



High point seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in sheep with abortion history in the central part of Türkiye

Ceylan Ceylan¹, Onur Ceylan^{2*}, Shinuo Cao³, Hasan Hüseyin Hadimli⁴, Xuenan Xuan⁵ and Ferda Sevinc²

¹Siirt University, Faculty of Veterinary Medicine, Department of Parasitology, Siirt, Türkiye

²Selcuk University, Faculty of Veterinary Medicine, Department of Veterinary Parasitology, Konya, Türkiye

³State Key Laboratory of Veterinary Biotechnology, Harbin Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Harbin, People's Republic of China

⁴Selcuk University, Faculty of Veterinary Medicine, Department of Microbiology, Konya, Türkiye

⁵Obihiro University of Agriculture and Veterinary Medicine, National Research Center for Protozoan Diseases, Obihiro, Japan

CEYLAN, C., O. CEYLAN, S. CAO, H. H. HADİMLİ, X. XUAN, F. SEVİNC: High point seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in sheep with abortion history in the central part of Türkiye. *Vet. arhiv* 95, 411-421, 2025.

ABSTRACT

Small ruminant breeding plays a vital role in livestock farming in Türkiye but is adversely affected by many infectious pathogens, including *Toxoplasma (T.) gondii* and *Neospora (N.) caninum*. This study aimed to investigate the seroprevalence of ovine toxoplasmosis and neosporosis in sheep with varying abortion histories in the Central Anatolia Region of Türkiye. Two hundred and forty-three sheep blood samples were taken from different farms, 115 of these samples from sheep with abortion problems, and the others were from sheep herds with no abortion history. Recombinant TgSAG2 and NcSAG1-based indirect ELISA were used to determine specific antibodies against *T. gondii* and *N. caninum*. The study revealed a high *T. gondii* and *N. caninum* prevalence in the herd suffering from abortions. The *T. gondii* and *N. caninum* seroprevalences in sheep with an abortion history were 66.1% and 50.4%, respectively. However, it was 26.6% for *T. gondii* and 10.9% for *N. caninum* in randomly selected sheep without an abortion history. This study reports the highest *N. caninum* seroprevalence both overall (29.63%) and in sheep with an abortion history (50.4%) in Türkiye. Although neosporosis is the primary abortifacient protozoan infection in cattle, the high *N. caninum* seroprevalence detected in the sheep with an abortion history indicates that this disease should also be considered in ovine abortion cases. The present study revealed that ovine toxoplasmosis and neosporosis are prevalent, and may contribute to abortions in sheep from the sampling area in the Central Anatolia Region of Türkiye.

Key words: ELISA; neosporosis; ovine abortion; protozoal infection; toxoplasmosis; Türkiye

* Corresponding author:

Onur Ceylan Assoc. Prof., Selcuk University, Faculty of Veterinary Medicine, Department of Preclinical Sciences, Department of Veterinary Parasitology, 42130, Konya, Türkiye, phone:+905062550038, e-mail: onurceylan@selcuk.edu.tr

Introduction

Toxoplasma gondii and *Neospora caninum* are intracellular apicomplexan parasites with a global distribution that share structural, genetic, and immunological characteristics (DUBEY, 2003; DUBEY, 2009). Although these two protozoan parasites have similar life cycles and infect various animal species, toxoplasmosis primarily impacts small ruminants, while neosporosis mainly affects cattle (DUBEY, 2009; HEMPHILL and GOTTSTEIN, 2000). *T. gondii*'s definitive hosts are cats and other members of the Felidae family, whereas *N. caninum*'s definitive hosts are canines such as dogs, coyotes, wolves, and dingoes (DUBEY and SCHARES, 2011). *T. gondii* has a broad intermediate host spectrum, including humans, and can infect nearly all warm-blooded animals (DUBEY et al., 2020). However, it is unclear whether *N. caninum* causes human infections or not (DUBEY et al., 2007). In a study, the DNA of *N. caninum* was not identified in human tissues, but anti-*N. caninum* specific antibodies were observed in human serum samples. This situation may suggest that *N. caninum* can cause human infections (TRANAS et al., 1999).

Toxoplasmosis is a reproductive disorder in sheep, and the infection is generally characterized by abortion, neonatal death, embryonic death and resorption, stillbirth, fetal death, and mummification, or congenitally infected apparently healthy lambs (DUBEY, 2009). Ovine toxoplasmosis, which causes economic losses, is also a significant public health problem because the infection may be transmitted to humans through the consumption of raw or undercooked infected mutton (TENTER et al., 2000; DUBEY et al., 2020). On the other hand, neosporosis primarily affects cattle and is one of the leading causes of bovine abortions. The infection may cause neuromuscular disorders in the definitive hosts (DUBEY et al., 2007). Compared to toxoplasmosis, the number of epidemiological and clinical studies on ovine neosporosis is lower. However, some studies have reported that neosporosis in sheep is characterized by symptoms similar to ovine toxoplasmosis and increases economic losses by causing abortions in sheep (GONZALES-WARLETA et al., 2014).

Small ruminant breeding plays a vital role in livestock farming, and sheep constitute approximately 60% of farm animals in Türkiye. Sheep breeding, which significantly contributes to the Turkish economy, is adversely affected by many causes, especially parasitic diseases (CEYLAN et al., 2021). Ovine toxoplasmosis and neosporosis, which cause loss of offspring, are two of the most critical diseases. This study aimed to investigate the seroprevalence of ovine toxoplasmosis and neosporosis in sheep farms with/without an abortion history in the Central Anatolia Region of Türkiye.

Materials and methods

Sampling site and serum collection. The material of this study consisted of 243 serum samples collected from sheep (Anatolian merinos) with abortion problems from a farm in Ankara, and one herd from each of the Konya, Karaman, and Aksaray provinces in the Central Anatolia Region of Türkiye, without an abortion history. One hundred and fifteen blood samples were collected from a farm with abortion problems in the Ankara province, while 128 sheep belonging to three herds without an abortion history in Konya (n=50), Karaman (n=50), and Aksaray (n=28) provinces were sampled. The sheep were individually bled through the jugular vein, and a 5 mL blood sample was collected into gel vials using sterile needles for each animal. All collected blood samples were held in a cooler box and then centrifuged (2000 rpm for 20 minutes) to yield serum at the laboratory of Selcuk University, Department of Veterinary Parasitology. After separation, serum samples were frozen and kept at -20°C until used in an indirect ELISA test. A detailed representation of the places where serum samples were collected in the study is given in Fig. 1.

Anamnesis. A long-term anamnesis of abortion in pregnant sheep on the farm in Ankara province was obtained, along with information indicating the presence of numerous carnivorous animals, especially cats, on the farm and the fact that these animals had not been treated with antiparasitic drugs. Samples from the affected herd with an abortion history were tested for a range of known abortifacient pathogens, including *Brucella melitensis*,

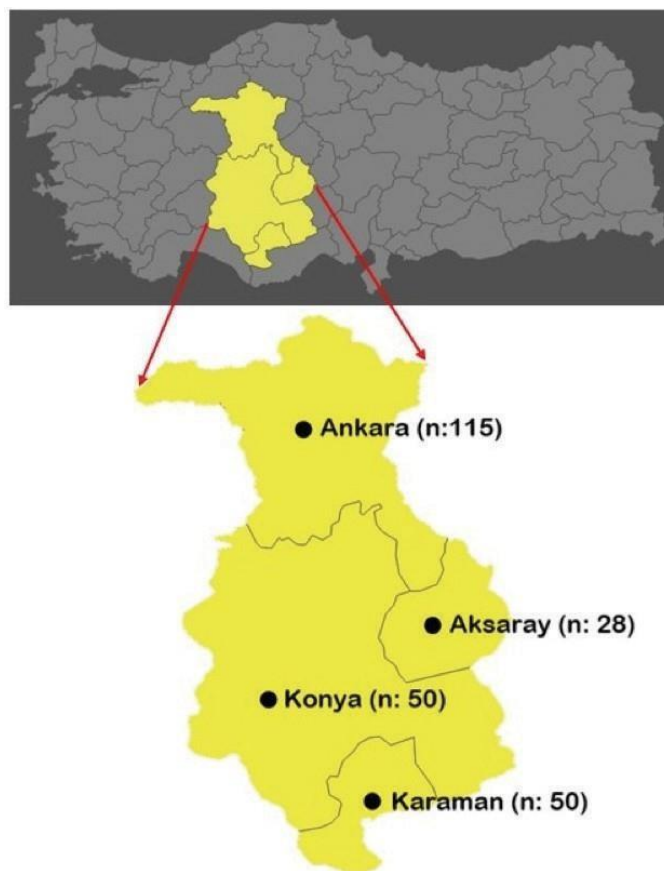


Fig. 1. Türkiye map, sampling area, and the number of samples collected from these areas

Chlamydia abortus, *Salmonella abortus ovis*, *Coxiella burnetii*, *Campylobacter fetus subspecies fetus*, and Border disease virus. These analyses were performed at the Department of Microbiology in the Faculty of Veterinary Medicine, as well as at a specialized private laboratory in Konya. Despite extensive testing, no definitive causative pathogen was identified, leading to suspicion of ovine toxoplasmosis and neosporosis as possible underlying factors. Additional information from the anamnesis revealed that the flock consisted of approximately 1000 ewes with an abortion rate of nearly 9%. For the study, representatives of the herd with this abortion history were enrolled for further investigation.

Indirect enzyme-linked immunosorbent assay. To detect the presence of specific antibodies in the serum samples, an indirect ELISA test using recombinant *T. gondii* SAG2 (*TgSAG2*) and *N. caninum* SAG1 (*NcSAG1*) proteins as antigens was used (ZHOU et al., 2017; LI et al., 2021; CEYLAN et

al., 2024). The study also used Glutathione S-transferase (GST) as the control antigen. In the indirect ELISA test, *TgSAG2*, *NcSAG1*, and GST proteins were reconstituted with carbonate-bicarbonate buffer (pH 9.6, 0.05 M) at 2 µg/ml concentration. The ELISA plate wells were coated with 100 µl of antigen and incubated at 4°C overnight. After pouring the coating solution and washing with PBST, the plates were blocked for 1 hour at 37°C with 3% skimmed milk solution containing 1X PBS. The microplate wells were rewashed using PBST, and the 1:100 diluted serum samples (with 3% skim milk solution) were added and incubated for 1 hour at 37°C. The washing processes were performed with PBST six times after this step. Binding antibodies were visualized with horseradish peroxidase-conjugated anti-sheep IgG secondary antibody (Bethyl, Montgomery, AL, USA) (1:4000) and ABTS [2,2'-azinobis (3-etilbenziazolinsülfonik asit)] substrate (Sigma, ABD, Louis, MO, ABD).

The absorbance was observed at room temperature, and 100 µl of stop solution (2 M Sulfuric acid) was added to inactivate the horseradish peroxidase enzyme. Optical density (OD) was measured with an ELISA microplate reader (Rayto Microplate Reader, Model: RT-2100C) at 415 nm.

Evaluation of the ELISA findings. The OD₄₁₅ value of the GST protein was subtracted from the OD₄₁₅ value of rTgSAG2 and rNcSAG2 for each sample to evaluate the ELISA results. The threshold for positive samples (cut-off value) was calculated by adding twice the standard deviation to

the mean OD₄₁₅ value of the negative sheep sera. If the OD₄₁₅ value of the sample was higher than the

cut-off value, the sample was considered positive. In this study, *Toxoplasma gondii* and *N. caninum* positive and negative sheep serum samples were obtained from our previous study (ZHOU et al., 2017). Recombinant TgSA2 and rNcSAG1 proteins were obtained from the Obihiro University of Agriculture and Veterinary Medicine, National Research Center for Protozoan Diseases, Obihiro, Japan.

Statistical analysis. The statistical program SPSS version 25 (IBM Corp. Released 2017. IBM SPSS Statistics for Windows, Version 25.0. Armonk, NY: IBM Corp.) was used to analyze all

data. *P* values were calculated to determine the potential statistical significance among different animal statuses. A *P*-value below 0.05 was regarded as statistically significant. The prevalence values were calculated by dividing the number of positive cases by the total number of individuals in a population over a specified period, and expressed as a percentage.

Results

In the study, among the serum samples col-

lected from sheep with an abortion history, anti-*T. gondii* IgG antibodies alone (*T. gondii* mono-infection) and anti-*N. caninum* IgG antibodies alone

(*N. caninum* mono-infection) were determined as 24.35% (28/115) and 8.7% (10/115), respectively. Specific IgG antibodies that formed against both protozoa (co-infection) were detected at 41.74% (48/115). The overall *T. gondii* and *N. caninum* seroprevalences in sheep with an abortion history were 66.1% and 50.4%, respectively. Anti-*T. gondii* IgG antibodies alone, anti-*N. caninum* IgG antibodies alone and co-infection with both species were respectively found in 25% (32/128), 9.38% (12/128), and 1.56% (2/128) in randomly selected sheep without an abortion history. The overall *T.*

Table 1. The seroprevalence of *T. gondii*, *N. caninum*, co-infection rates, and the number of sheep by provinces

Animal status	Locality (districts)	n	<i>T. gondii</i> infection	<i>N. caninum</i> infection	Co-infection	<i>T. gondii</i> overall infection	<i>N. caninum</i> overall infection
Sheep from the herd with abortion history	Ankara	115	28 (24.35%)	10 (8.7%)	48 (41.74%)	76 (66.09%)	58 (50.43%)
	Konya		24	5	-	24	5
Sheep from the herd without abortion history	(Sızma, Güvenc, Emirgazi)	50	5 (10%)	5 (10%)	1 (2%)	6 (12%)	6 (12%)
	Karaman	50	3 (10.71%)	2 (7.14%)	1 (3.57%)	4 (14.29%)	3 (10.71%)
	Aksaray (Sultanhanı)	28	3 (10.71%)	2 (7.14%)	1 (3.57%)	4 (14.29%)	3 (10.71%)
Total		243	60	22	50	110	72

Table 2. The results of statistical analysis

		Condition		Total	P
		Sheep with abortions	Sheep without abortions		
<i>Tg</i>	Negative	39	94	133	0,001
	Positive	76	34	110	
Total		115	128	243	
<i>Nc</i>	Negative	57	114	171	0,001
	Positive	58	14	72	
Total		115	128	243	

**Tg*: *Toxoplasma gondii*, *Nc*: *Neospora caninum*

gondii and *N. caninum* seroprevalences in randomly selected sheep without an abortion history were 26.56% (34/128) and 10.94% (14/128), respectively. Table 1 provides further details. The *T. gondii* and *N. caninum*-specific antibody prevalences in animals with a history of abortion were statistically higher than those in sheep without an abortion history ($P < 0.001$). More detailed results are shown in Table 2.

Discussion

Türkiye is a suitable country for small ruminant breeding due to its geographical structure, vegetation, and socio-economic and cultural characteristics (CEYLAN et al., 2021). The Central Anatolia Region is home to a significant proportion of Türkiye's small ruminant population, and makes a significant contribution to the country's meat and milk production (GUNLU and MAT, 2021). One of the indispensable elements of animal husbandry is to obtain healthy offspring regularly every year and to ensure the continuation of the herd (UTUK and ESKI, 2022). Abortion is a common clinical problem that significantly affects productivity in sheep herds. Causes of abortion include a variety of factors, such as stress, nutritional disorders, poisoning, hormonal imbalances, genetic factors, and infectious agents, among others (NOAKES et al., 2021). In healthy sheep herds, the rate of ewe abortion is generally less than 2%. When the abortion rate is between 2% and 5%, it may indicate the endemic presence of a disease. An abortion rate above

5% or a cluster of abortion cases within a short period of time or at a specific location suggests the presence of a pathogenic invasion and warrants an aggressive diagnostic investigation (MENZIES, 2011). In this study, after eliminating several primary bacterial and viral abortifacient agents (*Brucella melitensis*, *Chlamydia abortus*, *Salmonella abortus ovis*, *Coxiella burnetii*, *Campylobacter fetus subspecies fetus*, and Border disease virus), *T. gondii* and *N. caninum*, which are the most notable abortifacient protozoan agents of livestock animals, were investigated in a sheep herd with almost 9% abortion cases and herds comprising apparently healthy sheep. The present study on rTgSAG2 and rNcSAG1-based indirect ELISA revealed a high *T. gondii* and *N. caninum* seropositivity.

Toxoplasma gondii and *N. caninum* are well-recognized infectious pathogens leading to abortions and neonatal mortalities in small ruminants worldwide (BUXTON et al., 2007; DUBEY, 2009). Seroepidemiological studies have revealed the prevalence of ovine toxoplasmosis ranging between 2.8 and 98.9% in Türkiye (KARATEPE et al., 2001; MOR and ARSLAN, 2007; CICEK et al., 2011; ZHOU et al., 2017; CAKMAK and KARATEPE, 2017; ESKI et al., 2018; ASLAN CELIK et al., 2020; CEYLAN et al., 2024). In a survey conducted by CICEK et al. (2011), *T. gondii* seroprevalence was found to be the highest in Türkiye (98.9%) by the Sabin Feldman dye test. *T. gondii* overall seroprevalences were found as 66.1% in sheep with abortions and 26.6% in sheep without

an abortion history in this study. It was observed that *T. gondii* seroprevalence values in sheep from Konya, Karaman, Aksaray, and Ankara provinces were within the lower and upper limits obtained in studies conducted in Türkiye ([MOR and ARSLAN, 2007](#); [CICEK et al., 2011](#); [CAKMAK and KARATEPE, 2017](#); [ESKI et al., 2018](#)) but higher than some studies which use the same diagnostic techniques ([CAKMAK and KARATEPE, 2017](#); [ESKI et al., 2018](#); [ASLAN CELIK et al., 2020](#)). The seroprevalence of toxoplasmosis in aborted sheep in Türkiye was determined as 35.18% in Kayseri ([INCI et al., 1999](#)), 46.84% in Elazig ([AKTAS et al., 2000](#)), and 53.34% in Hatay ([KAMBURGIL et al., 2001](#)). In a study conducted in Kars, serum samples were taken from a total of 78 sheep, which were determined to have aborted during the research, and 97.4% of them were determined as seropositive by ELISA ([MOR and ARSLAN, 2007](#)). *T. gondii* seroprevalence was detected at 66.09% (76/115) in sheep who suffered abortions in the present study. Some researchers reported that the diagnostic techniques, the presence of cats in the environment, climate, age, and gender may have influenced the observed seroprevalence of ovine toxoplasmosis ([SAWADOGO et al., 2005](#); [CLEMENTINO et al., 2007](#)). Moreover, [VESCO et al. \(2007\)](#) reported that *Toxoplasma*-specific IgG antibodies transmitted in colostrum and milk could affect the seroprevalence in sheep under one year old. In light of these factors, the high seroprevalence detected in the aborted sheep may be attributed to the infected cat population around the farm, which likely plays a role in the shedding of *T. gondii* oocysts and the lack of anti-*T. gondii* treatment ([DUBEY and PROWELL, 2013](#)) in these cats. High ovine toxoplasmosis seroprevalence and abortion cases are expected in the herd, constantly in contact with the contaminated environment. Studies regarding neosporosis are also mostly on cattle, and a limited number of studies have revealed that the seroprevalence of ovine neosporosis is between 0 and 12.4% in Türkiye ([GOKCE et al., 2015](#); [ZHOU et al., 2017](#); [CAKMAK and KARATEPE, 2017](#); [ESKI et al., 2018](#); [KARATEPE and KARATEPE, 2020](#)). While *N. caninum* infection has been linked to abortion in sheep in recent years

([HECKER et al., 2019](#)), a high seroprevalence and incidence have also been reported in herds and individual ewes without a history of abortion or infertility ([ROSSI et al., 2011](#); [FILHO et al., 2017](#); [TIROSH-LEVY et al., 2022](#)).

The overall seroprevalence of *N. caninum* was detected in 29.63% of sheep sampled in the study, but it reached 50.4% in sheep suffering abortions. This study reports the highest ovine *N. caninum* seroprevalence in Türkiye; however, this value is consistent with some seroprevalence values reported from other countries ([PANADERO et al., 2010](#); [GUIMARÃES et al., 2015](#); [TAFNER FERREIRA et al., 2016](#); [DUBEY et al., 2017](#)). Studies on the seroprevalence of *N. caninum* in aborted sheep are limited worldwide ([HELMICK et al., 2002](#); [EZATPOUR et al., 2015](#); [MEIXNER et al., 2020](#)). The sampling region, the type of serological diagnostic technique, animal breed, the existence of dogs around the sampling area, and the number of animals studied can lead to differences in neosporosis seroprevalence ([FIGLIUOLO et al., 2004](#)). The prevalence of *N. caninum* in the herd with an abortion history is believed to result from the constant contamination of the environment by carnivores (especially shepherd dogs) and the exposure of the sheep to sporulated oocysts for a long time due to their age. Furthermore, the horizontal transmission of the infection from individual to individual may have contributed to this high seroprevalence. The view that the risk of infection increases with age, and vertical transmission of the disease with the advancement of age in animals also supports this situation ([NAYERI et al., 2022](#)).

The sheep owners in Ankara reported the presence of cats and dogs around the farm. This factor, along with issues such as inadequate hygienic practices in the facility management and improper disposal of carcasses and fetal-placental waste, may contribute to increased transmission of *T. gondii* and *N. caninum*. Some studies have shown that contact with cats and dogs increases the risk of toxoplasmosis and neosporosis in sheep ([ANDRADE et al., 2012](#); [MENDONCA et al., 2013](#); [GOKCE et al., 2015](#)). In this case, taking precautions to prevent the access of carnivores to carcasses and fetal/placental waste after lambing/abortion, and to avoid

contamination of the sheep's water and food by cat feces, can reduce the risk of *T. gondii* transmission to animals and decrease reproductive dysfunction in the herd (FILHO et al., 2017; HECKER et al., 2019; TIROSH-LEVY et al., 2022). In warmer climates, environmental factors, including temperature and humidity, can affect the survival and distribution of *T. gondii* and *N. caninum* oocysts in soil and water, potentially increasing exposure risk for animals (YAN et al., 2016). Such conditions may contribute to higher seroprevalence rates of these pathogens in regions with warmer climates, such as the Central Anatolian region of Türkiye. In a previous study conducted by ZHOU et al. (2017) in Konya and Karaman provinces, *T. gondii* seroprevalence was 24% and 9.2%, and *N. caninum* was 2.7% and 0.8%, and the mixed infection rate was 0.7% and 0% respectively. In the present study, the seroprevalence rates of both apicomplexan protozoa and mixed infection were higher than in the previously mentioned Konya and Karaman provinces, and *T. gondii* and *N. caninum* seroprevalences in animals with a history of abortion were statistically different from sheep without an abortion history ($P < 0.001$). Our results indicate a notably higher seroprevalence of *T. gondii* and *N. caninum* in sheep from Ankara, particularly in animals with a history of abortion, compared to previous studies in nearby Konya and Karaman provinces (ZHOU et al., 2017). The association between *T. gondii* and *N. caninum* seropositivity and abortion history suggests that these pathogens may be critical contributors to abortion cases in sheep in the sampling area. This association is supported by the presence of carnivores on the farms, as well as the environmental and management factors mentioned, which may facilitate the spread of oocysts and increase the risk of infection.

In studies investigating *T. gondii* and *N. caninum* seroprevalence in sheep worldwide, the seroprevalences of co-infections were found to be 0.5-9.03% (PANADERO et al., 2010; GUIMARÃES et al., 2015; TAFNER FERREIRA et al., 2016). The overall co-infection seroprevalence was 20.58%, whereas it reached 41.7% in sheep with an abortion history in the present study. Coinfection with *T. gondii* and *N. caninum* has been shown to exacerbate adverse clinical outcomes, notably increasing

the risk of abortion and embryonic death compared to single-pathogen infections (CIUCA et al., 2020). In this study, it is thought that the high abortion rate may be attributed to the high co-infection prevalence, as also reported by CIUCA et al. (2020). It is also recommended to consider the impact of co-infection on both transmission and recovery, so these factors can be integrated into future treatment strategies and disease control programs.

In conclusion, two crucial parasitic abortifacient agents, *T. gondii* and *N. caninum*, were investigated in sheep from Ankara, Konya, Karaman, and Aksaray provinces in the Central Anatolian Region. As a result of the study, it was observed that *T. gondii* infection is common in sheep in Konya, Karaman, Aksaray, and Ankara provinces. However, the widespread presence of *N. caninum* in sheep herds indicated that its economic impact should not be underestimated in the sheep industry of the region. It should not be overlooked that sheep infected with *T. gondii* and *N. caninum* may serve as reservoirs for sheep and cattle infections in the investigated provinces, and abortions may occur in herds with *T. gondii* and *N. caninum* seropositive sheep. As control measures, immediate action is necessary to reduce the impact of *T. gondii* and *N. caninum* infections in sheep with an abortion history. Implementing sanitary strategies in animal husbandry, including rigorous cleaning, improved waste management, and restricted access of cats and dogs to livestock areas, can mitigate the spread of these pathogens. Control programs should also secondarily consider antiprotozoal treatments for cats and dogs to reduce oocyst shedding and the overall risk of environmental contamination. In addition, educating farm personnel on the transmission risks and best practices for hygiene around animals can enhance these measures, helping to protect livestock health and productivity.

Ethical approval

All experimental procedures followed the ethical guidelines of the Veterinary Faculty of Selcuk University, Experimental Animals Production and Research Center Ethics Committee (SU-VDAMEK-2023/019).

Declaration of competing interest

The authors declare that they have no known competing professional, financial or non-financial, or personal conflict that could have appeared to influence the work reported in this paper.

Acknowledgments

The authors would like to thank Assist. Prof. Harun Yonar for the statistical analysis.

References

- AKTAS, M., N. DUMANLI, C. BABUR, Z. KARAER, H. ONGOR (2000): Determination of seropositivity for *Toxoplasma gondii* infection in pregnant and aborted sheep in Elazığ and vicinity by Sabin-Feldman (SF) Test. *Turk. J. Vet. Anim. Sci.* 24, 239-241.
- ANDRADE, G. S., F. R. P. BRHUNA, M. B. M. ROCHA, A. S. GUIMARÃES, A. M. G. GOUVEIA, A. M. GUIMARÃES (2012): Seroprevalence and risk factors for *Neospora caninum* in sheep in the state Minas Gerais, southeastern Brazil. *Vet. Parasitol.* 188, 168-171.
<https://doi.org/10.1016/j.vetpar.2012.03.006>
- ASLAN CELIK, B., O. Y. CELIK, N. MOR, K. IRAK (2020): Investigation of seroprevalence of *Toxoplasma gondii* in sheep and goats in Siirt province in Turkey. *Dicle Univ. Vet. Fak. Derg.* 13, 144-148.
<https://doi.org/10.47027/duvetfd.754668>
- BUXTON, D., S. W. MALEY, S. E. WRIGHT, S. RODGER, P. BARTLEY, E. A. INNES (2007): Ovine toxoplasmosis. Transmission, clinical outcome, and control. *Parassitologia* 49, 219-221.
- CAKMAK, D. O., B. KARATEPE (2017): Seroprevalence of *Toxoplasma gondii* in sheep from Nevşehir province in Turkey. *Türkiye Parazitolo. Derg.* 41, 148-151.
<https://doi.org/10.5152/tpd.2017.5245>
- CEYLAN, C., F. SEVINC, O. CEYLAN (2024): Serostatus of small ruminant toxoplasmosis and neosporosis throughout the southeastern Anatolia region of Türkiye. *Pak. Vet. J.* 44, 917-923.
<https://doi.org/10.29261/pakvetj/2024.247>
- CEYLAN, O., X. XUAN, F. SEVINC (2021): Primary tick-borne protozoan and rickettsial infections of animals in Turkey. *Pathogens* 10, 231.
<https://doi.org/10.3390/pathogens10020231>
- CICEK, H., C. BABUR, M. ESER (2011): Seroprevalence of *Toxoplasma gondii* in Pırlak sheep in the Afyonkarahisar province of Turkey. *Türkiye Parazitolo. Derg.* 35, 137-139.
<https://doi.org/10.5152/tpd.2011.34>
- CIUCA, L., G. BORRIELLO, A. BOSCO, L. D'ANDREA, G. CRINGOLI, P. CIARAMELLA, M. P. MAURELLI, A. DI LORIA, L. RINALDI, J. GUCCIONE (2020): Seroprevalence and Clinical Outcomes of *Neospora caninum*, *Toxoplasma gondii* and *Besnoitia besnoiti* Infections in Water Buffaloes (*Bubalus bubalis*). *Animals (Basel)* 10, 532.
<https://doi.org/10.3390/ani10030532>
- CLEMENTINO, M. M., M. F. SOUZA, V. F. ANDRADE NETO (2007): Seroprevalence and *Toxoplasma gondii* IgG avidity in sheep from Lajes, Brazil. *Vet. Parasitol.* 146, 199-203.
<https://doi.org/10.1016/j.vetpar.2007.02.036>
- DUBEY, J. P. (2009): Toxoplasmosis in sheep-The last 20 years. *Vet. Parasitol.* 163, 1-14.
<https://doi.org/10.1016/j.vetpar.2009.02.026>
- DUBEY, J. P. (2003): Review of *Neospora caninum* and neosporosis in animals. *Korean J. Parasitol.* 41, 1-16.
<https://doi.org/10.3347/kjp.2003.41.1.1>
- DUBEY, J. P., F. H. A. MURATA, C. K. CERQUEI-RA-CEZAR, O. C. H. KWOK, C. SU (2020): Economic and public health importance of *Toxoplasma gondii* infections in sheep: 2009-2020. *Vet. Parasitol.* 286, 109195.
<https://doi.org/10.1016/j.vetpar.2020.109195>
- DUBEY, J. P., A. HEMPHILL, R. CALERO-BERNAL, G. SCHARES (2017): Neosporosis in sheep. In: Neosporosis in Animals. (Dubey J. P., A. Hemphill, R. Calero-Bernal, G. Schares, Eds), 1st ed., CRC Press.
- DUBEY, J. P., M. PROWELL (2013): Ante-mortem diagnosis, diarrhea, oocyst shedding, treatment, isolation, and genetic typing of *Toxoplasma gondii* associated with clinical toxoplasmosis in a naturally infected cat. *J. Parasitol.* 99, 158-160.
<https://doi.org/10.1645/GE-3257.1>
- DUBEY, J. P., G. SCHARES (2011): Neosporosis in animals-the last five years. *Vet. Parasitol.* 180, 90-108.
<https://doi.org/10.1016/j.vetpar.2011.05.031>
- DUBEY, J. P., G. SCHARES, L. M. ORTEGA-MORA (2007): Epidemiology and control of neosporosis and *Neospora caninum*. *Clin. Microbiol. Rev.* 20, 323-367.
<https://doi.org/10.1128/cmr.00031-06>
- ESKI, F., P. DEMIR, C. BABUR, A. E. UTUK (2018): A research on the seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in sheep in Adana province of Turkey. *Etlık Vet. Mikrobiyol. Derg.* 29, 19-23. (in Turkish)
- EZATPOUR, B., M. ALIREZAEI, A. HASSANVAND, M. ZIBAEI, M. AZADPOUR, F. EBRAHIMZADEH (2015): The first report of *Neospora caninum* prevalence in aborted and healthy sheep from west of Iran. *Comp. Clin. Pathol.* 24, 19-22.
<https://doi.org/10.1007/s00580-013-1846-x>

- FIGLIUOLO, L. P., N. KASAI, A. M. RAGOZO, V. S. DE PAULA, R. A. DIAS, S. L. P. SOUSA, S. M. GENNARI (2004): Prevalence of anti-*Toxoplasma gondii* and anti-*Neospora caninum* antibodies in ovine from Sao Paulo State, Brazil. *Vet. Parasitol.* 123, 161-166.
<https://doi.org/10.1016/j.vetpar.2004.06.006>
- FILHO, P., J. M. B. OLIVEIRA, M. R. ANDRADE, J. G. SILVA, P. C. P. KIM, J. C. ALMEIDA, W. J. N. PORTO, R. A. MOTA (2017): Incidence and vertical transmission rate of *Neospora caninum* in sheep. *Comp. Immunol. Microbiol. Infect. Dis.* 52, 19-22.
<https://doi.org/10.1016/j.cimid.2017.05.006>
- GOKCE, G., N. MOR, A. H. KIRMIZIGUL, K. BOZUKLUHAN, E. E. ERKILIC (2015): The first report of seropositivity for *Neospora caninum* in sheep from Turkey. *Isr. J. Vet. Med.* 70, 40-44.
- GONZALEZ-WARLETA, M., J. A. CASTRO-HERMIDA, J. REGIDOR-CERRILLO, J. BENAVIDES, G. ALVAREZ-GARCIA, M. FUERTES, L. M. ORTEGA-MORA, M. MEZO (2014): *Neospora caninum* infection as a cause of reproductive failure in a sheep flock. *Vet. Res.* 45, 88.
<http://www.veterinaryresearch.org/content/45/1/88>
- GUIMARÃES, A., J. M. RAIMUNDO, L. MORAES, A. T. SILVA, H. A. SANTOS, M. S. PIRES, R. Z. MACHADO, C. D. BALDANI (2015): Occurrences of anti-*Toxoplasma gondii* and anti-*Neospora caninum* antibodies in sheep from four districts of Tocantins state, Brazilian Legal Amazon Region. *Pesq. Vet. Bras.* 35, 110-114.
- GUNLU, A., B. MAT (2021): The Place and Importance of Sheep-Goat Breeding in the Turkish Economy. In: *Sheep Goat Health and Breeding in the Prevention of Lamb and Capricorn Loss.* (Erdem, H., E. Ciftci, M. K. Isik, M. U. Yorgancılar, Eds.), Akademisyen Publishing House, pp 12-13.
- HECKER, Y. P., E. L. MORREL, M. A. FIORENTINO, I. GUAL, E. RIVERA, F. FIORANI, M. A. DORSCH, M. L. GOS, L. L. PARDINI, M. V. SCIOLI, S. MAGARINOS, F. A. PAOLICCHI, G. J. CANTON, D. P. MOORE (2019): Ovine abortion by *Neospora caninum*: First case reported in Argentina. *Acta Parasitol.* 64, 950-955.
<https://doi.org/10.2478/s11686-019-00106-z>
- HELMICK, B., A. OTTER, J. MCGARRY, D. BUXTON (2002): Serological investigation of aborted sheep and pigs for infection by *Neospora caninum*. *Res. Vet. Sci.* 73, 187-189.
[https://doi.org/10.1016/s0034-5288\(02\)00093-0](https://doi.org/10.1016/s0034-5288(02)00093-0)
- HEMPHILL, A., B. GOTTSTEIN (2000): A European perspective on *Neospora caninum*. *Int. J. Parasitol.* 30, 877-924.
[https://doi.org/10.1016/s0020-7519\(00\)00072-2](https://doi.org/10.1016/s0020-7519(00)00072-2)
- INCI, A., N. AYDIN, C. BABUR, Y. CAM, C. AKDOGAN, S. KUZAN (1999): Seroepidemiological studies on toxoplasmosis and brucellosis in cattle and sheep around Kayseri. *Pendik Vet. Mikrobiyol. Derg.* 30, 41-46. (in Turkish)
- KAMBURGIL, K., R. DURGUT, E. HANDEMIR (2001): Seroprevalence of toxoplasmosis in flocks that have aborted sheep in Hatay province. *Veterinarium* 12, 1-4. (in Turkish)
- KARATEPE, M., B. KARATEPE (2020): Prevalence of anti-*Neospora caninum* antibodies in sheep in Nevşehir province, Turkey. *Isr. J. Vet. Med.* 75, 168-171.
- KARATEPE, M., C. BABUR, B. KARATEPE (2001): Seroprevalence of *Toxoplasma gondii* detected by the Sabin-Feldman Dye Test in sheep in the region of Gümüşhacıköy (Amasya). *Türkiye Parazitol. Derg.* 25, 110-112. (in Turkish)
- LI, G., W. ZHENG, J. YANG, T. QI, Y. HE, W. CHEN, H. MA, Y. SUN, Y. LI, M. KANG, J. LI (2021): Seroprevalence and epidemiology of *Toxoplasma gondii* in animals in the Qinghai-Tibetan Plateau Area, China. *Pathogens* 10, 432.
<https://doi.org/10.3390/pathogens10040432>
- MEIXNER, N., M. F. SOMMER, N. SCUDA, K. MATIASEK, M. MULLER (2020): Comparative aspects of laboratory testing for the detection of *Toxoplasma gondii* and its differentiation from *Neospora caninum* as the etiologic agent of ovine abortion. *J. Vet. Diagn. Invest.* 32, 898-907.
<https://doi.org/10.1177/1040638720962110>
- MENDONÇA, C. E., S. L. B. BARROS, V. A. A. GUIMARÃES, A. S. FERRAUDO, A. D. MUNHOZ (2013): Prevalence and risk factors associated to ovine toxoplasmosis in northeastern Brazil. *Rev. Bras. Parasitol. Vet.* 22, 230-234.
<https://doi.org/10.1590/S1984-29612013000200042>
- MENZIES, P. I. (2011): Control of essential causes of infectious abortion in sheep and goats. *Vet. Clin. North Am. Food Anim. Pract.* 27, 81-93.
<https://doi.org/10.1016/j.cvfa.2010.10.011>
- MOR, N., M. O. ARSLAN (2007): Seroprevalence of *Toxoplasma gondii* in Sheep in Kars province. *Kafkas Univ. Vet. Fak. Derg.* 13, 165-170.
<https://doi.org/10.9775/kvfd.2007.33-A>
- NAYERI, T., S. SARVI, M. MOOSAZADEH, A. DARYANI (2022): The global prevalence of *Neospora caninum* infection in sheep and goats that had an abortion and aborted fetuses: A systematic review and meta-analysis. *Front. Vet. Sci.* 9, 870904.
<https://doi.org/10.3389/fvets.2022.870904>
- NOAKES, D. E., T. J. PARKINSON, G. C. W. ENGLAND (2001): Infertility in the ewe and doe (female goat). In: *Arthur's Veterinary Reproduction and Obstetrics.* (Noakes, D.E., N., T. J. Parkins, G. C. W. England, G. H. Arthur, Eds.), 8th ed., Bailliere Tindall, London, pp. 570-575.
<https://doi.org/10.1016/B978-0-7020-2556-3.X5001-4>

- PANADERO, R., A. PAINCEIRA, C. LÓPEZ, L. VÁZQUEZ, A. PAZ, P. DIAZ, V. DACAL, S. CIENFUEGOS, G. FERNANDEZ, N. LAGO, P. DIEZ-BANOZ, P. MORRONGO (2010): Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in wild and domestic ruminants sharing pastures in Galicia (Northwest Spain). *Res. Vet. Sci.* 88, 111-115.
<https://doi.org/10.1016/j.rvsc.2009.05.010>
- ROSSI, G. F., D. D. CABRAL, D. P. RIBEIRO, A. C. PAJUBA, R. R. CORREA, M. Q. MOREIRA, T. W. P. MINEO, J. R. MINEO, D. A. O. SILVA (2011): Evaluation of *Toxoplasma gondii* and *Neospora caninum* infections in sheep from Uberlandia, Minas Gerais state, Brazil, by different serological methods. *Vet. Parasitol.* 175, 252-259.
<https://doi.org/10.1016/j.vetpar.2010.10.017>
- SAWADOGO, P., J. HAFID, B. BELLETE, R. TRAN MANH SUNG, M. CHAKDI, P. FLORI, H. RABERIN, I. BENT HAMOUNI, A. CHAIT, A. DALAL (2005): Seroprevalence of *T. gondii* in sheep from Marrakech, Morocco. *Vet. Parasitol.* 130, 89-92.
<https://doi.org/10.1016/j.vetpar.2005.03.025>
- TAFNER FERREIRA, M. S., F. S. FLORES VOGEL, L. A. SANGIONI, SKREBSKY, A. CEZAR, F. REZER DE MENEZES (2016): *Neospora* spp. and *Toxoplasma gondii* infection in sheep flocks from Rio Grande do Sul, Brazil. *Semin. Cienc. Agrar.* 37, 1397-1406.
- TENTER, A. M., A. R. HECKEROTH, L. M. WEISS (2000): *Toxoplasma gondii*: from animals to humans. *Int. J. Parasitol.* 30, 1217-1258.
[https://doi.org/10.1016/S0020-7519\(00\)00124-7](https://doi.org/10.1016/S0020-7519(00)00124-7)
- TIROSH-LEVY, S., R. WOLKOMIRSKI, I. SAVITSKY, G. KENIGSWALD, O. FRIDGUT, S. BARDENSTEIN, S. BLUM, M. L. MAZUZ (2022): *Neospora*-related abortions in sheep in Israel-A serological diagnostic challenge in an endemic area. *Vet. Parasitol. Reg. Stud. Rep.* 36, 100809.
<https://doi.org/10.1016/j.vprsr.2022.100809>
- TRANAS, J., R. A. HEINZEN, L. M. WEISS, M. M. MCALLISTER (1999): Serological evidence of human infection with the protozoan *Neospora caninum*. *Clin. Diagn. Lab. Immunol.* 6, 765-767.
<https://doi.org/10.1128/CDLL6.5.765-767.1999>
- UTUK, A. E., F. ESKI (2022): *Toxoplasma gondii* and *Neospora caninum* Infections in Small Ruminants, Epidemiology, Monitoring, Prevention and Control Strategies. In: *Epidemiology, Monitoring and Eradication Strategies for Diseases of Primary Importance in Sustainable Herd Health in Sheep and Goat Farms.* (Yaşar, E., Ed.), Türkiye Klinikleri, pp. 102-113.
- VESCO, G., W. BUFFOLANO, S. LA CHIUSA, G. MANCUSO, S. CARACAPPA, A. CHIANCA, S. VILLARI, V. CURRO, F. LIGA, E. PETERSEN (2007): *Toxoplasma gondii* infections in sheep in Sicily, southern Italy. *Vet. Parasitol.* 146, 3-8.
<https://doi.org/10.1016/j.vetpar.2007.02.019>
- YAN, C., L. J. LIANG, K. Y. ZHENG, X. Q. ZHU (2016): Impact of environmental factors on the emergence, transmission and distribution of *Toxoplasma gondii*. *Parasit. Vectors.* 9, 137.
<https://doi.org/10.1186/s13071-016-1432-6>
- ZHOU, M., S. CAO, F. SEVINC, M. SEVINC, O. CEYLAN, M. LIU, G. WANG, P. F. A. MOUMOUNI, C. JIRAPATTHARASATE, H. SUZUKI, Y. NISHIKAWA, X. XUAN (2017): Enzyme-linked immunosorbent assays using recombinant TgSAG2 and NcSAG1 to detect *Toxoplasma gondii* and *Neospora caninum*-specific antibodies in domestic animals in Turkey. *J. Vet. Med. Sci.* 78, 1877-1881.
<https://doi.org/10.1292/jvms.16-0234>

Received: 3 September 2024

Accepted: 5 December 2024

Online publication: 30 April 2025

CEYLAN, C., O. CEYLAN, S. CAO, H. H. HADİMLİ, X. XUAN, F. SEVİNC: Visoka prevalencija parazita *Toxoplasma gondii* i *Neospora caninum* u ovaca s anamnezom pobačaja uzgajanih u središnjem dijelu Turske. Vet. arhiv 95, 411-421, 2025.

SAŽETAK

Uzgoj malih preživaca ima ključnu ulogu u stočarstvu u Turskoj, no pod utjecajem je različitih infektivnih patogena, uključujući parazite *Toxoplasma (T.) gondii* i *Neospora (N.) caninum*. Cilj rada bio je istražiti seroprevalenciju toksoplazmoze i neosporoze u ovaca s različitom pojavnošću pobačaja. Ovce su uzgajane u Središnjoj Anatoliji u Turskoj. Uzeta su ukupno 243 uzorka krvi ovaca s različitih farmi, među kojima je 115 uzoraka potjecalo od ovaca s problemom pobačaja, dok su ostali uzorci uzeti od ovaca bez povijesti pobačaja. Za određivanje specifičnih protutijela na *T. gondii* i *N. caninum* upotrijebljen je indirektni ELISA test temeljen na *TgSAG2* i *NcSAG1*. Istraživanje je pokazalo visoku prevalenciju parazita *T. gondii* i *N. caninum* u skupini ovaca s problemom pobačaja. Seroprevalencija navedenih parazita u tih ovaca bila je 66,1% za *T. gondii* i 50,4% za *N. caninum*. Seroprevalencija u nasumično odabranih ovaca, bez pobačaja u anamnezi, iznosila je 26,6% za *T. gondii* i 10,9% za *N. caninum*. Rezultati su pokazali najvišu seroprevalenciju parazita *N. caninum* u Turskoj i to kad se u obzir uzme ukupni broj pretraženih ovaca (29,63%) i kad se u obzir uzmu samo ovce s poviješću pobačaja (50,4%). Uzimajući u obzir da je neosporoza primarni protozoalni uzročnik pobačaja u goveda, visoka seroprevalncija parazita *N. caninum* u ovaca ukazuje da bi kod dijagnostike njihova pobačaja u obzir trebalo uzeti i tog uzročnika. Istraživanje je pokazalo da su toksoplazmoza i neosporoza rasprostranjene i mogu pridonijeti pojavi pobačaja ovaca uzgajanih u Središnjoj Anatoliji u Turskoj.

Ključne riječi: ELISA; neosporoza; pobačaj u ovaca; protozoalna infekcija; toksoplazmoza; Turska
