



Oxidative Stress and Antioxidants: Biological Response Modifiers of Oxidative Homeostasis in Cancer

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Abstract

Oxidative stress is defined as a misbalance in cell redox reactions resulting in the increase of reactive oxygen species (ROS) and/or decreased antioxidant defence. Crucial part of oxidative stress, which avoids protective effects of antioxidants, is lipid peroxidation (LPO). Since LPO comprises several chain reactions allowing the spread of ROS-caused damage to the macromolecules (proteins, nucleic acids and lipids) it is important in pathogenesis of various diseases such as cardiovascular diseases, neurodegenerative diseases, diabetes mellitus and cancer. On the other hand, research on physiology and pathology of lipid peroxidation revealed that not only ROS but also the LPO products such as 4-hydroxynonenal (HNE) are involved in physiological homeostasis of various tissues. This lead to conclusion that LPO and oxidative stress and not only pathological but also physiological processes. Accordingly, HNE was revealed as biomarker, growth regulating factor and signalling molecule.

In this light, both natural as well as synthetic antioxidants could be considered as »biological response modifiers« maintaining oxidative homeostasis. Accordingly, some antioxidants might eventually become important components of advanced individual and integrative biomedicine.

OXIDATIVE STRESS AND REACTIVE OXYGEN SPECIES

Oxygen is essential for aerobic organisms, but it can also be harmful because of formation of reactive oxygen species (ROS), and thereby oxidative stress. Oxidative stress is defined as a misbalance in cell redox reactions (1) which can be the result of either ROS overproduction or decreased antioxidant defence. As ROS are produced in cells under physiologic conditions, they induced evolution of different antioxidative mechanisms of cellular defence against oxidative stress (Figure 1).

ROS and Lipid Peroxidation

Reactive oxygen species include radical species, such as superoxide anion ($\bullet\text{O}_2^-$), hydroxyl radical ($\bullet\text{OH}$), and also non-radical species, such as hydrogen peroxide (H_2O_2), singlet oxygen ($^1\text{O}_2$), hypochloric acid (HOCl) and ozone (O_3). Radicals are produced either in controlled or uncontrolled manner. One of the main sources of ROS is leakage of electrons from the respiratory chain (2). Complex I and III of the respiratory chain are the source of superoxide anion (3). Similar reactions

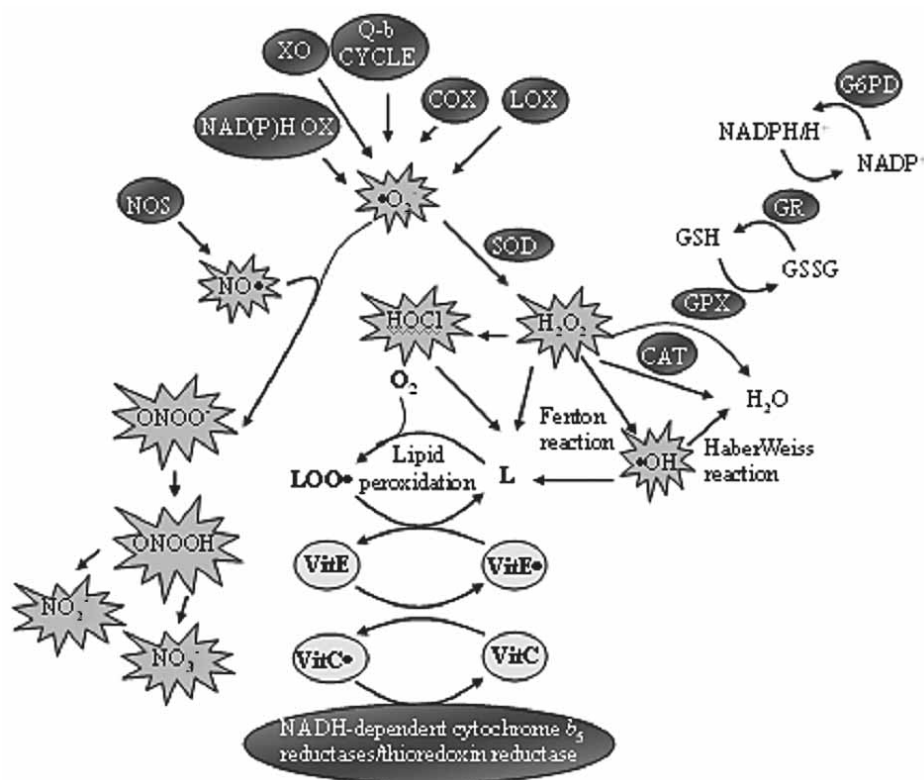


Figure 1. Redox cycle of the cell and antioxidant defence from radical attack. Superoxide radical (O_2^-) is produced by NAD(P)H oxidase (NAD(P)H-OX), cyclooxygenase (COX), lipoxygenase (LOX), xanthine oxidase (XO), and by mitochondrial ubiquinone-cytochrome b (Q-b) cycle. Other radicals, such as ONOO $^-$, are formed in the cell and are derived from other reactive oxygen or nitrogen species like nitric oxide (NO) and superoxide radical. NO is formed by NO synthase (NOS). O_2^- is then transformed by superoxide dismutase (SOD) to hydrogen peroxide (H_2O_2), which can be further neutralized to water by catalase (CAT), glutathione peroxidase (GPX) or can undergo Fenton reaction. Oxidized glutathione (GSSG) from GPX reaction is regenerated by glutathione reductase (GR), which cycles with glucose-6-phosphate dehydrogenase (G6PD). Reactive oxygen species cause lipid peroxidation which is stopped in membrane by vitamin E, consequently regenerated through vitamin C cycle.

are generated by cytochrome P450. Group of cytochrome P450 enzymes (P450 or CYP) contain haeme in their active site, and are located in endoplasmic reticulum of hepatocytes (4). Cytochrome P450 is known as one of the major enzymes involved in the metabolism of various drugs, and thereby metabolising and activating carcinogens (5). Cytoplasm is potential source of radicals due to high content of iron and other transition metals.

Iron participates in several biological reactions and is essential for living organisms. Iron-containing proteins of the respiratory chain are involved in electron transport to provide the energy for cellular functional activities (6). Iron is also required for cell growth and multiplication. However, the complex mechanism of the influence of iron on cell proliferation is not entirely understood. It is known that iron plays an important role in the activity of ribonucleotide reductase, a key-enzyme in DNA synthesis responsible for the reduction of ribonucleotides to deoxyribonucleotides (6). This enzyme turns over rapidly and needs a continuous supply of iron to maintain its activity. On the other hand, metal ions, in particular those of iron, are required for formation of $\bullet OH$ which might mediate DNA damage induced by H_2O_2 and $\bullet O_2^-$. Unlike these uncontrolled radical production, rad-

icals are produced in »oxidative burst« of leukocytes (7), iodide oxidation in thyroid gland (8), and in prostaglandin synthesis from arachidonic acid (9).

Oxidative stress affects all cell macromolecules, including DNA, causing mutations, proteins, causing inactivation, and lipids causing lipid peroxidation. Polyunsaturated fatty acids are especially sensitive to oxidative degradation caused by free radical reactions in cells (11). This process results in the production of highly reactive aldehydes which are proposed to be »second toxic messengers« for the primary free radicals which initiated lipid peroxidation (11). The highly reactive aldehyde 4-hydroxynonenal (HNE) appears to be one of the major cytotoxic products of lipid peroxidation that could be found in various tissues even under normal, physiological conditions (12). Unlike free radicals or the other ROS, HNE has the unique feature to remain stable, not metabolised, by binding to macromolecules (such as proteins). Such HNE-macromolecular conjugates might be even required for the biological activities of HNE and could be detected by the use of monoclonal antibodies (13). Furthermore, some of the results suggest that the modification of cell growth *in vitro* in presence of physiological concentrations of HNE (1 μM) could further de-

pend on the presence of certain serum factors, but not serum albumin which is known to couple with HNE, thus decreasing the cytotoxic effects of the aldehyde (14). Today, HNE is known to be important in signalling pathways for cell cycle arrest, differentiation, regulation of gene expression (p53 and c-fos especially), and apoptosis (10, 15). Consequentially, HNE acts as bifunctional (stimulating as well as inhibiting) regulator of the c-fos gene transcription thereby having an essential role in «turning inflammation against cancer». In agreement with this are the findings of Poli and co-workers who showed a decrease of HNE and TGF-beta in patients with colon cancer, while Chron's disease (an aggressive inflammatory disease of the intestine) was associated with an increase of HNE and TGF-beta production (16). These unique regulatory features explain presence of HNE in many diseases, such as cancer, neurodegenerative diseases, and autoimmune diseases (17). Taken together, these findings are of general importance for the understanding of the mechanisms of the biological effects of not only HNE but of ROS and oxidative stress in general.

Antioxidant Mechanisms

Due to deleterious effects of oxidative stress and lipid peroxidation, cells developed different mechanisms to cope with the challenge. Generally, mechanisms of antioxidative defence are grouped in two major groups: non-enzymatic and enzymatic systems.

Enzymatic mechanisms of ROS detoxification are intensively studied. These enzyme systems can be divided into two groups. One group reacts directly with ROS, while other act as redox regulators resulting in redox balance (18). In addition, there are separate enzymatic systems with identical activity regarding different cellular compartments. Duplication of enzymatic systems ensures optimal protection against deleterious effects of oxidative stress, as well as different regulation and site specific antioxidant mechanisms. Non-enzymatic antioxidative systems are not as specific as enzymatic, but nevertheless, they are the first line of antioxidative defense, and therefore are certainly not negligible.

Superoxide dismutases (SOD) are the first line of defense against superoxide anions. SOD catalyses disproportion of superoxide anion to hydrogen peroxide and molecular oxygen. SOD has two isoforms, cytoplasmic, Zn/Cu SOD, and mitochondrial, MnSOD. The essential role of SOD is seen in diseases with disturbed SOD activity. For example, lowered activities of CuZnSOD are often, but not always, seen in tumors, suggesting that decreased antioxidant protection accompanied by increased ROS production could be not only essential steps in carcinogenesis, but many of the properties of the cancer cells in general (19).

Hydrogen peroxide, generated by SOD, still represents a danger for the cell, and is therefore, substrate for another enzyme catalase. Catalase decomposes hydrogen peroxide in two ways: one acting as catalyst with water and oxygen as end products, and other acting as

peroxidase (20). Catalase is one of the most efficient enzyme, with reaction rate limited by the collision rate of enzyme with substrate (20). This enzyme is present in all prokaryotes and eukaryotes, and is predominantly located in peroxisomes in mammalian cells. Overexpression of catalase renders cells and organisms (transgenic mice) less sensitive to oxidative challenge (21). Interestingly, catalase deficiency yielded normal phenotype, which was described by Dr. Takahara in 1946. He described that acatalasemic patients had increased tendency in developing oral ganagrene, presumably as a result of tissue damage by H₂O₂ produced by bacteria (22). Also, there are reports that PEGylated catalase suppressed the growth of metastatic tumor cells after tumor removal by decreasing cytokines and their receptors such as EGF and EGFR. These findings indicate modulation of signal transduction by interfering with H₂O₂, which is thought to trigger the proliferation of dormant tumor cells (23).

Peroxidases are group of enzymes involved in reduction of inorganic and organic peroxides. Unlike SOD and catalases, these enzymes are not dependent on metal ions in their active site, but have cysteine instead. Therefore, peroxidases require electron donor from thiol groups. Taking this into consideration, two groups of peroxidases are distinguished: glutathione peroxidases (GSH as electron donor) and thioredoxin peroxidases (TRX as electron donor) (18). GSH peroxidase uses GSH as electron donor resulting in oxidised GSH (GSSG), which is then regenerated by glutathione reductase. Thioredoxin peroxidase reduces hydroperoxides by using thioredoxin as electron donor, which is regenerated by thioredoxin reductase (uses NADPH) (24).

Non enzymatic antioxidants are small hydro- and liposoluble molecules which protect against free radicals in aqueous as well as lipid cell compartments. These small molecules act as unspecific radical scavengers reacting as oxidants to ROS and thereby detoxifying them. Glutathione (GSH) is one of the most ubiquitous small molecules of antioxidant defence. Glutathione, γ -L-glutamyl-L-cysteinylglycine, reacts with its cysteine residue with oxidants thereby forming oxidised glutathione (GSSG) (25). Other well-known small antioxidant molecules are vitamins, vitamin C, L-ascorbic acid, which protect the aqueous cell compartments, and vitamin E, α -tocopherol, which protect lipid compartments. Vitamin C, ascorbate, quenches radicals and forms ascorbyl radical, a stable radical which causes little oxidative damage (26). Ascorbate regenerates through redox cycling, and this fact was used to selectively induce tumor cell death. Namely, menadione and ascorbate created a redox cycle which generated oxidative stress in tumor cells (27). This approach is supported by following facts: cancer cells lack antioxidant enzymes, and thereby are more sensitive to oxidative stress; over-expression of GLUT transporters in cancer cells causes increased uptake of vitamin C, cancer cells show universal glycolytic phenotype, which is required for survival and invasion of surrounding tissue (27). Eventually, oxidative stress caused by redox cycling of ascorbate and menadione results in

impairment of glycolysis and finally to cell death (28). Vitamin E is a generic name given to a group of tocopherol and tocotrienols, of which α -tocopherol is the most abundant in human tissue and is of highest biological activity (29). Vitamin E protects lipid compartments of cell by terminating the lipid peroxidation chain reaction or by inactivation of ROS (30). Also, there are reports of pro-oxidant properties depending on cell environment. Vitamin E was shown to be involved in signal transduction by modulating specific enzymes such as protein kinase C (PKC), protein phosphatase 2A (PP2A), protein tyrosine phosphatase (PTP), protein tyrosine kinase (PTK), diacylglycerol kinase (DAGK), 5-, 12- and 15-lipoxygenases (5-, 12-, and 15-LOX), phospholipase A2 (PLA2), cyclooxygenase-2 (COX-2), and the mitogen activated protein kinase (MAPK) signal transduction pathway (31), and also transcription factors like NF κ B. Modulation of these proteins occurs via direct binding or by the interference with enzyme activation and enzyme redox regulation (31). Finally, this alters cellular functions, such as apoptosis, necrosis, survival, adhesion, differentiation.

Antioxidants or agents metabolised to become antioxidants may function by variety of mechanisms: 1) quenching the formation of singlet oxygen, e.g. β -carotene, retinol; 2) scavenging ROS, e.g. polyphenolics; 3) scavenging or reducing lipid free radicals, e.g. α -tocopherol; 4) scavenging prooxidant metals, e.g. polyphenolics, flavonoids; 5) oxidizing ferrous iron, e.g. caeruloplasmin, apoferritin; 6) inhibiting prooxidant enzymes, e.g. allopurinol; 7) inducing or enhancing protective enzymatic defense against oxygen or oxydants, e.g. butylated hydroxyanisole; 8) sparing or renewing intracellular antioxidants, e.g. ascorbate, N-acetylcysteine; 9) stabilising membranes against lipid peroxidation, e.g. cholesterol, 17-beta-estradiol, tamoxifen; 10) reducing oxidatively stressed cells, e.g. ethanol, sorbitol, xylitol (NADH generators); 11) inhibiting enzymes that mediate gene expression as a result of oxidative stress, e.g. tamoxifen, methoxybenzamide. As mentioned above, antioxidants can also act as modulators of the cell proliferation and differentiation (32). Of special interest is their induction of phase II enzymes in cancer development (34): 1) glutathione transferases which conjugate mostly hydrophobic electrophiles with GSH; 2) NAD(P)H: quinone reductase (DT-diaphorase) which promotes two-electron reduction of quinones to hydroquinones; 3) UDP glucuronyltransferases which conjugate xenobiotics with glucuronic acid enhancing their extraction; 4) epoxide hydrolase which inactivates epoxides by hydration to diols. The most common plant antioxidant that is capable of inducing GSH-S-transferase both in vitro and in vivo (in particular in the liver) is ellagic acid which is present in strawberries, raspberries and grapes (34), and could attenuate harmful effects of tobacco nitrosamines (35). That might be particularly relevant if considered in the light of recent findings on the high relevance of GSH-S-transferases in cancer prevention and in general on the role of GSH in human pathology based on oxidative stress (36).

The efforts to use natural pure antioxidants were assumed to offer a good option for the use of antioxidants in cancer prevention. Unfortunately, these ended in well known ATBC study (Alpha-Tocopherol Beta-Carotene) in 90' which showed that the use of beta carotene with the aim to decrease the incidence of lung cancer in smokers resulted in opposite findings of increased incidence of cancer in subjects using beta carotene (37). Nowadays, it is clear that the use of beta carotene in the ATBC study resulted in the overload of beta carotene, which would on one hand cause misbalance of overall antioxidant mechanisms, while on the other beta carotene is decomposed under ROS attack (as in case of cigarette smoke) into mutagenic and potentially carcinogenic products (38).

Oxidative Stress, Antioxidants and Cancer

Various factors related to oxidative stress and antioxidants could play important role in development as well as in cancer therapies. Some of them are summarised in Table 1.

While many of such factors could have dual roles acting both as beneficial and as harmful factors, synthetic antioxidants that could act as desirable antioxidant and anticancer agents are almost entirely neglected. Synthetic antioxidants were in a way »collateral victims« of the studies that have shown that pure natural antioxidants could have undesirable side effects, mostly because under oxidative stress antioxidants could be metabolised in novel, cytotoxic free radicals, thus allowing the chain reactions and the spread of oxidative stress. Therefore, the efforts were raised to synthesize antioxidants that could under ROS attack from stable radicals ending oxidative stress. This feature gives a new insight in antioxidant function and therefore they could be considered as »biological response modifiers« maintaining oxidative homeostasis.

The example of oxidative stress/homeostasis in normal physiology is exercise. During exercise ROS and RNS (Reactive Nitrogen Species) are generated in muscle (39, 40). As oxidative stress was considered to be harmful, the efforts were made to investigate the role of oxidative stress during exercise, especially during heavy exercise (for professional athletes). The numerous studies reported that antioxidant supplementation during exercise decrease oxidative stress parameters (41, 42), but this did not turn out to be beneficial. In fact, it was shown that oxidative stress generated by exercise cause hormetic response leading to adaptation to oxidative stress and increasing organism's tolerance to stress (43). These discoveries pointed out the necessity to maintain the natural oxidative homeostasis of the organism and the importance to help the organism to keep this homeostasis in illness, especially in tumor therapy. Tumor therapy is often based on oxidative stress and also, it is commonly accepted that there is persistent oxidative stress in cancer (44). There are two general theories on the origin of cancer, and there are also two general theories on the carci-

TABLE 1

Some factors related to oxidative stress affecting cancer development.

FACTOR	TARGET ORGANS (ORGANIC SYSTEMS)	ACTIVITY PRINCIPLE FOR:	
		Harmful effects (<i>carcinogenesis</i>)	Beneficial effects (cancer prevention or therapy)
Polluting Smoke (cigarette, car & industry pollution, etc.)	respiratory system, genito-urinary system	complete chemical carcinogens and mechanical irritation by tar particles, hypoxia, immune suppression	none
Minerals (asbestosis, pro-oxidant pollutants)	respiratory system, gastrointestinal system	oxidative activity of iron, silicon oxide combined with mechanical irritation	possible free radicals scavenging activity of some minerals or miscellaneous anticancer effects due to the uncertain mechanisms
Ionising radiation (UV, X-rays)	skin, bone marrow, genito-urinary system	direct and indirect DNA damage	cancer radiotherapy due to the toxicity (DNA damage particularly of the proliferating cells)
Toxic chemicals (dyes, etc.)	skin, gastrointestinal system, respiratory system, genito-urinary system	multiple, involving pro-oxidants and complete carcinogens	possible increase of the ROS detoxifying systems – uncertain
Diet (food restriction or abuse)	gastrointestinal system, genito-urinary system	mostly related to some plant tumor promoting components, mucose irritation and metabolic imbalances (fat, meat or glucose abuse)	plant antioxidants – possible way for the natural chemoprevention and therapy
Medicaments (drugs, complementary treatments, etc.)	gastrointestinal system, genito-urinary system, bone marrow	mostly affecting liver cytochrome P450, chemotherapeutic agents (cytostatic drugs)	chemotherapy of cancer due to the DNA damage, phototherapy of cancer, possible activity principle of some plant extracts
Hereditary factors (xeroderma pigmentosum, Down's syndrome)	skin, bone marrow	lower DNA repair system after mutagenic effects of ROS, lowered antioxidative capacity of the cells	uncertain
Inflammation* (chronic)	unspecific	cellular production of ROS combined with tissue damage	suppression of the tumor progression due to the inhibiting effects of suppressive cytokines (TGF-beta, TNF-alpha), unspecific immune response against cancer, changing tumor stroma – fibrosis

* uncertain and not completely understood, but should be related to the mechanical irritation or response to particular tissue damage (such as inflammatory response to the tissue damage caused of radiation, asbestosis, smoke abuse, etc.)

nogenic effects of oxidative stress and anticancer activities of antioxidants (45).

The genetic theory of cancer implies particular oncogene mutations as the cause of cancer and the epigenetic theory assumes that the structure of the active genes is normal, while the regulation of cell growth is altered in direction of less matured aggressive cells due to the activation of genes which should be quiescent and/or suppression of the tumor-suppressing genes. In case of somatic mutations, carcinogens are often metabolically activated by cytochrome P-450 (designated as phase 1 enzyme) causing oxidative damage of the DNA which is not lethal for the cells, but is irreparable and inherited by the progeny of the altered cell (mutation) (46). Antioxidants should in this situation act as scavengers of ROS that could cause the damage of the genome. In case of epigenetic bases of cancer antioxidants should not only act as scavengers of ROS, but could, similar to oxidants and ROS, themselves act as modulators of the cell proliferation and differentiation (47, 48, 49, 50, 51).

In both genetic and epigenetic bases of cancer development antioxidants should primarily act in the initiation stage of carcinogenesis influencing the phase 1 enzymes. There are also phase 2 enzymes that could be induced by antioxidants (43). They include: 1) glutathione transferases which conjugate mostly hydrophobic electrophiles with GSH; 2) NAD(P)H: quinone reductase (DT-diaphorase) which promotes two-electron reduction of quinones to hydroquinones; 3) UDP glucuronyltransferases which conjugate xenobiotics with glucuronic acid enhancing their extraction; 4) epoxide hydrolase which inactivates epoxides by hydration to diols.

Although there are many efforts to find as much as possible novel plant antioxidants, the problem of their instability during isolation or in physiologic conditions (52) are the major obstacle. Therefore, efforts are made to synthesize novel antioxidants, stable and with enhanced antioxidant activity. Some of the other character-

istics of these novel antioxidants are: antitumor activity – curcumin derivatives (52), antidiabetic and cardioprotective effects – pyridoindole derivative (53), calcium channel antagonists – diludin derivatives (54, 55). Combination of antioxidants and other complementary treatment supporting conventional therapies could thus raise powerful tool in treatment of different diseases with disrupted oxidative homeostasis.

Conclusions

Oxidative stress was for long considered as cause of different illnesses, but new findings indicate its importance in cell homeostasis. One of the most popular parts of redox biology and biochemistry in common life are antioxidants. Although antioxidants are beneficial they should be handled with care under physiological conditions and homeostasis. Oxidative homeostasis is disrupted in disease, especially in cancer. Chemo/radiotherapy is also based on oxidative stress, therefore, there is a need for novel antioxidants which would have antioxidant as well as tumor suppressing activity. There is rising number of antioxidants that are promising and their properties need to be thoroughly studied together with their mechanisms. These substances and their activity are of great importance in treating cancer and improving life quality.

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REFERENCES

- SIES H 1985 Oxidative stress. Academic Press, Orlando.
- TURRENS J F, ALEXANDRE A, LEHNINGER A L 1985 Ubisemiquinone is the electron donor for superoxide formation by complex III of Heart mitochondria. *Arch Biochem Biophys* 237: 408–414
- RAHA S, ROBINSON BH 2000 Mitochondria, oxygen free radicals, disease and ageing. *Trends Biochem Sci* 25: 502–508
- GUENGERICH F P 1992 Characterization of human cytochrome P450 enzyme. *FASEB J* 6: 745–748
- SOUTHORN P A, POWIS G 1988 Free radicals In: Medicine II: Involvement in human disease. *Mayo Clin Proc* 63: 390–407
- WILLIAMS R J P 1974 Haem-proteins and oxygen. In: Jacobs A, Worwood M (eds), Iron in Biochemistry and Medicine. Academic Press, London, p 183–198
- ŽIVKOVIĆ M, POLJAK-BLAŽI M, ŽARKOVIĆ K, MIHALJEVIĆ D, SCHAUR RJ, ŽARKOVIĆ N 2007 Oxidative burst of neutrophils against melanoma B16-F10. *Cancer Lett* 246: 100–108
- SONG Y, DRIESSENS N, COSTA M, DE DEKEN X, DETOURS V, CORVILAIN B 2007 Roles of hydrogen peroxide in thyroid physiology and disease. *J Clin Endocrinol Metab* 92: 3764–3773
- KOZAK K R, MARNETT L J 2002 Oxidative metabolism of endocannabinoids, *Prostaglandins Leukot Essent Fatty Acids* 66: 211–220
- ESTERBAUER H, SCHAUR R J, ZOLLNER H 1991 Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med* 11: 81–128
- ESTERBAUER H, ZOLLNER H, SCHAUR R J 1990 Aldehydes formed by lipid peroxidation: mechanisms of formation, occurrence, and determination. In: Vigo-Pelfrey C. (ed.), Membrane Lipid Oxidation. CRC Press, Boca Raton, p 239–268
- SCHAUR R J, ZOLLNER H, ESTERBAUER H 1991 Biological effects of aldehydes with particular attention to 4-hydroxynonenal and malonaldehyde. In: Vigo-Pelfrey C (ed), Membrane lipid oxidation. CRC Press, Boca Raton, Florida, p 141–163
- ŽARKOVIĆ N, ŽARKOVIĆ K, SCHAUR RJ, ŠTOLC S, SCHLAG G, REDL H, WAEG G, LONČARIĆ I, JURIĆ G, HLAVKA V 1998 4-Hydroxynonenal as a second messenger of free radicals and growth modifying factor. *Life Sci* 65: 1901–1904
- KREUZER T, ŽARKOVIĆ N, GRUBE R, SCHAUR RJ 1997 Inhibition of HeLa cell proliferation by 4-Hydroxynonenal is associated with enhanced expression of the c-fos oncogene. *Cancer Biotechnol Biochem* 12: 131–136
- UCHIDA K 2003 4-Hydroxy-2-nonenal: a product and mediator of oxidative stress. *Prog Lipid Res* 42: 318–343
- CHIARPOTTO E, SCAVAZZA A, LEONARDUZZI G, CAMAN-DOLA S, BIASI F, TEGGIA P M, GARAVOGLIA M, ROBECCHI A, RONCARI A, POLI G 1997 Oxidative damage and transforming growth factor beta-1 expression in pretumoral and tumoral lesions of human intestine. *Free Radic Biol Med* 22: 889–894
- VALKO M, LEIBFRITZ D, MONCOL J, CRONIN MTD MAZUR M 2007 Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 39: 44–84
- HERRERO E, ROS J, BELLÍ G, CABISCOL E 2008 Redox control and oxidative stress in yeast cells. *BBA General subjects* 1780: 1217–1235
- OBERLEY L W, OBERLEY T D 1984 The role of superoxide dismutase and gene amplification in carcinogenesis. *J Theor Biol* 106: 403–422
- MICHELIS C, RAES M, TOUSSAINT O, REMACLE J 1994 Importance of SE-glutathione peroxidase, catalase, and CU/ZN-SOD for cell survival against oxidative stress. *Free Radic Biol Med* 17: 235–248
- ERZURUM S C, LEMARCHAND P, ROSENFELD M A, YOO J H, CRYSTAL R G 1993 Protection of human endothelial cells from oxidant injury by adenovirus-mediated transfer of the human catalase cDNA. *Nucleic Acids Res* 21: 1607–1612
- SATO K, ITO K, KOHARA H, YAMAGUCHI Y, ADACHI K, ENDO H 1992 Negative regulation of catalase gene expression in hepatoma cells. *Mol Cell Biol* 12: 2525–2533
- HYOUDOU K, NISHIKAWA M, KOBAYASHI Y, UMEYAMA Y, YAMASHITA F, HASHIDA M 2006 PEGylated catalase prevents metastatic tumor growth aggravated by tumor removal. *Free Radic Biol Med* 41: 1449–1458
- KOWALTOWSKI A J, DE SOUZA-PINTO N C, CASTILHO R F, VERCESI A E 2009 Mitochondria and Reactive Oxygen Species. *Free Radic Biol Med* (in press)
- JELLINGER K A 2000 Cell death mechanisms in Parkinson's disease. *J Neural Transm* 107: 1–29
- DAVIES M B, AUSTIN J, PARTRIDGE D A 1991 Vitamin C: its chemistry and biochemistry, royal society of chemistry.
- BENITES J, ROJO L, VALDERRAMA J A, TAPER H, CALDERO P B 2008 Part 1: Effect of vitamin C on the biological activity of two euryfurylbenzoquinones on TLT, a murine hepatoma cell line. *Eur J Med Chem* 43: 1813–1817
- VERRAX J, STOCKIS J, TISON A, TAPER HS, BUC CALDERON P 2006 Oxidative stress by ascorbate/menadiene association kills K562 human chronic myelogenous leukaemia cells and inhibits its tumour growth in nude mice. *Biochem. Pharm* 72: 671–680
- WOLF R, WOLF D, RUOCCO V 1998 Vitamin E: the radical protector. *J Eur Acad Dermatol Venereol* 10: 103–117
- BEZERRA F S, VALENÇA S S, LANZETTI M, PIMENTA W A, CASTRO P, KOATZ V L K, PORTO L C 2006 α -Tocopherol and ascorbic acid supplementation reduced acute lung inflammatory response by cigarette smoke in mouse. *Nutrition* 22: 1192–1201
- ZINGG J M 2007 Modulation of signal transduction by vitamin E. *Mol Asp Med* 28: 481–506
- RIMBACH G, MINIHANE A M, MAJEWICZ J, FISCHER A, PALLAUF J, VIRGLI F, WEINBERG P D 2002 Regulation of cell signalling by vitamin E. *Proc Natl Acad Sci* 99: 415–425
- BARRERA G, DI MAURO C, MURACA R, FERRERO D, CAVALLI G, FAZIO V, PARADISI L, DIANZANI M U 1991 Induction of differentiation of human HL-60 cells by hydroxynonenal, a product of lipid peroxidation. *Exp Cell Res* 297: 148–152
- O'BRIEN P J 1994 Antioxidants and cancer: Molecular mechanisms. In: Armstrong D. Ed.; Free radicals in diagnostic medicine. Plenum press, New York, p 215–239
- ZHANG D X, OKADA S, YU Y Y, ZHENG P D, YAMAGUCHI R, KASAI H, ZHANG Z, HAMILTON S M, STEWART C, STROTHERS A, TEEL R W 1993 Inhibition of microsomal cyto-

- chrome P450 activity and metabolism of the tobacco specific nitrosamine NNK by allagic acid. *Anticancer Res* 13: 2341–2346
36. SIEMS W G, KRÄMER K, GRUNE T 1996 Störungen im Glutathionsystem und klinische Konsequenzen. *Pharmazeutische Zeitung* 14: 11–22
 37. MARANTZ P R, KRITCHEVSKY D, GOLDSTEIN M R, PRYOR W A, LEO M A, LIEBER C S, BALLMER P E, STAHELIN H B, HEINONEN O P, HUTTUNEN J K, ALBANES D, TAYLOR P R 1994 The Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group, Hennekens C. H., Buring J. E., Peto R.; Beta Carotene, Vitamin E, and Lung Cancer. *N Engl J Med* 331: 611–614
 38. ALIJA A J, BRESGEN N, SOMMERBURG O, LANGHANS C D, SIEMS W, ECKL P M 2005 Cyto- and genotoxic potential of beta-carotene and cleavage products under oxidative stress. *Biofactors* 24: 159–63
 39. CHILD R, BROWN S, DAY S, DONNELLY H, ROPER H, SAXTON J 1999 Changes in indices of antioxidant status, lipid peroxidation and inflammation in human skeletal muscle after eccentric muscle actions. *Clin Sci* 96: 105–115
 40. MAIORANA A, O'DRISCOLL G, TAYLOR R, GREEN D 2003. Exercise and the nitric oxide vasodilator system. *Sports Med* 33: 1013–1035
 41. BLOOMER R J 2008 Effect of exercise on oxidative stress biomarkers. *Adv Clin Chem* 46: 1–50.
 42. URSO M J, CLARKSON P M 203 Oxidative stress, exercise, and antioxidant supplementation. *Toxicol* 189: 41–54
 43. JI L L, RADAK Z, GOTO S 2008 Hormesis and Exercise: How the Cell Copes with Oxidative Stress. *Am J Pharm Toxic* 3: 41–55
 44. JOHNSON D, TRAVIS J 1979 The oxidative inactivation of human alpha-1-proteinase inhibitor. Further evidence for methionine at the reactive center. *Biol Chem* 254: 4022–4026
 45. O'BRIEN P J 1994 Antioxidants and cancer: Molecular mechanisms. In: Armstrong D (ed.), Free radicals in diagnostic medicine. Plenum press, New York, p 215–239
 46. MILLER E C, MILLER J A, BROWN R R, MACDONALD J C 1958 On protective action of certain polycyclic aromatic hydrocarbons against carcinogenesis by aminoazo dyes and acetylaminofluorene. *Cancer Res* 18: 469–473
 47. ŽARKOVIĆ N, SCHAUR R J, PUHL H, JURIN M, ESTERBAUER H 1994 Mutual dependence of growth modifying effects of 4-hydroxy-nonenal and fetal calf serum *in vitro*. *Free Radic Biol Med* 16: 877–884
 48. ŽARKOVIĆ K, ŽARKOVIĆ N, SCHLAG G, REDL H, WAEG G 1997 Histological aspects of sepsis-induced brain changes in a baboon model. In: Schlag G, Redl H, Traber D L (eds), Shock, Sepsis and Organ Failure, 5th Wiggers Bernard Conference (Springer-Verlag, Heidelberg, p 146–160
 49. BARRERA G, DI MAURO C, MURACA R, FERRERO D, CAVALLI G, FAZIO V, PARADISI L, DIANZANI M U 1991 Induction of differentiation of human HL-60 cells by hydroxynonenal, a product of lipid peroxidation. *Exp Cell Res* 297: 148–152
 50. FAZIO V M, RINALDI M, CIAFRE S, BARRERA G, FARACE M G 1993 Control of neoplastic cell proliferation and differentiation by restoration of 4-hydroxynonenal physiological concentrations. *Mol Aspects Med* 14: 217–228
 51. KODAMA M, KANEKO M, AIDA M, INOUE F, NAKAYAMA T, AKIMOTO H 1997 Free radical chemistry of cigarette smoke and its implication in human cancer. *Anticancer Res* 17: 433–437
 52. BASILEA V, FERRARIB E, LAZZARIB S, BELLUTIA S, PIGNEDOLIB F, IMBRIANO C 2009 Curcumin derivatives: Molecular basis of their anti-cancer activity. *Biochem Pharmacol* 78: 1305–1315
 53. HORAKOVA L, STOLC S 1998 Antioxidant and pharmacodynamic effects of pyridoindole stobadine. *Gen Pharmacol* 30: 627–638
 54. BOROVIĆ S, TIRZITIS G, TIRZITE D, ČIPAK A, KHOSHSORUR G A, WAEG G, TATZBER F, SCUKANEC-SPOLJARM, ŽARKOVIĆ N 2006 Bioactive 1,4-dihydroisonicotinic acid derivatives prevent oxidative damage of liver cells. *Eur J Pharm* 537: 12–19
 55. TIRZITIS G, TIRZITE D, HYVONEN Z 2001 Antioxidant activity of 2,6-dimethyl- 3,5-dialkoxycarbonyl-1,4-dihydropyridines in metal-ion cat.