

# Exploration of new platforms and potentials in regenerative medicine

Nikola Štoković<sup>1,2</sup>, Natalia Ivanjko<sup>1,2</sup>, Marko Pećin<sup>3</sup>, Tatjana Bordukalo – Nikšić<sup>1,2</sup>, Vera Kufner<sup>1,2</sup>, Igor Erjavec<sup>1,2</sup>, Tamara Božić<sup>4</sup>, Marina Milešević<sup>1,2</sup>, Viktorija Rumenović<sup>1,2</sup>, Ivona Matić Jelić<sup>1,2</sup>, Valentina Blažević<sup>1,2</sup>, Ana Smajlović<sup>3</sup>, Hrvoje Capak<sup>5</sup>, Zoran Vrbanac<sup>5</sup>, Ana Javor<sup>5</sup>, Dražen Vnuk<sup>3</sup>, Hermann Oppermann<sup>4</sup>, Dora Adanić<sup>4</sup>, Mihaela Perić<sup>2,6</sup>, Dražen Matičić<sup>3</sup>, Slobodan Vukičević<sup>1,2</sup>

<sup>1</sup> Laboratory for Mineralized Tissues, Centre for Translational and Clinical Research. University of Zagreb School of Medicine, Zagreb, Croatia

<sup>2</sup> Center of Excellence for Reproductive and Regenerative Medicine, Zagreb, Croatia

<sup>3</sup> Clinics for Surgery, Orthopedics and Ophthalmology, Faculty of Veterinary Medicine, University of Zagreb, Zagreb, Croatia

<sup>4</sup> Genera Research, Kalinovica, Croatia

<sup>5</sup> Department of Radiology, Ultrasound Diagnostics and Physical Therapy, Faculty of Veterinary Medicine, University of Zagreb, Zagreb, Croatia

<sup>6</sup> Department for Intracellular Communication, Centre for Translational and Clinical Research, University of Zagreb School of Medicine, Zagreb, Croatia

OPEN ACCESS

**Correspondence:**  
Slobodan Vukičević  
vukicev@mef.hr

This article was submitted to RAD  
CASA - Medical Sciences  
as the original article

**Conflict of Interest Statement:**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Received:** 30 May 2024

**Accepted:** 3 June 2024

**Published:** 25 June 2024

**Citation:**

Štoković N, Ivanjko N, Pećin M, Bordukalo – Nikšić T, Kufner V, Erjavec I, Božić T, Milešević M, Rumenović V, Matić Jelić I, Blažević V, Smajlović A, Capak H, Vrbanac Z, Javor A, Vnuk D, Oppermann H, Adanić D, Perić M, Matičić D, Vukičević S. Exploration of new platforms and potentials in regenerative medicine 563=66-67 (2024): 48-56  
DOI: 10.21857/m3v76t1j6y

Copyright (C) 2024 Štoković N, Ivanjko N, Pećin M, Bordukalo – Nikšić T, Kufner V, Erjavec I, Božić T, Milešević M, Rumenović V, Matić Jelić I, Blažević V, Smajlović A, Capak H, Vrbanac Z, Javor A, Vnuk D, Oppermann H, Adanić D, Perić M, Matičić D, Vukičević S. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## ABSTRACT

Regenerative medicine is focused on the regeneration of various tissues and organs through the development of innovative therapeutic strategies and devices. Center of Excellence for Reproductive and Regenerative Medicine (CERRM) was established to provide necessary resources for conducting the state-of-the-art research in the field and to foster translation of developed therapeutic solutions to clinical practice. CERRM subunit for regenerative medicine has developed Osteogrow-C, a novel osteoinductive device for bone regeneration, which was successfully tested in various preclinical models including rabbit and sheep posterolateral spinal fusion (PLF) models as well as rabbit ulnar segmental defect model. Furthermore, CERRM has developed anti-BMP1.3 antibody-based therapy for various diseases that share fibrosis as a key pathological mechanism. This review paper provides the most important contemporary findings on the development of novel BMP-based devices for bone regeneration as well as the development of antifibrotic therapies.

**KEYWORDS:** Osteogrow, Osteogrow-C, rhBMP6, bone regeneration, posterolateral spinal fusion, regenerative medicine

## SAŽETAK

### ISTRAŽIVANJE NOVIH PLATFORMI I POTENCIJALA U REGENERATIVNOJ MEDICINI

Regenerativna medicina je usmjerena na regeneraciju različitih tkiva i organa kroz razvoj inovativnih terapijskih strategija i novih lijekova. Centar izvrsnosti za reproduktivnu i regenerativnu medicinu (CERRM) osnovan je kako bi pružio potrebne resurse za provođenje suvremenih pretkliničkih istraživanja i potaknuo translaciju razvijenih terapijskih rješenja u kliničku praksu. Podjedinica CERRM-a za regenerativnu medicinu je razvila Osteogrow-C, novi osteoinduktivni lijek za regeneraciju kostiju, koji je uspješno testiran u različitim pretkliničkim modelima uključujući modele posterolateralne spinalne fuzije kod kunića i ovaca te model segmentalnog defekta lakatne kosti kod kunića. Nadalje,

CERRM je razvio terapiju temeljenu na anti-BMP1.3 protutijelima za liječenje različitih bolesti koje dijele fibrozu kao ključni patološki mehanizam. Ovaj rad daje pregled razvoja i kliničkih studija novih terapija za koštanu regeneraciju temeljenu na koštanim morfogenskim proteinima te pregled razvoja inovativnih antifibrotičkih terapija.

**KEYWORDS:** Osteogrow, Osteogrow-C, rhBMP6, regeneracija kostiju, posterolateralna spinalna fuzija, regenerativna medicina

## 1. INTRODUCTION

Regenerative medicine is among the most propulsive scientific fields focused on the regeneration of various tissues and organs employing different strategies and approaches. In order to position Croatia as one of the leading EU countries in the field of regenerative medicine, Center of Excellence for Reproductive and Regenerative Medicine (CERRM) was established as a collaboration between leading academic institutions in Croatia including the University of Zagreb School of Medicine, University of Zagreb Faculty of Veterinary Medicine, University Hospital Center Zagreb, University Hospital Dubrava, University of Rijeka School of Medicine, Ruđer Bošković Institute as well as private companies Genera Research, Fidelta, and Smart Medico. In the past period (2017-2023) CERRM has conducted a major project „Reproductive and Regenerative Medicine – Exploration of new platforms and potentials (KK.01.1.1.01.0008)” that has resulted in unprecedented success in the development of novel diagnostic and therapeutic solutions in the field. Specifically, the CERRM subunit for Regenerative Medicine has developed Osteogrow-C, a novel osteoinductive device for bone regeneration, which was tested in various preclinical models with current efforts for initiation of clinical trials (1-8). Furthermore, CERRM has developed BMP1.3 antibody-based therapy for various diseases that share fibrosis as a key pathological mechanism (9-12). This review aims to provide an overview of the recent advancement in the development of novel devices for bone regeneration and BMP1.3-based anti-fibrotic therapy.

## 2. DEVELOPMENT OF NOVEL BMP-BASED THERAPEUTIC STRATEGIES FOR BONE REGENERATION

Bone morphogenetic proteins (BMPs) are growth factors and members of the TGF $\beta$  superfamily which are possessing potent osteoinductive properties (13-19). Due to their ability to induce bone formation, osteoinductive BMPs have been widely investigated as a key component of novel therapeutic solutions for bone regeneration and as a substitution for autologous bone graft (ABG) which is currently the gold standard for promoting spinal fusion as well as treatment of segmental defects and bone

fracture nonunions (20-28). Effective BMP application requires carrier which is biocompatible, easily manufactured with desired biomechanical properties, enabling vascular and cellular invasion, and in the end, provide good retention of an osteoinductive molecule of interest (29-34). BMP carriers are divided into synthetic polymers, natural polymers, and inorganic materials (35, 36). Synthetic polymers (e.g., polylactic acid-PLA, polyglycolic acid-PGA, polyethylene glycol-PEG, poly-E-caprolactone-PCL, polypropylene fumarate-PPF) and inorganic materials (e.g., calcium phosphate ceramics, bioglass, calcium sulfate cement) have been evaluated as a BMP carrier in numerous preclinical trials, however they have not been introduced to clinical practice (35, 37-39). On the other hand, natural polymers (e.g., collagen, hyaluronic acid, fibrin, chitosan, gelatin) have been the most commonly used BMP carriers in preclinical studies which ultimately led to the approval of currently only commercially available BMP-based osteoinductive device, Infuse, which uses collagen as a carrier in a form of absorbable collagen sponge in combination with recombinant human Bone morphogenetic protein (rhBMP2) (13, 31, 40-45). Infuse is currently approved for anterior lumbar interbody fusion (ALIF), acute tibial fracture treatment, and maxillofacial reconstructions (13). However, its *off-label* applications in various spinal fusion procedures have led to severe side effects, necessitating the development of safe, effective, and affordable autologous bone graft substitutes (ABGS) (46-49). Addressing this need involves ongoing research of novel BMP carriers, focusing on exploring combinations of natural or synthetic polymers with inorganic materials to optimize BMP delivery and consequently bone regeneration (13, 19, 29). Currently, one of the most promising BMP-based therapeutic solutions for bone regeneration is Osteogrow, an autologous bone graft substitute (ABGS) developed at the Laboratory for Mineralized Tissues at the University of Zagreb School of Medicine (1, 2). Osteogrow consists of recombinant human Bone Morphogenetic Protein 6 (rhBMP6) delivered within autologous blood coagulum (ABC) as a carrier (1, 2). In comparison to other osteoinductive BMPs, rhBMP6 showed superior ability

to promote osteoblast differentiation *in vitro* and induce bone regeneration *in vivo* when compared to BMP2 and BMP7 (50-52). Additionally, ABC serves as an ideal physiological carrier, suppressing foreign body responses, facilitating tight rhBMP6 binding with plasma proteins within the fibrin meshwork, and allowing sustained *in vitro* release of rhBMP6 (53-55).

Following successful preclinical studies, Osteogrow underwent Phase I/II trials in patients with distal radial fractures (DRF; EudraCT 2014-005101-21) and patients undergoing high tibial osteotomy (HTO; EudraCT 2015-001691-21) (56, 57). For applications where compressive forces are present, Osteogrow requires supplementation with a compression-resistant matrix (CRM). Initially, Osteogrow-A was developed, consisting of rhBMP6/ABC with the addition of allograft particles as CRM (58). Osteogrow-A underwent successful evaluation in rabbit and sheep posterolateral spinal fusion (PLF) models (2) and a Phase II clinical trial (EudraCT number 2017-000860-14) as part of the Horizon2020 project OSTEOproSPINE (2018-2023; GA 779340). However, the use of allograft is associated with several disadvantages, including immunogenicity and regulatory issues in different markets (24, 59, 60). Hence, our focus shifted towards identifying an alternative to allograft, leading us to explore calcium phosphate (CaP) synthetic ceramics as a viable substitute (60, 61). CaP ceramics offer versatility in shape, size, and chemical composition, with tricalcium phosphate (TCP) and hydroxyapatite (HA) being the most utilized variants (38, 62-75). Their key distinction lies in resorbability post-implantation, with TCP being highly resorbable while HA remains stable (76). Additionally, biphasic calcium phosphate (BCP) combines TCP and HA in varying ratios to achieve optimal resorbability (66, 76). Our efforts culminated in the development of novel Osteogrow formulation with addition of synthetic ceramics as a CRM - Osteogrow-C. Extensive animal studies were conducted to optimize implant properties and determine safety and efficacy in relevant preclinical models (3-7, 77, 78). Osteogrow-C was evaluated in several animal models ranging from initial to advanced models (3-7, 30, 53, 77-81). Rodent models (rats, mice) serve as the initial evaluation stage, facilitating rapid assessment of novel therapies potency and elucidating osteogenesis mechanisms (30, 80, 82-84). Promising therapies identified in rodent models progress to clinically relevant intermediate (rabbit) and larger animal models (sheep, dogs, non-human primates - advanced stage). In rat subcutaneous model, Osteogrow-C induced bone formation, with studies indicating that smaller particle sizes of rhBMP6 resulted in a higher bone volume (3). Moreover, the chemical composition of particles showed no significant impact on bone induction at the rat ectopic site (3).

In the subsequent phase of preclinical trials (Figure 1), optimal formulations of Osteogrow-C were selected for evaluation in pertinent models of posterolateral spinal fusion (PLF) and segmental defects (4, 5, 7). Specifically, chosen Osteogrow-C

formulations underwent assessment in a rabbit PLF model with follow-up periods of 7, 14, and 27 weeks (5). Early observations indicated that Osteogrow-C facilitated rapid spinal fusion between adjacent transverse processes (5). Moreover, extended follow-up revealed significant differences in resorption between TCP ceramics and biphasic ceramics containing high HA proportion. Nonetheless, these differences in residual ceramic amounts did not affect the biomechanical properties of the newly formed bone for spinal fusion. Building upon the success of rabbit PLF studies, the safety and efficacy of Osteogrow-C were further validated in sheep PLF models (7). In these studies, Osteogrow-C with 74-420  $\mu\text{m}$  and 1000-1700  $\mu\text{m}$  ceramic particles induced radiographic solid fusion 9 weeks following implantation. Moreover, spinal fusion and osseointegration with native bone were confirmed at the end of the follow-up period (14, 27 and 40 weeks following surgery) by microCT and histological analyses as well as biomechanical testing (7). Additionally, Osteogrow-C underwent evaluation in a rabbit ulnar segmental defect model, showcasing superiority over both Osteogrow and Infuse. Finally, the safety and efficacy of Osteogrow-C in treating large segmental defects were demonstrated in a case involving a dog with a significant gunshot defect of the humerus, successfully restored through the application of Osteogrow-C implants containing TCP ceramics (8).

### 3. DEVELOPMENT OF NOVEL ANTIFIBROTIC THERAPY BASED ON ANTI-BMP1.3 ANTIBODIES

Fibrosis is a key pathogenetic mechanism in many crucial diseases including chronic kidney disease, liver cirrhosis, and ischemic heart disease (85, 86). It is defined as the excessive accumulation of fibrous connective tissue in diverse organs and tissues. (86). Unfortunately, therapeutic interventions for fibrotic ailments remain scarce, often accompanied by grim prognoses (86, 87). Bone morphogenetic protein 1 (BMP1) was originally isolated from bone alongside other BMPs due to its heparin affinity but it diverges from the conventional BMP protein family and belongs to the astacin/BMP1/tolloid (TLD)-like family of zinc metalloproteinases (88-90). Recent advancements have unveiled multiple splice variants of BMP1 stemming from the same gene, denoted by sequential suffixes ranging from BMP1.1 to BMP1.7 (91-93). Both BMP1.1 and its isoform BMP1.3 play pivotal roles in converting various extracellular matrix (ECM) precursors into functional proteins. These include pro-collagens C I-III, small leucine-rich proteoglycans (such as decorin and biglycan), laminin, collagen VII, and perlecan within the basal membrane (94). Additionally, BMP1 isoforms facilitate the maturation of BMP antagonist chordin and pro-lysyl oxidases, crucial mediators of collagen crosslinking (95, 96). This process is indispensable for the proper assembly of insoluble collagen within the ECM and subsequent scar formation. Previously, employing liquid chromatography-mass spectrometry we have identified

the presence of BMP6, GDF15, and the BMP1.3 isoform of the Bmp1 gene in plasma samples collected from both healthy individuals and patients afflicted with chronic kidney disease (12). Subsequently, we isolated the endogenous BMP1-3 protein and provided evidence of its circulation as an active enzyme (12). Due to its role in ECM assembly, BMP1.3 emerged as a potential target for the prevention of fibrosis. To test the role of BMP1.3 in fibrosis we have conducted studies employing a rat model of chronic kidney disease (12). Administration of a BMP1.3 neutralizing antibody reduced renal fibrosis (CKD), preserved organ function and subsequently increased survival of rats with CKD. On the other hand, the administration of recombinant BMP1.3 increased renal fibrosis and reduced survival. Antifibrotic effects of BMP1.3 neutralizing antibody have been subsequently evaluated in rats with carbon tetrachloride (CCl<sub>4</sub>)-induced liver fibrosis (10). Results of this study revealed that administration of monoclonal BMP1.3 antibodies significantly decreased fibrosis and therefore might be a therapeutic solution for decreasing the progress of liver cirrhosis (10). Recently, we investigated BMP1.3 antibodies as the first antibody-based treatment for ischemic heart disease (9). Acute myocardial infarction is an extremely common disease with a high mortality rate for which there is currently no sufficiently effective therapeutic solution (97, 98). Our studies on rodents revealed that BMP1.3 inhibition reduces collagen deposition, decreases scar formation, and supports cardiomyocyte survival after myocardial infarction (9). Mechanistically, we have demonstrated that BMP1.3 monoclonal antibody inhibits the Transforming Growth Factor  $\beta$  (TGF $\beta$ ) pathway. Furthermore, CERRM has explored anti-BMP1.3 antibodies as a therapy for merosin-deficient congenital muscular dystrophy type 1A (MDC1A) mice which is the second most prevalent congenital muscular dystrophy (CMD). Dystrophy is caused by a mutation in the lama2 gene encoding the laminin- $\alpha$ 2 protein that contributes to merosin, an important protein present in the skeletal muscle basement membrane (99-102). To evaluate the potential therapeutic effects of antifibrotic therapy we evaluated anti-BMP1.3 antibodies in a mouse model of congenital muscular dystrophy with mutation in the laminin gene (103). The application of BMP1.3 antibodies had a stunning effect on mice with CMD and significantly increased their survival, mobility, body mass, and decreased fibrotic muscle area (Figure 2) (103). Promising results on the mouse CMD model indicate that BMP1.3 inhibition might be an effective therapy for patients suffering from this rare but extremely severe disease. In conclusion, antifibrotic therapy based on anti-BMP1.3 antibodies represents a potential therapeutic solution for a wide range of diseases in which fibrosis is a key pathogenetic mechanism, including but not limited to chronic kidney disease, liver cirrhosis, acute myocardial infarction, and congenital muscular dystrophy.

#### 4. CONCLUSIONS

The development of regenerative medicine and novel therapeutic devices provides hope to improve the life quality of millions of patients worldwide. The establishment of the Croatian Center for Reproductive and Regenerative Medicine (CERRM) provided necessary resources for conducting the state-of-the-art research in the field of regenerative medicine and translation of developed therapeutic solutions to clinical practice. CERRM members have conducted extensive preclinical studies to develop Osteogrow-C, a novel osteoinductive device for bone regeneration as well as novel anti-BMP1.3 based antifibrotic therapy (3-7, 9, 10, 12, 77, 78, 81). Moreover, several Osteogrow formulations (Osteogrow, Osteogrow-A) have also been evaluated in Phase I/II clinical trials (56, 57). All phases of Osteogrow development have been conducted or coordinated by the University of Zagreb School of Medicine which is a unique example worldwide that an academic institution has carried out the development of a drug from discovery and preclinical research to advanced stages of clinical trials. The aforementioned studies have resulted in more than 30 papers published in prestigious scientific papers including Nature Communications, Nature Microbiology, and Nature Scientific Reports (Figure 3).

Furthermore, CERRM scientists have presented their work at numerous international conferences (Figure 4) including the Tissue Engineering and Regenerative Medicine Society (TERMIS) Conference and the European Calcified Tissue Society (ECTS) Congress. Moreover, CERRM has significantly contributed to the internationalization of research through numerous established co-operations with leading scientists in the field and by the organization of the 13th International Conference on Bone Morphogenetic Proteins in Dubrovnik (October 2022) (18). The 13th International BMP Congress gathered world-leading experts in the BMP field who delivered keynote lectures and young scientists who had the opportunity to present and discuss their work. Finally, an integral part of CERRM's mission and vision is the education of young scientists in the field of regenerative medicine. Therefore, CERRM has provided optimal conditions for the development and education of young scientists and as part of the project, eight students have completed or will soon complete their doctoral dissertations (104, 105).

To summarize, the establishment and implementation of CERRM's activities have had a transformative impact on the development of regenerative medicine in Croatia, resulting in significant scientific discoveries, the development of new therapeutic devices, the education of young scientists, and the establishing of new collaboration with different institutions and experts in the field. The established network of CERRM scientists and institutions will continue their endeavor to develop and bring to clinical practice cutting-edge therapies for bone regeneration and treatment of fibrosis.

### DECLARATION OF COMPETING INTERESTS

S.V. is a coordinator of the EU HORIZON 2020 grant OSTEOproSPINE funding clinical studies of the new drug for bone repair (patent WO2019076484A1) and a founder of Genera Research.

### ACKNOWLEDGEMENTS

This program was funded by the FP7 Health Program (FP7/2007-2013) under grant agreement HEALTH-F4-2011-279239 (Osteogrow), H2020 Health GA 779340 (OSTEOproSPINE), and European Regional Development Fund - Scientific Center of Excellence for Reproductive and Regenerative Medicine (project "Reproductive and regenerative medicine - exploration of new platforms and potentials," GA KK.01.1.1.01.0008 funded by the EU through the ERDF).

### REFERENCES

- Vukicevic S, Oppermann H, Verbanac D, Jankolija M, Popcek I, Curak J, et al. The clinical use of bone morphogenetic proteins revisited: a novel biocompatible carrier device OSTEOGROW for bone healing. *Int Orthop* 2014;38:635-647.
- Vukicevic S, Grgurevic L, Erjavec I, Pecin M, Bordukalo-Niksic T, Stokovic N, et al. Autologous blood coagulum is a physiological carrier for BMP6 to induce new bone formation and promote posterolateral lumbar spine fusion in rabbits. *J Tissue Eng Regen Med* 2020;14:147-159.
- Stokovic N, Ivanjko N, Erjavec I, Milosevic M, Oppermann H, Shimp L, et al. Autologous bone graft substitute containing rhBMP6 within autologous blood coagulum and synthetic ceramics of different particle size determines the quantity and structural pattern of bone formed in a rat subcutaneous assay. *Bone* 2020;141:115654.
- Stokovic N, Ivanjko N, Pecin M, Erjavec I, Karlovic S, Smajlovic A, et al. Evaluation of synthetic ceramics as compression resistant matrix to promote osteogenesis of autologous blood coagulum containing recombinant human bone morphogenetic protein 6 in rabbit posterolateral lumbar fusion model. *Bone* 2020;140:115544.
- Stokovic N, Ivanjko N, Pecin M, Erjavec I, Smajlovic A, Milesevic M, et al. Long-term posterolateral spinal fusion in rabbits induced by rhBMP6 applied in autologous blood coagulum with synthetic ceramics. *Sci Rep* 2022;12:11649.
- Stokovic N, Ivanjko N, Rumenovic V, Breski A, Sampath KT, Peric M, et al. Comparison of synthetic ceramic products formulated with autologous blood coagulum containing rhBMP6 for induction of bone formation. *Int Orthop* 2022;46:2693-2704.
- Ivanjko N, Stokovic N, Pecin M, Vnuk D, Smajlovic A, Ivkic N, et al. Calcium phosphate ceramics combined with rhBMP6 within autologous blood coagulum promote posterolateral lumbar fusion in sheep. *Sci Rep* 2023;13:22079.
- Pecin M, Stokovic N, Ivanjko N, Smajlovic A, Kreszinger M, Capak H, et al. A novel autologous bone graft substitute containing rhBMP6 in autologous blood coagulum with synthetic ceramics for reconstruction of a large humerus segmental gunshot defect in a dog: The first veterinary patient to receive a novel osteoinductive therapy. *Bone Rep* 2021;14:100759.
- Vukicevic S, Colliva A, Kufner V, Martinelli V, Moimas S, Vodret S, et al. Bone morphogenetic protein 1.3 inhibition decreases scar formation and supports cardiomyocyte survival after myocardial infarction. *Nat Commun* 2022;13:81.
- Grgurevic L, Erjavec I, Grgurevic I, Dumic-Cule I, Brkljacic J, Verbanac D, et al. Systemic inhibition of BMP1-3 decreases progression of CCl(4)-induced liver fibrosis in rats. *Growth Factors* 2017;35:201-215.
- Bordukalo Niksic T, Kufner V. BMP1.3 protein as potential target in treatment of fibrosis. *Rad CASA - Medical Sciences* 2021;548:56-69.
- Grgurevic L, Macek B, Healy DR, Brault AL, Erjavec I, Cipic A, et al. Circulating bone morphogenetic protein 1-3 isoform increases renal fibrosis. *J Am Soc Nephrol* 2011;22:681-692.
- El Bialy I, Jiskoot W, Reza Nejadnik M. Formulation, Delivery and Stability of Bone Morphogenetic Proteins for Effective Bone Regeneration. *Pharm Res* 2017;34:1152-1170.
- Gautschi OP, Frey SP, Zellweger R. Bone morphogenetic proteins in clinical applications. *ANZ J Surg* 2007;77:626-631.
- Katagiri T, Watabe T. Bone Morphogenetic Proteins. *Cold Spring Harb Perspect Biol* 2016;8.
- Granjeiro JM, Oliveira RC, Bustos-Valenzuela JC, Sogayar MC, Taga R. Bone morphogenetic proteins: from structure to clinical use. *Braz J Med Biol Res* 2005;38:1463-1473.

17. Vukicevic S, Stavljenic A, Pecina M. Discovery and clinical applications of bone morphogenetic proteins. *Eur J Clin Chem Clin Biochem* 1995;33:661-671.
18. Stokovic N, Ivanjko N, Matic Jelic I, Milesevic M, Rumenovic V, Blazevic V, et al. State-of-the-art of the Bone Morphogenetic Protein research field: 13th International BMP Conference, Dubrovnik 2022. *Rad CASA – Medical Sciences* 2022;553: 84-99.
19. Vukicevic S, Peric M, Oppermann H, Stokovic N, Ivanjko N, Erjavec I, et al. Bone morphogenetic proteins: From discovery to development of a novel autologous bone graft substitute consisting of recombinant human BMP6 delivered in autologous blood coagulum carrier. *Rad CASA - Medical Sciences* 2020;544:26-41.
20. Sandhu HS. Bone morphogenetic proteins and spinal surgery. *Spine (Phila Pa 1976)* 2003;28:S64-73.
21. Dumic-Cule I, Peric M, Kucko L, Grgurevic L, Pecina M, Vukicevic S. Bone morphogenetic proteins in fracture repair. *Int Orthop* 2018;42:2619-2626.
22. Even J, Eskander M, Kang J. Bone morphogenetic protein in spine surgery: current and future uses. *J Am Acad Orthop Surg* 2012;20:547-552.
23. Krishnakumar GS, Roffi A, Reale D, Kon E, Filardo G. Clinical application of bone morphogenetic proteins for bone healing: a systematic review. *Int Orthop* 2017;41:1073-1083.
24. Baldwin P, Li DJ, Auston DA, Mir HS, Yoon RS, Koval KJ. Autograft, Allograft, and Bone Graft Substitutes: Clinical Evidence and Indications for Use in the Setting of Orthopaedic Trauma Surgery. *J Orthop Trauma* 2019;33:203-213.
25. Bibbo C, Nelson J, Ehrlich D, Rougeux B. Bone morphogenetic proteins: indications and uses. *Clin Podiatr Med Surg* 2015;32:35-43.
26. Bragdon B, Moseychuk O, Saldanha S, King D, Julian J, Nohe A. Bone morphogenetic proteins: a critical review. *Cell Signal* 2011;23:609-620.
27. Carreira AC, Zambuzzi WF, Rossi MC, Astorino Filho R, Sogayar MC, Granjeiro JM. Bone Morphogenetic Proteins: Promising Molecules for Bone Healing, Bioengineering, and Regenerative Medicine. *Vitam Horm* 2015;99:293-322.
28. De Biase P, Capanna R. Clinical applications of BMPs. *Injury* 2005;36 Suppl 3:S43-46.
29. Minamide A, Kawakami M, Hashizume H, Sakata R, Tamaki T. Evaluation of carriers of bone morphogenetic protein for spinal fusion. *Spine* 2001;26:933-939.
30. Stokovic N, Ivanjko N, Maticic D, Luyten FP, Vukicevic S. Bone Morphogenetic Proteins, Carriers, and Animal Models in the Development of Novel Bone Regenerative Therapies. *Materials* 2021;14:3513.
31. Agrawal V, Sinha M. A review on carrier systems for bone morphogenetic protein-2. *J Biomed Mater Res B Appl Biomater* 2017;105:904-925.
32. Hutmacher DW, Schantz JT, Lam CXF, Tan KC, Lim TC. State of the art and future directions of scaffold-based bone engineering from a biomaterials perspective. *J Tissue Eng Regen Med* 2007;1:245-260.
33. Haidar ZS, Hamdy RC, Tabrizian M. Delivery of recombinant bone morphogenetic proteins for bone regeneration and repair. Part B: Delivery systems for BMPs in orthopaedic and craniofacial tissue engineering. *Biotechnol Lett* 2009;31:1825-1835.
34. Bessa PC, Casal M, Reis RL. Bone morphogenetic proteins in tissue engineering; the road from laboratory to clinic, part II (BMP delivery). *J Tissue Eng Regen Med* 2008;2:81-96.
35. Lee SH, Shin H. Matrices and scaffolds for delivery of bioactive molecules in bone and cartilage tissue engineering. *Adv Drug Deliv Rev* 2007;59:339-359.
36. Yang S, Leong KF, Du Z, Chua CK. The design of scaffolds for use in tissue engineering. Part I. Traditional factors. *Tissue Eng* 2001;7:679-689.
37. De Witte TM, Fratila-Apachitei LE, Zadpoor AA, Peppas NA. Bone tissue engineering via growth factor delivery: from scaffolds to complex matrices. *Regen Biomater* 2018;5:197-211.
38. Ginebra MP, Espanol M, Maazouz Y, Bergez V, Pastorino D. Bioceramics and bone healing. *EFORT Open Rev* 2018;3:173-183.
39. Vukicevic S, Stokovic N, Pecina M. Is ceramics an appropriate bone morphogenetic protein delivery system for clinical use? *Int Orthop* 2019;43:1275-1276.
40. Geiger M, Li RH, Friess W. Collagen sponges for bone regeneration with rhBMP-2. *Adv Drug Del Rev* 2003;55:1613-1629.
41. Boden SD, Martin GJ, Horton WC, Truss TL, Sandhu HS. Laparoscopic anterior spinal arthrodesis with rhBMP-2 in a titanium interbody threaded cage. *J Spinal Disord* 1998;11:95-101.
42. Boden SD, Martin GJ, Jr., Morone MA, Ugbo JL, Moskovitz PA. Posterolateral lumbar intertransverse process spine arthrodesis with recombinant human bone morphogenetic protein 2/hydroxyapatite-tricalcium phosphate after laminectomy in the nonhuman primate. *Spine (Phila Pa 1976)* 1999;24:1179-1185.
43. Boden SD, Zdeblick TA, Sandhu HS, Heim SE. The use of rhBMP-2 in interbody fusion cages. Definitive evidence of osteoinduction in humans: a preliminary report. *Spine (Phila Pa 1976)* 2000;25:376-381.
44. Boyne PJ, Lilly LC, Marx RE, Moy PK, Nevins M, Spagnoli DB, et al. De novo bone induction by recombinant human bone morphogenetic protein-2 (rhBMP-2) in maxillary sinus floor augmentation. *J Oral Maxillofac Surg* 2005;63:1693-1707.
45. Sandhu HS, Toth JM, Diwan AD, Seim HB, 3rd, Kanim LE, Kabo JM, et al. Histologic evaluation of the efficacy of

- rhBMP-2 compared with autograft bone in sheep spinal anterior interbody fusion. *Spine (Phila Pa 1976)* 2002;27:567-575.
46. Epstein NE. Complications due to the use of BMP/INFUSE in spine surgery: The evidence continues to mount. *Surg Neurol Int* 2013;4:S343-352.
  47. McKay WF, Peckham SM, Badura JM. A comprehensive clinical review of recombinant human bone morphogenetic protein-2 (INFUSE Bone Graft). *Int Orthop* 2007;31:729-734.
  48. James AW, LaChaud G, Shen J, Asatrian G, Nguyen V, Zhang X, et al. A Review of the Clinical Side Effects of Bone Morphogenetic Protein-2. *Tissue Eng Part B Rev* 2016;22:284-297.
  49. Carragee EJ, Hurwitz EL, Weiner BK. A critical review of recombinant human bone morphogenetic protein-2 trials in spinal surgery: emerging safety concerns and lessons learned. *Spine J* 2011;11:471-491.
  50. Song K, Krause C, Shi S, Patterson M, Suto R, Grgurevic L, et al. Identification of a key residue mediating bone morphogenetic protein (BMP)-6 resistance to noggin inhibition allows for engineered BMPs with superior agonist activity. *J Biol Chem* 2010;285:12169-12180.
  51. Mizrahi O, Sheyn D, Tawackoli W, Kallai I, Oh A, Su S, et al. BMP-6 is more efficient in bone formation than BMP-2 when overexpressed in mesenchymal stem cells. *Gene Ther* 2013;20:370-377.
  52. Vukicevic S, Grgurevic L. BMP-6 and mesenchymal stem cell differentiation. *Cytokine Growth Factor Rev* 2009;20:441-448.
  53. Ivanjko N, Stokovic N, Milesevic M, Rumenovic V, Windhager R, Sampath KT, et al. rhBMP6 in autologous blood coagulum is a preferred osteoinductive device to rhBMP2 on bovine collagen sponge in the rat ectopic bone formation assay. *Biomed Pharmacother* 2023;169:115844.
  54. Fan Q, Bai J, Shan H, Fei Z, Chen H, Xu J, et al. Implantable blood clot loaded with BMP-2 for regulation of osteoimmunology and enhancement of bone repair. *Bioact Mater* 2021;6:4014-4026.
  55. Fan Q, Ma Q, Bai J, Xu J, Fei Z, Dong Z, et al. An implantable blood clot-based immune niche for enhanced cancer vaccination. *Sci Adv* 2020;6.
  56. Durdevic D, Vlahovic T, Pehar S, Miklic D, Oppermann H, Bordukalo-Niksic T, et al. A novel autologous bone graft substitute comprised of rhBMP6 blood coagulum as carrier tested in a randomized and controlled Phase I trial in patients with distal radial fractures. *Bone* 2020;140:115551.
  57. Chiari C, Grgurevic L, Bordukalo-Niksic T, Oppermann H, Valentinitsch A, Nemecek E, et al. Recombinant Human BMP6 Applied Within Autologous Blood Coagulum Accelerates Bone Healing: Randomized Controlled Trial in High Tibial Osteotomy Patients. *J Bone Miner Res* 2020;35:1893-1903.
  58. Grgurevic L, Erjavec I, Gupta M, Pecin M, Bordukalo-Niksic T, Stokovic N, et al. Autologous blood coagulum containing rhBMP6 induces new bone formation to promote anterior lumbar interbody fusion (ALIF) and posterolateral lumbar fusion (PLF) of spine in sheep. *Bone* 2020;138:115448.
  59. Tomford WW. Bone allografts: past, present and future. *Cell Tissue Bank* 2000;1:105-109.
  60. Mroz TE, Joyce MJ, Lieberman IH, Steinmetz MP, Benzel EC, Wang JC. The use of allograft bone in spine surgery: is it safe? *Spine J* 2009;9:303-308.
  61. Vaccaro AR, Chiba K, Heller JG, Patel T, Thalgot JS, Truumees E, et al. Bone grafting alternatives in spinal surgery. *Spine J* 2002;2:206-215.
  62. Alam MI, Asahina I, Ohmamiuda K, Takahashi K, Yokota S, Enomoto S. Evaluation of ceramics composed of different hydroxyapatite to tricalcium phosphate ratios as carriers for rhBMP-2. *Biomaterials* 2001;22:1643-1651.
  63. Best SM, Porter AE, Thian ES, Huang J. Bioceramics: Past, present and for the future. *J Eur Ceram Soc* 2008;28:1319-1327.
  64. Dorozhkin SV. Bioceramics of calcium orthophosphates. *Biomaterials* 2010;31:1465-1485.
  65. Lee JH, Ryu MY, Baek HR, Lee KM, Seo JH, Lee HK, et al. Effects of porous beta-tricalcium phosphate-based ceramics used as an E. coli-derived rhBMP-2 carrier for bone regeneration. *J Mater Sci Mater Med* 2013;24:2117-2127.
  66. LeGeros RZ, Lin S, Rohanizadeh R, Mijares D, LeGeros JP. Biphasic calcium phosphate bioceramics: preparation, properties and applications. *J Mater Sci Mater Med* 2003;14:201-209.
  67. Ghanaati S, Barbeck M, Orth C, Willershausen I, Thimm BW, Hoffmann C, et al. Influence of beta-tricalcium phosphate granule size and morphology on tissue reaction in vivo. *Acta Biomater* 2010;6:4476-4487.
  68. Glassman SD, Dimar JR, Carreon LY, Campbell MJ, Puno RM, Johnson JR. Initial fusion rates with recombinant human bone morphogenetic protein-2/compression resistant matrix and a hydroxyapatite and tricalcium phosphate/collagen carrier in posterolateral spinal fusion. *Spine (Phila Pa 1976)* 2005;30:1694-1698.
  69. Habraken W, Habibovic P, Epple M, Böhner M. Calcium phosphates in biomedical applications: materials for the future? *Mater Today*. 2016;19:69-87.
  70. Jung UW, Choi SY, Pang EK, Kim CS, Choi SH, Cho KS. The effect of varying the particle size of beta tricalcium phosphate carrier of recombinant human bone morphogenetic protein-4 on bone formation in rat calvarial defects. *J Periodontol* 2006;77:765-772.

71. Liu B, Lun DX. Current application of beta-tricalcium phosphate composites in orthopaedics. *Orthop Surg* 2012;4:139-144.
72. Parker RM, Malham GM. Comparison of a calcium phosphate bone substitute with recombinant human bone morphogenetic protein-2: a prospective study of fusion rates, clinical outcomes and complications with 24-month follow-up. *Eur Spine J* 2017;26:754-763.
73. Strobel LA, Rath SN, Maier AK, Beier JP, Arkudas A, Greil P, et al. Induction of bone formation in biphasic calcium phosphate scaffolds by bone morphogenetic protein-2 and primary osteoblasts. *J Tissue Eng Regen Med* 2014;8:176-185.
74. Tazaki J, Murata M, Akazawa T, Yamamoto M, Ito K, Arisue M, et al. BMP-2 release and dose-response studies in hydroxyapatite and beta-tricalcium phosphate. *Biomed Mater Eng* 2009;19:141-146.
75. Wu Y, Hou J, Yin M, Wang J, Liu C. Enhanced healing of rabbit segmental radius defects with surface-coated calcium phosphate cement/bone morphogenetic protein-2 scaffolds. *Mater Sci Eng C Mater Biol Appl* 2014;44:326-335.
76. El-Ghannam A. Bone reconstruction: from bioceramics to tissue engineering. *Expert Rev Med Devices* 2005;2:87-101.
77. Stokovic N, Ivanjko N, Erjavec I, Breski A, Peric M, Vukicevic S. Zoledronate Bound to Ceramics Increases Ectopic Bone Volume Induced by rhBMP6 Delivered in Autologous Blood Coagulum in Rats. *Biomedicines* 2021;9.
78. Stokovic N, Ivanjko N, Milesevic M, Matic Jelic I, Bakic K, Rumenovic V, et al. Synthetic ceramic macroporous blocks as a scaffold in ectopic bone formation induced by recombinant human bone morphogenetic protein 6 within autologous blood coagulum in rats. *Int Orthop* 2020;45:1097-1107.
79. Peric M, Dumic-Cule I, Grcevic D, Matijasic M, Verbanac D, Paul R, et al. The rational use of animal models in the evaluation of novel bone regenerative therapies. *Bone* 2015;70:73-86.
80. Sandhu HS, Khan SN. Animal models for preclinical assessment of bone morphogenetic proteins in the spine. *Spine (Phila Pa 1976)* 2002;27:S32-38.
81. Stokovic N, Ivanjko N, Milesevic M, Sampath K, Vukicevic S. A simple rodent subcutaneous assay for identification of new osteoinductive molecules: The key method for screening of novel bone regeneration implants. *Rad CASA - Medical Sciences* 2022;553 40-53.
82. Garcia P, Histing T, Holstein JH, Klein M, Laschke MW, Matthys R, et al. Rodent animal models of delayed bone healing and non-union formation: a comprehensive review. *Eur Cell Mater* 2013;26:1-14.
83. McGovern JA, Griffin M, Huttmacher DW. Animal models for bone tissue engineering and modelling disease. *Dis Model Mech* 2018;11.
84. Sommer NG, Hahn D, Okutan B, Marek R, Weinberg AM. Animal Models in Orthopedic Research: The Proper Animal Model to Answer Fundamental Questions on Bone Healing Depending on Pathology and Implant Material. In: Tvrdá E, Yeniseetti CS, editors. *Animal Models in Medicine and Biology*: IntechOpen; 2020.
85. Wynn TA, Ramalingam TR. Mechanisms of fibrosis: therapeutic translation for fibrotic disease. *Nat Med* 2012;18:1028-1040.
86. Ueha S, Shand FH, Matsushima K. Cellular and molecular mechanisms of chronic inflammation-associated organ fibrosis. *Front Immunol* 2012;3:71.
87. Antar SA, Ashour NA, Marawan ME, Al-Karmalawy AA. Fibrosis: Types, Effects, Markers, Mechanisms for Disease Progression, and Its Relation with Oxidative Stress, Immunity, and Inflammation. *Int J Mol Sci* 2023;24.
88. Kessler E, Takahara K, Biniaminov L, Brusel M, Greenspan DS. Bone morphogenetic protein-1: the type I procollagen C-proteinase. *Science* 1996;271:360-362.
89. Hulmes DJ, Mould AP, Kessler E. The CUB domains of procollagen C-proteinase enhancer control collagen assembly solely by their effect on procollagen C-proteinase/bone morphogenetic protein-1. *Matrix Biol* 1997;16:41-45.
90. Hopkins DR, Keles S, Greenspan DS. The bone morphogenetic protein 1/Tolloid-like metalloproteinases. *Matrix Biol* 2007;26:508-523.
91. Mohrlen F, Hutter H, Zwilling R. The astacin protein family in *Caenorhabditis elegans*. *Eur J Biochem* 2003;270:4909-4920.
92. Janitz M, Heiser V, Bottcher U, Landt O, Lauster R. Three alternatively spliced variants of the gene coding for the human bone morphogenetic protein-1. *J Mol Med (Berl)* 1998;76:141-146.
93. Park JO, Pan J, Mohrlen F, Schupp MO, Johnsen R, Baillie DL, et al. Characterization of the astacin family of metalloproteases in *C. elegans*. *BMC Dev Biol* 2010;10:14.
94. Anastasi C, Rousselle P, Talantikite M, Tessier A, Cluzel C, Bachmann A, et al. BMP-1 disrupts cell adhesion and enhances TGF-beta activation through cleavage of the matrix-cellular protein thrombospondin-1. *Sci Signal* 2020;13.
95. Maruhashi T, Kii I, Saito M, Kudo A. Interaction between periostin and BMP-1 promotes proteolytic activation of lysyl oxidase. *J Biol Chem* 2010;285:13294-13303.
96. Trackman PC. Diverse biological functions of extracellular collagen processing enzymes. *J Cell Biochem* 2005;96:927-937.
97. Sutton MG, Sharpe N. Left ventricular remodeling after myocardial infarction: pathophysiology and therapy. *Circulation* 2000;101:2981-2988.
98. Fan D, Takawale A, Lee J, Kassiri Z. Cardiac fibroblasts, fibrosis and extracellular matrix remodeling in heart disease. *Fibrogenesis Tissue Repair* 2012;5:15.



99. Butterfield RJ. Congenital Muscular Dystrophy and Congenital Myopathy. *Continuum (Minneapolis)* 2019;25:1640-1661.
100. Mann CJ, Perdiguero E, Kharraz Y, Aguilar S, Pessina P, Serrano AL, et al. Aberrant repair and fibrosis development in skeletal muscle. *Skeletal Muscle* 2011;1:21.
101. Nguyen Q, Lim KRQ, Yokota T. Current understanding and treatment of cardiac and skeletal muscle pathology in laminin-alpha2 chain-deficient congenital muscular dystrophy. *Appl Clin Genet* 2019;12:113-130.
102. Zambon AA, Muntoni F. Congenital muscular dystrophies: What is new? *Neuromuscul Disord* 2021;31:931-942.
103. Matic Jelic I, Vukicevic S. Regenerative and antifibrotic potential of anti-BMP1.3 in mouse model of congenital muscular dystrophy with mutation in the laminin gene 13th International BMP Conference; Dubrovnik, Croatia, 2022.
104. Stokovic N. Ectopic bone induction by osteoinductive device composed of recombinant human bone morphogenetic protein 6 (rhBMP6), autologous blood coagulum and biphasic bioceramics. Dissertation. Zagreb: University of Zagreb; 2022.
105. Ivanjko N. Comparison of the dynamics of ectopic bone formation using bone morphogenetic protein 2 on a collagen sponge carrier and bone morphogenetic protein 6 in an autologous blood coagulum. Dissertation. Zagreb: University of Zagreb; 2022.

