

SHORT COMMUNICATION

## DECOMPOSITION OF FISH SAMPLES FOR DETERMINATION OF MERCURY

LJERKAPRESTER, DIANA JUREŠA, AND MAJABLANUŠA

*Institute for Medical Research and Occupational Health, Zagreb, Croatia*

Received 12 November 1998

The aim of the study was to compare the efficiency of acid and alkaline decomposition of biological materials using an open and a closed system for total mercury determination. Acid digestion was performed with concentrated HNO<sub>3</sub> in tubes at 80 °C and lasted five hours. Alkaline digestion was performed with a 45% NaOH and a 1% cysteine, heated at 120 °C for 20 minutes.

Total mercury was measured by atomic absorption spectrometry using the cold vapour technique (CVAAS). The average recovery obtained for analysis of certified reference material in closed tubes for acid digested sample was superior to the alkaline one, 103±4% vs. 70±3%, respectively. In addition, the recoveries through the open system acid digestion (90±8%) and the open system alkaline digestion (57±2%) were lower than through the respective closed system digestions. Reproducibility of the acid decomposition method was superior to the alkaline one.

*Key words:*  
acid digestion, alkaline digestion, biological material, closed system, cold vapour atomic absorption spectrometry, mercury recovery, open system

The analysis of total mercury in biological samples requires two separate processes: (i) the conversion of all forms of mercury to ionic mercury (Hg<sup>2+</sup>) by various digestion/oxidation techniques and (ii) the reduction of Hg<sup>2+</sup> to its atomic form (Hg<sup>0</sup>) for analysis (1). Different methods for digestion of biological materials for quantitative determination of mercury are described in the literature. Some of these methods used alkaline (2, 3) and others acid digestion technique (4-6). So far, the results obtained

using the two techniques on the same sample have not been compared. The aim of this study was to compare the efficiency of alkaline and acid decomposition of biological material (fish), carried out in an open and a closed digestion system.

## MATERIAL AND METHODS

### *Reagents*

All reagents (Kemika, Zagreb) were analytical grade and were selected for their low mercury content. Water was deionised to the conductivity of 0.06  $\mu\text{S}/\text{cm}$ .

Stock solution of mercury (500  $\mu\text{g Hg}^{2+}/\text{ml}$ ) was prepared by dissolving 0.06767 g of mercuric chloride in 100 ml of 5%  $\text{H}_2\text{SO}_4$ . From this stock solution, 0.2  $\mu\text{g Hg}^{2+}/\text{ml}$  solution was prepared for making working solutions of 2–20 ng  $\text{Hg}^{2+}/\text{ml}$ . Working solutions were prepared daily. Reduction agent was made by mixing  $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$  (2.4%) and  $\text{CdCl}_2$  (0.5%), and by adding L-cysteine (0.2%) and NaCl (1%) to  $\text{H}_2\text{SO}_4$  (2.25 M). Sodium hydroxide (35%) was used to reduce mercury in the alkaline medium.

### *Acid digestion*

About 1 g of fish homogenate was first digested with 2 ml of concentrated  $\text{HNO}_3$  in borosilicate glass tubes overnight at room temperature and then the next day at 80 °C for five hours in a programmed digestion system (DS-40, Tecator, Sweden). Digestion was performed in open or in closed tubes. After digestion samples were cooled to room temperature and the volume adjusted to 10 ml with deionised water, an aliquot of 0.5 ml was taken for mercury determination.

### *Alkaline digestion*

About 1 g of fish homogenate was added 2 ml NaOH (45%) and 1 ml cysteine (1%), and heated in a glass tube at 120–150 °C for 20 minutes. Digestion was performed in open or in closed tubes. Samples were cooled to room temperature and the volume adjusted to 10 ml with deionised water. An aliquot of 0.5 ml was taken for mercury determination.

### *Sample preparation and method verification*

Before digestion, a sample of fish muscle (hake, *Merluccius merluccius*) was prepared as a 50% homogenate in a 1% saline solution with Kinematica homogeniser (Switzerland). The accuracy of both digestion techniques was verified using the reference material (horse kidney H-8, IAEA, Austria) and expressed as percentage of recovery. Five replicates of 0.04 g lyophilised reference material were digested either with 0.5 ml of  $\text{HNO}_3$  or with 0.5 ml of 45% NaOH and 0.25 ml of 1% L-cysteine. Samples were digested in open and in closed tubes as described in the fish digestion. To determine the reproducibility of the method, the whole procedure of digestion and mercury measurement was carried out with 9 replicates of the fish homogenate. Relative stan-

standard deviation (RSD) was calculated as percentage of standard deviation from arithmetic mean.

### Mercury determination

Total mercury was analysed by cold vapour atomic absorption spectrometry (CVAAS) on Mercury Monitor LDC (Milton Roy, Riviera Beach, FL) after reduction with stannous and cadmium chloride in an alkaline medium. This method was described as sensitive and reliable for determination of total mercury in aqueous solutions (2, 3, 7, 8). The detection limit of the method was calculated from 10 repeated measurements of the lowest standard solution (2 ng/ml).

## RESULTS AND DISCUSSION

Table 1 shows the reproducibility of acid and alkaline digestion methods. Mercury was analysed in 9 replicates of the same fish homogenate. Calculated as relative standard deviation, the precision of the acid digestion method performed in a closed or an open system was 10% and 13%, respectively. The precision of the alkaline digestion performed in an open system was 14%.

Table 1 *Reproducibility of the method for mercury determination in a 50% fish homogenate digested using acid or alkaline technique (in a closed or an open system)*

Digestion	Acid (ng Hg/g)	RSD (%)	Alkaline (ng Hg/g)	RSD (%)
Closed	319 ± 31	10	–	–
Open	112 ± 14	13	150 ± 21	14

Results are presented as arithmetic mean ± standard deviation of 9 replicates. RSD is relative standard deviation.

The accuracy of the methods after each digestion procedure was verified by measuring the certified reference material (Table 2). The average recovery for the sample digested with nitric acid in a closed and an open system was 103 ± 4% and 90 ± 8%, respectively. Alkaline digestion yielded lower recovery of only 70 ± 3% and 57 ± 2% for the respective closed and open systems. The detection limit of CVAAS method was 6 ng Hg/g of fresh tissue.

The results show that digestion of biological material in an open system results in considerable loss of mercury. Our study corroborates the results of *Jones and co-workers* (1) who recently found considerable losses of organic mercury in an open-vessel digestion. The above findings render questionable many earlier reports on mercury determination in undigested or open digested system of biological material (2, 3, 7).

Table 2 The effect of acid or alkaline digestion methods (carried out in closed or in open system) on the accuracy of mercury analyses using standard reference material (horse kidney H-8, IAEA) containing  $910 \pm 70$  ng Hg/g.

Digestion	ng Hg/g	Recovery (%)
Acid		
closed	$934 \pm 38$	$103 \pm 4$
open	$815 \pm 6$	$90 \pm 8$
Alkaline		
closed	$641 \pm 28$	$70 \pm 3$
open	$519 \pm 13$	$57 \pm 2$

Results are presented as arithmetic mean  $\pm$  standard deviation of 5 replicates.

Furthermore, our results show that alkaline digestion method yields lower recovery of mercury (Table 1 and 2), even if carried out in a closed system. The reason either lies in its loss during digestion or in its incomplete reduction in the reaction vessel. Alkaline digestion is probably unable to decompose methylmercury and, consequently, all mercury present in solution cannot be reduced in the reduction vessel (2). In general, acidic digestion yields much higher recovery than the alkaline.

There are many methods for the digestion of biological samples. Most of them involve an open-vessel digestion system and use a variety of acids (HCl, HNO<sub>3</sub>, and H<sub>2</sub>SO<sub>4</sub>) (9–12). Strong oxidants such as KMnO<sub>4</sub>, K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> in acid mixture of HNO<sub>3</sub> and H<sub>2</sub>SO<sub>4</sub> were used by some investigators (11, 13). These methods appear to result in complete digestion of biological material, but they are lengthy, may contaminate samples, exhibit high blank values, and give poor precision. Closed ampule digestion/oxidation technique prevents loss of organic-mercury compounds by volatilisation. A closed vessel also allows heat and pressure to break down the carbon-mercury bond in one step. Our experience confirms that once the samples are prepared in a closed vessel, they are stable with respect to the loss of mercury (1).

In the last few years, a simple method was developed using only one acid (concentrated nitric acid) in closed test tubes (1, 6, 12). In this study, we also applied a single (nitric) acid in digestion of samples. Our results show that acid digestion method using only nitric acid gives very good accuracy and precision (Table 1 and 2). Acid digestion performed in a closed system yields higher recovery than in the open one.

To conclude, our results indicate that acid digestion is superior to the alkaline in terms of accuracy and precision. A closed digestion system is also mandatory for quantitative mercury determination.

#### Acknowledgement

The authors are grateful to Ms. Marija Ciganović for excellent technical assistance.

## REFERENCES

1. Jones RD, West-Thomas J, Arfstrom C. Closed-ampule digestion procedure for the determination of mercury in soil and tissue using cold vapour atomic fluorescence spectrometry. *Bull Environ Contam Toxicol* 1997;59:29–34.
2. Magos L. Selective atomic-absorption determination of inorganic mercury and methylmercury in undigested biological samples. *Analyst* 1971;96:847–53.
3. Farant JP, Brissette D, Moncion L, Bigras L, Chartrand A. Improved cold-vapour atomic absorption technique for the microdetermination of total and inorganic mercury in biological samples. *J Anal Toxicol* 1981;5:47–51.
4. Horvat M, Zvonarić T, Stegnar P. Optimization of a wet digestion method for the determination of mercury in blood by cold vapour atomic absorption spectrometry (CVAAS). *Vest Slov Kem Drus* 1986;33:475-7.
5. Bergdahl IA, Schutz A, Hansson GA. Automated determination of inorganic mercury in blood after sulphuric acid treatment using cold vapour atomic absorption spectrometry and an inductively heated gold trap. *Analyst* 1995;120:1205–9.
6. Prester Lj, Blanuša M, Ciganović M. Analysis of mercury in fish from the Adriatic. In: XIII Meeting of Chemists of Croatia. Abstracts. Zagreb: Croatian Society of Chemical Engineers and Technologists, 1993;S-2 410 (in Croatian).
7. Ngim CH, Foo SC, Phoon WO. Atomic absorption spectrophotometric microdetermination of total mercury in undigested biological samples. *J Anal Toxicol* 1988;12:132–5.
8. Jureša D. Comparison of methods for decomposition of biological material in analysis of mercury. (B.Sc. Thesis). Zagreb: University of Zagrebu, 1997. 46 p. (in Croatian).
9. Nguyen TH, Boman J, Leermakers M, Baeyens W. Mercury analysis in environmental samples by EDXRF and CV-AAS. *Fres J Anal Chem* 1998;360(2):199–204.
10. Van Delft W, Vos G. Comparison of digestion procedures for the determination of mercury in soils by cold-vapour atomic absorption spectrometry. *Anal Chim Acta* 1988;209:147–56.
11. Szakacs O, Lasztity A, Horvath ZS. Breakdown of organic mercury compounds by hydrochloric acid-permanganate or bromine monochloride solution for the determination of mercury by cold vapour atomic absorption spectrometry. *Anal Chim Acta* 1980;121:219–24.
12. Horvat M. Studies and development of methods for determination of low concentrations of mercury in biological environmental samples. (PhD Thesis). Ljubljana: University of Ljubljana, 1989. 164 p. (in Slovene).
13. Scheuhammer AM, Bond AD. Factors affecting the determination of total mercury in biological samples by continuous-flow cold vapour atomic absorption spectrophotometry. *Biol Trace Elem Res* 1991;31:119–29.

*Sažetak***RAZARANJE UZORAKA RIBE ZA ANALIZU ŽIVE**

Svrha ovog rada bila je ispitati učinkovitost dviju metoda razaranja biološkog materijala pri analizi žive. Razaranje u kiselom mediju s koncentriranom  $\text{HNO}_3$  primijenjeno je pri  $80\text{ }^\circ\text{C}$  tijekom 5 sati. Razaranje uzorka u alkalnom mediju primijenjeno je s 45% NaOH i 1% cisteinom pri  $120\text{ }^\circ\text{C}$  tijekom 20 minuta. Obje metode razaranja provedene su u otvorenim ili zatvorenim staklenim epruvetama. Ukupna živa određena je metodom atomske apsorpcijske spektrometrije tehnikom razvijanja hladnih para (CVAAS). Iskorištenje razaranja uzorka ispitano je na referentnom uzorku (konjski bubreg H-8, IAEA, Austrija). Rezultati su pokazali bolje iskorištenje određivanja žive u zatvorenim epruvetama u kiselom mediju ( $103\pm 4\%$ ) negoli u alkalnom mediju ( $70\pm 3\%$ ). Iskorištenje u otvorenim epruvetama je manje negoli u zatvorenima te iznosi za razaranje uzorka u kiselom mediju  $90\pm 8\%$  i samo  $57\pm 2\%$  u alkalnom mediju. Ponovljivost metode (koeficijent varijabilnosti izražen kao postotak standardne devijacije od aritmetičke sredine) ispitana je na uzorcima oslića (*Merluccius merluccius*). Metoda razaranja biološkog materijala u kiselom mediju preciznija je od metode razaranja u alkalnom mediju.

*Ključne riječi:*

atomska apsorpcijska spektrometrija razvijanjem hladnih para, biološki materijal, oslobađanje žive, otvoreni proces, razgradnja uzoraka kiselinom, razgradnja uzoraka lužinom, zatvoreni proces

Requests for reprints:

Ljerka Prester, Ph.D.  
Institute for Medical Research and Occupational Health  
Ksaverska cesta 2, P.O. Box 291, HR-10001 Zagreb, Croatia  
E-mail: [ljprester@imi.hr](mailto:ljprester@imi.hr)