

## **IN VITRO GENOTOXICITY OF WASTEWATERS FROM THE TOWN OF SETTAT, MOROCCO**

Jaouad EL ASSLOUJ<sup>1</sup>, Loubna AMAHDAR<sup>2</sup>, Kamal GLOUIB<sup>2</sup>, Sanae KHOLTEI<sup>1</sup>,  
Namira EL AMRANI PAAZA<sup>3</sup>, Luc VERSCHAEVE<sup>4</sup>, and Abderraouf HILALI<sup>2</sup>

*Laboratory of Environmental Metrology<sup>1</sup>, Research Group on Toxicogenetics and Mutagenesis, Laboratory of Agrofood and Health<sup>2</sup>, Laboratory of Applied Geology<sup>3</sup>, University Hassan I, FST Settata, Morocco, Scientific Institute of Public Health, Department of Epidemiology and Toxicology, Brussels, Belgium<sup>4</sup>*

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In recent years, the town of Settata has seen a considerable industrial growth, which has resulted in increased environmental pollution. This includes pollution by household and industrial wastewaters, which are released into the Boumoussa River without any preliminary treatment. The river valley crosses the community of Mzamza 8 km to the north of the town. Years of drought forced members of the community to use this polluted ground water for irrigation and put themselves and the environment at risk.

The aim of this study was to determine the physicochemical and metal profile of Settata wastewaters and to assess their impact on the water table. The second objective was to investigate the genotoxic potential of wastewater on human peripheral blood lymphocytes *in vitro*, using the micronucleus test and cellular proliferation index.

This study demonstrated significant pollution of Boumoussa valley groundwater and of the local wells. Sampled water induced a clear increase in the frequency of micronucleated cells and a lower cell proliferation in human peripheral blood lymphocytes *in vitro*.

**KEY WORDS:** *Boumoussa, heavy metals, micronuclei, physicochemical profile, proliferation index*

In Morocco, wastewaters generated by various economic activities are used for many applications. However, their use for irrigation poses an enormous threat to the environment and humans. Nevertheless, this is exactly what is being done near the town of Settata, located in the heart of Morocco, 70 km to the south of Casablanca. In recent years, this town has undergone a considerable industrial growth, and household and industrial wastewaters have been released in the Boumoussa River without any preliminary treatment. The river valley has become an open sewer. It crosses the community of Mzamza 8 km to the north of the town (zone of study). The plateau of Settata is characterised by a semi arid climate. The temperature is about 20 °C on average,

and precipitation is about 400 mm a year (1). This plateau has a very important water potential, and is the only water resource for the community of Mzamza (2). Years of drought, however, have forced members of the community to use this water for irrigation and to put themselves and the environment at risk posed by contaminated groundwater.

We determined the physicochemical profile and the content of heavy metals (Pb, Al, Zn, Cd and Cr) of the wastewater and water of the wells located in the vicinity of the Boumoussa valley. We also studied *in vitro* the genotoxic effects of this water on human peripheral blood lymphocytes using the micronucleus test and calculating the cell proliferation index.

## MATERIALS AND METHODS

### Study area

We studied the area of household and industrial wastewater discharge from the town of Settât. We sampled wastewaters and water from wells located in the Boumoussa valley and its vicinity (Figure 1, Table 1). Sampling station B was somewhat distant from the Boumoussa valley (Figure 1) and sampling included household and industrial wastewater as well as rainwater leaving the main drain. Station C was located in the Boumoussa valley, at a little distance

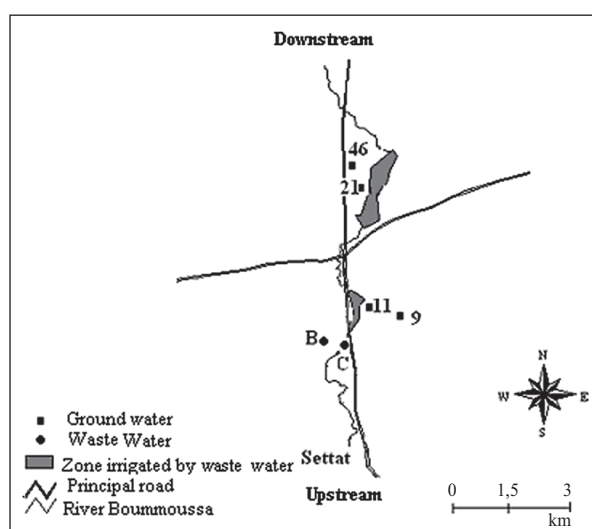


Figure 1 Geographic location of the sample sites

from the confluence. The wells that were included in this study were designated as wells 9, 11, 21, and 46. Well 9 was distant from the Boumoussa valley, whereas wells 11, 21, and 46 were near the valley.

### Water analysis

Waste and well water were collected in polyethylene bottles and transported in portable refrigerators at 4 °C. Physicochemical profiling was based on the AFNOR 2001 (3) and Rodier 1996 (4) standards. For heavy metal analysis, we mineralised waste and well water using nitric, hydrochloric, and perchloric acid (Merck, supra pure quality). Chromium, aluminium, lead, zinc, and cadmium content were measured using inductively coupled plasma atomic emission spectroscopy (ICP-AES). Mercury was analysed using a mercury AMA254 analyser.

### Micronucleus test

Blood was sampled in a sterile heparin test tube from a non-smoking healthy and sober 27-year-old man, who gave informed consent for participation in the study. Cell cultures were made according to the standard protocol for the micronucleus test as described by Fenech and Morley (5). Briefly, 0.5 mL of total venous blood was incubated in glass tubes containing 5 mL RPMI 1640 growth medium supplemented with 15 % foetal calf serum (Sigma-Aldrich Chemie GmbH), 1 % of phytohaemagglutinine CAS #14930-96-2, Sigma-Aldrich Chemie GmbH),

Table 1 Water sampling locations and distance of the wells from pollution sources

Stations	Depth / m	Altitude / m	Distance to sources of pollution / m	Observations	Type of irrigation
9	24.0	310	1150	Unprotected	Irrigation by rain water
11	19.0		400	Unprotected	Irrigation by rain water
21	18.1	273	475	Unprotected	Irrigation by waste water
46	11.9	500	500	Unprotected	Irrigation by waste water
Waste water B	Domestic and industrial waste water as well as rain water leaving the main drain. At some distance from the Boumoussa River				
Waste water C	A mix of waste water B and the water of the Boumoussa River				

and 1 % penicilline/streptomycine (Sigma-Aldrich Chemie GmbH). Culture tubes were incubated in the presence of 50 µL, 100 µL, 200 µL, or 400 µL of tap water, wastewater, or well water. Tap water was used as negative control, since it is the most consumed drinking water. The tubes were then placed horizontally in an incubator at 37 °C. Cytochalasin-B (CAS #9008-97-3, Sigma-Aldrich Chemie GmbH) was added after 44 hours (0.1 mL per tube to achieve the concentration of 3 µg mL<sup>-1</sup> in culture). At 72 h of incubation, cell suspensions were centrifuged at 1000 rpm for 10 minutes and subjected to a hypotonic shock with 0.075 mol L<sup>-1</sup> KCl. The cells were then fixed with a 1:3 acetic acid : methanol solution and spread over clean microscope slides. The slides were air-dried, stained with 5 % Giemsa in phosphate buffer (pH = 6.8) and coded. Each slide was examined for the presence of micronuclei (MN) using a light microscope (400X and 1000X). For MN identification we used the criteria as described by Fenech and Morley (5). Altogether, 1000 cells per treatment were scored and the code was broken only after all readings were performed.

*Proliferation index*

Cell proliferation index (PI) is an indirect measurement of cell cycle duration, and is calculated according to the following formula

$$PI = \frac{(1 \times N_1) + (2 \times N_2) + (3 \times N_3) + (4 \times N_4)}{1000 \text{ cells analysed}}$$

where N<sub>1</sub> to N<sub>4</sub> is the number of cells with 1 to 4 nuclei (6).

*Statistical analysis*

The statistically significant difference between control and treated samples was determined using the ANOVA II test without repetition.

RESULTS AND DISCUSSION

*Physicochemical and metal profiling*

Table 2 gives a brief physicochemical profile of control, waste, and well water. Wastewater collected at the point of discharge or at the Boumoussa valley had very low dissolved oxygen levels, not exceeding 9 %. Well water had dissolved oxygen below the standard limits (except for well 9). Water from stations B and C also had 1.466 mg L<sup>-1</sup> and 1.379 mg L<sup>-1</sup> of nitrite ions, respectively and 105 mg L<sup>-1</sup> and 84.6 mg L<sup>-1</sup> of ammonium, which are usually of household origin. Nitrates exceeded or approached the standard levels in all well waters except in well 9, which is the most

**Table 2** Physicochemical profile of waste and well

Stations	Water level* / m	PO <sub>4</sub> / mg L <sup>-1</sup>	pH	Cl / mg L <sup>-1</sup>	Conductivity at 20°C / µS cm <sup>-1</sup>	NO <sub>3</sub> / mg L <sup>-1</sup>	NO <sub>2</sub> / mg L <sup>-1</sup>	NH <sub>4</sub> / g L <sup>-1</sup>	O <sub>2</sub> / %
B	0	4.020	9.65	1065	2385.32	10.09	1.466	105	6.40
C	0	3.544	9.59	894.6	2285.71	17.07	1.379	84.6	9.00
21	4.78	0.041	7.72	1065	3505.15	92.63	0.046	0.02	59.5
46	4.00	0.044	7.28	795.2	3265.31	90.10	0.041	0.02	52.0
11	17.80	0.009	7.62	881.4	2857.14	47.33	0.017	0.03	53.7
9	19.21	0.009	7.06	463.0	1616.16	8.920	0.008	0.003	77.4
Tap water	0	0	7.08	433.0	1100.00	10.55	0	0	76.0
Moroccan standards (2002)	-----	0.700	6.50 to 8.50	750.0	2700.00	50.00	0.100	0.50	70.0
WHO standards (1996)	-----	-----	6.50 to 8.50	250.0	1200.00	50.00	3.000	----	75.0

\* compared to the ground

distant from the Boumoussa valley. At stations B and C nitrate concentrations were above standard (50 mg L<sup>-1</sup>). Lower nitrate concentrations at other places are most probably due to reduction by organic matter (7).

At wells 21, 46, and 11 conductivity was higher than the Moroccan standard (8) and higher than conductivity measured for wastewater. This is probably due to the presence of carbonated rocks in contact with ground waters (9). Except for well 9 water, chlorides exceeded the national standard.

Orthophosphates in wastewater were of household origin and were related to faeces and detergents (10). The observed values of 4.02 mg L<sup>-1</sup> and 3.54 mg L<sup>-1</sup> at points B and C, respectively are considerably higher than the values for well and tap water.

Groundwater pH was neutral to slightly alkaline, with a maximum of 7.72, whereas wastewater pH was alkaline, with a maximum of 9.65 at station B.

Wastewaters taken at stations B and C had Pb, Cr, Cd, Al, Zn and Hg levels considerably higher than those observed in well and tap water (Table 3). Except for Cd and Hg, all other levels were higher than the Moroccan and WHO (11) standards. It is important to point out that wastewater from stations B and C contained particularly high concentrations of Al, Zn, and Pb. These high levels can be due to specific use of land due to important industrial activities in the area, those related to surface treatments in particular.

Heavy metal concentrations measured in well and tap water were markedly lower than the Moroccan or WHO standards, and all well water concentrations were of the same order of magnitude as of tap water.

It should be noted that Al levels in wells 21, 46, and 11 approach the WHO limit. This may be related to

its mobility which is greater than of other metals (12), and also to its natural occurrence as aluminosilicate, which is the major constituent of clay (13).

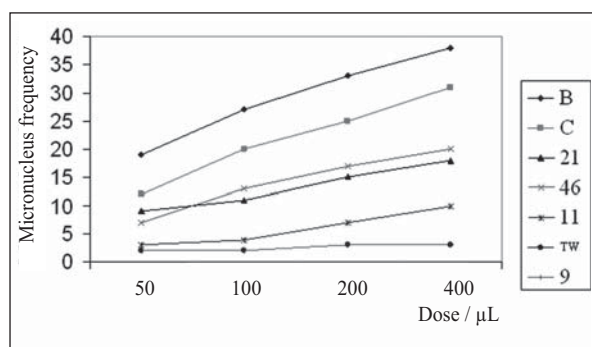
#### *Micronucleus test and proliferation index (PI)*

In this study, we used the micronucleus test to investigate the genotoxicity of the waste and well waters because it is a well-known and validated test system that was recently found to be predictive of cancer risk at the population level (14). This test detects chromosome breaks (clastogenicity) and aneuploidy (e.g. as a result of disturbances in the mitotic spindle). Furthermore, it can be more sensitive than the bacterial Ames assay, especially when heavy metal contamination is anticipated (15, 16). Recently, an inter-laboratory comparison study has also demonstrated that the *in vitro* micronucleus test is indeed very suitable for routine wastewater testing (17).

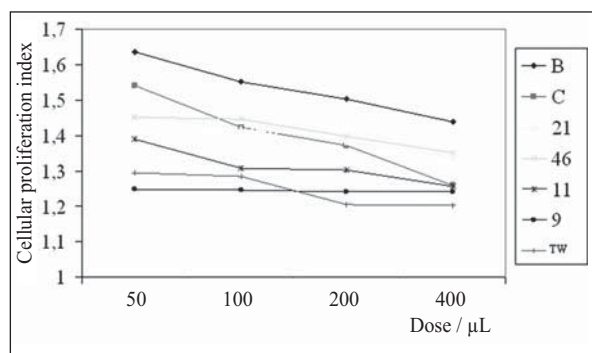
Data on micronucleus frequency are given in Figure 2 and Table 4. The frequencies of micronucleated cells increased with the amount of water added to cultures. However, this dose-related increase was statistically significant only for the wells 46 and 21 ( $p < 0.05$ ). When the micronucleus frequencies were compared between control (tap water) and exposed cultures (waste and well water), statistically significant increases were found for both stations and wells 21 and 46. Micronucleus frequency in the control culture was 3 micronuclei per 1000 binucleated cells, which is in accordance with literature and our own historical data (5, 18). This frequency (and an absence of the dose-effect relationship) was also found for well 9 water, which was the most distant from the Boumoussa valley (both graph lines on Figure 2 are overlapping).

Wells 21 and 46 showed the most significant effect among the wells, which is probably due to their vicinity to stations B and C and likely infiltration of pollutants from these wastewaters into the ground water.

Figure 3 and Table 4 show the proliferation index (PI). A significant increase ( $p < 0.05$ ) was found in cells treated with water from wells 21 and 46 compared to control (tap water). The difference was highly significant between the B and C stations and the tap water ( $p < 0.001$ ). With addition of more water PI would decrease, but not significantly. This dose-dependent decrease in proliferation may be indicative of the toxic effect of pollutants in the samples. It is not due to addition of increased volumes of water, as this had already been ruled out by tests which showed



**Figure 2** *Micronucleus frequency in human peripheral blood lymphocytes in relation to wastewater (B, C), well water (9, 11, 21, 46), or tap water (TW) doses (μL) added to cell cultures*



**Figure 3** Peripheral blood lymphocyte proliferation index in relation to wastewater (B, C), well water (9, 11, 21, 46), or tap water (TW) doses ( $\mu\text{L}$ ) added to cell cultures

that adding volumes of up to 800  $\mu\text{L}$  did not influence cell cultures and did not induce a toxic response, e.g., as a result of changed osmolarity (19).

Our results clearly demonstrate a high pollution level of the wastewaters in the Boumoussa valley and adjacent wells, which in turn involves significantly higher genotoxicity. This finding is in accordance with earlier investigations. Among recently published studies, reference can be made to Durgo et al. (16) who investigated wastewater polluted by fertilisers from a phosphoric gypsum depot near a factory in Croatia. Using the alkaline comet assay, they found a significant DNA damaging potential of wastewater on human leukocytes, but no effect in the Ames test. Aleem and Malik (20) investigated untreated wastewater from industrial and household sources that is used to irrigate agricultural crops in Aligarh, India. Extracts from irrigated soils were tested with different bacterial test systems and found to be clearly genotoxic. The same was found for wastewater from the Indian

city of Ghaziabad (21). Significant genotoxicity was also found with four different genotoxicity tests of untreated wastewater from different industries. It was also shown that genotoxicity could partially or completely be eliminated by wastewater treatment (22). Another example is a study by Swaileh et al. (23), who found genotoxicity in oat plants irrigated with wastewater. Of particular reference to our study is the study by Kholtei, who demonstrated a significant increase in DNA breaks in cells exposed to wastewater from the town of Settat (2). Other investigations have confirmed genotoxic effects in humans (18) and animals (24) who drank the Boumoussa water and showed elevated micronucleus frequencies in their white blood cells. So far, no follow up is available on the health status of the investigated animals and humans, but these studies clearly suggest that their health may be impaired in the long run.

Our and the above mentioned investigations made no attempt to determine whether elevated micronucleus frequencies were related to clastogenic and/or aneugenic effects of one or more pollutants. We also did not attempt to identify the compounds that were actually contributing to the observed genetic damage. Some of the heavy metals found in concentrations above limits, are the likely culprits such as lead, chrome, aluminium, and cadmium, as they may in certain forms be mutagenic. In general, metal genotoxicity is caused by indirect mechanisms, of which three are predominant (25): a) interference with cellular redox regulation and induction of oxidative stress, which may cause oxidative DNA damage [also see (26)]; b) inhibition of major DNA repair systems, resulting in genomic instability and accumulation of critical mutations [also see (26)]; and

**Table 3** Heavy metals in wastewater (B, C), well water (9, 11, 21, 46) or tap water samples

Stations	Concentration / $\text{mg L}^{-1}$					
	Pb	Cr	Cd	Al	Zn	Hg
B	4.7921	0.2622	0.0030	22.6815	5.8048	0.0336
C	4.5356	0.2206	0.0025	22.2053	5.7504	0.0136
21	0.0156	0.0014	0.0009	0.0589	0.0429	0.0005
46	0.0130	0.0016	0.0008	0.0458	0.0292	0.0001
11	0.0036	0.0011	0.0001	0.0434	0.0129	0.0000
9	0.0009	0.0004	0.0001	0.0120	0.0012	0.0000
Tap water	0.0030	0.0000	0.0000	0.0040	0.0021	0.0000
Morrocian standard (2002)	0.0500	0.0500	0.0050	0.2000	5.0000	1.0000
WHO standard (1996)	0.0100	0.0500	0.0030	0.0500	3.0000	1.0000

**Table 4** Results of statistical analyses for micronucleus frequencies and proliferation index determined in human peripheral blood lymphocytes treated with wastewater (B, C), well water (9, 11, 21, 46) or tap water in vitro

	Micronuclei		Proliferation Index (PI)	
	F*	Significance / p	F*	Significance / p
Station B/C	811.00	<0.001	57.14	<0.05
Station B/ well 9	48.58	<0.001	51.92	<0.05
Station B/ Well 11	78.39	<0.05	188.11	<0.001
Station B/ Well 21	54.86	<0.05	63.36	<0.05
Station B/ Well 46	135.00	<0.05	31.16	<0.05
Station B/ Drinking Water	48.58	<0.001	164.20	<0.001
Station C/ Well 9	26.68	<0.0001	7.33	NS
Station C/ Well 11	39.38	<0.0001	6.961	NS
Station C/ Well 21	17.42	<0.05	5.46	NS
Station C/ Well 46	38.44	<0.05	0.11	NS
Station C/ Drinking Water	26.68	<0.001	14.91	<0.05
Well 9/ Well 11	7.00	NS	7.10	NS
Well 9/ Well 21	37.73	<0.05	42.41	<0.05
Well 9/ Well 46	21.04	<0.05	59.26	<0.05
Well 11/ Well 21	229.36	<0.0001	75.96	<0.001
Well 11/ Well 46	33.00	<0.05	38.85	<0.05
Well 11/ Drinking Water	7.00	NS	13.97	<0.05
Well 21/ Well 46	1.00	NS	3.83	NS
Well 21/ Drinking Water	37.73	<0.001	212.36	<0.0001
Well 46/ Drinking Water	21.04	<0.05	278.40	<0.0001

\*Compared with theoretical F of 10.1279645

NS=statistically not significant

c) deregulation of cell proliferation by induction of signalling pathways or inactivation of growth controls such as tumour suppression genes. In addition, specific metal compounds exhibit unique mechanisms such as interruption of cell-cell adhesion by cadmium, direct DNA binding of trivalent chromium produced from hexavalent chromium [also see (27)], or perturbation of the mitotic spindle or inhibition of its formation. The latter holds true for Cr<sup>+6</sup> products (Na<sub>2</sub>CrO<sub>4</sub> and CaCrO<sub>4</sub>) and aluminium chloride (28, 29).

It is important to investigate deeper into these mechanisms in the future. In the meantime, our and other studies demonstrate the importance of environmental monitoring and the need for water treatment stations in the afflicted areas.

#### Acknowledgement

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**Sažetak****IN VITRO ISTRAŽIVANJE GENOTOKSIČNOG POTENCIJALA OTPADNIH VODA GRADA SETTATA (MAROKO)**

Nagli industrijski razvoj u gradu Settatu posljednjih je godina rezultirao povećanim onečišćenjem okoliša, osobito putem otpadnih voda iz kućanstava i industrije. Te se otpadne vode bez ikakve prethodne obrade odlažu u dolinu rijeke Boumoussa na području zajednice Mzamza, 8 km sjeverno od grada. Uzastopne sušne sezone lokalnoj su zajednici nametnule potrebu za uporabom tih voda za navodnjavanje, čime su i zajednica i okoliš postali izloženi povećanim rizicima od onečišćenja tla i izvora podzemnih voda. Svrha ovog istraživanja bila je provesti fizikalno-kemijsku karakterizaciju i utvrđivanje sadržaja metala u otpadnim vodama grada Settata te procijeniti njihove učinke na gornji sloj podzemnih voda. Ujedno je istraživana genotoksični potencijal otpadnih voda u uvjetima *in vitro* na limfocitima periferne krvi čovjeka primjenom mikronukleusnog testa i proliferacijskog indeksa. Istraživanje je potvrdilo visok stupanj onečišćenja vodâ u dolini Boumoussa, kao i vodâ iz bunara smještenih u neposrednoj blizini. Štetni su učinci onečišćenja potvrđeni i na osnovi povišene učestalosti stanica s mikronukleusima te snižene vrijednosti proliferacijskog indeksa u limfocitima periferne krvi u uvjetima *in vitro*.

**KLJUČNE RIJEČI:** *Boumoussa, fizikalno-kemijska karakterizacija, mikronukleus, proliferacijski indeks, teški metali*

**CORRESPONDING AUTHOR:**

Professor Abderraouf Hilali  
University Hassan I, Faculty of Sciences and Technics  
Settat, Km 3, Route de Casablanca  
B.P. 577, 26000 Settat, Morocco  
E-mail: [Hilalia@hotmail.com](mailto:Hilalia@hotmail.com)