservation of milk and milk products

Result

Reference

Treatment

Sr. No.

Type of

4	Whey lime	The packaged samples of whey-lime	HPP processing preserved the higher antioxidant	Bansal et
	beverage	beverage were processed at 500	capacity (54.2%), colour, content of phenolics	al., 2019
		MPa/10 min/ 25 °C and stored at 4	(60.2%), and low non-enzymatic browning index	
		^o C for 120 d. A control sample was	(0.181 \pm 0.03) compared to the control.	
		given the treatment of 90 °C for 60		
		seconds.		
5	Mozzarella	Two weeks after manufacture, 2	The results indicated that 600-MPa treatment	Ozturk et
	cheese	groups of cheese samples were	helped minimize typical age-related changes in the	al., 2018
		treated with HHP at 500 or 600 MPa	performance of the reduced-Na LMPS Mozzarella	
		for 3 min and then returned to	cheeses. The study demonstrated that the shelf life	
		storage at approx. 4°C.	of LMPS Mozzarella cheeses can be extended from	
			the traditional 4 to 6 wk to 20 wk of refrigerated	
			storage by the application of HHP at 600 MPa for 3	
			min.	
6	Milk	Milk was given HHP treatment at	The firmness of the curd was higher than that of	Liepa et
		400 - 600 MPa/ 15 min/room	raw or pasteurized milk. The RCT of milk treated at	al., 2017
		temperature.	400 MPa was at par with raw milk.	
7	Bovine	Milk inoculated with E. coli,	High-pressure processing of colostrum is effective	Foster et
	colostrum	Salmonella Dublin, or MAP, bovine	in the reduction of native aerobic bacteria, E. coli,	al., 2016
		herpes virus type 1, and feline	Salmonella Dublin, and both enveloped and	
		calicivirus were pressure processed	nonenveloped viruses, but MAP was resistant to	
		at 300 MPa. (30, 45, and 60 min)	the effects of HPP. Calves fed with pressure-	
		and 400 MPa (10, 15, and 20 min).	processed colostrum had similar serum IgG but	
			lower efficiency of absorption than calves fed with	
			heat-treated colostrum.	
8	Milk	The effect of HPP (200, 400, and 600	The 600 MPa combined with heat enhanced the	Evelyn and
		MPa for up to 40 min at 70 °C) on <i>B.</i>	spore inactivation in milk, requiring a temperature	Silva, 2015
		cereus spores was investigated.	lower than 20 °C to achieve the same spore	
		Thermal treatment was given at 70,	inactivation.	
		80, and 90 °C.		
		Pulsedelec	tric field (PEF)	
1	Milk	The milk received pre-heat	The different treatment of samples showed shelf	Indumathi
		treatment of 40 $^\circ C$ and 50 $^\circ C$	life between 16.33 \pm 0.21 to 30.83 \pm 0.17 (mean	et al., 2018
		followed by PEF of 10KV/cm,	value).	
		20KV/cm, and 30KV/cm at two		
		different treatment times (3 and 6		
		minutes). The samples were stored		
		at refrigeration temperature.		
2	Milk	Electric field strengths of 18-28	Controlled pre-heating and PEF treatment (23	Sharma et
		kV/cm were applied for 17-235 μs to	kV/cm, 17-101 $\mu s)$ with intermediate cooling	al., 2014a
		milk at different temperatures (4-55	during PEF has the potential to provide the	
		°C) for 24 s.	equivalence to thermal pasteurization conditions.	

3	Milk	E. coli 916 (ATCC 25922) and L.	Combined treatment (at 55 °C) for 24 s and 15.9-	Sharma et	
		innocua 3024 (ATCC 33090)	26.1 kV/cm for 34-101 μs effectively reduced the	al., 2014b	
		inoculated milk was pre-heated at	organism'snumber below the detection limit of 2		
		55 °C for 24 s followed by PEF in a	log cfu/mL. Pre-heat treatment and PEF at 26.1		
		continuous mode at electric field	kV/cm for 34 $\mu s,$ reduced the activity of plasmin,		
		intensities of 15.9-26.2 kV/c. for 17-	xanthine oxidase, and lipolysable fat by 12%, 32%,		
		101 µs.	and 82%, respectively, compared to raw milk.		
4	Milk	Milk was treated at 46.15 kV/cm /	The treated samples showed minor physico-	Bermúdez-	
		20 to 60 °C/ 30 pulses (2 μs each).	chemical changes. The samples stored at 4 $^{\circ}\mathrm{C}$ were	Aguirreet	
		Then, the samples were stored at 4	stable for 33 d while samples stored at 21 $^{\circ}\mathrm{C}$ were	al., 2011	
		°C and 21 °C for 35 d.	stable for up to the 5 th d of storage.		
5	Milk	Milk subjected to PEF consisted of 5	The shelflife of milk was extended by a minimum	Sepulveda	
		pulses of 35 kV/cm and pulse width	of 24 days.	et al., 2009	
		of around 2.3 $\mu s,$ applied to milk at			
		65 °C. The samples were stored at 4			
		°C for 50 d.			
		Irra	diation		
1	Cheese	X-rays at intensities of 0.5, 2, and 3	The artisanally and industrially manufactured	Ricciardi et	
	(Ricotta)	kGy were used for sanitizing cheese	cheese showed a shelf life of 20 and 84 d,	al., 2019	
		manufactured artisanally and	respectively, compared to the control.		
		industrially. Then, samples were			
		stored at 4 °C for 24 and 84 d.			
2	Cheese	Cheese samples containing	A germicidal effect of about 1–2 log cycles on	Lacivita et	
	(Fiordilatte)	Pseudomonass pp were exposed to	Pseudomonas spp. and Enterobacteriaceae was	al., 2016	
		surface UV-C light treatment at 0.1,	observed during storage. An 80% shelf-life		
		0.6, 1.2, and 6.0 kJ/ m ² for 5, 30, 60,	extension was achieved by selecting adequate		
		and 300 s, respectively. The samples	processing conditions of exposure of Fiordilatte		
		were then stored at 9 °C.	cheese to UV-C light (6.0 kJ/ m ²).		
3	Cheese	Cheese samples were irradiated (10	The treatment has significantly (P<0.05) increased	Huo et al.,	
	(Mozzarella)	MeV electron beam at doses of 0-	the shelf life of the product.	2013	
		2.5 kGy at 30 °C) and stored at 10 °C			
		for 90 d.			
4	Cheese	Milk inoculated with three different	Irradiation at a dose of 2 kGy was sufficient for the	Badr, 2011	
		species of Mycobacterium (M.	complete inactivation.		
		bovis, M. paratuberculosis, and M.			
		tuberculosis) used to prepare			
		cheese was exposed to gamma			
		radiation at intensities of 0, 2, and 4			
		and stored at 4 <u>+</u> 1 °C for 15 d.			
	Cold plasma				

1	Cheese	Fresco cheese (QFC) and cheese	After direct HVACP treatment, a reduction of 3.5	Wan et al.,
		model (CM) inoculated (L. innocua)	and 1.6 log10 CFU/g was observed for CM and QFC.	2019
		samples were treated with HVACP	Direct plasma treatment was more effective than	
		(100KV/ 60 Hz/ 5 min.) under the	indirect.	
		direct and indirect modes of		
		exposure in dry air gas environment		
		for 5 min.		
2	Milk	Sterilization of raw cow milk was	3kV/ 3 min/500 Hz frequency had completely killed	Aslan,
		performed by using the dielectric	the bacteria in raw milk.	2016
		barrier discharge (DBD) plasma		
		method.		
3	Milk	Plasma was generated in the milk (E.	No viable cells were detected in milk treated at 4	Ponraj et
		coli inoculated pasteurized and raw)	kHz (inoculated with <i>E. coli</i>) and stored for up to 4	al., 2015
		by a nanosecond pulse generator	weeks after treatment.	
		(18 kV, argon) at two different		
		frequencies: i) 2.5 kHz, and ii) 4 kHz		
		for 2 minutes. The samples were		
		stored at 4 °C for 4 weeks.		
4	Cheese	Sliced cheddar cheese was	The level of these microorganisms on sliced	Yong et al.,
		inoculated with E. coli O157:H7, L.	cheddar cheese in response to 10-min plasma	2015a
		monocytogenes, and S.	treatment was significantly decreased by 3.2, 2.1,	
		typhimurium and exposed to	and 5.8 Log CFU/g, respectively.	
		flexible thin-layer dielectric barrier		
		discharge (DBD) plasma.		
5	Cheese	Cheese slices were inoculated with	The number of E. coli, S. typhimurium, and L.	Yong et al.,
		E. coli O157:H7, S. typhimurium, and	monocytogenes inoculated on cheese slices	2015b
		L. monocytogenes treated with	decreased by 2.67, 3.10, and 1.65 at 60 s, 45 s, and	
		encapsulated DBD plasma produced	7 min. Further reduction was observed during	
		at 250 W/ bipolar 15 kHz/1-15 min.	post-treatment storage. Thus, encapsulated DBD	
			can be used to increase the post-treatment storage	
			of cheese.	
6	Milk	Milk was inoculated with E. coli, S.	The total aerobic count was not detected in plasma	Kim et al.,
		typhimurium, and L.	treated sample whereas the samples inoculated	2015
		monocytogenes treated with	with pathogenic strains showed a reduction of 2.64	
		encapsulated DBD plasma produced	log cfu/ml in case of 10 min.	
		at 250 W/ bipolar 15 kHz/5-10 min.	The encapsulated DBD with less than a 10 10-	
		, p	minutetreatment period can be applied to milk to	
			increase the shelf life with minimum physico-	
			chemical changes.	
7	Cheese	Cheese slices were inoculated with	The number of <i>E. coli</i> inoculated on cheese slices	Lee et al.
	Slices	E. coli and S. aureus treated with	decreased by 0.09. 0.47. 1.16. and 1.47 log cycles	2012
	000		with helium (4 liters/min [lom]) and 0.05, 0.87	

		DBD plasma produced at 3.5 KV / 50	1.89, and 1.98 log cycles with He/O_2 mixture (4	
		kHz/ 1-15 min.	lpm/15 standard cubic centimeters per minute),	
			after being treated with plasma for 1, 5, 10, and 15	
			min, respectively. Significant reductions were also	
			observed in S. aureus inoculated onto cheese slices	
			ranging from 0.05 to 0.45 log cycles with He and	
			from 0.08 to 0.91 log cycles with He/O_2 -treated	
			samples, respectively.	
8	Milk	Low-temperature atmospheric	A significant 54% reduction in the population of E.	Gurol et
		pressure plasma treatment (9 kV	coli cells after only 3 min was observed regardless	al., 2012
		AC/ 0-20 min) was given to E. coli	of the fat content of the milk. The levels of E. coli	
		ATCC 25922 inoculated UHT milk	for 20 min plasma applied samples were	
		(whole, semi-skimmed and	undetectable after one day of storage and	
		skimmed milk) samples followed by	remained thereafter at the end of a 6 week period.	
		storage at $4-7$ °C for 6 weeks.		
			rasound	
1	Milk	Milk samples were sonicated at	Counts in pasteurized controls and C-S milk	lim et al
T	IVIIIK	2 200 W/ 20 kHz/10.60 c. Thermo	did not exceed 2.00 log cfu/mL for up to 50 di	2010
		sonisation was performed at	and not exceed 5.00 log cid/me for up to 50 d,	2019
		The solution was performed at	Counts in 1-5 milk exceeded 5.00 cru/mL by a 36.	
		72.5 <u>+</u> 0.3 °C (mean <u>+</u> SD) after	Neither C-S hor I-S were appropriate techniques	
		pasteurization $(72.5\pm0.3$ °C)	for reducing bacterial count in fluid milk beyond	
		(mean <u>+</u> SD), whereas cold-	standard pasteurization and, in fact, increased	
		sonication was carried out at	counts of spore-forming spoilage bacteria.	
		12.5 <u>+</u> 5°C (mean <u>+</u> SD) before		
		pasteurization. Then, milk was		
		refrigerated up to 50 d.		
2	Milk	Pasteurized and UHT milk	Optimization of the inactivation of microbes was	Ganesan et
		(inoculated) received high-intensity	found to be at 84.8°C, 216 μm amplitude, and 5.8	al., 2015
		ultrasound treatment (range from 0	min.	
		to 84 °C, amplitude range from 0 to		
		216 $\mu m,$ and time range from 0.17		
		to 5 min).		
3	Milk	The milk samples inoculated with a	The test organisms exhibited biphasic inactivation	Gabriel et
		cocktail of L. monocytogenes strains	curves in all milk samples. The correctedD-value	al., 2015
		(two) were exposed to ultrasonic	was shortest in full-cream milk at 24.81 min,	
		oscillations where frequencies were	followed by those in nonfat and low-fat milk at	
		switched to 28, 45, and 100 kHz at 1	29.17 and 30.64 min, respectively.	
		ms time intervals.		
4	Queso	Raw milk was sonicated at 400 W/	Curdling time was reduced considerably, cheese	Bermúdez-
	fresco	24 kHz/ 120 μ m and heated at two	yield (20.6%) was almost doubled. and the	Aguirreand
	cheese	different time-temp combination:	luminosity of cheese was increased (L^*) . Cheese	Barbosa-
		63 °C/10-30 min or 72 °C/0.15-1	processed at 63 °C/ 120 um/30 min had the best	*
		.,	,	

		min. Samples were stored at 4 °C for	quality. Shelf life was extended considerably and	Canovas,
		23 d.	the product had higher quality.	2010
5	Milk	Milk (UHT) inoculated with E. coli, L.	Viable counts of E. coli, P. fluorescens and L.	Cameron
		monocytogenes, and P. fluorescens	monocytogeneswere reduced by 100% after 10.0,	et al., 2009
		received treatment at 750 W, 20	100% after 6.0 min, and by 99% after 10.0 min.	
		kHz, 124 μm/ 2.5-10 min.		

High-pressure processing (HPP)

HPP is considered as one of the most emerging techniques in the last few decades. First time Hite demonstrated the shelf life extension of raw milk by applying HPP. Later on in the 20th century, Japan successfully manufactured and marketed HPP-treated fruit and jams. It can inactivate the spoilage of foods by delaying the onset of enzymatic and chemical deteriorative processes. It works on two principles; 1) Le Chatelier's: pressure favours all structural reactions and changes that involve a decrease in volume and 2) Isostatic principle: the distribution of pressure is proportional in all parts of a foodstuff irrespective of their shape and size. The food is exposed toa high pressure in the range of 100-1000 Mpa. (Voigt et al., 2015). Water or a mixture of oil or alcohol can be used as a pressure-transmitting medium. The temperature is increased at a tune of 2-3 °C per 100 MPa. The treatment below 420 MPa produces an effect similar to pasteurization and above 700 MPa similar to sterilization. Inactivation of spores required much higher pressure and temperature than vegetative cells. In Figure 2, general structures/organelles present in a bacterial cell are depicted. In the majority of thermal and non-thermal methods, the cell wall components and cell membrane of bacteria usually get affected. Specifically, the proposed mechanism of microbial inactivation is related to the loss of cell membrane permeability (Figure 3) and the breaking down of cell membranes, cell walls, etc.

Pulsed electric field (PEF)

PEF has been successfully applied to alter the genomic material amongst microorganisms by inducing electroporation (perforating cell membrane). In the 19th century, it was used toinactivate the enzymes and MOs in food products. The food is placed between two electrodes and short pulses (1-10 μ s) are generated through a high voltage (20-80 kV) pulse generator. Pasteurization of milk is generally performed with square-wave. A number of functional and structural changes takeplace in cell membranesdue to high-voltage treatment-led microbial inactivation. Critical transmembrane potential or Critical electric field (*Ec*) (kV/cm) indicates the maximum potential difference that the cell membrane can withstand. A higher external electric field than *Ec* forms irreversible pores whereas a lower external electric field than Ec forms reversible pores. The pore formation leads to cell death (Figure 4). Both, static and continuous systems are used to process food (Sampedro and Rodrigo, 2015). Example of companies manufacturing PEF processing units are Diversified Technologies Inc. and PurePulse Technologies Inc. in the USA and ScandiNova Systems AB in Sweden with overall flow rates ranging from 400 to 6000 I/h. The process is affected by various factors such as process time, voltage applied, physiological state of cells (log phase is more susceptible), conductivity, and pH of the product.



Figure 2. Bacterial cell structures and sectioning of cellwall of Gram positive bacteria



Figure 3. Cell membrane damage by high-pressure processing (HPP) (modified from Naik et al., 2013)

Cold plasma

Plasma is a Greek word meaning mouldable substances, first explained by Irving Langmuir in 1920s (Baghya and Narayanan, 2019). It is referred to as the fourth state of matter, electrically neutral, and produced in the presence of energy sources (i.e. electricity) and gas. It is found electrically neutral. Based on the method of generation, pressure, and relative temperature, plasma is classified into two groups; a) Thermal- produced under high pressure and power where gas species and electrons are in thermodynamical equilibrium in nature, and b) Cold or Non-thermal- produced under reduced pressure (at atmospheric) and power where gas species and electrons are thermodynamically non-equilibrium in

nature. Cold plasma is found suitable for the application in food industry because the ions or uncharged molecules gain only little energy and remain at a low temperature. The products of plasma include N_2 , NO, NO_2 , nitric oxide radical NO^+ , atomic oxygen (O), ozone (O₃), ions, neutrons, protons and reactive oxygen and nitrogen species, and hydroxyl radicals (OH⁺) (Coutinho et al., 2019;2021; Nikmaram and Keener, 2022).



Figure 4. The microbial cell destruction by Pulsed Electric Fields (PEF) (modified from Sampedro and Rodrigo, 2015; Sharma et al., 2014 a, b)

Cold plasma is generated by various methods such as Dielectric barrier discharge (DBD) method, atmospheric plasma jet discharge, corona discharge, and microwave-driven discharge. MW-driven discharge uses microwaves instead of electrical fields. Air or nitrogen or a mixture of noble gases like argon, neon, and helium are used in the presence of the electric field. The most commonly used method in dairy industry is DBD. It has been successfully applied to destroy pathogenic as well as spoilagecausing organisms in cheese, raw milk, cheese slices, etc. The effectiveness is significantly affected by Relative Humidity (RH), type of species generated, length of treatment, and power level used for generation. The RH is directly correlated with the reactive species generated, e.g. the higher the RH, the more the peroxyl acid groups and OH there will be. The radical bombardment causes several phenomena on/in a microbial cell, i.e. formation of lesions, mutational damage to DNA and/or RNA by breakage of chemical bonds, lipid oxidation or peroxidation in the cell membrane, denaturation of cell protein or enzymes as depicted in Figure 5 (Coutinho et al., 2019). Therefore, one or more of such effects led to cell injury or viability. It is an energy efficient method and is applied on both solid and liquid food at low temperatures. However, the lipid oxidation and the low penetration power are the limitations for the application of cold plasma in dairy products since dairy products are rich in triglycerides-natural milk fat together with a colloidal form of protein (casein) that protect the microbial cells against the potential damage caused by cold plasma method.



Figure 5. Various modes of bacterial inactivation by cold plasma (modified from Coutinho et al., 2019)

Ultrasonication

The method refers to the application of sound waves at a frequency (>16 kHz) greater than the upper limit of human hearing through liquid, solid, or gases which causes vibration, acoustic streaming, and formation of small bubbles (known as cavitation) due to pressure variation. Two types of cavitations occur; a) transient (20 kHz) and b) stable (>200 kHz). The size of the bubble increases and when it attains a volume at which itcan no longer absorb energy, it implodes violently at higher-intensity waves. This generates mechanical, physical, and chemical effects, such as shockwave formation and turbulent motion. Both, the temperature and pressure are high during implosion (Ashokkumar et al., 2010). The treatment involves direct (using ultrasonic probe) and indirect mode (ultrasonic bath). It has been found to be an important tool for various applications, i.e. drying, crystallization, extraction, filtration, emulsification, cleaning, etc.

The combination of heat and ultrasonication is more lethal (known as thermosonication). Another approach includes pressure+ultrasonication (Manosonication) and heat+pressure+sonication (Manothermosonication) (Zisu and Chandrapala, 2015). High-intensity ultrasonication tends to develop undesirable off-flavour. On the other hand, mild-level ultrasonication helps in improving quality, reducing fermentation time and enzyme productivity. Cell inactivation occurs by disrupting both cell wall structure and function through cavitation in an ultrasonicator.

Irradiation

The substantial reduction of microorganisms at ambient temperature without affecting the vitamins, flavour, and colour makes this technique appropriate for the processing of milk and milk products. Food

is exposed to ionizing radiation in the form of Gamma (Cesium 137 or cobalt 60), X-rays, ultraviolet light and electron beams. Radiation dose is measured by a Dosimeter device and expressed in terms of Gray (Gy). WHO has endorsed the use of irradiation doses up to 10 kGy in foods. UV light can potentially reduce the microbial count without affecting other properties of food (Roberts, 2016). Based on wavelength, UV rays are classified as UV-A (315 and 400 nm), UV-B (280–315nm), and UV-C (200-280 nm) (Datta et al., 2015). UV-C is most effective in food processing.

The composition and transparency of food affect the efficacy of UV light. As the milk is opaque in nature due to colloidal and suspended solids, two approaches have been suggested to obtain complete penetration; a) turbulent flow of milk exposes all surfaces to UV light and also reduces the path length, and b) laminar flow that forms thin layer of milk on UV irradiated surfaces. It tends to mutate the bacterial cell by forming pyrimidine dimmers (Figure 6) that block DNA transcription and replication causingbacterial cell death. The irradiation dose of bacterial inactivation is much lower than required for algae, viruses, and fungi. The radiation is successfully applied for food application on a commercial scale in over 26 countries. The irradiated products are being identified through the "*Radura*" logo labeled on packed food. Irradiation of cheese significantly increases the shelf life along with the inactivation of microbes (Badr, 2011; Ricciardi et al., 2019).



Figure 6. DNA damage by irradiation (adapted and modified from Huang and Zhou, 2020)

Ohmic heating

In the 19th century, James Joule, hence named as Joule heating, revealed the heat transfer upon passage of electric current passed through food. Electrical resistance is found responsible for heatproduced inside the food and the unit of measurement of resistance is Ohm (Ω), therefore, also referred to as Ohmic heating (OH). In the year 1919, Anderson and Finkelstein were first to recommend OH for milk. The initial successful commercial technique was known as "Electro Pure". The movement of ions in the liquid causes collision, which in turn, results in creating resistance and generation of heat (Sakr and Liu, 2014; Kumar, 2018). Factors like electrode type, conductivity, concentration of ions, field strength, etc. greatly influence the effectiveness of OH in food. OH of milk products improves the texture as well as the shelf life of products (Sun et al., 2008; Kumar et al., 2014; Parmar et al., 2018).

Similar to this, Light-emitting diodes (LEDs) are another eco-friendly light source with wide-spectrum antimicrobial activity against bacteria, fungi, viruses, planktonic cells, and endospores (Yu et al., 2022).

Bio-preservation

Bio-preservation makes the use of metabolites/substances that are produced by microorganisms or entire cells or present naturally in milk. Bacteriocins, lysozyme, lactoferrin, natamycin, bacteriophage, and endolysins are the different kinds of biopreservatives, that obtained the designation Generally Recognized as Safe (GRAS), and can be employed to enhance the shelf life of milk and milk products (Table 3) (Conte et al., 2011). Nisin and pediocin are well known examples of bacteriocins, which are proteinaceous compounds, produced by LAB to inhibit the growth of similar or closely related bacterial strain(s). These compounds, either alone or in combination with other antimicrobial agents, (like sorbic acid, EDTA) have been shown to inhibit the growth of spoilage and pathogenic bacteria (*E.coli, Bacillus sp., L. monocytogenes, Staph. aureus*, etc.) in several milk products (Conte et al., 2011; Chawla et al., 2015). Natamycin (E235) is an antifungal compound produced as a secondary metabolite by some species of *Streptomyces*; it is effective at very low levels (MIC is less than 10 ppm) for most moulds. It has been successfully used to preserve various types of cheeses, sour cream, yoghurt, and packaged salad mixes (Saad et al., 2015).

Sr.	Type of Food	Treatment	Result	Reference
No.				
		Bio-preserva	tive agents	
1	Pasteurized	Milk was inoculated with MRSA	In milk stored at 4 °C and 25°C, significant	Chang et al.,
	milk	CCARM 3089 cells (2 x 105 CFU/mL)	inhibitory effects (P<0.05) were shown	2017a
		and LysSA11 at 0, 1.125, 2.25, 3.375,	within 15 min by 3.375 μM of LysSA11.	
		4.5, and 9 $\mu M.$ The milksample was	Moreover, viable cells were reduced to	
		then incubated at 4 °C or 25 °C for an	undetectable levels at 1 h (4°C) and 30 min	
		additional hour.	(25°C) by treatment with 9 μM of LysSA11.	
2	White mould-	The treatments include, partial	The concentration at 8 mg/ ml was the	Khider, 2017
	ripened cheese	purified bacteriocin-like substances	most suitable one to minimize the growth	
		(PPBLS),	of P. candidum and hence extended the	
		and bacteriocin producer isolate LAB	shelf life of Camembert cheese.	
		100.		
3	Pasteurized	Milk samples were inoculated with S.	The number of live S. aureus cells	Chang et al.,
	milk	aureus RN4220, LysSA97, and	decreased below the detection limit using	2017b
		carvacrol at 10 ⁵ CFU/mL, 1.88 μ M,	the cocktail of LysSA97 and carvacrol conc.	
		and 6.66 mM. The samples were	over 3 h.	
		kept at room temperature for 3h.		

Table 3 Use of bio-preservatives and essential oils and herbs for milk and milk products preservation

4	Milk	L. monocytogenes and Bacillus	In both skimmed and whole milk, free and	Martinez et
		spores contaminated milk (Skim and	encapsulated Nisaplin® combined (0.5	al., 2016
		Whole) was supplemented with free	mg/L each) exhibited the strongest	
		and encapsulated commercial nisin	antibacterialeffect, although Lm-resistant	
		(0.25-1.0 mg/L, alone and combined)	cells were observed. Free and	
		and stored at 6±1 °C for 21 d.	encapsulated commercial nisin (0.25	
			mg/L) were highly effective against Bc	
			spores germination and for the pathogen	
			inhibition in both types of milk, improving	
			the food product microbiological safety.	
5	Yoghurt	Yoghurt incorporated with 25 RU/ml	Application of nisin for shelf-life extension	Sarkar, 2016
		nisin was stored at 10-15°C for 40 d.	of stirred yoghurt made from cow milk is	
			suggested.	
6	ESL Milk	The milk contaminated with L.	The combination of FWLLm1 and coagulin	Rodríguez-
		monocytogenes 2000/47 treated	C23 reduced the count of L.	Rubio et al.,
		with phage FWLLm1 added at 5 $ imes$ 10 6	monocytogenes 2000/47 below detection	2015
		PFU/ml, FWLLm3 at 5 × 10 ⁵ PFU/ml,	limits (less than 10 CFU/ml) from day 4	
		and bacteriocin coagulin C23 at 584	until the end of the experiment.	
		AU/ml was stored at 4 °C for 10 days.		
7	Processed	Processed cheese sauces with	The combination of nisin + natamycin +	Saad et al.,
	cheese sauces	different preservative systems;	potassium sorbate mixture was the most	2015
		Nisin, Nisin + Natamycin, Nisin +	effective one for the product shelf life.	
		Potassium sorbate, or Nisin +		
		Natamycin + Potassium sorbate		
		were prepared. The melted		
		processed cheese sauce was purred		
		into glass jars (150g) and stored at		
		room temperature (25 \pm 2 °C) for 3		
		months.		
8	Pasteurized	Pasteurized S. aureus ATCC33591	Compared with the raw milk, the viable	Guo et al.,
	milk	inoculated milk supplemented with	counts of S. aureus were reduced by 10^{5} -	2016
		overnight cultures of L. lactis	fold in the cheese inoculated with the	
		CCTCCAB20102111 and L. casei	engineered L. casei strain during the	
		BL/pBLysdb were used to prepare	fermentation process, and the pathogenic	
		cheese.	bacterial numbers remained at a low level	
			(10^4 CFU/g) after 6 weeks of ripening at 10	
			°C.	
9	Doda burfi	Pediocin (0.12%), microgard- 100	The shelflife of 27 days of the treated	Chawla et al.,
		(0.5%), along with chemical	product counterpart to 12 days of control	2015
		preservatives such as potassium	product kept at 30 °C was recorded.	
		sorbate (0.1%) and sodium EDTA as		
		chelating agent (20mM) were added		

life of the product. Radha, 2014 10 Pasteurized Nisin was added at the rate of 50, nulk In the case of nisin added samples, overall ad 300 U/ml at about one hour before pasteurization. After pasteurization, samples were stored at 4±1°C for 16 d. Radha, 2014 11 Milk pudding Nisin A was added to milk pudding to final concentrations of 40, 80, 120, and 240 IU/ml against spores from <i>B. thuringiensis, B. cerus, and P.</i> <i>jomilae.</i> The samples were incubated at 15, 20, and 30°C for 29 days. >120 IU/g in 7.5% fat) was found to be al., 2014 Acceltal of the three phages completely McLean McLean effective as a natural preservative to and EC11) was tested and added to inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> and EC11) was tested and added to milk samples incubated with <i>E. coli</i> 0217/H6 in both UHT and raw milk at 25° C and at 5-9°C. A cocktail containing EC6 OS:H- and <i>E. coli</i> O127:H61. The samples were incubated either at 25 , an enterohemorrhagic strain, in UHT milk refrigerator (3-9°C) for 168 h. and EC10 completely inhibited <i>E. coli</i> 05:H- samples were incubated either at 25 , an enterohemorrhagic strain, in UHT milk refrigerator (3-9°C) for 168 h. Conte et al., and 50 m/N Na;-EDTA were added to the cheese and packed under MAP. Uspazyme. The samples were kept at 8°C for 9 d. Conte et al., achieved by the combined activity of both autimicrobials (after 6 h of incubation). 2010 13 Pasteurized milk 10° c fu/mil and 10° Cr /m g. and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. The use of nisin extended the shelf-life of incubation).			in combination to enhance the shelf		
10 Pasteurized milk Nisin was added at the rate of 50, 100, 200, and 300 IU/mi at about one hour before pasteurization. After pasteurization, samples were stored at 4±1°C for 16 d. In the case of nisin added samples, overall acceptability scores remained within the acceptable limit until 16 days of storage. Radha, 2014 11 Milk pudding Nisin A was added to milk pudding to final concentrations of 40, 80, 120, and 240 IU/mi against spores from <i>B. thuringiensis, B. cereus</i> , and <i>P.</i> Nisin A (at 2 80 IU/g in 5.0% fat and at piorilize. Oshima et al., 2014 12 UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, and EC11) was tested and added to milk samples inoculated with <i>E. coli</i> otsriains (<i>E. coli</i> ATCC 25922, <i>E. coli</i> atta 5.9 °C. A cocktail containing EC6 and EC completely inhibited <i>E. coli</i> O127:H6i. The samples were incubated ether at 25, °C for 24 h, or in a domestic screitor (5-9° C) for 168 h. A cocktail containing EC6 and EC3 completely inhibited <i>E. coli</i> 05:H- and EC3 completely inhibited <i>E. coli</i> 05:H- and S0 MM Na ₂ :EDTA were added to the cheses and packed under MAP. The shelf life of cheses was increased conte et al., specifically with the highest content of 2011 14 Pasteurized milk 10° cfu/ml and 10° cfu/ml of soft-acid curdi Mix 10° cfu/ml and 0.75 µg /mi, endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Clearance of the pathogen was only achieved by the combined activity of both 2010 Kykkidou et antimicrobials (after 6 h of incubation). 15 Galotyri (Greek soft-acid curdi) different concentrations (N			life of the product.		
milk 100, 200, and 300 IU/milat about on hour before pasteurization. After pasteurization, samples were stored at 41°C for 16 d. acceptability scores remained within the acceptable limit until 16 days of storage. 11 Milk pudding Nisin A was added to milk pudding to final concentrations of 40, 80, 120, and 240 IU/ mil against spores from <i>B. thuringiensis, B. cereus, and P. gimiles.</i> The samples were incubated at 15, 20, and 30°C for 29 days. Nisin A (at 2 80 IU/g in 7.5% fat) was found to be and 240 IU/mil against spores from <i>B. thuringiensis, B. cereus, and P. gimiles.</i> The samples were shelf life (4 weeks). Mile pudding Mile against and EC11) was tested and added to milk samples inoculated with <i>E. coli</i> A cocktail of the three phages completely and at 5-9°C. A cocktail of the three thages completely mile samples inoculated with <i>E. coli</i> McLean et al. 2013 12 UHT and Raw A cocktail of <i>E. coli</i> ATCC 25922, <i>E. coli</i> and EC11) was tested and added to milk samples inoculated with <i>E. coli</i> A cocktail of the three phages completely mile samples inoculated with <i>E. coli</i> McLean et al. 2013 13 Cheese (Burrata) Locyarme (150, 250, and 500 ppm), the cheese and packed under MAP. No samples were kept at 8°C Gord The samples and backed under MAP. No samples were kept at 8°C Gord Contre et al., achieved by the combined activity of both 2010 2011 14 Pasteurized milk Pasteurized milk was inoculated with Clearance of the pathogen was only Garda et al., 2007 Chera et al., achieved by the combined activity of both antimicrobials (after	10	Pasteurized	Nisin was added at the rate of 50,	In the case of nisin added samples, overall	Radha, 2014
hour before pasteurization. After pasteurization, samples were stored at 4±1°C for 16 d. acceptable limit until 16 days of storage. 11 Milk pudding Milk pudding At 12°C for 16 d. Nisin A was added to milk pudding to Ad 90. 120. 2 ±20 IU/g in 5.5% fat) was found to be al., 2014 and 240 IU/mil against spores from at 240 IU/mil against spores from and 240 IU/mil against spores from incubated at 15.20, and 30°C for 29 days. Statural preservative to Shelf life (4 weeks). 12 UHT and Raw Acceltail of <i>E. coli</i> phage (EC6, EC9, A cocktail of the three phages completely McLean et and EC11) was tested and added to milk samples inoculated with <i>E. coli</i> ATCC 25922. E coli and at E -9 °C. A cocktail containing EC6 Statis and EC 2000 pastely inhibited <i>E. coli</i> ATCC 25922. and <i>E. coli</i> al., 2013 milk samples inoculated with <i>E. coli</i> and a E -9 °C. A cocktail containing EC6 Statis and EC 2000 completely inhibited <i>E. coli</i> ATCC 25922. and <i>E. coli</i> ada 25°C Statis and EC 2000 completely inhibited <i>E. coli</i> ada, 2013 milk samples were incubated either at 25 °C and at E -9 °C. A cocktail containing EC6 Statis and EC 2000 completely inhibited <i>E. coli</i> ada, 2013 milk samples were incubated either at 25 receive and acceive and packed under MAP. The samples were incubated to the face and EC 2000 completely inhibited <i>E. coli</i> ada 2000 content ada 2000 completely inhibited <i>E. coli</i> ada 2000 content ada 2000		milk	100, 200, and 300 IU/ml at about one	acceptability scores remained within the	
pasteurization, samples were stored at 4±1°C for 16 d. Oshima et 441°C for 16 d. 11 Milk pudding Nisin A was added to milk pudding to and 240 IU/g in 7.5% fat) was found to be al, 2014 and 240 IU/m lagainst spores from effective as a natural preservative to 8. thuringiensis, B. cereus, and P. control spoilage bacteria and extend its jamilae. The samples were shell if (4 weeks). incubated at 15, 20, and 30°C for 29 days. Control spoilage bacteria and extend its shell life (4 weeks). incubated at 15, 20, and 30°C for 29 days. 12 UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, A cocktail of the three phages completely McLean et and EC11) was tested and added to inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> and 25°C strains (<i>E. coli</i> C1227:H6 in both UHT and raw milk at 25°C strains (<i>E. coli</i> O127:H6). The samples inoculated with <i>E. coli</i> O127:H6 in both UHT and raw milk at 25°C strains (<i>E. coli</i> C159 °C) for 168 h. regrowth was observed. Conte et al., an enterohemorrhagic strain, in UHT milk regrowth was observed. 13 Cheese Lysozyme (150, 250, and 500 ppm). The shell life of cheese was increased to the cheese and packed under MAP. The samples were kept at 3°C for 9 d. Conte et al., achieved by the combined activity of both 2010 antimicrobials (after 6 h of incubation). (0.37 µg/ mil and 0.75 µg /mil), endolysin (7.5 U/mil and 15° U/mil), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. The use of nisin extended the shelf-life of Kykkidou et arebically under refrigeration (N1, 50 14 Pasteurized milk was added to cheese at two soft-acid curd different concentrations (N1, 50 The use of nisin extended the shelf-life of sof			hour before pasteurization. After	acceptable limit until 16 days of storage.	
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11 Milk pudding Nisin A was added to milk pudding to final concentrations of 40, 80, 120, and 240 IU/ mi against spores from B. thuringiensis, B. cereus, and P. jamilae. The samples were incubated at 15, 20, and 30°C for 29 days. > 120 IU/ g in 7.5% fat) was found to be al, 2014 oshina et al, 2014 12 UHT and Raw A cocktail of E. coli phage (EC6, EC9, and EC11) was tested and added to milk samples inculated with E. coli Strains (E. coli ATCC 25922, and E. coli and EC11) was tested and added to milk samples inculated with E. coli OS:H- and E. coli O127:H6 in both UHT and raw milk at 25 °C strains (E. coli ATCC 25922, E. coli and EC9 completely inhibited E. coli O5:H- samples were incubated either at 25 °C for 24 h, or in a domestic refrigerator (5-9°C) for 168 h. A cocktail of the three phages connel tely inhibited E. coli O5:H- and EC9 completely inhibited E. coli O5:H- and E50 mM Na ₂ :EDTA were added to the cheese and packed under MAP. Its completely and 10° for 9.4. Conte et al, specifically with the highest content of the cheese and packed under MAP. Its samples were kept at 8°C for 9.4. Clearance of the pathogen was only achieved by the combined activity of both antimicrobials (after 6 h of incubation). O2010 14 Pasteurized milk 10° Clu/mil of S. achieved by the combined activity of both achieved by the combined activity of both added. The samples were kept at 37 °C for 10 h. The use of nisin extended the shelf-life of for shold with cheese achieved by the combined activity of both antimicrobials (after 6 h of incubation). Sorti act al, althib miltion a			at 4±1°C for 16 d.		
inal concentrations of 40, 80, 120, ≥ 120 IU/ g in 7.5% fat) was found to be al., 2014 and 240 IU/ ml against spores from effective as a natural preservative to B. thuringiensis, B. cereus, and P. control spoilage bacteria and extend its jamilae. The samples were incubated at 15, 20, and 30°C for 29 days. control spoilage bacteria and extend its shell life (4 weeks). 12 UHT and Raw A cocktail of E. coli phage (EC5, EC9, A cocktail of the three phages completely and EC11) was tested and added to milk samples inoculated with E. coli O127:H6 in both UHT and raw milk at 25 °C strains (E. coli ATCC 25922, E. coli and at 5-9 °C. A cocktail containing EC6 OS:H and E. coli 0127:H6). The and EC9 completely Inhibited E. coli OS:H and E. coli 0127:H6). The and EC9 completely Inhibited E. coli OS:H and E. coli 0127:H6 in both UHT and raw milk at 25 °C strains (E. coli ATCC 25922, E. coli and at 5-9 °C. A cocktail containing EC6 OS:H and E. coli 0127:H6). The and EC9 completely Inhibited E. coli OS:H and E. coli 0127:H6). The and EC9 completely Inhibited E. coli OS:H and E. coli 0127:H6 in both UHT and raw milk at 25 °C for 24 h, or in a domestic refrigerator (5-9°C) for 168 h. regrowth was observed. conte et al., and ED9 mM Na ₂ -ED7 A were added to specifically with the highest content of the cheese and packed under MAP. lysozyme. The samples were kept at 8°C for 9 d. 2011 14 Pasteurized Pasteurized milk was inculated with G. Su antiturerobials (after 6 h of incubation). (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mitture of both, were also added. The samples were kept at 37 °C for 10 h. Conte et al., achieved by the combined activity of both activity of both activity of both, were also added. The samples were kept at 37 °C for 10 h. The use of nisin extended the	11	Milk pudding	Nisin A was added to milk pudding to	Nisin A (at \geq 80 IU/ g in 5.0% fat and at	Oshima et
 and 240 IU/ ml against spores from effective as a natural preservative to <i>B. thuringiensis, B. cereus,</i> and <i>P. control spolage bacteria and extend its jamilae.</i> The samples were incubated at 15, 20, and 30°C for 29 days. UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, A cocktail of the three phages completely McLean et inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 and EC9 completely inhibited <i>E. coli</i> O127:H6 in both UHT and raw milk at 25 °C and at 5–9 °C. A cocktail containing EC6 O5:H- and <i>E. coli</i> O127:H6). The samples were incubated either at 25 regretation (5–9 °C) for 168 h. regrowth was observed. C for 24 h, or in a domestic refrigerator (5–9 °C) for 168 h. regrowth was observed. Cheese Lysoryme (150, 250, and 500 ppm). The shelf life of cheese was increased content of 2011 the cheese and packed under MAP. Usyayme. The samples were kept at 8°C for 9 d. Pasteurized Pasteurized milk was inoculated with 10° cfu/ml of 5. <i>aureus</i> 549. Immediately after, nisin added. The samples were kept at 8°C for 9 d. Pasteurized Milk and 0.75 µg /mI), endolysin (7.5 U/ml and 10° cfu/ml of 5. added. The samples were kept at 37 °C for 10 h. Galotyri (Greek Nisin was added to cheese at two soft-acid curd) different concentrations (NI, 50 rds) and stored are of nisin extended the shelf-life of kykkidou et areobically under refrigerator for a maintaining good sensory characteristics. period of 42 days. 			final concentrations of 40, 80, 120,	\geq 120 IU/ g in 7.5% fat) was found to be	al., 2014
B. thuringiensis, B. cereus, and P. control spoilage bacteria and extend its jamilae. The samples were shelf life (4 weeks). incubated at 15, 20, and 30°C for 29 days. 12 UHT and Raw A cocktail of E. coli phage (EC6, EC9) A cocktail of the three phages completely McLean et and EC11) was tested and added to inhibited E. coli ATCC 25922 and E. coli al., 2013 al., 2013 milk samples inoculated with E. coli O127:H61 in both UHT and raw milk at 25 °C and at 5–9 °C. A cocktail containing EC6 al., 2013 05:H- and E. coli O127:H61. and et 5–9 °C. A cocktail containing EC6 control spoilage bacteria and extend its and et 5–9 °C. A cocktail containing EC6 content and EC9 completely inhibited E. coli O5:H- and et 5–9 °C. A cocktail containing EC6 content and EC9 completely inhibited E. coli O5:H- and et 5–9 °C. A cocktail containing EC6 content and EC9 completely inhibited E. coli O5:H- and et 5–9 °C. A cocktail containing EC6 content and EC9 completely inhibited E. coli O5:H- and et 5–9 °C. A cocktail containing EC6 content al., (Burrata) and 50 m/M Nay-EDTA were added to specifically with the highest content of 2011 14 Pasteurized milk was incluated with Clearance of the pathogen was only Garcie et al., adi			and 240 IU/ ml against spores from	effective as a natural preservative to	
jamilae. The samples were incubated at 15, 20, and 30°C for 29 days. McLean et inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 12 UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, and EC11) was tested and added to inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 McLean et inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 12 UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, and EC11) was tested and added to inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> and EC3 completely inhibited <i>E. coli</i> OS:H- and <i>E. coli</i> O127:H6 in both UHT and raw milk at 25 °C 3 C for 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. regrowth was observed. 13 Cheese Lysozyme (150, 250, and 500 ppm), the shelf life of cheese was increased content of the cheese and packed under MAP. The samples were kept at 8°C for 9 d. Clearance of the pathogen was only García et al., achieved by the combined activity of both 2010 14 Pasteurized milk was inoculated with for 9 d. Clearance of the pathogen was only García et al., achieved by the combined activity of both 2010 15 Galotyri (Greek sole). Nisin was added to cheese at two soft-acid curd) The use of nisin extended the shelf-life of thy kikidou et different concentrations (N1, 50) 15 Galotyri (Greek are 00000000000000000000000000000000000			B. thuringiensis, B. cereus, and P.	control spoilage bacteria and extend its	
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12 UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, a cocktail of the three phages completely McLean et inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 12 UHT and Raw A cocktail of <i>E. coli</i> phage (EC6, EC9, a cocktail of the three phages completely McLean et inhibited <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 13 Cheese Cost at <i>C</i> of 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. an enterohemorrhagic strain, in UHT milk at both temperatures, whereas in raw milk regrowth was observed. 13 Cheese Lysozyme (150, 250, and 500 ppm), the cheese and packed under MAP. The samples were kept at 8°C for 9 d. The samples content of the pathogen was only García et al., achieved by the combined activity of both 2011 14 Pasteurized milk was inoculated with milk Clearance of the pathogen was only García et al., achieved by the combined activity of both 2010 2010 14 Pasteurized milk was and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. The use of nisin extended the shelf-life of Kykkidou et fresh Galotyri cheese stored at 4 °C by ca. al., 2007 15 Galotyri (Greek Nisin was added to cheese at two thresh Galotyri cheese stored at 4 °C by ca. al., 2007 10/g, N2 150 IU/g) and stored are brickly under refrigeration for a period of 42 days. The use of nisin extended the shelf-life of Kykkidou et minimization good sensory characteristics.			days.		
 and EC11) was tested and added to milk samples inoculated with <i>E. coli</i> ATCC 25922 and <i>E. coli</i> al., 2013 milk samples inoculated with <i>E. coli</i> Strains (<i>E. coli</i> ATCC 25922, <i>E. coli</i> and at 5–9 °C. A cocktail containing EC6 O5:H- and <i>E. coli</i> O127:H6). The samples were incubated either at 25 °C for 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. Cheese Lysozyme (150, 250, and 500 ppm), (Burrata) Cheese and packed under MAP. The samples were kept at 8°C for 9 d. Pasteurized Pasteurized Pasteurized Pasteurized milk was inoculated with nilk 10² cfu/ml and 10⁵ cfu/ml of <i>S.</i> <i>aureus</i> Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Galotyri (Greek soft-acid curd) Galotyri (Greek soft-acid curd) Galotyri (Greek acrobically under refrigeration (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for period of 42 days. 	12	UHT and Raw	A cocktail of <i>E. coli</i> phage (EC6, EC9,	A cocktail of the three phages completely	McLean et
 milk samples inoculated with <i>E. coli</i> O127:H6 in both UHT and raw milk at 25 °C strains (<i>E. coli</i> ATCC 25922, <i>E. coli</i> and at 5–9 °C. A cocktail containing EC6 O5:H- and <i>E. coli</i> O127:H6). The and EC9 completely inhibited <i>E. coli</i> O5:H- and <i>E. coli</i> O127:H6). The samples were incubated either at 25 , an enterohemorrhagic strain, in UHT milk "C for 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. regrowth was observed. Cheese Lysozyme (150, 250, and 500 ppm), (Burrata) and 50 m/ Na₂-EDTA were added to the cheese and packed under MAP. Iysozyme. The samples were kept at 8°C for 9 d. Pasteurized Pasteurized milk was inoculated with 0¹⁰ cfu/ml and 10⁵ cfu/ml of <i>S. achieved</i> by the combined activity of both 2010 antimicrobials (after 6 h of incubation). (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Galotyri (Greek Nisin was added to cheese at two soft-acid curd) different concentrations (N1, 50 [U/g], N2 150 [U/g] and stored acrobically under refrigeration for a areobically under refrigeration for a period of 42 days. 			and EC11) was tested and added to	inhibited E. coli ATCC 25922 and E. coli	al., 2013
 strains (E. coli ATCC 25922, E. coli O5:H- and E. coli O127:H6). The samples were incubated either at 25 'C for 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. and at 50 m/M Na₂-EDTA were added to the cheese and packed under MAP. (Burrata) and 50 m/M Na₂-EDTA were added to the cheese and packed under MAP. lysozyme. The samples were kept at 8°C for 9 d. Pasteurized Pasteurized milk was inoculated with 0.37 µg/ ml and 10° cfu/ml of S. aureus Sa9. Immediately after, nisi and a mixture of both, were also added. The samples were kept at 37 'C for 10 h. Galotyri (Greek soft-acid curd) Galotyri (Greek soft-acid curd) Misin was added to cheese at two tildefrequencies for 10 h. Salotyri (Greek soft-acid curd) Galotyri (Greek soft-acid curd) Misin was added to cheese at two aerobically under refrigeration for a period of 42 days. Essential oils and herbs 			milk samples inoculated with E. coli	O127:H6 in both UHT and raw milk at 25 $^\circ \! C$	
 O5:H- and <i>E. coli</i> 0127:H6). The samples were incubated either at 25 'a nenterohemorrhagic strain, in UHT milk at both temperatures, whereas in raw milk refrigerator (5-9 °C) for 168 h. Cheese Lysozyme (150, 250, and 500 ppm), Beeifically with the highest content of 2011 the cheese and packed under MAP. Iysozyme. The samples were kept at 8°C for 9 d. Pasteurized Pasteurized milk was inoculated with Clearance of the pathogen was only aureus Sa9. Immediately after, nisin autimicrobials (after 6 h of incubation). (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Galotyri (Greek Nisin was added to cheese at two soft-acid curd) different concentrations (N1, 50 fresh Galotyri cheese stored at 4°C by ca. al., 2007 1U/g, N2 150 IU/g) and stored activity chaese stored at 4°C by ca. al., 2007 7 days (N1) and 21 days (N2) with cheese areobically under refrigeration for a period of 42 days. 			strains (E. coli ATCC 25922, E. coli	and at 5–9 °C. A cocktail containing EC6	
 samples were incubated either at 25 "C for 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. regrowth was observed. Cheese Lysozyme (150, 250, and 500 ppm), (Burrata) and 50 m/ Na₂-EDTA were added to the cheese and packed under MAP. The samples were kept at 8°C for 9 d. Pasteurized milk 10² cfu/ml and 10⁵ cfu/ml of 5. <i>aureus</i> Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Soft-acid curd) Galotyri (Greek soft-acid curd) Salotyri (Greek soft-acid curd) Ciseration (N1, 50 [U/g, N2 150 IU/g) and stored aerobically under refrigeration for a aerobically under refrigera			O5:H- and E. coli O127:H6). The	and EC9 completely inhibited E. coli O5:H-	
°C for 24 h, or in a domestic refrigerator (5-9 °C) for 168 h. at both temperatures, whereas in raw milk regrowth was observed. 13 Cheese Lysozyme (150, 250, and 500 ppm), (Burrata) The shelf life of cheese was increased Conte et al., specifically with the highest content of 14 Pasteurized Pasteurized milk was inoculated with milk Clearance of the pathogen was only aureus Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. The use of nisin extended the shelf-life of fresh Galotyri (Greek Kykkidou et different concentrations (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. The use of nisin extended the shelf-life of faver samples and period Kykkidou et al., 2007			samples were incubated either at 25	, an enterohemorrhagic strain, in UHT milk	
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13 Cheese (Burrata) Lysozyme (150, 250, and 500 ppm), and 50 m/M Na ₂ -EDTA were added to the cheese and packed under MAP. The samples were kept at 8°C for 9 d. The shelf life of cheese was increased Conte et al., 2011 14 Pasteurized milk Pasteurized milk was inoculated with 10 ² cfu/ml and 10 ⁵ cfu/ml of <i>S</i> . <i>aureus</i> Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. The use of nisin extended the shelf-life of fresh Galotyri cheese stored at 4 °C by ca. al., 2007 Kykkidou et al., 2007 15 Galotyri (Greek soft-acid curd) Nisin was added to cheese at two different concentrations (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. The use of nisin extended the shelf-life of aunitaining good sensory characteristics. Kykkidou et al., 2007			refrigerator (5-9 °C) for 168 h.	regrowth was observed.	
(Burrata) and 50 m/M Na2-EDTA were added to the cheese and packed under MAP. The samples were kept at 8°C for 9 d. lysozyme. 14 Pasteurized milk Pasteurized milk was inoculated with 10 ² cfu/ml and 10 ⁵ cfu/ml of S. aureus Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Clearance of the pathogen was only aureus Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Kykkidou et al., 2007 15 Galotyri (Greek soft-acid curd) Nisin was added to cheese at two different concentrations (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. The use of nisin extended the shelf-life of resh Galotyri cheese stored at 4 °C by ca. al., 2007 Kykkidou et al., 2007	13	Cheese	Lysozyme (150, 250, and 500 ppm),	The shelf life of cheese was increased	Conte et al.,
the cheese and packed under MAP. Iysozyme. The samples were kept at 8°C for 9 d. 14 Pasteurized milk 10 ² cfu/ml and 10 ⁵ cfu/ml of S. achieved by the combined activity of both 2010 aureus Sa9. Immediately after, nisin achieved by the combined activity of both 2010 (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), antimicrobials (after 6 h of incubation). (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. °C for 10 h. Kykkidou et 15 Galotyri (Greek Nisin was added to cheese at two The use of nisin extended the shelf-life of Kykkidou et 15 Galotyri (Greek Nisin was added to cheese at two fresh Galotyri cheese stored at 4 °C by ca. al., 2007 10/g, N2 150 IU/g) and stored 7 days (N1) and 21 days (N2) with cheese aerobically under refrigeration for a maintaining good sensory characteristics. period of 42 days. period of 42 days. Essential oils and herbs Herbs		(Burrata)	and 50 mM Na ₂ -EDTA were added to	specifically with the highest content of	2011
The samples were kept at 8°C for 9 d. 14 Pasteurized Pasteurized milk was inoculated with milk Clearance of the pathogen was only achieved by the combined activity of both 2010 García et al., achieved by the combined activity of both 2010 aureus Sa9. Immediately after, nisin (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. 15 Galotyri (Greek soft-acid curd) Nisin was added to cheese at two IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. The use of nisin extended the shelf-life of resh Galotyri cheese Kykkidou et ainitaining good sensory characteristics.			the cheese and packed under MAP.	lysozyme.	
14 Pasteurized Pasteurized milk was inoculated with Clearance of the pathogen was only García et al., milk 10 ² cfu/ml and 10 ⁵ cfu/ml of S. achieved by the combined activity of both 2010 aureus Sa9. Immediately after, nisin antimicrobials (after 6 h of incubation). (0.37 µg/ ml and 0.75 µg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. °C for 10 h. The use of nisin extended the shelf-life of Kykkidou et all, 2007 IU/g, N2 150 IU/g) and stored 7 days (N1) and 21 days (N2) with cheese al., 2007 repriod of 42 days. Essential oils and herbs maintaining good sensory characteristics. maintaining good sensory characteristics.			The samples were kept at 8°C for 9 d.		
 milk 10² cfu/ml and 10⁵ cfu/ml of S. achieved by the combined activity of both 2010 aureus Sa9. Immediately after, nisin (0.37 μg/ ml and 0.75 μg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. 15 Galotyri (Greek soft-acid curd) U/g, N2 150 IU/g) and stored fresh Galotyri cheese stored at 4 °C by ca. al., 2007 IU/g, N2 150 IU/g) and stored fresh Galotyri cheese stored at 4 °C by ca. aerobically under refrigeration for a maintaining good sensory characteristics. period of 42 days. 	14	Pasteurized	Pasteurized milk was inoculated with	Clearance of the pathogen was only	García et al.,
aureus Sa9. Immediately after, nisin antimicrobials (after 6 h of incubation). (0.37 μg/ ml and 0.75 μg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. °C for 10 h. 15 Galotyri (Greek Nisin was added to cheese at two The use of nisin extended the shelf-life of Kykkidou et different concentrations (N1, 50 fresh Galotyri cheese stored at 4 °C by ca. IU/g, N2 150 Berobically under refrigeration for a maintaining good sensory characteristics. period of 42 days.		milk	10^2 cfu/ml and 10^5 cfu/ml of S.	achieved by the combined activity of both	2010
 (0.37 μg/ ml and 0.75 μg /ml), endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Galotyri (Greek soft-acid curd) Nisin was added to cheese at two different concentrations (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. Essential oils and herbs 			aureus Sa9. Immediately after, nisin	antimicrobials (after 6 h of incubation).	
 endolysin (7.5 U/ml and 15 U/ml), and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Galotyri (Greek soft-acid curd) Misin was added to cheese at two different concentrations (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. 			(0.37 μg/ ml and 0.75 μg /ml),		
 and a mixture of both, were also added. The samples were kept at 37 °C for 10 h. Galotyri (Greek soft-acid curd) Misin was added to cheese at two different concentrations (N1, 50 IU/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. 			endolysin (7.5 U/ml and 15 U/ml),		
added. The samples were kept at 37 °C for 10 h. 15 Galotyri (Greek soft-acid curd) Nisin was added to cheese at two soft-acid curd) The use of nisin extended the shelf-life of fresh Galotyri cheese stored at 4 °C by ca. IV/g, N2 150 IU/g) and stored aerobically under refrigeration for a period of 42 days. Essential oils and herbs			and a mixture of both, were also		
 °C for 10 h. 15 Galotyri (Greek soft-acid curd) Nisin was added to cheese at two different concentrations (N1, 50 fresh Galotyri cheese stored at 4 °C by ca. al., 2007 IU/g, N2 150 IU/g) and stored 7 days (N1) and 21 days (N2) with cheese aerobically under refrigeration for a maintaining good sensory characteristics. period of 42 days. 			added. The samples were kept at 37		
15 Galotyri (Greek soft-acid curd) Nisin was added to cheese at two different concentrations (N1, 50 The use of nisin extended the shelf-life of fresh Galotyri cheese stored at 4 °C by ca. al., 2007 10/g, N2 150 IU/g) and stored acrobically under refrigeration for a period of 42 days. The use of nisin extended the shelf-life of fresh Galotyri cheese stored at 4 °C by ca. al., 2007 Essential oils and herbs			°C for 10 h.		
soft-acid curd) different concentrations (N1, 50 fresh Galotyri cheese stored at 4 °C by ca. al., 2007 IU/g, N2 150 IU/g) and stored 7 days (N1) and 21 days (N2) with cheese aerobically under refrigeration for a maintaining good sensory characteristics. period of 42 days.	15	Galotyri (Greek	Nisin was added to cheese at two	The use of nisin extended the shalf-life of	Kykkidou ot
IU/g, N2 150 IU/g) and stored 7 days (N1) and 21 days (N2) with cheese aerobically under refrigeration for a maintaining good sensory characteristics. period of 42 days.	15	soft-acid curd)	different concentrations (N1 50	fresh Galotyri cheese stored at 4 °C by ca	al 2007
aerobically under refrigeration for a maintaining good sensory characteristics. period of 42 days.			U/g, N2 150 U/g) and stored	7 days (N1) and 21 days (N2) with cheese	, 2007
period of 42 days. Essential oils and herbs			aerobically under refrigeration for a	maintaining good sensory characteristics	
Essential oils and herbs			period of 42 days.		
			Essential oils	and herbs	

1	Peda	Peda was incorporated with 1%	The shelf life of herbal pedawas 48 days at	Panday et al.,
		black pepper and 1% turmeric and	7 °C.	2018
		stored at (7±1 °C) for 48 d.		
2	Fresh acid-	Cheese was fortified with (2, 3, and 4	Ethanolic extract of Moringa oleifera	Mohamed et
	curd soft	%) Moringa oleifera extract and	leaves can be used to add nutritional value	al., 2018
	cheese	supplemented with L. plantarumand	and to extend the shelf life of cream	
		L. mesenteroides (1:1). The samples	cheese.	
		were stored at5 °C ± 1 for 30 d.		
3	Labneh	The samples were prepared from UF	The productcan be considered a new	El-Sayed et
		retentate of buffalo's skim milk and	functional product with extended shelf	al., 2017
		fortified with Moringa oleifera oil	life.	
		using three different ratios:10, 15,		
		and 20% and L. acidophilus, too. The		
		samples were stored at 5 °C \pm 1 for		
		30 d.		
4	Ghee	Ghee was treated with curry leaves	The additionof curry leaves at the final	Kapadiaand
		(0.1%, 0.2%, 0.3%, and 0.4%) and	stage of heat clarification was found more	Aparnathi,
		stored at 80±1 °C for an accelerated	effective than at the initial stage of heat	2017
		storage stability test for 12 days.	clarification. The optimum rate of curry	
			leaves found was 0.3%.	
5	Cottage	Five different (parsley, dill, pepper,	The sensorial best result was obtained	Josipović et
	cheese	garlic, and rosemary) herbs were	with fresh sweet red pepper whereas dry	al., 2015
		incorporated in fresh and dried form	rosemary had the highest antioxidant and	
		at 0.5. 1.0 and 2.0%. The samples	antibacterial activity.	
		were stored at 4 $^{\circ}$ C ± 1 for 3 d.		
6	Flavouredmilk	Essential oils were added at 0.005%	Increased antioxidant activity of the	Samaddar et
		(eugenol and trans-cinnamaldehyde	product, ultimately enhances the shelf life	al., 2015
		enriched) in milk and stored at 4-7 °C	of the product.	
		for 7 d.		
7	Labneh	Labneh was supplemented with	It can be concluded that 0.3% of cinnamon	Thabet et al.,
		essential oils cinnamon, cumin, and	can be used to increase the shelf life of	2014
		mint oils, to a final conc. of 0.3, 0.5,	labneh for up to 24 days, with a higher	
		and 0.8%. The samples were stored	level of total volatile free fatty acid and	
		at 6±1 °C for 24 d.	therapeutic bacteria counts and a low level	
			of total viable, mould and yeast count.	
8	Paneer	Brine dipped (5%) and dry salted	The products were microbiologically safe	Rani et al.,
		(3%) masala paneer (coriander	and remained so for at least 6 days when	2014
		leaves (1%), mint leaves (1%) and	stored in a refrigerator.	
		green chilies (0.3%), roasted and		
		groundcumin seeds (0.3%) and black		
		pepper (0.3%)) were prepared and		
		stored at 4±1 °C for 8 d.		

9	Clarified	Ethanolic extract of Arjuna was	The shelf life of the Arjuna ghee samples	Parmar et al.,
	butterfat	added at 7% in butter fat. The	was 8 days at 80 \pm 1 °C as compared to 2	2013
		prepared ghee samples were stored	days in the control.	
		in a hot air oven at 80±1 °C for an		
		accelerated storage stability test and		
		analyzed at regular intervals of 0, 2,		
		4, 6, 8, and 10 days.		
10	Butter	2% of dried rosemary herb and sage	The results of chemical analyses suggest	Najgebauer-
		was added to butter. The samples	that addition of rosemary herb was more	Lejko et al.,
		were packed and stored at 4 °C for 5	effective in retardation of lipolysis than	2009
		months.	supplementation with sage. However,	
			both supplemented products were less	
			stable during storage than the control	
			sample.	
11	Labneh	Three essential oils, namely thyme,	Thyme, marjoram, or sage at 0.2 ppm can	Otaibi and
		marjoram, and sage, were added to	be used to increase the shelf life of labneh	Demerdash,
		concentrated yoghurt (labneh) at	for up to 21 d.	2008
		conc. of 0.2, 0.5, and 1.0 parts per		
		million (ppm) and stored at $5^{\circ}C \pm 1$		
		for 21 d.		

Lysozyme is a glycoside hydrolase that catalyzes the hydrolysis of 1, 4-beta-linkages between N-acetyl-D-glucosamine and N-acetylmuramic acid residues in peptidoglycan- the major component of Grampositive bacterial cell wall. Furthermore, the recent approaches deal with the application of phages infecting the bacteria known as "Bacteriophage". Being obligate parasites, they must first infect host cellsto multiply. Bacteriophages follow either the lysogenic or lytic cycle.

In addition, two types of proteins are encoded by phages, namely virion-associated peptidoglycan hydrolases (VAPGHs) and endolysin (Chang et al., 2017a; 2017b). VAPGHs are associated with the initial phage infection step while endolysins mediate bacterial lysis. Because of host specificity, phages have drawn the attention of food researchers and have been accepted as green technology to safeguard food. Phages have found many other applications other than bio-control, such as improving crop yields, sanitizer for farm facilities, animal and animal handling, treatment and prophylaxis in cattle, etc. Several commercial preparations (SalmoFresh[™] and PhageGuard S[™], PhageGuard Listex[™]) havebeen approved and granted GRAS status by FDA to prevent the growth of pathogens. Several studies haveshown the successful application of host-specific bacteriophages and endolysins which are hydrolytic enzymes synthesized by bacteriophages to cleave the host's cell wall during the final stage of the lytic cycle.

Herbs and essential oils

Herbs and spices have played remarkable roles not just as a food flavouring substances, but also as a medicine and preservative forcenturies. Indeed, there is a great linkage between diseases and food habits. India is known asthe "Botanical Garden of the World" being the largest producer of medicinal herbs. They are valued as equivalent to gold or jewels in many countries. Around 70-80% population in

developing countries and 60% world's population rely directly on herbs and plants for their medicinal uses.

Moreover, as per the report of the World Health Organization, modern or non-conventional medicines based on herbal sources occupy a significant part in primary healthcare of 70% world population (Oraon et al., 2017). According to Ayurveda, the herb/spice is used as a whole plant including the root/rhizomes (turmeric), leaves (tulsi, phudina), bark (cinnamon), flower (clove), fruit (cardamom, ashwagandha), etc., which specifically contains phytosterols, antioxidants, essential oils, vitamins, and other substances that help to inhibit the growth of microorganisms (Oraon et al., 2017).

On the other hand, essential oils are plant-derived aromatic oily liquors, that possess antimicrobial properties and are also known for their therapeutic properties (for instance diuretics, anti-inflammatory, antiseptic, carminative, antispasmodic, and tonic substances). EOs are produced by steam distillation and have gained the status of GRAS, too. Several studies have indicated shelf life extension of different milk products viz. kinds of milk, fermented milk and ghee using herbs and EO (Table 3).

Chemical preservatives

The incorporation of chemical agents in foodhaving antimicrobial activity has been practiced since ancient times. Benzoic acid or its sodium salt, benzoate wast he first molecule approved in the USA. Others include salt, sugar, inorganic salts, etc. (as per the Appendix- A, FSSR, 2011).

Another reason to be added todairy products may deal with the antioxidant properties of chemical preservatives. However, several chemical preservatives have been found to have negative and life-threatening effect son human health, particularly infants.

Packaging

Both, the packaging system and the packaging material of milk and milk products act as external means of preservation after processing, during transportation and distribution (Ščetar et al., 2019; Vieira et al., 2019). It performs various functions like protection, convenience, communication, containment, etc. In the 19th century, milk was packed using glass bottles. The customer's demand for fresh, convenient, easy-to-cook food and the industrial threat of wastage, non-recyclable packaging and global warming issues brought technological innovations in the form of Vacuum packaging, Modified Atmosphere Packaging (MAP), Active packaging, Edible coating, Intelligent or smart packaging, etc. Various packaging materials include LDPE, HDPE, LLPDE, Polystyrene, EV-OH, Aluminium foil, etc. The focus in the next section is given to various packaging systems rather than packaging material. The studies on various packaging systemsto enhance the shelf life of milk and milk products are depictedin Table 4.

Sr.	Type of	Treatment	Result	Reference
NO.	1000	Active packaging		
1	Macaroni and	Ready-to-eat macaroni and cheese filled in novel oxygen scavenger and metal oxide-coated high-	The results indicated that oxygen scavenger and high-barrier	Patel et al., 2019
	cheese	barrier polymer packages, were processed in pilot scale 915-MHz microwave-assisted thermal sterilization system (MATS). Also, aluminum foil packages were processed in the Allpax retort system	packaging can be used for ready-to- eat meals with extended shelf life for soldiers and astronauts.	

		to compare packaging performance. The samples were stored for 6 months at 37.8 °C.		
2	White fresh cheese	The sample was stored in PET waste-based active packaging films (containing chitosan and Ag: silver nanoparticles, at three storage temperatures (6, 25, and 40 °C), for various durations (0-30 days).	5% Ag–Cs–PET90:10 film is efficient enough to make cheese bacteria- free within 7 days at 40 °C. The PET waste-based active packaging film is found to be capable of a shelf life extension of white fresh cheese for up to 30 days.	Singh et al., 2018
3	UF cheese	<i>L. monocytogenes</i> inoculated cheese samples were put in polyethylene bags (cellulosic paper coated with chitosan-zinc oxide nanocomposite containing nisin (500 ppm and 1000 ppm)), and stored at 4 ± 1 °C for 14 days.	Films with 1000 µg/mL of nisin completely inactivated <i>L.</i> <i>monocytogenes</i> cheese after storage at 4 °C for 14 days.	Divsalar et al., 2018
4	Mozzarel la cheese	Cheese slices were inoculated with 10 ⁶ spores/ml of Alatoxigenic strains of <i>P. digitatum</i> CECT 2954 and <i>A. parasiticus</i> CECT 2681 packed with either of the following antimicrobial devices: (1) paper filter with AIT (; (2) AIT sticker; or (3) oriental mustard meal pouchand were placed in plastic bags or plastic trays composed of MAP material. The samples were kept at 4°C and were observed for 60 d. AIT= Allyl isothiocyanate	Doses ≥4 µL/L of AIT may significantly increase the shelf life and safety of sliced mozzarella cheese.	Tracz et al., 2018
5	Butter container	Two types of oxygen absorbers adhesive labels and sachets and two caps with and without adjustable closure were tested.	The best results were achieved with the oxygen absorber sachets and using caps with adjustable closure. Under these conditions, the oxygen concentration inside the container remained below 3% during 150 h.	Otero-Pazos et al., 2018
6	Cheese	Two active films based on chitosan (1.5% w/v) and methylcellulose (3% w/v) enriched with natamycin (0.01% (w/v)) were used to cover and store cheese at 20 °C for 7 days.	A significant reduction in yeast and mould was observed in cheese samples treated with chitosan films containing natamycin (p < 0.05).	Santonicola et al., 2017
7	Queso Blancoch eese	Cheese was inoculated with a cocktail of the <i>L.</i> monocytogenes strains wrapped in the foxtail millet starch (3.5%, w/v) and clove leaf oil (1.0%), and stored at 4 °C for 24 d.	The FMS films that contain clove leaf oil can be used as a new packaging material for enhancing the shelf life of Queso blanco cheese.	Yang et al. 2018
8	Milk	Milk (2% Fat, 131.1 °C for 2 s) was stored in HDPE packages consisting of TiO_2 at 3 levels (low: 0.6%; medium: 1.3%; high: 4.3%) at 3°C for up to 43 d. Light-protected (translucent, foil-wrapped) and light-exposed (translucent) HDPE packages served as controls.	The high TiO ₂ -HDPE package provided protection similar to the light-protected control package through 22 days of light exposure, with less consistent performance thanthe medium TiO ₂ package.	Johnson et al., 2015

9	Milk pomade sweet- Sherbet	The samples were packed in different packaging materials using several packaging technologies (MAP). For reduced oxygen packaging (ROP) creation $(O_2 - 0\%)$ in pouches, an iron-based oxygen scavenger sachets of 100 cc were used. The samples were stored at the room temperature of +21±1 °C for 12 weeks.	Met. BOPET/PE and Aluthen are considered the best material for extending the shelf-life.	Ungure et al. 2012		
10	Fior di Latte cheese	Three concentrations (10, 15, and 20 mg) of silver montmorillonite embedded in agar were used to pack the cheese during storage at 10 °C for 7 d.	The active packaging system markedly increased the shelf life of Fior di Latte cheese.	Incoronato et al., 2011		
11	UHT milk	Indirectly processed UHT milk was packaged in IntaseptTM aseptic pouches with (treatment) or without (control) oxygen-scavenging film and stored for 14 weeks at 26 \pm 0.3 °C.	Oxygen content reduced (p <0.05) by 23–28% as well as some volatile compounds contributing to flavor also reduced by 23–41% during storage. However, the consumer panel failed to detect a significant difference in odour between the treatment and control samples.	Perkins et al., 2007		
		Edible coating				
1	Gouda cheese	CNANI (water, natamycin, and nisin) and GNANI (Starch, glycerol, water, natamycin, and nisin) with final conc. of 12.5 ppm of nisin and 50 ppm of natamycin were kept for ripening at 10 ± 1 °C and at humidity 79 ± 7% controlled Chamber for 30 d.	It was observed that these coatings did not alter the physicochemical properties (pH, ash, protein, chloride, water activity, ripening index, and colour) and the development of Lactobacilli that takes place during the ripening of Gouda cheese. The GNANI applied on Gouda cheese resulted in an improved barrier against external contamination during ripening, compared to CNANI.	Berti et al., 2019		
Z	Soft cheese Kleo	packaging material was used to pack soft cheese either in vacuum (as control) or in MAP consisting of carbon dioxide CO_2 (E 290)-30% and nitrogen N_2 (E 941)-70%. The samples were stored at the temperature of 4.0±0.5 °C up to 32 d.	days, and good outside appearance and lactic acid aroma was observed.	Muizniece- Brasava et al., 2011		
3	Ricotta cheese	Cheese samples were dipped into a coating solution (0.8% w/v) of chitosan and 2.4% w/v lyophilized milk whey (11% WP) for 30 sec and packed under MAP (40% CO2/60% N2 mixture). Both samples; control and treated, werestored at $4\pm$ 0.5 °C for 30 d.	Findings suggest a potential utility of chitosan/whey protein coatings to extend fresh dairy product shelf- life based on physico-chemical and microbial parameters.	DiPierro et al., 2011		
4	Cheese	Galactomannan (0.5% (w/v) and nisin at 50 IU/ g used to coat <i>L. monocytogens</i> added dried cheese samples followed by storage at 4 ± 2 °C for 28 days in a cooled chamber at 75% relative humidity (RH).	Results showed that the cheese coated with nisin-added galactomannan film presented the best results in terms of microbial growth delay (p < 0.05).	Martins et al., 2010		
5	Cheese	Chitosan, galactomannan from <i>Gleditsia triacanthos</i> , and agar from <i>Glacilaria birdiae</i> were tested, at 0.5% and 1.5% (w/v) conc.	The uncoated cheese had an extensive mold growth at the surface when compared with the coated cheese.	Cerqueira et al., 2009		
	Modified atmospheric packaging (MAP)					

1 2 3	"Anthotr yros" (whey) cheese Yoghurt	MAP mixtures were 30%/70% CO ₂ /N ₂ (M1) or 70%/30% CO ₂ /N ₂ (M2), while VP was taken as the control sample. The samples were stored at 4 or 12 °C for 37 or 17 days, respectively. Fermentation was done under HP (0.1, 10, 20, 30, and 40 MPa at 43 °C) followed by storage at 4 °C for 23 d.	Both MAP conditions extended the shelf-life of fresh Anthotyros cheese stored at 4 °C by ca. 10 days (M1) or 20 days (M2) compared with VP and by ca. 2 days (M1) and 4 days (M2) at 12 °C, with cheese maintaining good sensory characteristics. Fermentation at higher pressure affectsthe LAB count, syneresis, and firmness. The pH remains unaffected throughout the storage. A combination of CO ₂ and N ₂ was	Vieira et al., 2019 Vieira et al., 2019 Geetha et al.,
	jalebi	potassium sorbate 800 ppm and packed under MAP (100% CO ₂ , 100% N ₂ , and 50-50% CO ₂ and N ₂). The samples were stored at 28±2 °C.	found more suitable for packaging. The shelf life of the product was 40 days.	2017
4	Fresh mozzarell a cheese	Different proportions of CO ₂ and N ₂ were used to pack the samples. All samples were stored at 4 °C for 6 weeks.	Atm 3 (packaged in 40% CO ₂ / 60% N ₂), exhibited the best sensory characteristics of the investigated cheese samples during the storage period. It also allowed a shelf-life extension of 46 days at 4 °C compared to vacuum.	Felfoul et al., 2017
5	Soft Surface mould ripened Cheese	MAP-A (0% O ₂ , 27 <u>+</u> 6% CO ₂) and MAP-B (2 <u>+</u> 1% O ₂ , 19 <u>+</u> 2% CO ₂) was studied at 12 °C for 14 days.	The shelf-life of surface mould ripened cheese with low levels of O_2 (1-3%) and relatively high levels of CO_2 (17-21%) extended up to 17 days.	Rodriguez- Aguilera et al., 2011
6	Fresh Stracciat ella cheese	Combination of CO ₂ :N ₂ :O ₂ 50:50:0 (M1), 95:5:0 (M2), 75:25:0 (M3), and 30:65:5 (M4) vol/vol, for packaging of cheese was used and then stored at 8 °C for 8 d.	M1 and M2, delayed the microbial growth of spoilage bacteria, without affecting the dairy microflora, and prolonged the sensorial acceptability limit.	Gammariello et al., 2009
7	Whey cheese; Lor cheeses	The productwas exposed to different MAP conditions and kept in the refrigerator at 4°C for 45 days.	Sensory evaluation (odour and taste) results showed that Lor cheese packaged under modified atmosphere packaging ($60\% \text{ CO}_2/$ $40\% \text{ N}_2$ and $70\% \text{ CO}_2/$ $30\% \text{ N}_2$) retained good characteristics for 45 days of storage compared to other samples.	Temiz et al., 2009
8	Ricotta	Three gas mixtures were used: $50:50 (CO_2:N_2)$, (MAP50), 70:30 (CO ₂ :N ₂) (MAP70), and $95:5 (CO_2:N_2)$ (MAP95), the control sample was packed under a normal atmosphere. All samples were stored at 4 °C for 8.	The longest shelf life was obtained with MAP containing 95% carbon dioxide compared to the control.	Del Nobile et al., 2009

Modified atmosphere packaging

Solubilization of gas into the water phase is the basis of surrounding food with a gas mixture. It deals with the modification/alteration of the gaseous environment during the packaging of food through the removal and replacement of gas or a mixture of gases. The major gases used are CO_2 , N_2 , and O_2 (Ščetar et al., 2019). The N_2 has no taste, acts as an inert gas, and prevents package collapse. The O_2 is used

almost negligible due to its negative role during the storage of non-respiring food and hence, replaced with CO_2 . O_2 is combined with CO_2 and N_2 only in case of respiring food products. The permeability of packaging material towards O₂, CO₂, and water is essential in determining the effective atmosphere inside the package. The atmosphere is modified either passively or actively in foods (Vieira et al., 2019). In the case of former, the rate of atmosphere modification is slow and happens due to interaction between food (respiration) and gases present in the surrounding environment. The phenomenon is to be regulated by packaging material. On the other side, active modification is faster and accomplished by creating a vacuum and flushing the desired mixture of gases in the package. However, it is more expensive than the passive method. MAP has been found effective against yeast and mould growth and subsequently production of aflatoxin in varieties of cheese (Jalilzadeh et al., 2015). Other gases like carbon monoxide, helium, neon, argon, ethanol vapour, sulfur dioxide, nitrous oxide, etc. can be applied at the commercial level on a restricted basis to improve the shelf life of various foods. MAP gas leak indicator additionally confers the integrity of the package during transportation and storage. The ultraviolet-activated visual oxygen indicators have been most popularized. These indicators are composed of an electron donor, semiconductor, and redox dye. The result is indicated in the form of a change in colour. It prevents or slowsdown the microbial or chemical deterioration of food. The dairy products packed (cheese, jalebi, yoghurt) under MAP showed longer shelf life (Felfoul et al., 2017; Geetha et al., 2017).

Active packaging

The condition of packaged food is changed to extend the shelf life by the addition of active component into the packaging material. It involves various components that act as active ingredients, i.e. O₂ scavengers, CO₂ emitting/absorbing, Ethanol emitters/scavengers, Anti-oxidants releasing, etc. (Ščetar et al., 2019) that react continuously with the inside environment of food. In addition to its function asactive packaging, smart/intelligent packaging, known as non-traditional packaging, provides information about the quality of food during transportation and storage through a device placed internally or externally on the package. Further, different types of methodology can be used in active packaging technology, e.g. releasing agents, absorbers (O₂, CO₂, moisture, taint), etc. iron-based scavengers under the trade name "Ageless" are produced by the leading Mitsubishi Gas Chemical Company, in Japan (Ganguly et al., 2017).

Generally, desiccants containing moisture absorbers and ethanol vapour generating material are used as active packaging for cheese. CO₂ absorbents are based on either physical (active carbon powder or zeolite) or chemical (calcium hydroxide/magnesium hydroxide) composite. Moisture (Drip) absorbent pad includes super absorbent polymers carboxymethyl cellulose, modified starches and polyacrylate salts.

Ethylene absorber is an example of an absorbent that scavenges off flavour-generating compounds. Aroma-Can[®] provides orange or lemon flavour in the food product (Haghighi-Maneshet al., 2017). Potassium permanganate, being a catalyst, oxidizes ethylene to water and CO₂.

The combination of charcoal and palladium helps in delaying the softening rate of fruits and vegetables by preventing ethylene accumulation. Sometimes the antimicrobial agents viz. organic acids, bacteriocins, phenolic compounds, plant extracts, etc. are embedded within the packaging material which is designed to be released slowly and to function at the surface of a food product. The incorporation of essential oils, nano-composites of Ag, Zinc oxide, and O₂ scavengers into packaging material markedly increase the shelf life of cheese and milk (Singh et al., 2018; Divsalar et al., 2018; Yang et al., 2018).

Edible coating

Biodegradable or edible packaging/coating of food opens up a new avenue for the industry. It is a thin layer made from edible material, that function ssimilarly to the conventional packaging material with aided benefits, i.e. carrier for active compounds, and potentially acts as a barrier for flavour compounds, water vapour, and gases. Based on the internal structural molecule, edible films are classified into three types; a) composites, a mixture of different hydrocolloids and lipids with each other, b) hydrocolloids, prepared from protein and polysaccharides and c) lipids.

However, the mechanical strength is the main drawback of edible coating, and therefore secondary nonedible packaging is still required for proper and hygienic handling of food. Chitosan and galactomannan are mainly used for cheese packaging from the polysaccharide group. The protein-based coating material attracts the interest of industry due to its strong mechanical and barrier properties (Ščetar et al., 2019). Examples are collagen, fish proteins, wheat gluten, casein, ovalbumin, soy protein isolate, whey protein isolate, etc.

Amongst all, milk proteins are found most suitable due to their nutritional value, emulsifying properties, solubility in water, and industrial surplus.

The plasticizers/surfactants are usually added. The neutralor non-reacting/interfering nature of coating material with the food is an important characteristic. Various compounds such as cinnamaldehyde, linalool, carvacrol, thymol, lauramide arginineethyl ester (LAE), cocoa extract, natamycin, nisin have been incorporated into different coating/film made up of chitosan, galactomannan, polylactic acid, whey protein and have extended the shelf life of various types of cheeses and other food products (Muizniece-Brasava et al., 2011; Ungure et al., 2012; Berti et al., 2019).

Sr. No.	Method/Technology	Advantages	Disadvantages	
1	Thermal treatment	 Highly efficient amongst all otherprocesses. Well proven in the dairy industry. 	 Destruction of heat-sensitive nutrients. Costly. Fouling rate of equipment is high. Supply chain must be operated under refrigeration temp. Complex process. 	
2	Microwave	 Faster heating rate. Requires shorter processing time. Better quality (nutrition) retention. Ease of operation. Requires less space and also consumes low energy. 	 Non-uniform temperature distribution. Not suitable for solid and semi-solid food. Surface heating more rapidly than the inner part. 	
3	НРР	 Rapid and uniform pressure distribution. Does not produce any significant physico-chemical changes. Retains flavour, colour, and nutritional value. Extends shelf life up to 2-3 fold. 	 Food must contain water. May not inactivate spores and enzymes. High installation cost. Structurally fragile product; needs special attention. 	
4	PEF	 Less treatment time and temperature. Retains nutritional quality. Used to pasteurize the milk. 	 High capital cost. Spores are found resistant. May not be suitable for solid foods. Air bubble interferes with the process. 	

Table 5. Advantages and disadvantages of the different shelf life enhancement technologies

5	Cold Plasma	•	Microbial inactivation at low temp.	•	Control of chemistry of gas plasma
		•	Negligible impact on the food		reactions.
			matrix.	•	Oxidation in high-fat products.
		•	Applied to both solid and liquid	•	Difficult with bulky and irregularly shaped
			food.		food.
		•	Energy efficient process.	•	Penetration power is lower.
6	Ohmic Heating (OH)	•	Rapid and uniform heating.	•	Lack of generalized information.
		•	Reduced fouling of equipment.	•	Required adjustment according to the
		•	Low maintenance cost.		conductivity of the product.
		•	Environmentally friendly process.	•	Difficult to monitor and control.
7	Ultrasonication	•	Inactivates enzymes and Mos.	•	Causes oxidation of products.
		•	Environmentally friendly process.	•	Specific frequency and time.
8	Irradiation	•	Used for both continuous and batch	•	Oxidation of protein (removal of H ⁺ from
			operation.		amino acids).
		•	Does not produce any chemical	•	Certain vitamins get affected.
			residue.	•	Opaque fluid and solid content reduce
		•	Low maintenance cost.		effectiveness.
		•	Environmentally friendly process.	•	Human contact results incancer.
9	MAP	•	Shelf life increasedby 50% to 400%.	•	Increase in processing cost.
		•	Reduced economic losses.	•	Gas combination optimization.
		•	Odourless.	•	Collapse pack due to CO ₂ .
		•	Little or no need forpreservatives.	•	Favours the growth of anaerobes.
10	Edible coating	•	Limits migration of aroma, flavour,	•	Less stability.
			and moisture.	•	Reaction between additives and packaging
		•	Gas barrier.		material.
		•	Protects food.		
		•	Controlled migration of additives.		
		•	Zero wastage.		

Conclusion and future prospective

Milk, yoghurt, cheese, curd or dahi, and other milk products have been consumed for thousands of years and are considered an important part of the human diet to date. The extension of shelf life is meant to preserve the nutritional quality of milk. Apart from preservation, the traditional methods of processing milk and milk products severely affect the nutrition contents, especially vitamins as well as taste, flavour, and colour. Industries are continuously engaged insearching for innovative approaches to satisfy the hunger of consumers for minimally processed and healthy food. Thus, the industry has focused on three major areas; non-thermal techniques, additives, and packaging, all of which are being continually researched. Many of them, including UHT, HPP, PEF, irradiation, and the use of bio preservative sare successfully adopted at the industry level to increase the shelf life of various types of milkand milk products. Microwave-assisted novel techniques like PEF, HHP, and Ultrasoncation have already been studied for improving the quality of food and processing efficiency producing promising results. Similarly, the combination of biopreservatives such as lysozyme bacteriocins, or EO with HHP or PEF showed tremendously encouraging results due to their synergistic effects.

The useof biopreservatives will become a great tool when it comes to prolonging shelf life in the upcoming time. However, each process has its benefits and limitations (discussed in Table 5) and therefore needs to be optimized according to the product profile. Even though cold plasma has poor penetration power such combinations can improve its effectiveness. Further, in-depth understanding would definitely help to change the mindset of consumers to facilitate the acceptance of new technologies in the future.

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